

Research Article

Hepatocyte Lysosomal Membrane Stabilization by Olive Leaves against Chemically Induced Hepatocellular Neoplasia in Rats

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Received 1 July 2010; Revised 29 September 2010; Accepted 17 October 2010

Academic Editor: Masahiko Hirota

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Extensive efforts are exerted looking for safe and effective chemotherapy for hepatocellular carcinoma (HCC). Specific and sensitive early biomarkers for HCC still in query. Present work to study proteolytic activity and lysosomal membrane integrity by hepatocarcinogen, trichloroacetic acid (TCA), in Wistar rats against aqueous olive leaf extract (AOLE). TCA showed neoplastic changes as oval- or irregular-shaped hepatocytes and transformed, vesiculated, and binucleated liver cells. The nuclei were pleomorphic and hyperchromatic. These changes were considerably reduced by AOLE. The results added, probably for the first time, that TCA-induced HCC through disruption of hepatocellular proteolytic enzymes as upregulation of papain, free cathepsin-D and nonsignificant destabilization of lysosomal membrane integrity, a prerequisite for cancer invasion and metastasis. AOLE introduced a promising therapeutic value in liver cancer, mostly through elevating lysosomal membrane integrity. The study substantiated four main points: (1) the usefulness of proteolysis and lysosomal membrane integrity in early prediction of HCC. (2) TCA carcinogenesis is possibly mediated by lysosomal membrane destabilization, through cathepsin-D disruption, which could be reversed by AOLE administration. (3) A new strategy for management of HCC, using dietary olive leaf system may be a helpful phytotherapeutic trend. (4) A prospective study on serum proteolytic enzyme activity may introduce novel diagnostic tools.

1. Introduction

The metabolism of functional proteins is a continuous process involving proteolysis, which is the destruction of individual protein molecules and their replacement through protein synthesis. Intracellular proteolysis includes either lysosomal (cathepsins) or nonlysosomal pathway, which is mostly executed by calpains and proteasomes [1–3]. The rate of degradation of both intracellular and extracellular proteins is greatly dependent on cathepsin activity.

The degradation rate of proteins can be monitored by measuring cathepsin activity oscillations [4]. Lysosomes are membrane-bound structures containing hydrolytic enzymes capable of degrading most of the cellular constituents. They also play a pivotal role in secretion and transport processes. Leakage of lysosomal enzymes accounts for many tissue derangements and target organopathies [5, 6]. Lysosomes are found in all animal cells and are more numerous in disease-fighting cells.

Lysosomal enzyme disorders contribute to several human diseases, either due to genetic defects in its enzyme expression or the escape of lysosomal enzymes (lysozymes) into extralysosomal medium [7]. Hepatocellular carcinoma (HCC) is considered as one of the worst prognostic cancers in the world. It develops mostly over chronic liver diseases of cirrhotic fate. HCC nodules are always encapsulated by excessive extracellular matrix (ECM) materials among a bed of cirrhotic tissue [8, 9]. ECM degradation by specific proteases was reported to play an important role in cancer invasion and metastasis [10]. It was noticed that some herbals could ameliorate anticancer-induced lysosomal abnormalities, conserving lysosomal integrity, probably, through an antioxidant mechanism [6].

Studies of the trichloroethylene (TCE) metabolites, dichloroacetic acid (DCA), trichloroacetic acid (TCA), and chloral hydrate suggested that both DCA and TCA are involved in TCE-induced liver tumorigenesis and that many

DCA effects are consistent with conditions that increase the risk of liver cancer [11–13].

Olive tree (*Olea europaea*, Oleaceae) is a longevous plant, anciently known in the Mediterranean basin [14]. Olive leaves are rich in active constituents showing medicinal value. They possess many water-soluble compounds of antioxidant and anti-inflammatory properties [15]. The most important bioactive compounds are oleuropein (13.4%) and rutin (0.18%), luteolin-7-glucoside, verbascoside, apigenin-7-glucoside, hydroxytyrosol, and some other hydrolytic products, both of glycosidic and flavonoid nature [16, 17]. The leaves also contain many terpenoids having hepatoprotective and cancer-preventive potential [18]. Besides, Guinda et al. [19] had described many liposoluble principles in olive leaf, as squalene, carotenoid, vitamin E, alcohols, and saturated hydrocarbons. Many other ingredients were studied in olive leaf of different medicinal and nutritional values [20]. It is now established that olive polyphenols exert chemopreventative effects in the large intestine by interacting with signaling pathways responsible for colorectal cancer development [21]. The present work was executed to explore the possible therapeutic value of an aqueous extract of olive leaves which possess polyphenolic flavonoids, taking into account the cost-effectiveness of olive leaves as a nonordinarily utilized part and to try its efficacy in hepatocarcinoma treatment, through monitoring the early variations in liver tissue proteolytic activity, as specific enzyme, pepsin, trypsin, papain, and cathepsin enzyme activities. The total/free cathepsin activity ratio was considered as an index to lysosomal membrane integrity.

2. Materials and Methods

2.1. Materials

2.1.1. Animals. Male Wistar rats weighing 190 ± 10 g were used in this experiment. These rats were given ordinary rodent diet and water ad libitum. They were housed in polyethylene cages in a humid room with controlled 12 hours light and 12 hours dark cycle. They were classified into 4 groups (8 rats/group).

2.1.2. Chemicals. All chemicals were supported from local suppliers, all were of analytical grade. Tyrosine was purchased from Merck.

2.1.3. Aqueous Olive Leaves Extract (AOLE). It was kindly given by Dr. Ashraf El-Bassiouny, Natural Product Chemistry Department, Institute of Education and Science, Beni Suf University, Egypt. It was prepared by drying for 8 hours, at 40°C , water-washed fresh olive leaves collected on February of 2007, milled, homogenized and then extracted by heating with distilled water with stirring at 40°C for 48 hours as 10 g powder of dried leaves/60 CC solvent, cooled, filtered, and the supernatant was kept at 4°C during the period of administration [22]. This extract is dark-green in color, clear, fragrant, and has a little sweet taste.

2.1.4. Drug Administration Schedules per Groups. The first group of rats served as control, in which animals were given 0.5 ml physiological saline by intraperitoneal route (IP), daily, for one month. The second group served as liver cancer group, given a daily IP dose of trichloroacetic acid (TCA) (Sigma-Aldrich, USA), for 5 consecutive days, by gavage as 500 mg/kg body weight, previously dissolved in distilled water, neutralized by NaOH to a pH of 5–7.5 [23]. Animals of the third group were given AOLE (softened by drying under vacuum) by gavage in a single daily dose for one month as 0.5 g/kg body weight [24]. This group was considered as positive drug control.

Animals of group four were given doses of TCA for 5 consecutive days, 24 hours after the last dose, AOLE doses were given as stated before. Twenty-four hours after each dosing period, animals were killed, livers were excised, washed with saline, frozen directly into liquid nitrogen and kept frozen at -80°C right at the day of analytical investigations.

2.2. Methods. (1) Extraction of the target enzymes from liver tissue: 300 mg of liver tissue was homogenized with 6 ml ice-cold distilled water as described elsewhere [25]. Total protein was determined in each enzyme substrate prior to enzyme assessment, by the method Lowry et al. [26].

(2) Determination of specific proteolytic enzyme activity was achieved in the tissue extract on a substrate containing 1% egg albumin as the substrate protein for enzyme actions [27]. Colorimetric estimation of tyrosine, as the product of proteolysis was conducted by diluted Folin-Ciocalteu (FC) reagent, [28] against tyrosine standard curve [29].

(3) Estimation of pepsin, trypsin, and papain proteolytic activity was accomplished through changing the pH of the protein substrate per each target enzyme as stated by Anson [27], then, tyrosine was estimated as mentioned earlier [25, 29].

(4) Estimation of lysosomal membrane integrity: this variable relied on determination of cathepsin proteolytic activity in the tissue extract by acidifying the protein substrate (free cathepsin-D activity) and after dilution of the extract, then, frequent cycles of freezing and thawing were applied (total cathepsin-D activity) [30–33]. Enzyme activity in all cases was considered as μg -released tyrosine/mg substrate protein. Total/free cathepsin D activity was considered as a measure of lysosomal membrane integrity [31, 32].

(5) Statistical analysis: the presented data were expressed as mean \pm SE. Statistical significance was examined by one-way analysis of variance (ANOVA), [34]. *P* values less than .05 were assumed to be statistically significant.

(6) Liver samples were stained by Hematoxylin and Eosin, examined microscopically to follow tissue changes in response to TCA and AOLE therapy.

3. Results

TCA administration showed loss of normal architecture with oval- or irregular-shaped hepatocytes. Many transformed liver cells of foci were substantially enlarged, largely vesiculated and frequently binucleated, which were clearly

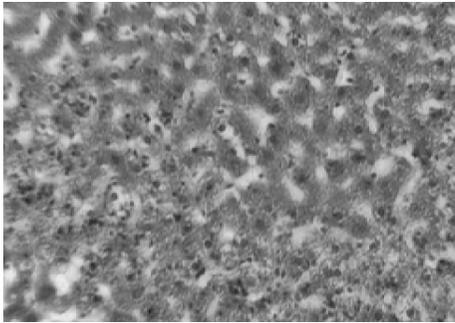


FIGURE 1: Liver section in control animals showing normal parenchymal cells with granulated cytoplasm and normal architecture.

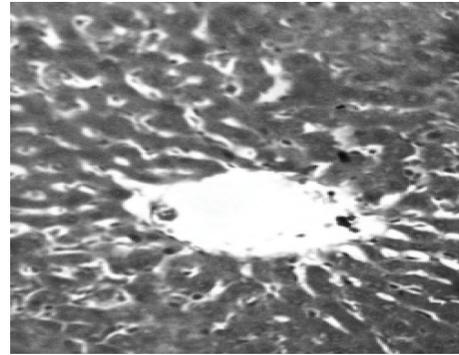
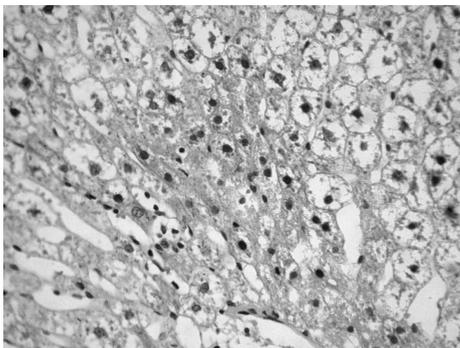
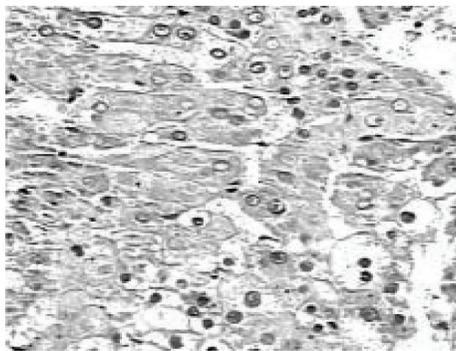


FIGURE 3: Liver section in AOLE-treated rats shows a normal central vein and blood sinusoids.



(a)



(b)

FIGURE 2: TCA-treated rats showing loss of normal architecture with oval- or irregular-shaped hepatocytes, denser nuclear chromatin, the ratio of nucleus to cytoplasm was increased, many transformed liver cells were frequently binucleated and hyperchromatic (basophilic).

distinguishable from the surrounding normal parenchyma. The nuclei were mostly found to be pleomorphic and hyperchromatic. These changes were reduced by the treatment with AOLE (Figures 1, 2, 3, and 4).

The biochemical results of the experiment are summarized in Table 1. As shown, TCA treatment significantly upregulated both papain and free cathepsin-D and non-significantly elevated specific enzyme, pepsin, trypsin, and

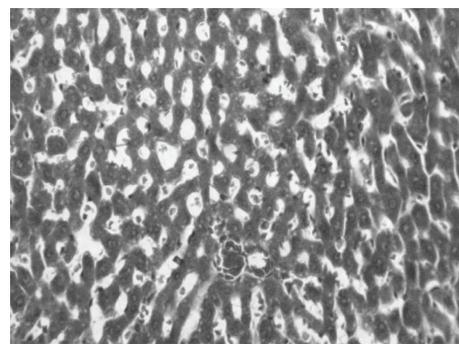


FIGURE 4: Liver section in TCA- and AOLE-treated rats showing a moderate improvement in hepatocellular structure, evidenced as a moderate improvement in vacuolation and compactness of hepatocytes compared to TCA-treated group but dilated sinusoids are still present.

total cathepsin-D activities, with nonsignificant reduction of lysosomal membrane integrity.

Administration of AOLE alone did not significantly affect the studied variables, but only significantly upregulated papain activity. Treatment of TCA-injured rats with AOLE significantly elevated trypsin and papain, with reduction of pepsin activities. It significantly decreased free cathepsin-D activity, leading to significant increase in lysosomal integrity index.

4. Discussion

Lysosomal proteolytic activity in liver cells plays a decisive role in protein metabolism, with cathepsin-A and cathepsin-D being the main enzymes involved in catabolic processes [35]. Tumor cells require specific proteolytic enzymes for invasion and metastasis, including lysosomal peptidases-cathepsin [36].

Recently, it was noticed that proteases play a critical role not only in tumor cell invasion, but also in the earliest stages of carcinogenesis and its associated changes: angiogenesis and metastasis. Proteases are also signaling molecules that modulate other molecules by underlying pathways in addition to their degradative role [37].

TABLE 1: Proteolytic enzyme activities (μg -released tyrosine/mg substrate protein) and lysosomal integrity (total/free cathepsin-D activity) in liver tissue of rats treated by aqueous olive leaf extract (for one month) after trichloroacetic acid carcinogenic challenge (for 5 days). Values are expressed as mean \pm S.E ($N = 8$).

Parameter	Group			
	Control	TCA (500 mg/kg)	AOLE (500 mg/kg)	TCA + AOLE (500 mg/kg + 500 mg/kg)
Specific enzyme activity	24.2 \pm 0.55	24.9 \pm 0.32	24.4 \pm 0.39	24.7 \pm 0.3
Pepsin activity	15.5 \pm 0.13	15.8 \pm 0.13	15.3 \pm 0.13	12.3** \pm 0.25
Trypsin activity	11.3 \pm 0.3	11.9 \pm 0.2	11.4 \pm 0.6	13.3** \pm 0.2
Papain activity	18 \pm 1.1	22* \pm 1.3	25** \pm 0.9	24** \pm 0.3
Free cathepsin-D activity	5.1 \pm 0.1	6.2** \pm 0.2	5.5 \pm 0.2	4.5* \pm 0.1
Total cathepsin-D activity	3.6 \pm 0.13	4 \pm 0.05	4 \pm 0.05	4.4** \pm 0.04
Lysosomal integrity	0.72 \pm 0.03	0.65 \pm 0.02	0.74 \pm 0.03	0.99** \pm 0.04

*Significantly different from control at ** $P < .05$. Significantly different from control at $P < .01$.

In the present work, TCA treatment depicted non-significant upregulations in specific enzyme, pepsin, trypsin, and total cathepsin-D activities, with decrease in lysosomal integrity; however, it significantly activated both papain and free cathepsin-D activity. Specific proteolytic enzymes, including endopeptidases are necessary prognostic indicators (prognosticators) after tumor resection. They always rise up in patients at risk of recurrence [38]. TCA is one of three metabolites of trichloroethylene, a common solvent known classically to pollute groundwater. They are nongenotoxic carcinogens [23]. TCA is also considered as a member of peroxisome proliferators, responsible for many mutations, leading to diverse-target organ cancers [3].

This early upregulatory impact of TCA on proteolytic enzymes may contribute to the mechanism of cancer induction and invasion by this substance, an action that was noticed in some previous publications [39, 40]. Administration of AOLE alone maintained some proteolytic enzyme activities within normal values, except an elevation of papain activity to a significant level. The modulatory action on both total and free cathepsin-D activities could preserve lysosomal membrane integrity, thus protecting cellular biomolecules. This lysosomal-stabilizing potential of AOLE is mostly referred to its potent antioxidant and anti-inflammatory properties [16].

It is clear in these results that disruption of lysosomes will lead to increased proteolytic activity, while preservation will keep cellular components and protect against neoplastic invasiveness [41]. In addition, AOLE, when given after TCA challenge, corrected the proteolytic action and restabilized lysosomal membrane to a significant value. It seems that the main therapeutic pathway for this leaf extract is mediated by pepsin and cathepsin-D variation in favour of lysosomal membrane stabilization. It is well established that cancer can induce enzyme synthesis to an extent of leakage into pericellular components [42]. However, addition of AOLE may induce increased proteolytic enzymes that might hydrolyse angiogenic proteins useful for de novo cancer progression [37].

The use of olive leaves introduces supplementary treatment in cancer management, instead of cytotoxicity and cytodestruction induced by chemotherapeutic agents. It may modulate the cellular phenotypic characteristics attend-stage

of adult cells leading to programmed cell death (apoptosis) [43]. Recent study proved the usefulness of olive leaf extract in inhibition of differentiation of human leukemia cell line [44].

It can be concluded that olive leaf consumption has health benefits in favouring liver cancer management. It can be recommended as a dietary supplement during liver cancer management and even after cancer protocol is stopped to guarantee recurrence. TCA can be considered as a carcinogen of proteolytic potential, by which it initiates hepatocarcinoma. Early changes in lysosomal membrane integrity, manifested by increased release of proteases may add a new line of research to figure out more specific and sensitive early biomarkers as diagnostic tools. This change in proteolytic activity necessitates further studies on blood to assess a possible escape of these studied enzymes into circulation. This may contribute to a new trend, both in early HCC assessment and prognosis.

References

- [1] D. E. Goll, W. C. Kleese, and A. Szpacinko, "Skeletal muscle proteases and protein turnover," in *Animal Growth Regulation*, D. R. Campion, G. J. Hausman, and R. J. Martin, Eds., pp. 141–182, Plenum Publishing, New York, NY, USA, 1989.
- [2] S. Johari, Y. Maeda, S. Okamoto, and T. Hashiguchi, "Comparison of calpain and calpastatin activities in skeletal muscle of broiler and layer chickens," *British Poultry Science*, vol. 34, no. 4, pp. 819–824, 1993.
- [3] M. G. Thompson and R. M. Palmer, "Signalling pathways regulating protein turnover in skeletal muscle," *Cellular Signalling*, vol. 10, no. 1, pp. 1–11, 1998.
- [4] D. J. Millward, P. C. Bates, and S. Rosochacki, "The extent and nature of protein degradation in the tissues during development," *Reproduction Nutrition Developpement*, vol. 21, no. 2, pp. 265–277, 1981.
- [5] N. Gebbia, G. Leto, and M. Gagliano, "Lysosomal alterations in heart and liver of mice treated with doxorubicin," *Cancer Chemotherapy and Pharmacology*, vol. 15, no. 1, pp. 26–30, 1985.
- [6] R. Subashini, A. Gnanapragasam, S. Senthilkumar, S. K. Yogeeta, and T. Devaki, "Protective efficacy of *Nardostachys jatamansi* (Rhizomes) on mitochondrial respiration and lysosomal hydrolases during doxorubicin induced myocardial

- injury in rats," *Journal of Health Science*, vol. 53, no. 1, pp. 67–76, 2007.
- [7] R. K. Murray, D. K. A. Granner, P. Mayes, and V. W. Rodwell, "Cell diseases resulting from faulty targeting of lysosomal enzymes and genetic deficiencies of glycoprotein lysosomal hydrolases cause diseases such as mannosidosis," in *Harper's Biochemistry*, pp. 691–692, Appleton and Lange, San Mateo, Calif, USA, 1999.
- [8] T. Torimura, T. Ueno, S. Inuzuka, M. Tanaka, H. Abe, and K. Tanikawa, "Mechanism of fibrous capsule formation surrounding hepatocellular carcinoma: immunohistochemical study," *Archives of Pathology and Laboratory Medicine*, vol. 115, no. 4, pp. 365–371, 1991.
- [9] I. O. L. Ng, E. C. S. Lai, M. M. T. Ng, and S. T. Fan, "Tumor encapsulation in hepatocellular carcinoma: a pathologic study of 189 cases," *Cancer*, vol. 70, no. 1, pp. 45–49, 1992.
- [10] W. G. Stetler-Stevenson, S. Aznavoorian, and L. A. Liotta, "Tumor cell interactions with the extracellular matrix during invasion and metastasis," *Annual Review of Cell Biology*, vol. 9, pp. 541–573, 1993.
- [11] C. H. Anna, R. R. Maronpot, M. A. Pereira, J. F. Foley, D. E. Malarkey, and M. W. Anderson, "Ras proto-oncogene activation in dichloroacetic acid-, trichloroethylene- and tetrachloroethylene-induced liver tumors in B6C3F1 mice," *Carcinogenesis*, vol. 15, no. 10, pp. 2255–2261, 1994.
- [12] A. Ferreira-Gonzalez, A. B. DeAngelo, S. Nasim, and C. T. Garrett, "Ras oncogene activation during hepatocarcinogenesis in B6C3F1 male mice by dichloroacetic and trichloroacetic acids," *Carcinogenesis*, vol. 16, no. 3, pp. 495–500, 1995.
- [13] J. C. Caldwell and N. Keshava, "Key issues in the modes of action and effects of trichloroethylene metabolites for liver and kidney tumorigenesis," *Environmental Health Perspectives*, vol. 114, no. 9, pp. 1457–1463, 2006.
- [14] L. Melillo, "Diuretic plants in the paintings of Pompeii degli Studi di Napoli, Italy," *American Journal of Nephrology*, vol. 14, no. 4–6, pp. 423–425, 1994.
- [15] F. Visioli and C. Galli, "Biological properties of olive oil phytochemicals," *Critical Reviews in Food Science and Nutrition*, vol. 42, no. 3, pp. 209–221, 2002.
- [16] A. Guinda, "Use of solid residue from the olive industry," *Grasas y Aceites*, vol. 57, no. 1, pp. 107–115, 2006.
- [17] E. Altioik, D. Baycin, O. Bayraktar, and S. Ulku, "Isolation of polyphenols from the extracts of olive leaves (*Olea europaea* L.) by adsorption on silk fibroin," *Separation and Purification Technology*, vol. 62, no. 2, pp. 342–348, 2008.
- [18] P. T. Albi, A. Guinda, and A. Lanzon, "Procedimiento de obtencion y determinacion de acidos terpenicos de la hoja del olivo (*Olea europaea*)," *Grasas y Aceites*, vol. 52, no. 5, pp. 275–278, 2001.
- [19] A. Guinda, A. Lanzon, J. J. Rios, and T. Albi, "Aislamiento y cuantificación de los componentes hoja del olivo. Extracto de hexano," *Grasas Y Aceites*, vol. 53, pp. 240–245, 2002.
- [20] L. Ok-Hwan, L. Hee-Bong, L. Junsoo et al., "Chemical properties of olive and bay leaves," *Journal of the Korean Society of Food Science and Nutrition*, vol. 34, no. 4, pp. 503–508, 2005.
- [21] G. Corona, M. Deiana, A. Incani, D. Vauzour, M. Assunta Dessi, and J. P. E. Spencer, "Inhibition of p38/CREB phosphorylation and COX-2 expression by olive oil polyphenols underlies their anti-proliferative effects," *Biochemical and Biophysical Research Communications*, vol. 362, no. 3, pp. 606–611, 2007.
- [22] R. Japon-Lujan and M. D. Luque de Castro, "Superheated liquid extraction of oleuropein and related biophenols from olive leaves," *Journal of Chromatography A*, vol. 1136, no. 2, pp. 185–191, 2006.
- [23] L. Tao, S. Yang, M. Xie, P. M. Kramer, and M. A. Pereira, "Effect of trichloroethylene and its metabolites, dichloroacetic acid and trichloroacetic acid, on the methylation and expression of c-Jun and c-Myc protooncogenes in mouse liver: prevention by methionine," *Toxicological Sciences*, vol. 54, no. 2, pp. 399–407, 2000.
- [24] M. Y. Hung, T. Y. C. Fu, P. H. Shih, C. P. Lee, and G. C. Yen, "Du-Zhong (*Eucommia ulmoides* Oliv.) leaves inhibits CCl₄-induced hepatic damage in rats," *Food and Chemical Toxicology*, vol. 44, no. 8, pp. 1424–1431, 2006.
- [25] S. Kumar, J. G. Sharma, and R. Chakrabarti, "Quantitative estimation of proteolytic enzyme and ultrastructural study of anterior part of intestine of Indian major carp (*Catla catla*) larvae during ontogenesis," *Current Science*, vol. 79, no. 7, pp. 1007–1011, 2000.
- [26] O. H. Lowry, N. J. Rosebrough, A. L. Farr, and R. J. Randall, "Protein measurement with the Folin phenol reagent," *Journal of Biological Chemistry*, vol. 193, pp. 265–275, 1951.
- [27] M. L. Anson, "The estimation of pepsin, trypsin, papain and cathepsin with hemoglobin," *The Journal of General Physiology*, vol. 22, pp. 79–89, 1938.
- [28] O. Folin and V. Ciocalteu, "On tyrosine and tryptophane determinations in proteins," *Journal of Biological Chemistry*, vol. 73, pp. 627–650, 1927.
- [29] M. M. Acharya and S. S. Katyare, "An improved micromethod for tyrosine estimation," *Zeitschrift fur Naturforschung. Section C.*, vol. 59, no. 11–12, pp. 897–900, 2004.
- [30] A. J. Barret and M. F. Heath, "Lysosomal enzymes," in *Lysosome: Laboratory Handbook*, J. T. Dingle, Ed., pp. 124–126, North Holland, Amsterdam, The Netherlands, 1977.
- [31] M. A. Nerurkar, J. G. Satav, and S. S. Katyare, "Insulin-dependent changes in lysosomal cathepsin D activity in rat liver, kidney, brain and heart," *Diabetologia*, vol. 31, no. 2, pp. 119–122, 1988.
- [32] M. A. Khandkar, D. V. Parmar, M. Das, and S. S. Katyare, "Is activation of lysosomal enzymes responsible for paracetamol-induced hepatotoxicity and nephrotoxicity?" *Journal of Pharmacy and Pharmacology*, vol. 48, no. 4, pp. 437–440, 1996.
- [33] M. M. Acharya, S. H. Khamesra, and S. S. Katyare, "Effect of repeated intraperitoneal exposure to picrotoxin on rat liver lysosomal function," *Indian Journal of Experimental Biology*, vol. 42, no. 8, pp. 808–811, 2004.
- [34] D. B. Duncan, "Multiple range and multiple F tests," *Biometrics*, vol. 11, pp. 1–42, 1955.
- [35] F. Marzo, E. Urdaneta, and S. Santidrian, "Liver proteolytic activity in tannic-fed birds," *Poultry Science*, vol. 81, pp. 92–94, 2002.
- [36] B. Werle, C. Kraft, T. T. Lah et al., "Cathepsin B in infiltrated lymph nodes is of prognostic significance for patients with nonsmall cell lung carcinoma," *Cancer*, vol. 89, no. 11, pp. 2282–2291, 2000.
- [37] D. Flores-Reséndiz, E. Castellanos-Juárez, and L. Benítez-Bribiesca, "Proteases in cancer progression," *Gaceta Medica de Mexico*, vol. 145, no. 2, pp. 131–142, 2009.
- [38] T. Lah, M. Čerček, A. Blejec et al., "Cathepsin B, a prognostic indicator in lymph node-negative breast carcinoma patients: comparison with cathepsin D, cathepsin L, and other clinical indicators," *Clinical Cancer Research*, vol. 6, no. 2, pp. 578–584, 2000.
- [39] N. M. Abdel-Hamid, "Premalignant variations in extracellular matrix composition in chemically induced hepatocellular

- carcinoma in rats," *The Journal of Membrane Biology*, vol. 230, no. 3, pp. 155–162, 2009.
- [40] N. M. Abdel-Hamid and M. A. Morsy, "Novel biochemical pathways for 5-fluorouracil in managing experimental hepatocellular carcinoma in rats," *Journal of Membrane Biology*, vol. 234, no. 1, pp. 29–34, 2010.
- [41] M. Yamashita and S. Konagaya, "Participation of cathepsin L into extensive softening of the muscle of chum salmon in spawning migration," *Comparative Biochemistry and Physiology*, vol. 95, no. 1, pp. 149–152, 1990.
- [42] J. P. Braun, A. Rico, and P. Bernard, "Changes in the distribution of gamma-glutamyl transferase in the organs of the mouse as a function of development Evolution de la distribution de la gamma-glutamyl transférase dans les organes de la Souris en fonction du développement," *Comptes Rendus Hebdomadaires des Seances de l'Academie des Sciences. Serie D*, vol. 286, no. 20, pp. 1483–1485, 1978.
- [43] H. M. Beere and J. A. Hickman, "Differentiation: a suitable strategy for cancer chemotherapy?" *Anti-Cancer Drug Design*, vol. 8, no. 4, pp. 299–322, 1993.
- [44] L. Abaza, T. P. N. Talorete, P. Yamada, Y. Kurita, M. Zarrouk, and H. Isoda, "Induction of growth inhibition and differentiation of Human Leukemia HL-60 cells by a Tunisian Gerbouli olive leaf extract," *Bioscience, Biotechnology and Biochemistry*, vol. 71, no. 5, pp. 1306–1312, 2007.



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