Research Article

Interactions of Lactobacilli with Pathogenic Streptococcus pyogenes

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Objective. To determine whether (1) a decreased concentration of Lactobacilli allows S. pyogenes to grow; (2) S. pyogenes is able to grow in the presence of healthy Lactobacillus concentrations; (3) S. pyogenes is capable of inhibiting Lactobacilli. Methods. One hundred fifty patient samples of S. pyogenes were mixed with four different concentrations of L. crispatus and L. jensenii. Colony counts and pH measurements were taken from these concentrations and compared using t-tests and ANOVA statistical analyses. Results. Statistical tests showed no significant difference between the colony counts of S. pyogenes by itself and growth when mixed with Lactobacilli, and no significant difference between the colony counts of S. pyogenes in the four different concentrations of Lactobacilli. Conclusion. The statistical data representing the growth of these two organisms suggests that Lactobacilli did not inhibit the growth of S. pyogenes. Also, S. pyogenes did not inhibit the growth of Lactobacilli.

1. Introduction

Lactobacillus bacteria (Lactobacilli) are large Gram positive rods that exist as nonpathogenic microbiota. Lactobacilli have been extensively studied due to their remarkable ability to inhibit the growth of other organisms through bactericidal activity and by producing lactic acid as a byproduct of metabolism [1, 2]. Lactic acid production, production of bacteriocins, and the production of hydrogen peroxide have led to an abundance of research involving the ability of Lactobacilli to inhibit pathogens. Lactobacillus species have proven effective at inhibiting the growth of bacterial and fungal pathogens which commonly cause vaginosis. Lactobacillus species, specifically L. crispatus and L. jensenii, are the predominant flora in the vagina, and thus minimize opportunities for infection [3–8]. Several common pathogens that Lactobacilli inhibit are: Candida albicans, Escherichia coli (including E. coli O157:H7); and Neisseria gonorrhoeae [1, 2, 5–9].

Due to their ability to inhibit other organisms, Lactobacilli are commonly used for probiotic therapy to enhance intestinal microbiota, as well as to treat vaginosis. The principle of this treatment is to increase the concentration of Lactobacilli, which will inhibit pathogens and allow the body’s immune system to overcome the infection without the use of antimicrobials [8, 9].

Streptococcus pyogenes, often referred to as Group A strep, is a Gram positive coccus which tends to group together in chains. S. pyogenes causes the infection commonly known as “strep throat” and is the cause of 90% of bacterial pharyngitis cases. It can cause impetigo, erysipelas (cellulitis), toxic shock syndrome, and necrotizing fasciitis (also known as “flesh-eating strep”). Untreated infections may lead to acute glomerulonephritis, scarlet fever, or rheumatic fever. It has many virulence factors that contribute to its pathogenicity, such as lipoteichoic acid, M protein, hyaluronidase, protease, streptokinase, DNase/RNase, C5a peptidase, and Streptolysins O and S. These allow the bacteria to hemolyze blood cells, spread throughout the body, adhere to surfaces, and necrotize tissues [10].

S. pyogenes can exist in the vagina [4], but it was not previously considered a cause of bacterial vaginosis [11–13]. However, the recorded incidence of bacterial vaginosis caused by S. pyogenes has increased during the past two
In order to minimize variation in Lactobacillus species that might be found in clinical specimens, strains of L. crispatus and L. jensenii were purchased from the American Type Culture Collection (ATCC). L. crispatus (ATCC 33197) and L. jensenii (ATCC 25258) were mixed in sterile Columbia broth (Fisher Scientific, Pittsburgh, PA) to concentrations of 10^6, 10^7, 10^8, and 10^9 CFU/mL [23]. The 10^6 is representative of the average Lactobacillus concentration in a healthy female [5]. A higher than average concentration was set up to represent the females who have more Lactobacillus, along with two lower concentrations to represent individuals who would be at a higher risk of infection. Columbia broth was chosen because it did not favor the growth of either organism, and it allowed the fluctuations in pH to be measured easily.

The source of bacterial vaginosis caused by S. pyogenes is suspected to be the throat, so 150 positive S. pyogenes throat screens were donated by Ogden Clinic in Ogden, Utah. Personal identification information of each patient was removed by the clinic before donating the samples, earning this study an exempt status from the Weber State University Institutional Review Board approval. Subcultures were performed in order to isolate and verify the identity of S. pyogenes. Research indicates that the average concentration of pathogens which cause bacterial vaginosis is about 10^9 CFU/mL [16]. However, the concentration of S. pyogenes in saliva has not yet been studied. A preliminary experiment was conducted to determine a concentration to use. For this preliminary experiment, concentrations of S. pyogenes at 10^2 and 10^3 CFU/mL were grown with Lactobacilli at 10^6 CFU/mL. A concentration of 10^3 CFU/mL was used, based on the colony counts from these concentrations.

A nephelometer, which is an instrument that measures the turbidity of liquid solutions, was used to measure the concentrations of the bacteria in the broth preparations, followed by serial dilutions to achieve the various desired concentrations. Three mL preparations of S. pyogenes at 10^9 CFU/mL were mixed with three mL of each of the four different concentrations of Lactobacilli. Each of these four mixtures was then plated on Columbia Sheep Blood Agar (SBA) (Fisher Scientific, Pittsburgh, PA) using calibrated 1.0μL loops. The broth mixtures and SBA plates were incubated in a carbon dioxide incubator at 37°C for 48 hours. After 48 hours, colonies were counted on the SBA plates, and pH measurements were taken from each broth mixture. Colonies were counted at 48 hours because the Lactobacillus colonies were larger and easier to count after 48 hours. There was no difference in colony counts or pH levels between 24 and 48 hours.

To validate the methodology used in this study, the same process was repeated using Escherichia coli, Streptococcus agalactiae (Group B strep), Staphylococcus epidermidis, and Staphylococcus aureus, which are known to be inhibited by Lactobacilli. Each of these was plated on an SBA plate before mixing with the Lactobacillus concentrations to compare to the growth in the presence of Lactobacilli. One hundred μL were taken from the broth preparations before mixing with the Lactobacilli and were incubated along with the mixtures. The pH of the broth without Lactobacilli was measured after incubation to compare to the pH of that pathogen mixed with Lactobacilli. The growth of each pathogen used was inhibited by the Lactobacilli, and the broth pH was lowered. Utilizing these pathogens as controls proves that the conditions used in the methodology allowed the Lactobacilli to inhibit other organisms as they would in the body.

### 3. Results and Discussion

In each of the samples, no inhibition of S. pyogenes by L. crispatus and L. jensenii was observed. Statistical tests (t-tests with an alpha level of α = 0.01) showed no significant difference between the colony counts of S. pyogenes by itself and growth when mixed with Lactobacilli. In the case of most pathogens, a higher concentration of Lactobacilli would cause more inhibition. However, statistical tests (ANOVA, α = 0.01) also showed no significant difference between the colony counts of S. pyogenes in the four different concentrations of Lactobacilli. The difference in colony-forming units for each S. pyogenes isolate was most likely due to technical error during the serial dilutions and/or random error due to the differing strains of S. pyogenes.
Table 1: Average growth and pH of the broth mixtures.

<table>
<thead>
<tr>
<th>Colony counts from broth mixtures</th>
<th>10^8 CFU/mL Lactobacilli</th>
<th>10^6 CFU/mL Lactobacilli</th>
<th>10^4 CFU/mL Lactobacilli</th>
<th>10^3 CFU/mL Lactobacilli</th>
</tr>
</thead>
<tbody>
<tr>
<td>Average colony counts of Lactobacilli</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>10</td>
<td>4</td>
</tr>
<tr>
<td>Average colony counts of S. pyogenes</td>
<td>13</td>
<td>14</td>
<td>13</td>
<td>11</td>
</tr>
<tr>
<td>Average pH</td>
<td>6.0</td>
<td>6.5</td>
<td>6.5</td>
<td>6.5</td>
</tr>
</tbody>
</table>

Figure 1: Percent of total growth of each organism comparing growth with Lactobacilli to growth without Lactobacilli. These data were taken from the results of this study.

The pH of the broth for each sample was reduced from the starting pH of 8.5 to a pH range of 4.5–7.0. The pH of the broth only containing S. pyogenes ranged from 5.5 to 6.5, suggesting that S. pyogenes grows optimally in an acidic environment [5]. Table 1 summarizes the average colony counts and pH of the mixtures of S. pyogenes and Lactobacilli.

Limitations of this study included the elevated pH of the broth medium and technical error associated with serial dilutions. The process of serial dilutions allowed the possibility of pipetting error which may have influenced the colony counts. The elevated pH of the broth medium did not allow any data at a pH lower than 4.5. It is also possible that the alkaline pH of the broth medium impacted inhibition of the Lactobacilli and/or S. pyogenes.

Additional studies that further investigate the prevalence and clinical significance of bacterial vaginosis caused by S. pyogenes would add to the current body of knowledge on this topic.

4. Conclusions

Figure 1 shows the growth patterns of the organisms used in this study. With the exception of S. pyogenes and S. epidermidis, the pathogens grew optimally in conditions where the Lactobacillus concentrations were decreased. S. epidermidis is normal skin flora and is inhibited to nonpathogenic levels by the Lactobacilli within the vagina. S. pyogenes grew optimally in the same conditions as the Lactobacilli. It is possible that higher concentrations of Lactobacilli may assist the growth of S. pyogenes, as shown in Figure 2.

It is suggested that medical personnel treat vaginosis caused by S. pyogenes using antimicrobial therapy, as they would treat other S. pyogenes infections. It is also suggested that Lactobacillus probiotic therapy not be used as the sole means to treat bacterial vaginosis caused by S. pyogenes. The statistical data representing the growth of these two organisms suggest that Lactobacilli did not inhibit the growth of S. pyogenes. Also, S. pyogenes did not inhibit the growth of Lactobacilli.

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References

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