The Oxytocin-Oxytocin Receptor System and Its Antagonists as Tocolytic Agents

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Oxytocin, a hormone involved in numerous physiologic processes, plays a central role in the mechanisms of parturition and lactation. It acts through its receptor, which belongs to the G-protein-coupled receptor superfamily, while Gq/phospholipase C (PLC)/inositol 1,4,5-triphosphate (InsP3) is the main pathway via which it exerts its action in the myometrium. Changes in receptor levels, receptor desensitization, and locally produced oxytocin are factors that influence the effect of oxytocin on uterine contractility in labor. Activation of oxytocin receptor causes myometrial contractions by increasing intracellular Ca²⁺ and production of prostaglandins. Since oxytocin induces contractions, the inhibition of its action has been a target in the management of preterm labor. Atosiban is today the only oxytocin receptor antagonist that is available as a tocolytic. However, the quest for oxytocin receptor antagonists with a better pharmacological profile has led to the synthesis of peptide and nonpeptide molecules such as barusiban, retosiban, L-368,899, and SSR-126768A. Many of these oxytocin receptor antagonists are used only as pharmacological tools, while others have tocolytic action. In this paper, we summarize the action of oxytocin and its receptor and we present an overview of the clinical and experimental data of oxytocin antagonists and their tocolytic action.

1. Introduction

Oxytocin (OT) is a nonapeptide synthesized by the magnocellular neurons located in the supraoptic and paraventricular nuclei of the hypothalamus and secreted to the circulation by the posterior pituitary and nerve terminals in response to various stimuli. The sequence of amino acids in the OT molecule is Cysteine-Tyrosine-Isoleucine-Glutamine-Asparagine-Cysteine-Proline-Leucine-Glycinamide, with a sulfur bridge between the two cysteines. OT and vasopressin have similar structures and differ only in two amino acids. Oxytocin is also synthesized in many peripheral tissues, for example, uterus, placenta, amnion, corpus luteum, testis, and heart [1].

Oxytocin exerts a variety of actions and is involved in a large number of physiological and pathological processes. These actions include the regulation of the hypothalamo-pituitary-adrenal axis in response to stress, pregnancy, luteal function, maternal behavior, cell proliferation, modulation of emotional relationships and sexual behavior, erectile function and ejaculation, antinociception, cardiovascular function, osteoporosis, and neuropsychiatric disorders [2–6]. However, its best-known and most well-established roles are stimulation of uterine contractions during parturition and milk release during lactation. In 1906, Sir Henry Dale found that an extract from the human posterior pituitary gland had a uterotonic effect, and Vincent du Vigneaud et al. achieved synthesis of oxytocin in 1953 [7]. Since oxytocin contributes to myometrial contractility, its receptor has been a target for tocolytic agents. While atosiban is an oxytocin receptor (OTR) antagonist used for the management of preterm labor [8], research is ongoing for the tocolytic properties of various other OTR antagonists.
2. The Oxytocin Receptor

The oxytocin receptor belongs to the rhodopsin-type class I G-protein-coupled receptor (GPCR) superfamily. The gene of the OTR contains 3 introns and 4 exons and is located in a single copy on chromosome 3p25. Apart from oxytocin, other molecules such as arginine vasopressin (AVP) and oxytocin agonists or antagonists are able, because of their similar structure, to bind to the receptor. Binding of OT to the cell surface transmembrane OTR activates the receptor which subsequently activates various intracellular signal pathways, this triggering the numerous effects of the hormone, including contraction (Figure 1). OTR is coupled to the Gq/11 a-class guanosine triphosphate (GTP) binding proteins. The Gq/phospholipase C (PLC)/inositol 1,4,5-triphosphate (InsP3) pathway is the major pathway mediating the signal of OTR after binding of OT to its receptor. Binding of OT activates, through Gaq11, PLC, which hydrolyzes phosphatidylinositol 4,5-bisphosphate (PIP2) to InsP3 and diacylglycerol (DAG) [9]. InsP3 releases Ca^{2+} ions from intracellular stores, while DAG activates protein kinases type C (PKC), which further phosphorylates other proteins. Ca^{2+} ions bind to calmodulin, and the Ca^{2+}-calmodulin complex activates the myosin light-chain kinase, thereby causing myometrial muscle contraction. The same mechanism additionally causes contraction of mammary myoepithelial cells and milk ejection [1]. However, OTR is also coupled with other G-proteins, Gs and Gi, which result in various cellular effects, among them the inhibition of cell growth [10].

OTR also acts on voltage-gated or receptor coupled channels, leading to membrane depolarization and the entry of extracellular Ca^{2+} to the cells, this further promoting smooth muscle contractility [11].

Moreover, OTR activates the mitogen-activated protein kinase (MAPK) and the Rho kinase pathways. Rho-associated protein kinases are implicated in many cellular phenomena, including cell contractility, cell migration, and cell cycle control [12]. Activations of OTR and MAPK both result in increased cytosolic phospholipase A2 (cPLA2) activity. cPLA2 hydrolyzes phospholipids and liberates arachidonic acid, which results in increased production of prostaglandins via cyclooxygenase-2 (COX-2), an enzyme that is upregulated by MAPK [13, 14]. The increased production of prostaglandins in conjunction with the increase in intracellular Ca^{2+} and the activation of the Rho and MAP kinase pathways results in the contractile effects of OT-OTR activation.

3. The Oxytocin-Oxytocin Receptor in Labor

The mechanism of transition from myometrial quiescence to labor initiation is a complex one in which numerous diverse factors contribute in conjunction to the process. Hormones such as progesterone and CRH have a crucial role in this transition [15–17]. Immune responses, triggered via release of cytokines and other mediators of inflammation, are also important, especially in cases of preterm labor [18–22]. Oxytocin additionally contributes to the mechanism of labor and interacts with many of the other contributing factors. However, circulating oxytocin does not seem to be necessary for the initiation and completion of parturition in humans, since normal labor can be achieved in the absence of oxytocin, as in cases of pituitary dysfunction [23]. Furthermore, in humans oxytocin circulation levels do not increase significantly during pregnancy or the beginning of labor. However, some changes in circulating oxytocin occur as its levels are increased at the expulsive stage and pulsatile changes occur in pregnant women at term. As mentioned above, the oxytocin locally produced in the placenta is likely to be more important than circulating oxytocin for the mechanism of labor. Changes to the OTR also occur towards labor. OTR is upregulated at the end of gestation and in both term and preterm labor, and sensitivity to oxytocin-induced contractions is greatly increased [24–26]. Though it is assumed that steroid hormones may also influence the number of OTRs, the mechanisms of regulation are complex and not as yet fully understood [27–31]. It is of interest that in rabbits, myometrium under progesterone dominance is refractory to oxytocin [15].

Coordinated synchronous myometrial contractions during labor are achieved through the formation of gap junctions among cells, which are created by multimers of the protein connexin 43 [32, 33]. Oxytocin increases connexin-43 levels and upregulates morphologic gap junctions [34, 35]. After parturition, OTR expression in the mammary glands remains high thus facilitating lactation, while OT-binding sites in the uterus decline rapidly [36, 37].

Continuous exposure to high doses of oxytocin leads to desensitization of the receptor [38]. Desensitization is a phenomenon that is observed in GPCR receptors and occurs via different mechanisms such as phosphorylation, internalization, or changes at the receptor mRNA levels. Desensitization, whose purpose is to protect cells from overstimulation after prolonged agonist stimulation, can occur very rapidly, within seconds or minutes, and takes place in two steps. The first step is the phosphorylation of the receptor that inhibits G-protein activation, and the second step is the binding of proteins called arrestins, which prevent G-protein activation and promote receptor internalization. Evidence appears to indicate that receptors recycle back to the cell surface after internalization [39].

OTR desensitization is a phenomenon that occurs after prolonged agonist stimulation and may last for hours or even days [36]. The mRNA of the OTR is also reduced after continuous OT treatment. In cultured human myometrial cells, treatment with OT for up to 20 hours causes OTR desensitization and great reduction of OT-binding sites without receptor internalization. OTR mRNA levels are reduced, though not the total amount of OTR protein [40]. In vivo, when oxytocin is used to augment labor in pregnant women, there is also a reduction in myometrial oxytocin binding sites and in OTR mRNA levels [41]. Oxytocin receptor downregulation has great clinical significance, as prolonged oxytocin infusion may fail to augment labor or can lead to postpartum uterine atony that cannot be managed with additional oxytocin infusion. However, as oxytocin is normally secreted in pulses, this pulsatile secretion may...
Figure 1: Oxytocin receptor linked signaling pathways resulting in myometrial contraction. Binding of OT to OTR activates Gaq/11 and then PLC, which hydrolyses PIP2 to InsP3 and DAG. InsP3 causes release of Ca\(^{2+}\) from the sarcoplasmic reticulum, while DAG activates PKC. Gaq/11 also causes activation of voltage-regulated Ca\(^{2+}\) channels and Ca\(^{2+}\) entry into the cells. Ca\(^{2+}\) binds to calmodulin, and the Ca\(^{2+}\)-calmodulin complex activates MLC kinase, resulting in myometrial contraction. Both OTR and PKC activate the MAPK cascade, while OTR and MAPK result in increased cPLA2 activity. Increased cPLA2 activity and MAPK activation result in prostaglandin production, which also contributes to the contractile effect. The activation of the RhoA-ROK cascade by the OTR is another pathway that results in increased MLC phosphorylation and myometrial contraction. OT: oxytocin, OTR: oxytocin receptor, PLC: phospholipase, PIP2: phosphatidylinositol 4,5-bisphosphate, InsP3: inositol 1,4,5-triphosphate, DAG: diacylglycerol, PKC: protein kinases type C, MLC: myosin light-chain, MAPK: mitogen-activated protein kinase, cPLA2: cytosolic phospholipase A2, ROK: RhoA associated protein kinase.

prevent desensitization from occurring. This might explain why lower doses of oxytocin are required to augment labor when it is administered in pulses and higher doses when it is infused continuously [42].

4. Oxytocin Agonists

OTR agonist molecules have been developed and studied as pharmacological tools or as potential drugs for the management of neuropsychiatric diseases including anxiety-related disorders, schizophrenia, and autism. OTR agonists may be peptides ([Thr⁴]OT, [HO¹][Thr⁴]OT, [Thr⁴,Gly⁷]OT, [HO¹][Thr⁴,Gly⁷]OT and other molecules) or nonpeptides (WAY-267464 and other compounds) [43–45]. WAY-267464 exerts oxytocinergic actions, such as anxiolytic effects in mice [46].

Synthetic oxytocin is used to augment labor and treat postpartum hemorrhage. Demoxytocin is an oxytocin analogue that has been used for labor induction, though it has been proven less effective compared to prostaglandins [47]. Carbetocin is a peptide synthetic oxytocin analogue for the prevention of uterine atony after cesarean section with spinal or epidural anesthesia: it has a longer half-life than oxytocin and possesses the advantage that it is administered in a single dose, intramuscularly or intravenously [48].

5. Oxytocin Antagonists as Tocolytic Agents

The therapeutic target in the treatment of preterm labor is currently the pharmacological inhibition of uterine contractions with the use of various tocolytic agents. Tocolytic agents are used to maintain pregnancy for 24–48 hours to allow corticosteroids administration to act and to permit the transfer of the mother to a center with a neonatal intensive care unit. Ritodrine (a beta-receptor agonist), nitric oxide donors (as glyceryl trinitrate), calcium-channel blockers, and COX-2 inhibitors such as indomethacin are some of the currently available tocolytics [4].

5.1. Peptide OTR Antagonists. Selective human oxytocin receptors antagonists have also been synthesized as tocolytic agents for the management of preterm labor. Atosiban is an oxytocin analogue (1-Deamino-2-D-Tyr-(O-ethyl)-4-Thr-8-ornoxytocin) based on modification of some amino acids in the structure of oxytocin at positions 1, 2, 4, and 8.
(Figure 2). It is a mixed vasopressin (V1a) and oxytocin receptor antagonist that blocks OT binding to OTR and is the only oxytocin antagonist used today for the treatment of preterm labor in Europe and other countries, though not in the USA. However, as it is also an antagonist of the vasopressin receptor V1a, this results in related undesirable effects. The onset of uterine relaxation after atosiban administration is rapid. Atosiban is given intravenously for up to 48 hours. Clinically, atosiban is as effective as β2-adrenergic agonists and with lesser adverse effects. When subcutaneously administered as maintenance therapy after a period of preterm labor, atosiban failed to reduce the incidence of preterm birth or improve neonatal outcome [49].

Clinically, atosiban is safer than beta-receptor agonists. Atosiban is comparable in clinical effectiveness to conventional beta-agonist therapy (ritodrine, salbutamol, or terbutaline), but it is associated with fewer maternal cardiovascular side effects and is better tolerated [50–53]. Compared to calcium channel blockers, atosiban has been shown equally effective as nifedipine and with fewer maternal side effects [54, 55]. Cyclooxygenase inhibitors also present significant side effects [56]. Conversely, there is at present insufficient evidence to recommend the use of nitric oxide donors as inhibitors of preterm delivery [57]. However, atosiban has not been proven to be superior in terms of neonatal outcome, as concerns have been described in some studies [58].

Unfortunately, atosiban displays some disadvantages when used as a tocolytic. It has limited bioavailability and requires parenteral administration and hospitalization, has low affinity of the OTR, and is also an antagonist for the V1a receptors that cause side effects. These limitations have led to efforts for the discovery of new peptide and nonpeptide OT antagonists for the management of preterm labor. Most of these substances are still being evaluated at an experimental level (Table 1), and clinical studies in most cases have not been successfully completed [43].

A number of highly selective OT peptidic antagonists have been designed and synthesized, like d(CH2)5[Tyr(Me)2]-OVT, desGly-NH2,d(CH2)5[Tyr(Me)2,Thr4]OVT, desGly-NH2, d(CH2)5[D-Tyr2,Thr4]OVT, d(CH2)5[D-Thi2,Thr4, Tyr-NH2]OVT, and desGly-NH2,d(CH2)5[D-Trp2,Thr4, Dap5]OVT. These molecules are both OTR and AVP receptor antagonists but are more potent as OT antagonists than as V1a antagonists. However, experiments have revealed striking differences between species concerning the affinity of many antagonists to the various receptors [59].

Some of the new peptide OT/AVP antagonists have higher affinity for human receptor than the peptide atosiban. These new peptides are desGly-NH2,d(CH2)5[D-2-Nal2,Thr4]OVT, desGly-NH2,d(CH2)5[2-Nal2,Thr4]OVT, d(CH2)5[D-2-Nal2,Thr4,Tyr-NH2]OVT, and d(CH2)5[2-Nal2,Thr4,Tyr-NH2]OVT and may be candidates as potential tocolytic agents [43].

Barusiban is a selective peptide oxytocin antagonist with a high affinity for the human OTR and low for the V1a receptor. It has a higher potency and a longer duration of action than atosiban. In contractility studies with isolated human myometrium, barusiban inhibits oxytocin-induced myometrial contractions of both preterm and term myometrium, and this action was at least as potent as the action of atosiban [60]. In nonhuman primates, barusiban also inhibits oxytocin-induced myometrial contractions [61, 62]. In pregnant monkeys, in which atosiban and barusiban were tested following induction of contractions by OT, the duration of action of barusiban was generally longer than that of atosiban (13–15 hours compared to 1.5–3 hours). For long-term treatment, continuous high-dose infusions of barusiban or the beta-2 agonist fenoterol were administered: barusiban moderated intrauterine pressure increase in response to daily OT challenge and prolonged pregnancy more effectively than fenoterol [63].

However, despite the promising results in nonhuman primates, in a recent study in pregnant women, barusiban was not more effective than placebo in stopping preterm labor. In a multicenter trial that was conducted at 21 participating centers in 6 different European countries, participants were randomly assigned to receive a single intravenous bolus dose of placebo or barusiban. There was no significant difference
Table 1: The most extensively studied OT antagonists and their experimental or clinical applications.

<table>
<thead>
<tr>
<th>Oxytocin antagonists</th>
<th>Organism</th>
<th>Experimental or therapeutic application</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>(I) Peptide</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1-Deamino-2-D-Tyr-(O-ethyl)-4-Thr-8-ornoxytocin (Atosiban)</td>
<td>Human</td>
<td>Clinically used tocolytic agent</td>
</tr>
<tr>
<td>d(CH2)5[Tyr(Me)2]OVT</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td>desGly–NH2,d(CH2)5[Tyr(Me)2,Thr4]OVT</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td>desGly–NH2,d(CH2)5[D-Tyr2,Thr4]OVT</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td>d(CH2)5,[D-Thi2,Thr4,Tyr–NH92]OVT</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td>desGly–NH2,d(CH2)5[D-2-Nal2,Thr4,Dap5]OVT</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td>desGly–NH2,d(CH2)5[2-Nal2,Thr4]OVT</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td>d(CH2)5,[2-Nal2,Thr4,Tyr–NH92]OVT</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td>FE 200 400 (Barusiban)</td>
<td>Monkeys/human myometrial tissue</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td><strong>(II) Nonpeptide</strong></td>
<td></td>
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</tr>
<tr>
<td>GSK221149A (Retosiban)</td>
<td>Rats</td>
<td>Tocolytic action. A phase II clinical trial for its use as a tocolytic in humans has not yet been completed</td>
</tr>
<tr>
<td>L-368,899</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tools in experimental studies of OT function. CNS effects. Inhibits spontaneous nocturnal uterine contractions in pregnant rhesus monkeys. Clinical studies discontinued</td>
</tr>
<tr>
<td>L-371,257</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td>WAY-162720</td>
<td>Animals/human cells and tissues, or both</td>
<td>Pharmacological tool in experimental studies of OT functions</td>
</tr>
<tr>
<td>SSR-126768A</td>
<td>Rats/human myometrial tissue</td>
<td>Tocolytic action in rats. Inhibits the response to OT in term human pregnant uterine sections</td>
</tr>
</tbody>
</table>

between the placebo group and any of the barusiban groups with regard to the percentage of women who did not deliver within 48 hours, nor was there any significant decrease in the number of uterine contractions between the barusiban groups and placebo group. Additionally, there were no statistical differences in maternal or neonatal adverse effects between placebo and barusiban groups [64].

5.2. Nonpeptide OTR Antagonists. Since peptide antagonists lack oral bioavailability, pharmaceutical companies have searched for an effective nonpeptide oxytocin antagonist. GSK221149A (2′-methyl-1′,3′-oxazol-4′-yl morpholine amide derivative 74), known as retosiban, is such a nonpeptide oxytocin antagonist. In vitro experiments using Chinese hamster ovary (CHO) cell membranes expressing human OT receptors or human vasopressin (V1a, V1b, V2) receptors and human endothelial kidney (HEK) cells expressing rat oxytocin receptors revealed that retosiban has a higher affinity for human and rat oxytocin receptors than for V1a and V2 receptors. In rats, when administered orally or intravenously, retosiban produced a dose-dependent decrease in oxytocin-induced uterine contraction. This effect was observed after either single or multiple dosing for 4 days. Also, spontaneous uterine contractions in late-term pregnant rats (at 19–21 days gestation) were significantly reduced by intravenous administration of retosiban [65]. Retosiban is more than
15-fold more potent compared to atosiban for the OTR [66]: it is at present on a Phase II Clinical trial investigating its action as a tocolytic, but the results have not as yet been published [67].

Another nonpeptide oxytocin antagonist is L-368,899 (1-(((7,7-Dimethyl-2(S)-2(S)-amino-4-(methylsulfonyl) butyramido) bicyclo [2.2.1]-heptan-1(S)-yl)methyl) sulfonyl)-4(2methylphenyl) piperazine). In pregnant rhesus monkeys, L-368,899 was shown to be a potent OT antagonist that inhibits spontaneous nocturnal uterine contractions. In women, L-368,899 also blocked OT-stimulated uterine activity postpartum with a potency similar to that observed in the pregnant rhesus monkey [68]. However, no further clinical evaluation was undertaken because of suboptimal oral bioavailability and pharmacokinetics [69]. L-368,899 is also brain penetrant, and experiments in one adult female monkey demonstrated that it altered maternal and sexual behavior [4].

SSR-126768A (4-Chloro-3-[(3R)-(+)5-chloro-1-(2,4-dimethoxybenzyl)-3-methyl-2-oxo-2,3-dihydro-1H-indol-3-yl]-N-ethyl-N-(3-pyridylmethyl)-benzamide, Hydrochloride), a nonpeptide oxytocin antagonist, produced a competitive antagonistic effect against OT in rat myometrial strips, as in conscious rat after oral administration. After oral administration in pregnant rats in labor, SSR-126768A significantly delayed parturition, similarly to ritodrine, and had a rapid onset of action, and its duration was still observed 24 hours after treatment. Experimentally, SSR-126768A inhibited the response to OT at-term human pregnant uterus sections, this effect being observed in a concentration-dependent manner [70].

As nonpeptide OTR antagonists display high selectivity for the OTR receptor, they may provide orally administered tocolytics for both the acute and long-term management of preterm labor, which would undoubtedly offer a significant improvement in neonatal outcomes.

In women who go into preterm labor, relcovaptan (which is a nonpeptide vasopressin V1a receptor antagonist) was reported to inhibit uterine contractions [71, 72]. In a study in 18 women in preterm labor between 32–36 weeks, 12 patients received at random a single oral dose of relcovaptan and 6 patients received placebo, while uterine contractions, were monitored up to 6 h after administration. Rilcovaptan inhibited uterine contractions, and, compared with placebo, the difference was statistically significant. Rilcovaptan has also revealed positive initial results in tests against Raynaud’s disease, dysmenorrhea, and experimentally in ischemic brain damage, although it has not yet been approved for clinical use [73–76].

6. Conclusion

Oxytocin is a neuropeptide which affects uterine contractility during labor. Its receptor is a transmembrane receptor belonging to the G-protein-coupled receptor superfamily. The main signaling pathway is the Gq/LPC/Ins3 pathway, but the MAPK and the RhoA/Rho kinase pathways are also activated, contributing to increased prostaglandin produc-

References


