

Research Article

Protective Effect of Selected Medicinal Plants against Hydrogen Peroxide Induced Oxidative Damage on Biological Substrates

Namratha Pai Kotebagilu, Vanitha Reddy Palvai, and Asna Urooj

Department of Studies in Food Science and Nutrition, University of Mysore, Manasagangothri, Mysore 570006, India

Correspondence should be addressed to Asna Urooj; asnaurooj@foodsci.uni-mysore.ac.in

Received 25 May 2014; Revised 2 October 2014; Accepted 16 October 2014; Published 12 November 2014

Academic Editor: Paola Fossa

Copyright © 2014 Namratha Pai Kotebagilu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Oxidative stress is developed due to susceptibility of biological substrates to oxidation by generation of free radicals. In degenerative diseases, oxidative stress level can be reduced by antioxidants which neutralize free radicals. Primary objective of this work was to screen four medicinal plants, namely, *Andrographis paniculata*, *Costus speciosus*, *Canthium parviflorum*, and *Abrus precatorius*, for their antioxidant property using two biological substrates—RBC and microsomes. The antioxidative ability of three solvent extracts, methanol (100% and 80%) and aqueous leaf extracts, was studied at different concentrations by thiobarbituric acid reactive substances method using Fenton's reagent to induce oxidation in the substrates. The polyphenol and flavonoid content were analyzed to relate with the observed antioxidant effect of the extracts. The phytochemical screening indicated the presence of flavonoids, polyphenols, tannins, and β -carotene in the samples. In microsomes, 80% methanol extract of *Canthium* and *Costus* and, in RBC, 80% methanol extract of *Costus* showed highest inhibition of oxidation and correlated well with the polyphenol and flavonoid content. From the results it can be concluded that antioxidants from medicinal plants are capable of inhibiting oxidation in biological systems, suggesting scope for their use as nutraceuticals.

1. Introduction

It is well known that oxidant by-products of normal metabolism such as free radicals and reactive oxygen species (ROS) in excess can cause extensive damage to DNA, proteins, and lipids [1]. Under stress, the body produces more ROS, such as superoxide anion and hydroxyl radical, which are highly reactive and potentially damaging transient chemical species. These are continuously produced in the human body, as they are essential for energy supply, detoxification, chemical signaling, and immune function. Overproduction of free radicals and ROS induced by exposure to external oxidant substances or a failure in the defense mechanisms causes oxidative stress in turn leading to various degenerative diseases of aging such as cancer, cardiovascular disease, cataracts, immune system decline, and brain dysfunction [2, 3].

Antioxidants reduce the oxidative stress in cells and are therefore useful in the treatment of many human diseases.

There are various endogenous and exogenous sources of antioxidants. The endogenous sources include antioxidant enzymes such as superoxide dismutase, catalase, glutathione peroxidase, and low-molecular weight antioxidants and exogenous sources such as food sources and medicinal plants.

Several *in vitro*, *ex vivo*, and *in vivo* methods are used to measure the antioxidant potential of plants and foods. It is well known that the antioxidant capacity of a sample will differ with the different oxidants it is measured against, the substrate used (food or biological substrate), and the analytical method used to measure the overall antioxidant capacity. It is also important to examine the effect of antioxidant on markers of oxidative stress. Antioxidant activity can be indirectly assessed by monitoring levels of oxidative stress. Several biomarkers can be used to assess oxidative damage to lipids, protein, and DNA. Thiobarbituric acid reactive substances (TBARS) method is used to measure the level of resistance to oxidation. It determines the resistance of lipids and proteins to oxidation in the presence of an antioxidant.

The malondialdehyde (MDA) generated through oxidative degradation of polyunsaturated fatty acids forms a complex with thiobarbituric acid (MDA-TBA complex) to give a pink color which can be measured spectrophotometrically [4]. These peroxidation products are measured to monitor levels of oxidative stress and are thus an indirect measure of antioxidant capacity.

Biological substrates such as brain, microsomes, low density lipoprotein (LDL), red blood cells (RBC), and cholesterol are susceptible to oxidation due to their composition. Microsomes and RBC are composed of proteins which on oxidation yield protein by-products. Denatured haemoglobin products, particularly free heme, can promote membrane damage and hemolysis through mechanisms including lipid peroxidation and oxidation of protein thiol groups [5]. The disruption of normal processes through injury or the presence of a xenobiotic may greatly enhance the microsomal source of free radicals, resulting in chemical modifications of surrounding structures [6].

Extracts from the stems, roots, bark, leaves, fruits, and seeds of many plants have antioxidant potential. Natural antioxidants represent a potentially side-effect-free alternative to synthetic antioxidants [2]. Phytomedicines are active ingredients exclusively derived from plants which are capable of preventing, curing, or managing a disease. Various medicinal plants have been studied and used for their capability in curing disease.

Literature reports the use of various parts of medicinal plants from the Western Ghats of Southern India in the treatment of several diseases. The whole plant of *Abutilon indicum* has been used in urinary diseases and inflammation, the leaves of *Alstonia scholaris* have been used in the treatment of asthma, beriberi, ulcers, and tumors, and the fruit of *Catunaregam spinosa* has been used in the treatment of diarrhea and dysentery while the root of *Asparagus racemosus* is reported to be beneficial in the treatment of piles, gastritis, cough, and diarrhea [7].

In our laboratory, biscuits prepared by incorporating the extracts of Amla powder (*Emblica officinalis*), drumstick leaves (*Moringa oleifera*), and raisins (*Vitis vinifera*) exhibited better shelf life and were acceptable in sensory characteristics [8]. Other medicinal plants, namely, *Morus indica* [9], *Moringa oleifera*, *Aegle marmelos* [10], *Raphanus sativus*, olive leaves [11], pomegranate peel [11], *Costus speciosus* [12], *Canthium parviflorum* [12], and *Abrus precatorius* [13] leaf extracts, have effectively inhibited the rate of oxidation in the edible oil emulsions.

The selected medicinal plants for the study, *Andrographis paniculata*, *Costus speciosus*, *Canthium parviflorum*, and *Abrus precatorius*, have been reported to exhibit antioxidant, antipyretic, antibacterial, antiviral, anticancer, hypoglycemic, hepatoprotective, gastroprotective, immunoprotective, and cardioprotective effect [14–19]. These four plants were selected to investigate their antioxidative ability in biological substrates based on our earlier investigations which revealed their ability to inhibit oxidation in food system [12, 13]. In this *ex vivo* study, the antioxidative ability of various solvent extracts of these four plants to prevent oxidative

damage against Fenton's reagent induced peroxidation in human erythrocytes and rat liver microsomes were explored.

2. Materials and Methods

2.1. Chemicals. All the chemicals used were of analytical grade. Thiobarbituric acid, triethanolamine, rutin, dithiothreitol, and EDTA were purchased from Hi Media Pvt. Ltd., Mumbai. DTNB was purchased from Sigma chemicals. Protein kit was purchased from Span Diagnostics Ltd., Gujarat.

2.2. Plant Materials. The plant samples selected for the study were *Andrographis paniculata* (AnP), *Costus speciosus* (CS), *Canthium parviflorum* (CP), and *Abrus precatorius* (AP) which were collected from the Western Ghats. The plant samples were identified by Dr. Janardhan, Department of Studies in Botany, University of Mysore, and voucher specimen was retained in the laboratory for future reference. The leaves were cleaned, washed, and dried in hot air oven at 55°C for 8–10 h. The dried leaves were ground to a fine powder and passed through 60 sieve mesh and stored in air tight containers until use.

2.3. Preparation of Extracts. Three different extracts were prepared from the dehydrated samples, that is, 100% methanol (ME), 80% methanol (methanol 80 : water 20) (80 ME), and 100% aqueous (AQ). 10 g of each sample was extracted in 100 mL of the solvent in a mechanical shaker for 12 hours and filtered with Whatman number 1 filter paper. The filtrate obtained from ME and 80 ME was evaporated to dryness in a rotary evaporator at 50°C (Superfit, Bangalore) and aqueous extract was freeze dried (Thermo Electron Corporation, Pune). The extracts were stored at 4°C until further use.

2.4. Estimation of Phytochemicals. The samples were analyzed for various antioxidant components using standard methods. Ascorbic acid was determined according to the titrimetric method using 2, 6-dichlorophenol-indophenol dye [20]. α -Tocopherol was extracted by direct saponification of dried sample and estimated based on formation of a red complex from reaction of α, α' -bipyridyl with ferrous ion due to reduction of ferric ion by tocopherol [21]. β -Carotene was quantified by open column chromatography, followed by measuring the absorbance of elute at 450 nm against standard β -carotene [20]. Reduced glutathione was determined based on the development of a yellow compound due to reaction of DTNB (5, 5-dithio(bis)nitrobenzoic acid) with compounds containing sulphhydryl groups [22]. Total phenols were extracted from a weighed portion (1 g) of dried sample with 5 mL of 1.2 M HCl in 50% aqueous methanol for 2 h and with 70% acetone shaken for 2 h and analyzed by Folin-Ciocalteu micromethod [23]. Results were expressed as mg or g of gallic acid equivalent. The flavonoid content was determined by pharmacopoeia method [24] using rutin as a reference compound. Alkaloids were analyzed by spectrophotometric method using atropine as reference standard [25]. Saponins and steroidal saponins were estimated using diosgenin as standard [26, 27].

2.5. Isolation of Substrates. Two substrates, namely, RBC and microsomes, were chosen to study the potency of selected samples in inhibiting oxidation. Blood (20 mL) was drawn from healthy human volunteers, in vials, and centrifuged at 500 g for 10 min at 4 °C and plasma was separated. RBC were washed with cell wash buffer in 1:2 ratio (RBC-1: Buffer-2) and centrifuged at 500 g for 10 min at 4 °C. The supernatant was removed and the procedure was followed 2-3 times [28]. The study was approved by the Human Ethics Committee of the University of Mysore (IHEC-UOM number 36 Res/2013-14, 16.04.2013).

Microsomes were isolated from the liver of healthy adult male rats from Central Animal House of the University of Mysore. Permission from Institutional Animal Ethics Committee of the University of Mysore was taken (UOM/IAEC/04/2013 dated 28-09-2013). The liver tissue was minced, homogenized, and centrifuged for 10 min at 12,000 g to remove cell debris and mitochondria. The supernatant solution was carefully removed to prevent contamination with mitochondria and recentrifuged for 10 min at 12,000 g to ensure removal of mitochondria; it was then centrifuged at 60,000 g for 60 min. The microsomal pellet was rinsed with buffer and frozen at -20 °C. Frozen microsomes were resuspended in 0.1 M triethanolamine buffer, pH 7.4 containing 0.02 M EDTA, and 10 mM dithiothreitol. It was allowed to stand for 60 min packed in ice and diluted with buffer to give a protein concentration of 5–10 mg/mL [29].

2.6. Estimation of Thiobarbituric Acid Reactive Substances (TBARS). Lipid peroxide formation was measured by the modified method of Ohkawa et al. [30]. Substrate (RBC and microsomes) equal to 1 mg protein was taken for the experiment. Plant extracts of different concentrations (300–500 µL of 10 mg/mL concentration) were added to the substrates. Fenton's reagent was added to the substrates to induce oxidation (500 µL of 20 mM FeSO₄ and 250 µL of 20 mM H₂O₂). In case of RBC, 1 mL trichloroacetic acid (TCA, 10%) was added to precipitate the RBC and both substrates were incubated for 2 h at 50 °C. After 2 h, the RBC were separated by centrifuging at 3000 rpm for 10 min at 5 °C. The supernatant was separated and 1 mL of thiobarbituric acid (TBA, 0.67%) was added. To the microsomes, 1 mL of TBA (0.67%) and 1 mL of TCA (10%) were added and kept in boiling water bath for 30 min and cooled. After cooling, 3 mL butanol was added to the tubes and vortex mixed. The pink color in butanol layer was measured at 532 nm [4]. IC₅₀ values were calculated using mean values of triplicates of the inhibition of oxidation by the extracts at 300–500 µL. Consider

% Inhibition of oxidation

$$= \frac{[\text{O.D of Control} - \text{O.D of sample}]}{[\text{O.D of control}]} \times 100. \quad (1)$$

2.7. Statistical Analysis. All the experiments were carried out in triplicate ($n = 3$).

The correlation was determined by Pearson's product-moment correlation coefficient. IC₅₀ was calculated using

linear regression. Data was subjected to one-way ANOVA using SPSS software, 2011 version ($P < 0.05$).

3. Results and Discussion

3.1. Phytochemical Composition. It is well known that oxidation of biological substrates leads to various diseases. Antioxidants and phytochemicals from medicinal plants are now being used to counteract oxidation at the biological level, which is a safer approach than using pharmaceutical drugs to reduce oxidation.

The plants investigated are used in traditional medicine for various medicinal and pharmacological effects [14–19]. Studies also report that they exhibit good antioxidant activity *in vitro* [13, 31] Table 1 depicts the phytochemical composition of the samples, where it was observed that CS and AnP were good sources of polyphenols, glutathione, β-carotene, total saponins, and steroidal saponins, whereas AP and CP were found to be good sources of flavonoids, alkaloids, α-tocopherol, vitamin C, and tannins.

3.2. Polyphenol and Flavonoid Content of the Extracts. The polyphenol and flavonoid content of the extracts are given in Table 1. In general, all the extracts of the samples were found to be fair sources of polyphenol and flavonoids; however, it was high in the extracts of AP. The polyphenol content in two extracts of AnP (AQ and ME) was higher than in CP and CS, while the polyphenols in 80 ME of CP and CS was higher than AnP. Similarly, the flavonoid content of AP was high in all the extracts, followed by AnP > CP > CS. It may be inferred that methanol and 80% methanol are solvents suitable for extraction of polyphenol and flavonoid in these samples. Similar trend was observed in earlier studies on *Morus indica* [32, 33] and *Moringa oleifera* [34].

3.3. Inhibition of Oxidation. Lipid peroxidation is a complex process wherein the initially formed lipid radicals are converted to TBARS [35]. The first event of lipid peroxidation is oxygen absorption [36]. In Fenton's reaction, the free radical produced is hydroxyl radical which is the most deleterious and reactive radical among the ROS with shortest half-life. The oxygen derived hydroxyl radicals in presence of transition metal ion Fe²⁺ causes the degradation to form malondialdehyde (MDA) which produces a pink chromogen with thiobarbituric acid. MDA produced due to oxidative damage of biological substrates is an indicator of free radical pathology [37]. In the present work, TBARS was used to evaluate the antioxidative activity of different medicinal plant extracts and inhibition of generation of hydroxyl radicals by Fenton reaction in two biological substrates, microsomes and RBC.

3.3.1. Microsomes. Microsomes due to their high PUFA content are susceptible to oxidation and hence are extensively used as a model for studying oxidative stress and antioxidant studies. The inhibition of oxidation at different concentrations (300, 400, and 500 µL) by different solvent extracts of the sample is given in Figure 1. The activity was found to be

TABLE 1: Phytochemical composition of selected medicinal plants.

Phytochemicals	AnP	CS	CP	AP
Polyphenols*	3.21 ± 0.10 ^b	3.21 ± 0.12 ^b	1.28 ± 0.32 ^a	1.06 ± 0.06 ^a
Flavonoids**	1.50 ± 0.16 ^a	1.17 ± 0.42 ^a	2.44 ± 0.81 ^b	1.32 ± 0.08 ^a
Glutathione***	824 ± 28.84 ^c	285.6 ± 33 ^a	311.55 ± 25 ^{ab}	375 ± 76.77 ^b
α-Tocopherol**	NA	90.9 ± 3.29 ^b	164.6 ± 22.10 ^c	36.6 ± 3.05 ^a
Tannins**	846 ± 0.15 ^b	1088 ± 0.03 ^c	605 ± 0.02 ^a	1160 ± 0.07 ^c
β-Carotene****	ND	1377 ± 2.51 ^b	1060 ± 0.6 ^a	1260 ± 0.05 ^b
Alkaloids**	44.6 ± 6.65 ^a	120 ± 0.03 ^b	400 ± 0.06 ^c	1100 ± 0.03 ^d
Total sap.**	0.300 ^c	0.282 ± 0.04 ^{bc}	0.200 ± 0.04 ^a	0.244 ± 0.01 ^{ab}
Steroidal sap.**	0.110 ^a	0.238 ± 0.038 ^c	0.148 ± 0.051 ^{ab}	0.186 ± 0.028 ^b
Vitamin C*	ND	0.0237 ± 0.00 ^a	0.0228 ± 0.00 ^a	0.643 ± 0.04 ^b

AnP: *Andrographis paniculata*, CS: *Costus speciosus*, CP: *Canthium parviflorum*, and AP: *Abrus precatorius*; *g/100 g, **mg/100 g, and ***mmol/100 g, ****μg/100 g dry basis. ND: not detected, sap.: saponins, and NA: not analyzed. Mean values with different superscripts a, b, c and d in rows, differ significantly ($P \leq 0.05$).

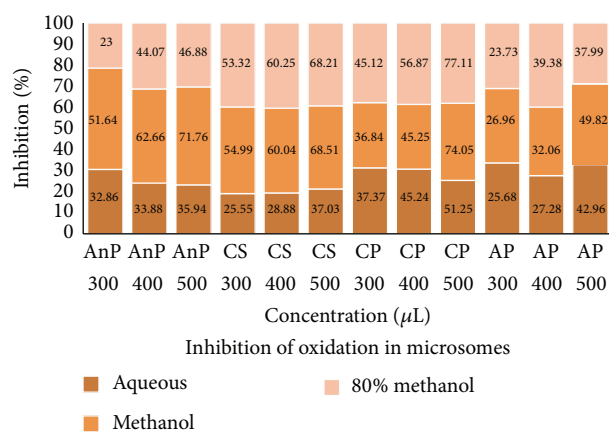


FIGURE 1: Inhibition of oxidation by different plant extracts in microsomes; AnP: *Andrographis paniculata*, CS: *Costus speciosus*, CP: *Canthium parviflorum*, and AP: *Abrus precatorius*. Values are the mean of triplicates ($n = 3$).

dose dependent in all the extracts. At higher concentration, the inhibition was greater in 80 ME (77%) and ME (74%) of CP, CS (ME, 68% and 80 ME, 68%), and AnP (ME, 71%). In AQ extract the activity was low in all the samples and maximum percent inhibition observed was $51 \pm 2.90\%$ in CP. The results indicate that CP and CS were highly effective in inhibiting oxidation. The results are in accordance with an earlier study in *Morus indica* where ME and 80 ME extracts were potent in inhibiting the oxidation in microsomes [33].

3.3.2. RBC. Erythrocytes have been extensively used as a simple cell model to study oxidative stress. The inhibition of oxidation at different concentrations (300, 400, and 500 μL) by different solvent extracts of the sample is given in Figure 2. The activity was found to be dose dependent in all the extracts of AP, whereas in the ME of CS and AnP, 80 ME of CP, and AQ of AnP the maximum activity was exhibited at 400 μL; however no notable difference was observed in the activity between 400 and 500 μL. Among all the extracts of

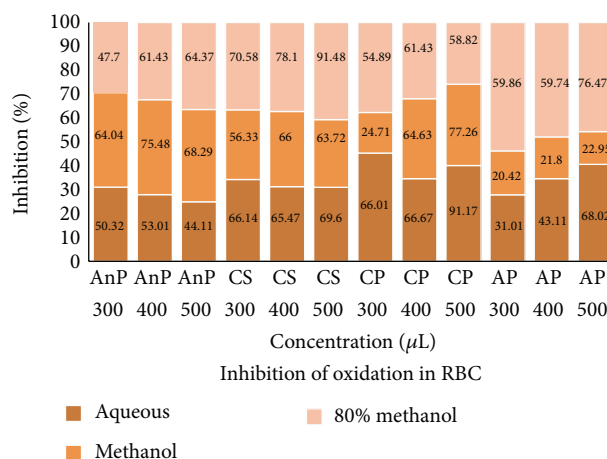


FIGURE 2: Inhibition of oxidation by different plant extracts in RBC. AnP: *Andrographis paniculata*, CS: *Costus speciosus*, CP: *Canthium parviflorum*, and AP: *Abrus precatorius*. Values are the mean of triplicates ($n = 3$).

the samples, maximum activity of more than 90% was shown by 80 ME of CS and AQ of CP.

As expected, TBARS values were effectively reduced by the plant extracts in biological substrates. This indicated that the extracts could efficiently prevent the nonenzymatic induced lipid peroxidation in various models [38]. The mode of action is primarily reducing oxidative damage by trapping the free radicals directly or scavenging them through a series of coupled reactions with antioxidant enzymes, contributing to stabilization of lipid peroxides [39].

Protective effect of *Ugni molinae* against oxidative damage of human erythrocytes is reported. HPLC-MS analysis of *Ugni molinae* leaves revealed the presence of polyphenols, flavonols and flavanols, myricetin, quercetin, and epicatechin [40]. In another study, the stem bark extract of *Semecarpus anacardium* showed significant protection against Fenton reaction induced lipid peroxidation in sheep liver tissue homogenate model and heat induced hemolysis in human

TABLE 2: Relationship (r value) with the antioxidant effect and polyphenol and flavonoid content of plant extracts in microsomes.

	AQ	r value	ME	r value	80ME	r value
Polyphenol (mg/g)						
AnP	63.33 ± 4.71	0.233	88.33 ± 2.35	0.889	41.66 ± 2.35	0.532
CP	25.00 ± 0	0.792	46.66 ± 4.71	0.959	55.00 ± 0	0.964
CS	55.00 ± 0	0.706	51.66 ± 2.35	0.716	63.33 ± 2.35	0.915
AP	81.66 ± 8.49	0.828	83.33 ± 9.42	0.951	120.00 ± 0	0.786
Flavonoid (mg/g)						
AnP	0.59 ± 0.02	0.232	1.87 ± 0.07	0.889	0.96 ± 0.11	0.532
CP	0.01 ± 0	0.790	0.59 ± 0.01	0.941	1.09 ± 0.01	0.957
CS	0.16 ± 0.03	0.669	0.04 ± 0	0.710	0.40 ± 0.06	0.914
AP	1.02 ± 0.08	0.828	2.58 ± 0.10	0.943	1.69 ± 0.02	0.786

Aq: aqueous extract; ME: methanol extract; 80ME: 80% methanol extract;

AnP: *Andrographis paniculata*, CS: *Costus speciosus*, CP: *Canthium parviflorum*, and AP: *Abrus precatorius*. Values are the mean of triplicates ($n = 3$).

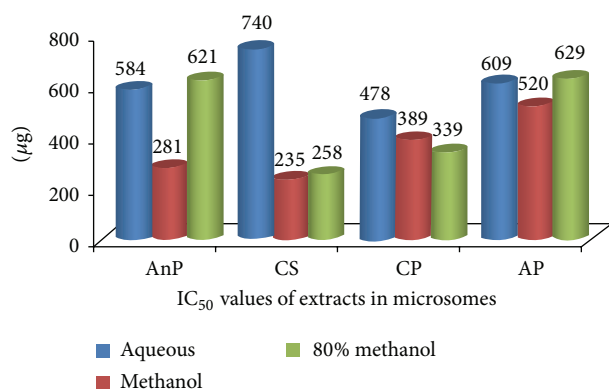


FIGURE 3: IC₅₀ values of different solvent extracts of selected medicinal plants in microsomes; AnP: *Andrographis paniculata*, CS: *Costus speciosus*, CP: *Canthium parviflorum*, and AP: *Abrus precatorius*.

RBC membrane model [37]. Hence, the antioxidant activity of these medicinal plants can be related to their phytochemical constituents.

3.4. *IC₅₀ of the Plant Extracts*. Linear regression was used to calculate IC₅₀ values. The IC₅₀ values of different solvent extracts of the samples are depicted in Figure 3 (microsomes) and Figure 4 (RBC). In microsomes, it can be observed that the IC₅₀ values of ME (235 µg) and 80 ME of CS were lower than other extracts followed by ME of AnP and 80 ME of CP. In RBC, the IC₅₀ value of 80 ME of CS (112 µg) and, AP and AQ of CP was lower than other extracts. In rest of the extracts, no remarkable difference was observed; however, a high IC₅₀ value (921 µg) was observed in ME of AP.

3.5. *Correlation between the Inhibition of Oxidation and Polyphenol and Flavonoids*. The correlation was measured in all the extracts in both substrates (given in Tables 2 and 3). ME ($r = 0.95$; $P \leq 0.01$) and 80 ME ($r = 0.96$; $P \leq 0.01$) of CP and CS and 80 ME ($r = 0.93$; $P \leq 0.01$) have shown a high correlation with both polyphenols and

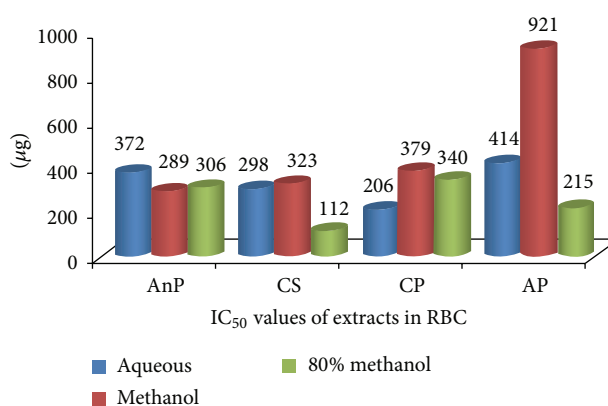


FIGURE 4: IC₅₀ values of different solvent extracts of selected medicinal plants in RBC. AnP: *Andrographis paniculata*, CS: *Costus speciosus*, CP: *Canthium parviflorum*, and AP: *Abrus precatorius*.

flavonoids indicating the protective effect of these phytochemicals on biological substrates. The inhibitory action might also be due to the presence of other phytochemicals such as glutathione, ascorbic acid. Literature reports the presence of alkaloids in these plants such as 2-amino-4-benzylthiomethyl-6-morpholino-1,3,5-triazine present in *Abrus precatorius* [41], the lactone andrographolide and flavonoids, 5,7,2',3'-tetramethoxyflavone and 5-hydroxy-7,2',3'-trimethoxyflavone in *Andrographis paniculata* [42], flavonoids and phenolic compounds present in *Canthium parviflorum* [43], and alkaloids and flavonoids in *Costus speciosus* [44] which also might have imposed a protective effect.

4. Conclusion

It can be inferred from the findings that *Andrographis paniculata*, *Abrus precatorius*, *Canthium parviflorum*, and *Costus speciosus* possess ability to inhibit oxidation of biological substrates, by virtue of the presence of flavonoid and polyphenols. It was also observed that the extractability depends primarily on the solvent systems used. Further studies to

TABLE 3: Relationship (r value) with the antioxidant effect and polyphenol and flavonoid content of plant extracts in RBC.

	AQ	r value	ME	r value	80ME	r value
Polyphenol (mg/g)						
AnP	63.33 ± 4.71	-0.578	88.33 ± 2.35	0.270	41.66 ± 2.35	0.900
CP	25.00 ± 0	0.725	46.66 ± 4.71	0.882	55.00 ± 0	0.366
CS	55.00 ± 0	0.235	51.66 ± 2.35	0.584	63.33 ± 2.35	0.937
AP	81.66 ± 8.49	0.970	83.33 ± 9.42	0.455	120.00 ± 0	0.790
Flavonoid (mg/g)						
AnP	0.59 ± 0.02	-0.55	1.87 ± 0.07	0.270	0.96 ± 0.11	0.900
CP	0.01 ± 0	0.779	0.59 ± 0.01	0.901	1.09 ± 0.01	0.394
CS	0.16 ± 0.03	0.227	0.04 ± 0	0.659	0.40 ± 0.06	0.931
AP	1.02 ± 0.08	0.970	2.58 ± 0.10	0.455	1.69 ± 0.02	0.790

Aq: aqueous extract; ME: methanol extract; 80ME: 80% methanol extract.

AnP: *Andrographis paniculata*, CS: *Costus speciosus*, CP: *Canthium parviflorum*, and AP: *Abrus precatorius*. Values are the mean of triplicates ($n = 3$).

isolate and identify the bioactive constituents are in progress for their optimal utilization.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

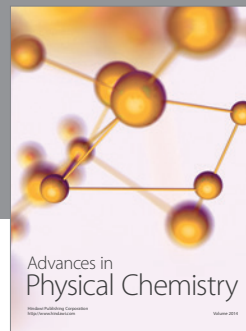
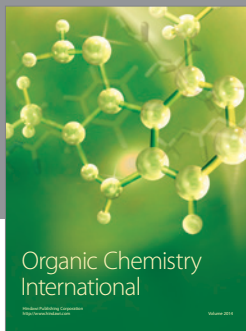
Acknowledgment

Authors acknowledge UGC-IOE (Institute of Excellence) for providing financial assistance.

References

- [1] C. V. Smith and J. P. Kehrer, "Free radicals in biology: sources, reactivities and roles in the etiology of human diseases," in *Natural Antioxidants in Human health and Diseases*, pp. 27–30, Academic press, San Diego, Calif, USA, 1994.
- [2] D. Krishnaiah, R. Sarbatly, and R. Nithyanandam, "A review of the antioxidant potential of medicinal plant species," *Food and Bioproducts Processing*, vol. 89, no. 3, pp. 217–233, 2011.
- [3] S. A. Sharif, N. Kasoju, A. Luthra, A. Singh, H. Sharanabasava, and A. Sahu, "Review on Indian medicinal herbs as sources of antioxidants," *Food Research International*, vol. 41, pp. 1–15, 2008.
- [4] I. J. Kade, E. O. Ibukun, C. W. Nogueira, and J. B. T. da Rocha, "Sun-drying diminishes the antioxidative potentials of leaves of *Eugenia uniflora* against formation of Thiobarbituric acid reactive substances induced in homogenates of rat brain and liver," *Experimental and Toxicologic Pathology*, vol. 60, no. 4–5, pp. 365–371, 2008.
- [5] T. Kanas and J. P. Acker, "Mechanism of hemoglobin-induced cellular injury in desiccated red blood cells," *Free Radical Biology and Medicine*, vol. 49, no. 4, pp. 539–547, 2010.
- [6] B. Frei and J. P. Kehrer, "Free radicals in biology: sources, reactivities and roles in the etiology of human diseases," in *Natural Antioxidants in Human health and Diseases*, pp. 33–37, Academic Press, San Diego, Calif, USA, 1994.
- [7] N. Shiddamallayya, A. Yasmeen, and K. Gopakumar, "Hundred common forest medicinal plants of Karnataka in primary healthcare," *Indian Journal of Traditional Knowledge*, vol. 9, no. 1, pp. 90–95, 2010.
- [8] V. Reddy, A. Urooj, and A. Kumar, "Evaluation of antioxidant activity of some plant extracts and their application in biscuits," *Food Chemistry*, vol. 90, no. 1–2, pp. 317–321, 2005.
- [9] L. G. Roy, S. A. Delouee, and A. Urooj, "Antioxidant efficacy of Mulberry (*Morus indica* L.) leaves extract and powder in edible oil," *International Journal of Food Properties*, vol. 13, no. 1, pp. 1–9, 2010.
- [10] P. V. Reddy, N. Sahana, and A. Urooj, "Antioxidant activity of *Aegle marmelos* and *Psidium guajava* leaves," *International Journal of Medicinal and Aromatic Plants*, vol. 2, no. 1, pp. 155–160, 2012.
- [11] M. Poodineh, *Effect of fruit peel, spice and herb extracts on the oxidative stability of Sunflower oil [Dissertation]*, Department of Food science and Nutrition, University of Mysore, Mysore, Karnataka, India, 2013.
- [12] M. Sowmya, *Phytochemical composition, antioxidant activity and stability of selected medicinal plants' extracts [Dissertation]*, Department of Food science and Nutrition, University of Mysore, Mysore, Karnataka, India, 2012.
- [13] P. V. Reddy, S. Mahalingu, and A. Urooj, "*Abrus precatorius* leaves: antioxidant activity in food and biological systems, pH and temperature stability," *International Journal of Medicinal Chemistry*, vol. 2014, Article ID 748549, 7 pages, 2014.
- [14] K. Vijaykumar, B. S. P. Murthy, S. Kannababu, B. Syamasundar, and G. V. Subbaraju, "Estimation of Andrographolide in *Andrographis paniculata* herb, extracts and dosage forms," *International Journal of Applied Science and Engineering*, vol. 5, no. 1, pp. 27–39, 2007.
- [15] V. Duraipandiyar, N. Abdullah Al-Harbi, S. Ignacimuthu, and C. Muthukumar, "Antimicrobial activity of sesquiterpene lactones isolated from traditional medicinal plant, *Costus speciosus*," *BMC Complementary and Alternative Medicine*, vol. 12, article 13, 2012.
- [16] J. Nehete, M. Bhatia, and M. Narkhede, "In-vitro evaluation of antioxidant activity and phenolic content of *Costus speciosus* (Koen) J.E. Sm," *Iranian Journal of Pharmaceutical Research*, vol. 9, no. 3, pp. 271–277, 2010.
- [17] T. P. Prabhu, S. S. Kumari, W. C. Atlee, R. Vijayakumar, and R. Suresh, "Antioxidant activity of ethanolic extract of *canthium parviflorum* lamk in alloxan induced diabetic rats," *International Journal of Ayurvedic and Herbal Medicine*, vol. 2, no. 5, pp. 766–770, 2012.
- [18] T. P. Prabhu, P. Panneerselvam, S. Selvakumari, and D. Sivaraman, "In vitro and In vivo anticancer activity of ethanolic extract

- of *Canthium Parviflorum* Lam on DLA and Hela cell lines," *International Journal of Drug Development and Research*, vol. 3, no. 4, pp. 280–285, 2011.
- [19] D. J. Taur and R. Y. Patil, "Effect of *Abrus precatorius* leaves on milk induced leukocytosis and eosinophilia in the management of asthma," *Asian Pacific Journal of Tropical Biomedicine*, vol. 1, no. 1, pp. S40–S42, 2011.
- [20] S. Ranganna, *Handbook of Analysis and Quality Control for Fruits and Vegetables Products*, McGraw-Hill, New Delhi, India, 2nd edition, 1999.
- [21] M. Freed, *Methods of Vitamin Assay*, Association of Vitamin Chemists, Interscience, New York, NY, USA, 3rd edition, 1966.
- [22] E. Beutler and B. M. Kelly, "The effect of sodium nitrite on red cell GSH," *Experientia*, vol. 19, no. 2, pp. 96–97, 1963.
- [23] K. Slinkard and V. L. Singleton, "Total phenol analysis, automation and comparison with manual methods," *American Journal of Enology and Viticulture*, vol. 28, pp. 49–55, 1967.
- [24] G. Miliauskas, P. R. Venskutonis, and T. A. Van Beek, "Screening of radical scavenging activity of some medicinal and aromatic plant extracts," *Food Chemistry*, vol. 85, no. 2, pp. 231–237, 2004.
- [25] F. Shamsa, H. Monsef, R. Ghamooshi, and M. Verdian-rizi, "Spectrophotometric determination of total alkaloids in some Iranian medicinal plants," *Thai Journal of Pharmaceutical Sciences*, vol. 32, pp. 17–20, 2008.
- [26] S. Hiai, H. Oura, and T. Nakajima, "Color reaction of some saponin and saponins with vanillin and sulfuric acid," *Planta Medica*, vol. 29, no. 2, pp. 116–122, 1976.
- [27] J. C. Baccou, F. Lambert, and Y. Sauvaire, "Spectrophotometric method for the determination of total steroidal saponin," *Analyst*, vol. 102, no. 1215, pp. 458–465, 1977.
- [28] M. S. Hanson, A. H. Stephenson, E. A. Bowles, M. Sridharan, S. Adderley, and R. S. Sprague, "Phosphodiesterase 3 is present in rabbit and human erythrocytes and its inhibition potentiates iloprost-induced increases in cAMP," *American Journal of Physiology—Heart and Circulatory Physiology*, vol. 295, no. 2, pp. H786–H793, 2008.
- [29] F. H. Hulcher and W. H. Oleson, "Simplified spectrophotometric assay for microsomal 3 hydroxy 3 methylglutaryl CoA reductase by measurement of coenzyme A," *Journal of Lipid Research*, vol. 14, no. 6, pp. 625–631, 1973.
- [30] H. Ohkawa, N. Ohishi, and K. Yagi, "Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction," *Analytical Biochemistry*, vol. 95, no. 2, pp. 351–358, 1979.
- [31] P. V. Reddy, S. Mahalingu, and A. Urooj, "*Canthium parviflorum* leaves: antioxidant activity in food and biological systems, pH, and temperature stability," *Chinese Journal of Biology*, vol. 2014, Article ID 813201, 7 pages, 2014.
- [32] S. Arabshahi-Delouee and A. Urooj, "Antioxidant properties of various solvent extracts of mulberry (*Morus indica* L.) leaves," *Food Chemistry*, vol. 102, no. 4, pp. 1233–1240, 2007.
- [33] P. V. Reddy and A. Urooj, "Proximate, phytochemical profile and antioxidant activity (*in vitro* and *ex vivo*) of *Morus indica* varieties," *International Journal of Pharmaceutical Sciences and Research*, vol. 4, no. 4, pp. 1626–1634, 2013.
- [34] P. V. Reddy and A. Urooj, "*Moringa oleifera*: antioxidant properties and stability of various solvent extracts," *International Journal of Pharmaceutical Sciences and Biotechnology*, vol. 1, no. 1, pp. 1–6, 2010.
- [35] J. A. Knight, R. K. Pieper, and L. McClellan, "Specificity of the thiobarbituric acid reaction: Its use in studies of lipid peroxidation," *Clinical Chemistry*, vol. 34, no. 12, pp. 2433–2438, 1988.
- [36] Y.-J. Cai, J.-G. Fang, L.-P. Ma, L. Yang, and Z.-L. Liu, "Inhibition of free radical-induced peroxidation of rat liver microsomes by resveratrol and its analogues," *Biochimica et Biophysica Acta—Molecular Basis of Disease*, vol. 1637, no. 1, pp. 31–38, 2003.
- [37] A. N. D. Kumar, G. B. Bevara, K. Laxmikoteswaramma, and R. R. Malla, "Antioxidant, cytoprotective and anti-inflammatory activities of stem bark extract of *Semecarpus anacardium*," *Asian Journal of Pharmaceutical and Clinical Research*, vol. 6, no. 1, 2013.
- [38] S. Bhattacharya, J. P. Kamat, S. K. Bandyopadhyay, and S. Chattopadhyay, "Comparative inhibitory properties of some Indian medicinal plant extracts against photosensitization-induced lipid damage," *Food Chemistry*, vol. 113, no. 4, pp. 975–979, 2009.
- [39] S. S. Sakat, A. R. Juvekar, and M. N. Gambhire, "In-vitro antioxidant and anti-inflammatory activity of methanol extract of *Oxalis corniculata* linn," *International Journal of Pharmacy and Pharmaceutical Sciences*, vol. 2, no. 1, pp. 146–155, 2010.
- [40] M. Suwalsky, P. Orellana, M. Avello, and F. Villena, "Protective effect of *Ugni molinae* Turcz against oxidative damage of human erythrocytes," *Food and Chemical Toxicology*, vol. 45, no. 1, pp. 130–135, 2007.
- [41] R. Pokharkar and R. Saraswat, "GCMS studies of *Abrus precatorius*," *Pharmacology Online*, vol. 2, pp. 1178–1189, 2011.
- [42] S. Akbar, "*Andrographis paniculata*: a review of pharmacological activities and clinical effects," *Alternative Medicine Review*, vol. 16, no. 1, pp. 66–77, 2011.
- [43] T. Sathish Kumar, S. Shanmugam, T. Palvannan, and V. M. Bharathi Kumar, "Evaluation of antioxidant properties of *Canthium parviflorum* Lam. leaves," *Natural Product Radiance*, vol. 7, no. 2, pp. 122–126, 2008.
- [44] C. Najma, K. J. Chandra, and H. Ansarul, "Effect of *Costus speciosus* Koen on Reproductive organs of female albino mice," *International Research Journal of Pharmacy*, vol. 3, no. 4, pp. 200–202, 2012.



Hindawi

Submit your manuscripts at
<http://www.hindawi.com>

