Clinical Study

Impact of Long-Acting Somatostatin Analogues on Glucose Metabolism in Acromegaly: A Hospital-Based Study

Ming Shen,1 Meng Wang,2 Wenqiang He,1 Min He,2 Nidan Qiao,1 Zengyi Ma,1 Zhao Ye,1 Qilin Zhang,1 Yichao Zhang,1 Yeping Yang,2 Yanjiao Cai,2 Yakupujiang ABuDuoReYiMu,3 Yun Lu,4 Bin Lu,2 Xuefei Shou,1 Yongfei Wang,1 Hongying Ye,2 Yiming Li,1 Shiqi Li,1 Yao Zhao,1 Xiaoyun Cao,1 and Zhaoyun Zhang2

1Department of Neurosurgery, Huashan Hospital, Fudan University, Shanghai 200040, China
2Department of Endocrinology and Metabolism, Huashan Hospital, Fudan University, Shanghai 200040, China
3Department of Endocrinology and Metabolism, The Second People’s Hospital of Kashi, Xinjiang Uygur Autonomous Region 844000, China
4Department of Nuclear Medicine, Huashan Hospital, Fudan University, Shanghai 200040, China

Correspondence should be addressed to Xiaoyun Cao; caoxiaoyun2594@163.com and Zhaoyun Zhang; zhaoyunzhang@fudan.edu.cn

Received 13 July 2017; Accepted 13 December 2017; Published 26 April 2018

Academic Editor: Mario Maggi

Copyright © 2018 Ming Shen et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Purpose. To evaluate the change in glucose tolerance in treatment-naïve patients with acromegaly after administration of SSA and to identify predictive factors of glucose impairment during SSA therapy. Methods. Oral glucose tolerance testing (OGTT) was performed on 64 newly diagnosed and treatment-naïve patients with acromegaly both at pretreatment and 3 months after initiation of treatment with long-acting SSA. Insulin resistance (IR) was assessed by homeostatic model assessment- (HOMA-) IR and ISOGTT. Insulin secretion was assessed by HOMA-β, INS/BG0, IGI (insulinogenic index), IGI/IR, ISSI2, and AUCINS/AUCBG. Receiver-operating characteristic (ROC) curves were generated to determine the optimal cutoffs to predict the impact of SSA on glucose metabolism. Results. Pretreatment, 19, 24, and 21 patients were categorized as having normal glucose tolerance (NGT), impaired glucose tolerance (IGT), and diabetes mellitus (DM), respectively. Posttreatment, IR, represented by ISOGTT, was significantly improved in all 3 groups. Insulin secretion, represented by HOMA-β, declined in the NGT and IGT groups, but was unaltered in the DM group. The glucose tolerance status deteriorated in 18 (28.1%) patients, including 13 patients in the NGT group and 5 patients in the IGT group. Deterioration was associated with lower baseline BG 120 (plasma glucose 120 min post-OGTT), less reduction of growth hormone (GH), and greater reduction of insulin secretion after SSA therapy. BG120 greater than 8.1 mmol/l provided the greatest sensitivity and specificity in predicting the stabilization and/or improvement of glucose tolerance status after SSA treatment (PPV 90.7%, NPV 66.7%, p < 0.001). Conclusions. The deterioration of glucose metabolism induced by SSA treatment is caused by the less reduction of GH and the more inhibition of insulin secretion, which can be predicted by the baseline BG120 during OGTT.

1. Introduction

Acromegaly is an insidious disease associated with a 1.72 times increased mortality risk [1]. Cardiovascular, respiratory, and metabolic complications are the main causes of death in acromegaly. Disturbances of carbohydrate metabolism are the major type of metabolic disorder [2]. Overt type 2 diabetes mellitus is reported in 19–56% and impaired glucose tolerance (IGT) in 16–46% of patients with acromegaly [3]. GH-mediated insulin resistance (IR) is the major cause of impaired glucose metabolism in active acromegaly [4]. Although transsphenoidal surgery is the first-line therapy for GH-secreting adenomas, for those who are not in remission after surgery or for whom surgery is contraindicated,
long-acting somatostatin analogues (SSA) are generally considered to be first-line therapy [5]. However, the impact of SSA on glucose metabolism has not been fully elucidated and previous results from small series are conflicting [6–9]. This may be due to the fact that SSA inhibits GH and glucagon secretion while also suppressing the release of insulin [10, 11]. The aim of our study was to investigate the effects of SSA on glucose homeostasis and to determine whether there are any variables that could predict the influence of SSA on glucose metabolism in patients with active acromegaly.

2. Subjects and Methods

2.1. Patients. This was a retrospective study of prospectively obtained data from patients seen between July 2012 and August 2014 at a tertiary referral center in the East of China. Sixty-four newly diagnosed and untreated patients with acromegaly (38 females and 26 males, mean age 41.7 ± 13.0 years) were recruited. Clinical and biochemical findings of the patients are summarized in Supplementary Tables 1–4. The diagnosis of active disease was based on the clinical features of acromegaly, failure of GH suppression to below 1 μg/l in response to a 75 g oral glucose tolerance test (OGTT), plasma IGF-1 levels above the age-appropriate reference range, and radiological evidence of a pituitary tumor. The mean GH (GHm) was obtained as the average level of 5 samples drawn within a 2 h period (every 30 min from 0700 to 0900 h) [12]. Before and after SSA treatment, glycosylated hemoglobin (HbA1c) was obtained. Glucose tolerance was evaluated by OGTT. Briefly, after an overnight fasting, blood samples were drawn for baseline blood glucose (BG) and insulin (INS). Then, 75 g of glucose was administered orally. Sampling for BG and insulin was performed 30, 60, 120, and 180 min later. Three months after initiation of long-acting SSA treatment, octreotide LAR, 20 mg every 120, and 180 min later. Three months after initiation of long-acting SSA treatment, octreotide LAR, 20 mg every 30 min to 180 min during the OGTT. INSmean and INSmean represent the mean insulin and glucose concentrations during the OGTT. AUC_{BG} = (BG_{0} + BG_{30}) × 15 + (BG_{30} + BG_{60}) × 15 + (BG_{60} + BG_{120}) × 30 + (BG_{120} + BG_{180}) × 30. AUC_{INS} = (INS_{0} + INS_{30}) × 15 + (INS_{30} + INS_{60}) × 15 + (INS_{60} + INS_{120}) × 30 + (INS_{120} + INS_{180}) × 30. HOMA-β = (20 × INS_{0})/(BG_{0} – 3.5) × 100%. ISOGTT (the OGTT insulin sensitivity index) = 10,000/SQRT (BG_{30} × INS_{0} × BGmean × INSmean). IGI (insulinogenic index) = (INS_{30} – INS_{0})/(BG_{30} – BG_{0}) = ΔINS_{30}/ΔBG_{30}. IGI/IR = IGI/HOME-IR. ISSI2 (the OGTT insulin secretion sensitivity index-2) = (AUC_{INS} / AUC_{BG}) × ISOGTT.

2.2. Evaluation of Insulin Resistance and β-Cell Function. Homeostatic model assessment (HOMA) (including HOMA-IR and HOMA-β) was used to estimate insulin resistance (IR) and β-cell function [14]. Insulin sensitivity was also assessed by calculating IS_{OGTT} (the OGTT insulin sensitivity index) [15]. INS_{0}/BG_{0}, IGI (insulinogenic index), IGI/IR, and ISSI2 (the OGTT insulin secretion sensitivity index-2) were also used to estimate β-cell function [14, 16–22]. The areas under the curve of glucose (AUC_{BG}) and insulin (AUC_{INS}) during OGTT were calculated using the trapezoidal rule [9, 23]. AUC_{INS}/AUC_{BG}, which is an indicator of insulin secretion, was also calculated [20].

2.3. Abbreviated Variables and Formulas. BG_{0} was the baseline blood glucose value during the OGTT. BG_{30}, BG_{60}, BG_{90}, BG_{120}, and BG_{180} were the blood glucose values from 30 min to 180 min during the OGTT. INS_{0}, INS_{30}, INS_{60}, INS_{120}, and INS_{180} were the insulin values from basal to 180 min during the OGTT. BGmean and INSmean represent the mean insulin and glucose concentrations during the OGTT. AUC_{BG} = (BG_{0} + BG_{30}) × 15 + (BG_{30} + BG_{60}) × 15 + (BG_{60} + BG_{120}) × 30 + (BG_{120} + BG_{180}) × 30. AUC_{INS} = (INS_{0} + INS_{30}) × 15 + (INS_{30} + INS_{60}) × 15 + (INS_{60} + INS_{120}) × 30 + (INS_{120} + INS_{180}) × 30. HOMA-β = (20 × INS_{0})/(BG_{0} – 3.5) × 100%. ISOGTT (the OGTT insulin sensitivity index) = 10,000/SQRT (BG_{30} × INS_{0} × BGmean × INSmean). IGI (insulinogenic index) = (INS_{30} – INS_{0})/(BG_{30} – BG_{0}) = ΔINS_{30}/ΔBG_{30}. IGI/IR = IGI/HOME-IR. ISSI2 (the OGTT insulin secretion sensitivity index-2) = (AUC_{INS} / AUC_{BG}) × ISOGTT.

2.4. Biochemical Measurements. GH was measured by a two-site chemiluminescent immunometric assay (AutoDELFIA® hGH, PerkinElmer Life and Analytical Sciences, Wallac Oy), intra-assay CV: 5.3–6.5%, inter assay CV: 5.7–6.2%, and sensitivity: up to 0.01 μg/l (0.026 mU/l).

IGF-1 was measured with the IMMULITE 2000 solid-phase, enzyme-labeled chemiluminescent immunometric assay (Siemens Healthcare Diagnostic Products Limited, UK); normal age-appropriate ranges are as follows: 1–6 years: 49–327 μg/l; 7–11 years: 57–551 μg/l; 12–13 years: 143–850 μg/l; 14–16 years: 220–996 μg/l; 17–18 years: 163–731 μg/l; 19–20 years: 127–483 μg/l; 21–35 years: 115–358 μg/l; 36–50 years: 94–284 μg/l; >50 years: 55–238 μg/l; intra-assay CV: 2.3–3.5%; interassay CV: 7.0–7.1%; and sensitivity: 20 μg/l. IGF-1 index = IGF – 1/upper limit of normal range (ULN) [24].

Insulin was measured by chemiluminescence immunoassay (ADVIA Centaur XP, Siemens, USA). BG was measured by a Hitachi 7600 Biochemical Analyzer (Tokyo, Japan). HbA1c was detected with high-performance liquid chromatography (Tosoh HLC-723 G8 HPLC Analyzer, Japan).

2.5. Statistics Analysis. Data are presented as mean ± SD (or median with interquartile range) for continuous variables normally (or not normally) distributed, respectively, and as frequency for categorical variables. Normality was tested using the Kolmogorov-Smirnov test. The change of variables between pre- and post-SSA treatment within one group was compared using the paired t-test when data distribution was normal or by the Wilcoxon rank-sum (Mann–Whitney) test when variables were not normally distributed. One-way ANOVA with LSD post hoc analysis (or the Kruskal-Wallis test followed by Bonferroni post hoc test) was used for comparisons among multiple groups. For categorical variables, differences were analyzed by the chi-square test. Univariate regression analysis was performed, and Spearman rank correlation coefficients are reported. After construction of receiver-operating characteristic (ROC) curves, Youden indicies were calculated to determine the optimal cutoffs for variables to predict the change in glucose metabolism after SSA treatment (sensitivity, specificity, PPV, and NPV). Statistical
analysis was performed with SPSS 16.0 statistical software. A two-tailed $p$ value $< 0.05$ was considered significant.

3. Results

3.1. Baseline Characteristics among NGT/IGT/DM Groups. Pretreatment, patients were categorized into three groups: normal glucose tolerance (NGT) group (19 patients, 8 females/11 males), impaired glucose tolerance (IGT) group (24 patients, 15 females/9 males), and diabetes mellitus (DM) group (21 patients, 15 females/6 males). 8 patients in the DM group were known to have diabetes and were treated with oral antidiabetic drugs prior to taking part in this study. For these patients, OGTT was only performed when fasting plasma glucose (FPG) was below 8 mmol/l (previously diagnosed diabetic patients with FPG above 8 mmol/l was excluded from this study). The other 13 diabetic patients were diagnosed at baseline OGTT along with the diagnosis of acromegaly. During the study, 10 patients were treated with oral antidiabetic drugs and 11 patients were given advice about lifestyle/dietary modifications. The baseline characteristics of the three groups are shown in Supplementary Table 5. Age, body mass index (BMI), $GH_m$, IGF-1 index, and HOMA-IR did not differ significantly among the three groups. $HbA_1c$ was higher in the DM group than in the NGT and IGT groups, while HOMA-$\beta$ was significantly lower in the DM group than in the other two groups.

No difference was found between females and males in age, BMI, $HbA1c$, $GH_m$, FPG, and $BG_{120}$. Females had significantly higher FPI, $INS_{0/0}$ HOME-$\beta$, $INS_B/G_B$, and HOMA-IR, with lower $IS_{O/GT}$ and lower IGF-1 index, than males had (Supplementary Table 6). Thus, females were prone to higher insulin resistance and higher $\beta$-cell function than males were.

3.2. Effect of SSA Treatment on BG and $HbA_1c$ Levels. Compared to pretreatment, $HbA_1c$ dropped significantly within the DM group ($8.35 \pm 2.47$ versus $6.88 \pm 1.00\%$, $p = 0.015$) after SSA treatment. In the entire cohort, NGT, and IGT groups, $HbA1c$ showed no change from pretreatment to posttreatment (Table 1). Compared to pretreatment, FPG increased significantly in the entire cohort, NGT, and IGT groups after SSA treatment. However, in the DM group, no changes were detected from pretreatment to posttreatment. From before to after SSA treatment, $BG_{120}$ increased in the NGT group and decreased in the DM group, while it was unaltered in the entire cohort and IGT group (Table 1).

3.3. Effect of SSA Treatment on Plasma Insulin Levels during OGTT. Compared to pretreatment, the posttreatment levels of fasting plasma insulin (FPI) declined in the group as a whole and in NGT and IGT groups. However, no change was detected within the DM group from pretreatment to posttreatment. Compared to pretreatment, after SSA treatment, $INS_{120}$ decreased in the group as a whole and in the IGT group, but remained unaltered in the NGT and DM groups (Table 1).

3.4. Effect of SSA Treatment on Insulin Resistance. After SSA treatment, HOMA-IR significantly decreased within the group as a whole, and in the NGT and IGT groups, but not in the DM group. Moreover, $IS_{O/GT}$ significantly increased in the group as a whole, as well as in the NGT, IGT, and DM groups (Table 1). After SSA treatment, HOMA-IR significantly decreased, while $IS_{O/GT}$ significantly increased, in both females and males (Supplemental Table 7).

3.5. Effect of SSA Treatment on Insulin Secretion. In the group as a whole and in the IGT group, there was a significant decline in $\beta$-cell function, including HOMA-$\beta$, $INS_B/BG_D$, IGI, IGI/IR, and $AUC_{INS}/AUC_{BG}$ after SSA treatment. However, no significant change was observed in ISSI2. In the NGT group, all variables reflective of $\beta$-cell function declined. However, in the DM group, no change was observed in any variables reflective of insulin secretion (Table 1). In females, all variables reflective of $\beta$-cell function declined except $AUC_{INS}/AUC_{BG}$. In males, all variables reflective of $\beta$-cell function declined except ISSI2 (Supplementary Table 7).

3.6. Effects of SSA Treatment on Glucose Tolerance. At the baseline, 29.7% (19/64) of patients had NGT, 37.5% (24/64) had IGT, and 32.8% (21/64) had DM. After SSA treatment for 3 months, 26.6%, 42.2%, and 31.2% of the patients, respectively, were categorized as NGT, IGT, and DM (Figure 1). After SSA treatment, in the NGT group ($n = 19$), 31.5% maintained the status quo, while 63.2% developed IGT and 5.3% became diabetic. In the IGT group ($n = 24$), 45.8% of the patients became NGT, 33.4% remained unchange, and 20.8% progressed to diabetes. In the DM group ($n = 21$), 66.7% continued to have diabetes mellitus while 33.3% improved to IGT. In summary, after SSA treatment, the distribution of glucose metabolism status was as follows: 43.8% (28/64) patients were stable, 28.1% (18/64) of the subjects improved, and 28.1% (18/64) of the subjects deteriorated (Figure 1).

After SSA therapy, subjects were classified into 3 groups according to the change of glucose tolerance category: Improved ($n = 18$, from IGT to NGT, from DM to IGT, or from DM to NGT), Stable ($n = 28$, from NGT to NGT, from IGT to IGT, or from DM to DM), and Deteriorated ($n = 18$, from NGT to IGT, from NGT to DM, or from IGT to DM). The baseline characteristics of these 3 groups are shown in Table 2. Patients in the Stable group were older than those in the other two groups ($p = 0.049$). The baseline $BG_{120}$ levels were significantly lower in the Deteriorated group than in the other two groups ($p < 0.001$).

The changes in glucose metabolism-related variables after SSA treatment are shown in Table 3. The reduction of $GH_m$ was much less in the Deteriorated group than in the other two groups ($p = 0.021$). The reduction of HOMA-$\beta$ was greater in the Deteriorated group than in the Stable group ($p = 0.043$) and Improved group ($p = 0.046$).

Patients were further divided into biochemically controlled ($n = 16$, posttreatment GH levels $< 2.5 \mu g/l$) group and uncontrolled ($n = 35$, posttreatment GH levels $\geq 2.5 \mu g/l$) group based on posttreatment GH levels. As shown in Supplementary Table 8, We found a trend toward a decrease...
<table>
<thead>
<tr>
<th>Table 1: Changes of variables in NGT, IGT, and DM groups from pretreatment to after SSA treatment.</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>The entire cohort (n = 64)</strong></td>
</tr>
<tr>
<td>--------------------------------</td>
</tr>
<tr>
<td><strong>HbA1c (%)</strong></td>
</tr>
<tr>
<td><strong>FPG (mmol/l)</strong></td>
</tr>
<tr>
<td><strong>BG120 (mmol/l)</strong></td>
</tr>
<tr>
<td><strong>FPI (mU/l)</strong></td>
</tr>
<tr>
<td><strong>INS120 (mU/l)</strong></td>
</tr>
<tr>
<td><strong>HOMA-IR</strong></td>
</tr>
<tr>
<td><strong>ISOGTT</strong></td>
</tr>
<tr>
<td><strong>HOMA-β (%)</strong></td>
</tr>
<tr>
<td><strong>INS/BG0</strong></td>
</tr>
<tr>
<td><strong>IGI</strong></td>
</tr>
<tr>
<td><strong>IGI/IR</strong></td>
</tr>
<tr>
<td><strong>ISS12</strong></td>
</tr>
<tr>
<td><strong>AUCINS/AUCBG</strong></td>
</tr>
</tbody>
</table>

NGT: normal glucose tolerance; IGT: impaired glucose tolerance; DM: diabetes mellitus; IGF-1 index: the ratio of the measured IGF-1 value to the upper limit of normal (ULN); FPI: fasting plasma glucose; BG120: plasma glucose 120 min during OGTT; AUCBG, the areas under the curve of glucose; FPI: fasting plasma insulin; INS120: plasma insulin 120 min during OGTT; AUCINS: the areas under the curve of insulin; HOMA-IR: indicator of insulin resistance; ISOGTT: the OGTT insulin sensitivity index; HOMA-β: homeostatic model assessment of pancreatic beta-cell function; IGI: insulinogenic index; ISS12: the OGTT insulin secretion sensitivity index. p values are for variations before and after SSA treatment; *p < 0.05.
on HbA1c (6.09 ± 1.32 versus 5.81 ± 0.64%), FPG (5.68 ± 1.75 versus 5.53 ± 0.45 mmol/l), and BG120 (8.75 versus 7.85 mmol/l) in the controlled group. As for the change in insulin resistance and secretion, we found that after treatment, insulin resistance, represented by ISOGTT, was significantly improved in both groups. And all variables reflective of insulin secretion except ISSI2 declined in both groups (Supplementary Table 8).

### 3.7 Correlation Studies

In the group as a whole, the reduction in HbA1c positively correlated with the reduction in GHm (r = 0.348, p = 0.018, Figure 2(a)) and negatively correlated with the reduction of ISSI2 (r = -0.408, p = 0.003, Figure 2(b)), IGI (r = -0.294, p = 0.032), and IGI/IR (r = -0.273, p = 0.048) after SSA treatment (Supplementary Table 9).

### 3.8 The Predictive Value of Baseline BG120 for the Effect of SSA Treatment on Glucose Metabolism

ROC curve analysis was performed to further estimate the predictive value of BG120 on the change of glucose tolerance status. The cutoff value of baseline BG120 was 8.1 mmol/l which demonstrated the greatest sensitivity and specificity in predicting the stability and/or improvement of glycemic status after SSA treatment, with a PPV of 90.7% and a NPV of 66.7% (sensitivity 84.8%, specificity 77.8%, AUC = 0.844, p < 0.001, Figure 3). Patients were categorized into two groups according to BG120 at baseline: group A (BG120 greater than 8.1 mmol/l) and group B (BG120 less than 8.1 mmol/l). First, we compared these two groups at baseline. We found that IGI (p = 0.001), IGI/IR (p < 0.001), and ISSI2 (p < 0.001) were higher in group B than in group A (Supplementary Table 10). Second, the changes in variables after SSA treatment were analyzed (Supplementary Table 11). We found that the reduction of GHm was less in group B than in group A (p = 0.019), while the reduction of HOMA-β, IGI, IGI/IR, and ISSI2 was more

**Figure 1:** Flowchart of prevalence of NGT, IGT, and DM at pretreatment and after SSA treatment, and the change of glucose metabolism status after SSA therapy. NGT: normal glucose tolerance; IGT: impaired glucose tolerance; DM: diabetes mellitus.

**Table 2:** Comparison of baseline characteristics of patients in the Improved/Stable/Deteriorated glucose tolerance status groups.

<table>
<thead>
<tr>
<th></th>
<th>Improved (n = 18)</th>
<th>Change in glucose status</th>
<th>Deteriorated (n = 18)</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Female [n/()]</td>
<td>11 (61.1)</td>
<td>17 (60.7)</td>
<td>10 (55.6)</td>
<td>0.927</td>
</tr>
<tr>
<td>Age (years)</td>
<td>38.4 ± 9.7</td>
<td>46.1 ± 13.2</td>
<td>38.1 ± 13.7</td>
<td>0.049*</td>
</tr>
<tr>
<td>BMI (kg/m2)</td>
<td>21.99 ± 8.32</td>
<td>24.79 ± 6.26</td>
<td>24.99 ± 7.79</td>
<td>0.378</td>
</tr>
<tr>
<td>GHm (pg/l)</td>
<td>40.72 (25.88–86.15)</td>
<td>27.98 (15.69–70.23)</td>
<td>22.61 (12.65–49.69)</td>
<td>0.172</td>
</tr>
<tr>
<td>IGF-1 index</td>
<td>2.87 ± 1.05</td>
<td>2.94 ± 0.89</td>
<td>2.58 ± 0.65</td>
<td>0.489</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>5.80 (5.50–6.25)</td>
<td>6.30 (5.60–9.18)</td>
<td>5.70 (5.60–5.88)</td>
<td>0.196</td>
</tr>
<tr>
<td>FPG (mmol/l)</td>
<td>5.50 (5.18–6.18)</td>
<td>5.75 (5.03–6.70)</td>
<td>5.30 (5.10–5.80)</td>
<td>0.254</td>
</tr>
<tr>
<td>BG120 (mmol/l)</td>
<td>9.25 (8.68–12.73)</td>
<td>11.40 (7.83–17.13)</td>
<td>6.80 (5.58–8.23)</td>
<td>&lt;0.001^</td>
</tr>
<tr>
<td>FPI (mU/l)</td>
<td>17.65 (11.55–26.38)</td>
<td>13.02 (8.66–23.53)</td>
<td>16.40 (12.13–30.58)</td>
<td>0.329</td>
</tr>
<tr>
<td>INS120 (mU/l)</td>
<td>174.05 (63.53–248.98)</td>
<td>69.80 (25.18–152.33)</td>
<td>73.65 (48.90–189.85)</td>
<td>0.166</td>
</tr>
</tbody>
</table>

IGF-1 index: the ratio of the measured IGF-1 value to the upper limit of normal (ULN); HbA1c: glycosylated hemoglobin; FPG: fasting plasma glucose; BG120: plasma glucose 120 min during OGTT; FPI: fasting plasma insulin; INS120: plasma insulin 120 min during OGTT; p values are for variations among the 3 groups; ^p < 0.05.
The correlation between the reduction of HbA1c and the reduction of ISSI2 remained stable in the DM group. BG120 increased in both NGT and IGT groups, with the baseline status of glucose metabolism in patients. The change in glucose metabolism-related variables after SSA treatment among the Improved/Stable/Deteriorated groups are shown in Table 3.

FIGURE 2: The reduction of HbA1c was positively correlated with the reduction of GHm (a) and negatively correlated with the reduction of IGF-1 (b) after SSA treatment in the entire cohort. (a) The correlation between the reduction of HbA1c and the reduction of GHm. (b) The correlation between the reduction of HbA1c and the reduction of ISSI2. Correlation coefficients and p values were shown for each correlation.

in group B than in group A (p = 0.037, 0.002, 0.008, and 0.046, resp.).

4. Discussion

In the present study, we demonstrated that the change in glucose metabolic status after SSA therapy strongly correlated with the baseline status of glucose metabolism in patients with acromegaly. FPG rose in both NGT and IGT groups, but remained stable in the DM group. BG120 increased in the NGT group, stabilized in the IGT group, and decreased in the DM group. Insulin resistance was improved in all 3 groups, while insulin secretion declined in the NTG and IGT groups and was unchanged in the DM group. The glucose tolerance status was improved in 28.1% patients, deteriorated in 28.1% patients, and stabilized in 43.8% patients. Deterioration was associated with lower baseline BG120, less reduction in GHm, and a greater reduction in insulin secretion after SSA therapy. The cutoff value of BG120 (8.1 mmol/l) at baseline predicted the stabilization and/or improvement of glucose metabolism during SSA treatment.

The impact of SSA on glucose metabolism has been studied, but the results are conflicting [6–9]. Several studies have reported no change of glucose levels after SSA treatment [6, 25]. A meta-analysis also indicated that SSA might have an overall minor impact on glucose homeostasis in patients with acromegaly [26]. However, others found that SSA significantly aggravated glucose tolerance in patients with acromegaly [23, 29–31], thus mandating glucose monitoring during SSA therapy. Interestingly, Ho et al. even reported that SSA has beneficial effects on carbohydrate metabolism in patients with acromegaly and glucose intolerance [30]. In

<table>
<thead>
<tr>
<th>Reduction (post-SSA)</th>
<th>Improved (n = 18)</th>
<th>Stable (n = 28)</th>
<th>Deteriorated (n = 18)</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>GHm (μg/l)</td>
<td>-28.01 (−53.71−10.80)</td>
<td>-15.55 (−32.89−6.50)</td>
<td>-5.89 (−16.55−6.69)</td>
<td>0.021†</td>
</tr>
<tr>
<td>IGF-1</td>
<td>-0.64 (−1.40−0.26)</td>
<td>-1.16 (−1.44−0.42)</td>
<td>-0.60 (−1.58−0.18)</td>
<td>0.353</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>-0.75 (−4.15−0.00)</td>
<td>-1.45 (−2.98−0.28)</td>
<td>-1.17 (−2.41−0.26)</td>
<td>0.830</td>
</tr>
<tr>
<td>ISOGTT</td>
<td>26.01 (5.16−54.78)</td>
<td>26.01 (5.23−53.45)</td>
<td>15.13 (0.54−51.43)</td>
<td>0.781</td>
</tr>
<tr>
<td>HOMA-β (%)</td>
<td>-56.15 (−103.77−4.03)*</td>
<td>-36.94 (−140.44−0.03)*</td>
<td>-85.52 (−206.70−65.53)</td>
<td>0.074</td>
</tr>
<tr>
<td>INS0/BG0</td>
<td>-0.78 (−2.65−0.14)</td>
<td>-0.64 (−1.66−0.13)</td>
<td>-1.16 (−3.14−0.81)</td>
<td>0.121</td>
</tr>
<tr>
<td>IGI/IR</td>
<td>-1.26 (−4.46−0.14)</td>
<td>-0.47 (−6.36−0.13)</td>
<td>-2.28 (−7.23−0.19)</td>
<td>0.844</td>
</tr>
<tr>
<td>ISSI2</td>
<td>36.16 (−118.62−113.34)</td>
<td>15.97 (−59.98−54.94)</td>
<td>84.97 (−158.72−100.10)</td>
<td>0.319</td>
</tr>
<tr>
<td>IGI</td>
<td>-17.10 (−28.02−5.78)</td>
<td>-1.79 (−30.96−0.07)</td>
<td>-18.69 (−41.77−5.24)</td>
<td>0.350</td>
</tr>
<tr>
<td>AUCINS/AUCBG</td>
<td>-4.76 (−10.44−1.96)</td>
<td>-2.42 (−8.32−0.43)</td>
<td>-4.40 (−13.00−1.93)</td>
<td>0.244</td>
</tr>
</tbody>
</table>

*IGF-1 index: the ratio of the measured IGF-1 value to the upper limit of normal (ULN); HOMA-IR indicator of insulin resistance; ISOGTT: the OGTT insulin sensitivity index; HOMA-β: homeostatic model assessment of pancreatic beta-cell function; INS0: fasting plasma insulin; BG0: fasting plasma glucose; IGI: insulinogetic index; AUCBG: the areas under the curve of glucose; AUCINS: the areas under the curve of insulin; ISSI2: the OGTT insulin secretion sensitivity index-2. p values are for variations before and after SSA treatment; †p < 0.05 versus the Deteriorated group; ‡p < 0.05.
our study, the predominant pattern of change in glucose tolerance status was deterioration in the baseline NGT group, stabilization in the baseline IGT group, and amelioration in the baseline NGT group. The central line indicates neutrality, and the arrow shows the baseline BG$_{120}$ 8.1 mmol/l during the OGTT with a PPV of 90.7% and a NPV of 66.7% (AUC = 0.844, $p < 0.001$).

Recently, pasireotide was approved for acromegaly and showed more efficacy in controlling GH and IGF-1 levels [32]. As for the effects on glucose metabolism, a head-to-head study has reported that compared with octreotide LAR, hyperglycemia-related adverse events were more common with pasireotide [33].

The change in glucose metabolism correlated strongly with the change of insulin resistance and insulin secretion after SSA treatment [34]. Ronchi et al. found that HOMA-IR significantly declined during SSA treatment [9]. Baldelli et al. found that insulin resistance was improved but the insulin secretion was 30 minutes delayed after 6 months of SSA therapy [27]. However, Steffin et al. found that SSA decreased $\beta$-cell function without affecting insulin resistance [35]. In the present study, we used not only HOMA but also various derivatives of the OGTT to evaluate insulin sensitivity and $\beta$-cell function. For insulin sensitivity, HOMA-IR decreased in the NGT and IGT groups and remained unaltered in the DM group, while IS$_{OGTT}$, another major parameter reflecting insulin resistance, improved in all groups. Matsuda et al. first developed the IS$_{OGTT}$ index and proved IS$_{OGTT}$ to be a reasonable and better approximation of whole-body insulin sensitivity in patients with diabetes mellitus than HOMA [15]. This might be applicable to patients with acromegaly. Variables reflective of $\beta$-cell function, such as HOMA-$\beta$, INS$_{c}$/BG$_{120}$, IGI/IR, and IGI, declined in both NGT and IGT groups, but remained unchanged in the DM group. The above results showed SSA decreased insulin secretion in NGT and IGT groups, but had no effect in the DM group.

Excess GH levels led to insulin resistance in both NGT and DM patients, and SSA therapy could significantly reduce GH levels, resulting in the decrease of insulin resistance both in NGT and DM groups. Meanwhile, insulin secretion was decreased after SSA treatment in the NGT group, but was not compromised in the DM group. Thus, the glucose metabolic status was generally improved after SSA administration in the DM group due to the alleviated degree of insulin resistance without compromise of insulin secretion. But in the NGT group, the glucose metabolic status might even deteriorate if the reduction of insulin secretion overcomes the improvement of insulin resistance. This may be the potential underlying mechanism for the different effects of SSA on glucose metabolism in patients with NGT and patients with DM.

Several studies revealed factors associated with the SSA-induced changes in glucose tolerance status. Koop et al. stated that female patients and those with higher baseline insulin levels were more likely to develop DM during SSA therapy [11]. Ho et al. found that improvement in glucose tolerance status was dependent on pretreatment BG concentrations [30]. Colao et al. found that deterioration of glucose metabolism was correlated with increased BMI, uncontrolled acromegaly during SSA therapy, and abnormal glucose tolerance at baseline [28, 29]. In the present study, we showed that the deterioration of glucose tolerance was associated with less reduction of GH and greater reduction in insulin secretion after SSA therapy. In addition, the reduction of HbA$_{1c}$ was positively correlated with the reduction of GH$_{m}$ and negatively correlated with the reduction of insulin secretion. Interestingly, we found that SSA administration can significantly improve insulin resistance with a compromise in insulin secretion, in patients with both biochemically controlled (posttreatment GH levels < 2.5 $\mu$g/l) and uncontrolled (posttreatment GH levels $\geq$ 2.5 $\mu$g/l) acromegaly, which was similar with Giordano et al. [36]. Some discrepancy (e.g., IGI) may be related to the different races and duration of SSA treatment (3 months in our study, ≥12 months in literature) between studies. When exploring potential baseline predictors, we found that the baseline BG$_{120}$ was significantly lower in patients whose glucose status deteriorated. Furthermore, for the first time, we generated ROC curves to obtain the most sensitive and specific cutoff values which predicted the change of glucose metabolism after SSA therapy. We showed that when the baseline BG$_{120}$ was higher than 8.1 mmol/l, there was a 90.7% chance of stabilized and/or improved glucose tolerance status. However, when the
baseline $BG_{120}$ was lower than 8.1 mmol/l, there was a 66.7% chance of deterioration in glucose tolerance status. To explore the potential mechanism, we examined the difference between patients with baseline $BG_{120}$ above 8.1 mmol/l and those with baseline $BG_{120}$ below 8.1 mmol/l. Interestingly, we found that patients with baseline $BG_{120}$ below 8.1 mmol/l had less of a reduction in $GH_m$ and a greater reduction in $\beta$-cell function. Less reduction of $GH_m$ led to less improvement in insulin resistance in patients with baseline $BG_{120}$ below 8.1 mmol/l. In addition to less improvement in insulin resistance, patients with baseline $BG_{120}$ below 8.1 mmol/l had a greater reduction in insulin secretion, which indicated that there was more chance of deteriorating glucose tolerance status after SSA treatment in these subjects. Thus, vice versa, baseline $BG_{120}$ higher than 8.1 mmol/l after OGTT may be considered as a beneficial predictive factor for glucose metabolism during SSA treatment. This seemed to be discordant with a previous study indicating that baseline glucose status was one of the major predictors of changing glucose status [29]. But actually, in our study, the percentage of improved glucose metabolism in the IGT group tended to be more than in the DM group [45.8% (11/24) versus 33.3% (7/21)], which was consistent with the study of Colao et al.

The limitation of the current study is that this study is not a blinded study from a patient’s point of view and patients who are diagnosed with diabetes mellitus or impaired glucose tolerance at pretreatment assessment may have lifestyle/dietary modification, which may have had an impact on the glucose metabolism results in the follow-up assessment.

In conclusion, the impact of SSA on the change in glucose metabolic status, insulin resistance, and $\beta$-cell function depends on the pretreatment glucose metabolism status in patients with acromegaly. Deterioration is associated with lower baseline $BG_{120}$, the less of a reduction in $GH_m$, and a greater reduction in insulin secretion after SSA therapy. $BG_{120}$ during OGTT can predict the impact of SSA treatment on glucose metabolism.

**Ethical Approval**

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. For this type of study, formal consent is not required.

**Consent**

Informed consent was obtained from all individual participants included in the study.

**Conflicts of Interest**

The authors declare that they have no conflict of interest.

**Authors’ Contributions**

Ming Shen and Meng Wang contributed equally to this work.

**Acknowledgments**

The authors thank Dr. Karen Klahr Miller at Massachusetts General Hospital for helpful comments and edits with the manuscript. This work was supported by the following grants: from the National Natural Science Foundation of China (nos. 81370938, 81172391, 81370884, and 81602191), Shanghai Rising-Star Tracking Program (12QH1400400), and Shanghai Municipal Commission of Health and Family Planning (nos. XYQ20111002, XYQ2013120, XYQ20134280, and XYQ201640058).

**Supplementary Materials**

Supplementary Table 1: baseline characteristics of the 64 patients. Supplementary Table 2: patients’ data before and after SSA therapy. Supplementary Table 3: OGTT and glucose tolerance status before and after SSA therapy. Supplementary Table 4: insulin levels during OGTT before and after SSA therapy. Supplementary Table 5: comparison of pretreatment variables among NGT/IGT/DM groups. Supplementary Table 6: the baseline characteristics of the female group and male group. Supplementary Table 7: changes of variables in female and male groups from pretreatment to after SSA treatment. Supplementary Table 8: changes of variables in “controlled” and “uncontrolled” patients from pretreatment to after SSA treatment. Supplementary Table 9: correlation between the changes of HbA1c and glucose metabolism-related variables after SSA treatment. Supplementary Table 10: comparison of baseline characteristics between group A and B. Supplementary Table 11: comparison of the change in variables after SSA treatment between groups A and B. (Supplementary Materials)

**References**


International Journal of Endocrinology


