Research Article
Prevalence and Antibiotic Resistance Patterns of Ocular Bacterial Strains Isolated from Pediatric Patients in University Hospital of Campania “Luigi Vanvitelli,” Naples, Italy

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Eye infections caused by bacteria are a serious public health problem among pediatric patients. These diseases, if not properly treated, can cause blindness and impaired vision. The study aimed to evaluate the antimicrobial resistance profiles of the main pathogens involved in eye infections. This study involved pediatric patients enrolled at the “Luigi Vanvitelli” University Hospital of Campania in Naples, Italy, between 2017 and 2019. Of a total of 228 pediatric patients, 73 (32%) tested positive for bacterial infection. In terms of strain distribution, 85% were Gram-positive bacteria, while 15% were Gram-negative bacteria. The most frequently isolated strains were coagulase-negative Staphylococci (60.4%), followed by Staphylococcus aureus (16.4%). The isolated bacteria showed a significant percentage of resistance to multiple antibiotics. Therefore, the identification of the causal bacteria and antimicrobial sensitivity tests are mandatory to select the effective drug for the treatment of eye infections and prevent the development of antibiotic-resistant bacteria.

1. Introduction

Ocular infections and their complications represent an important public health problem [1]. These diseases are associated with a high degree of visual morbidity and blindness worldwide [2]. The ocular infections distribution in the population is conditional on many factors: (i) the use of contact lenses; (ii) surgery; (iii) trauma; (iv) previous eye infections; (v) obstruction of the nasolacrimal duct; (vi) age; and (vii) dry eye [3, 4]. These infections are commonly observed in pediatric patients, affecting infants and preschool-aged children of both genders [5]. Bacteria are the main cause of ocular infection, although viruses, fungi, and parasites may be involved in the origin of this infection [6, 7]. These microorganisms contribute to 32–74% of eye infections, globally [8]. Bacteria are associated with different types of eye surface infections including keratitis, dacryocystitis, blepharokeratoconjunctivitis, and conjunctivitis [9, 10]. The most common bacterial pathogens, involved in pediatric ocular infection are coagulase-negative Staphylococci (CoNS), Staphylococcus aureus (S. aureus), Streptococcus pneumoniae, Pseudomonas aeruginosa (P. aeruginosa), and Haemophilus influenzae [11]. Gram-positive bacteria are primarily responsible for pediatric ocular infection [12]. A prospective study conducted in the United States has revealed that 65% of children have ocular infections caused by Gram-
positive bacteria [13]. According to the guidelines, the diagnosis of ocular bacterial infection is based on the examination of the patient’s clinical symptoms and laboratory testing [14]. Cultural analysis and antibiotic susceptibility testing are ideal for guiding therapy. Although the guidelines for the treatment of these infections recommend the laboratory procedures, empirical broad-spectrum antibiotics treatment is initially used [15]. This contributes to the development of antimicrobial resistance (AMR) among ocular pathogens, which has increased dramatically in recent decades [16]. A national surveillance study started in 2009 (ARMOR) has monitored the resistance profiles among bacterial species that most commonly cause eye infections: Staphylococcus species, *S. pneumoniae*, *P. aeruginosa*, and *H. influenzae*. The study reported high rates of AMR, particularly among the Staphylococci species [17]. Ocular diseases, if not treated properly, can cause irreversible damage to the structures of the eye, leading to visual impairment and blindness [18]. Drug-resistant bacteria and the high prevalence of ocular bacterial infections in pediatric patients stress the importance of knowing the causative microorganisms and antimicrobials susceptibility profile. Therefore, the goal of our study was to evaluate the etiology and antimicrobial resistance profiles of ocular infection pathogens isolated from pediatric patients in the University Hospital of Campania “Luigi Vanvitelli,” Naples.

2. Materials and Methods

2.1. Sample Collection. Our retrospective study was conducted on 228 pediatric patients with clinical diagnoses of ocular infections, at the University Hospital of Campania “Luigi Vanvitelli” (UOC) in Naples, Italy, between July 2017 and November 2019. Each patient had undergone a conjunctival sampling. This procedure consisted of rolling a thin cotton swab over the lower fornix of the conjunctival sac. The eye swab was inserted into the transport media and delivered to the bacteriology laboratory and processed.

2.2. Bacterial Culture and Identification. The samples were transferred into 5 ml of Brain-Heart Infusion broth (Oxoid, Hampshire, UK) and incubated overnight at 37°C. The broth was inoculated on blood agar, chocolate agar, MacConkey agar, mannitol salted agar, modified Thayer-Martin agar, and Sabouraud glucose agar (Oxoid, Hampshire, United Kingdom). All plates were incubated overnight at 37°C. The chocolate and Thayer-Martin agar were maintained in the presence of CO₂. After 24 hours of incubation, each plate was examined, and negative plates were incubated for an additional 24 hours. Bacterial identification was obtained via Matrix-Assisted Laser Desorption Ionization-Time of Flight Mass Spectrometry (MALDI-TOF MS) (Bruker Dal-tonics, Germany). Identifications were performed according to the manufacturer’s instructions. A score higher than 2 allowed a reliable identification of the species [19].

2.3. Antibiotic Susceptibility Test. Confirmation of identification obtained through MALDI-TOF MS and antibiotic sensitivity tests was performed through the Phoenix BD (Becton Dickinson, United States). Identifications and antibiotic sensitivity tests were performed according to the manufacturer’s instructions. The examined antimicrobials in this study were: ampicillin, ciprofloxacin, gentamycin, fosfomycin, trimethoprim-sulfamethoxazole. fusidic acid, cefoxitin, ceftaroline, daptomycin, erythromycin, imipenem, linezolid, moxifloxacin, oxacillin, rifampicin, teicoplanin, tetracycline, tigecycline, mupirocin, vancomycin, nitrofurantoin, amidacin, amoxicillin-clavulanic acid, cefepime, cefotaxime, ceftazidime, cefuroxime, colistin, ertapenem, fosfomycin, meropenem, levofloxacin, piperacillin, piperacillin-tazobactam, tigecycline, and tobramycin ((Becton Dickinson, United States) [20]. Reference strains of *Escherichia coli* ATCC 25922 and *S. aureus* ATCC 25923 were used as quality control measure for identification criteria and antimicrobial susceptibility tests.

2.4. Data Analysis. Data were analyzed using IBM SPSS software (version 22.0; IBM SPSS Inc., New York, USA). Descriptive statistics were computerized for the study, and variables such as sex and pathogenic bacteria were isolated from the study population. The tables show the frequency of isolated ocular bacteria and also compare the resistance percentage of different antibiotics. For the categorical variables, the chi-square test values <0.05 were considered significant [21].

2.5. Ethical Consideration Statement. Ethical approval by the Human Research Ethics Committee was not requested for this study. The resignation was given as our study used laboratory management data and clinical information on patients, collected from databases. This is a retrospective study and not directly associated with patients. This study was consistent with the principles of the Helsinki Declaration.

3. Results

3.1. Incidence of Ocular Infections in Pediatric Patients. In this study, 228 ocular samples, obtained from pediatric patients, were processed. Ocular infections were diagnosed based on the patient’s clinical symptoms, redness with mucopurulent discharge. As reported in Table 1, 32% of patients were positive for bacterial growth, while 68% were negative (Table 1). Among the isolated strains, 85% was Gram-positive bacteria, while 15% was Gram-negative bacteria (Table 1). Our study showed a high frequency of ocular infections in males compared to females (31.5%) (Table 1). In addition, we observed most cases over a 12-month group (Figure 1).

Bacterial species, which appertain to 9 genera, were identified by 73 positive cultures. For Gram-positive bacteria, isolates of CoNS (*Staphylococcus epidermidis, Staphylococcus haemolyticus, Staphylococcus warneri, Staphylococcus hominis*, and *Staphylococcus lugdunensis*) were the most commonly
isolated bacteria, followed by S. aureus, Enterococcus species, Streptococcus salivarius, and Bacillus megatherium (Table 2).

For Gram-negative bacteria, Enterobacter cloacae and Serratia marcescens have represented the bacteria frequently encountered, followed by Klebsiella pneumoniae, Escherichia coli, and Pseudomonas aeruginosa (Table 3).

3.2. Antimicrobial Resistance Profile of Bacterial Isolates.
In this study, the antimicrobial resistant pattern of Gram-positive and Gram-negative ocular bacterial isolates was evaluated. The most of Gram-positive isolates had shown greater resistance ($R > 62.1\%$) to ampicillin, amoxicillin-clavulanic acid, cefoxitin, erythromycin, and oxacillin. These strains had exhibited a higher rate of antibiotic sensitivity ($S > 62.1\%$) to ceftaroline, daptomycin, linezolid, moxifloxacin, rifampicin, teicoplanin, tigecycline, mupirocin, vancomycin, trimethoprim-sulfamethoxazole, and nitrofurantoin (Table 4). Among the strains of S. aureus, 50% were methicillin-resistant (MRSA) and had inducible MLSB resistance phenotypes. Among the CoNS isolates, 72% were methicillin-resistant (MR-CoNS), and of these, only 41.4% had shown inducible MLSB resistance phenotypes.

The Gram-negative isolates had presented greater resistance ($R > 63.6\%$) to amikacin, cefotaxime, ceftazidime, ciprofloxacin, colistin, ertapenem, gentamycin, meropenem, imipenem, levofloxacin, piperacillin-tazobactam, and trimethoprim-sulfamethoxazole (Table 5). Resistance profile of the most represented bacterial isolates for common drugs of choice for empirical therapy is shown in Table 6. Bacterial strains most resistant to the common treatments of ocular infections were Enterococcus spp. for Gram-positive and K. pneumoniae for Gram-negative.
4. Discussion

The bacterial ocular infections are commonly diagnosed in pediatric patients, affecting infants and preschool-aged children of both genders [22]. These diseases affect about 1 in every 8 children every year [23]. Although most cases are self-limiting, in others, about three weeks may be needed to remove the infection [24]. The gold standard for the treatment of bacterial ocular infection should be the identification of the agent and antibiotic susceptibility testing. In order to reduce antibiotic resistance, surveillance data of resistance profiles can guide the choice of appropriate empirical therapy in the absence of culture and sensitivity data [25]. The current analysis shows the incidence of ocular infections among pediatric patients, evaluating the pathogens involved in the infection and the related resistance profiles. In this study, 228 pediatric patients with supposed ocular surface infection were enrolled. Of these, 32% were suffering from an ocular infection. Similar proportions had been observed in India (34.5%), Japan (32.2%), and Iran (37.5%) [26–28]. A higher incidence was recorded in Ethiopia (74.7%) and Jordan (54.2%) [29, 30]. Sociodemographic and geographic aspects could explain these differences [31]. The high frequency of ocular infections was observed in the 12-month group. The elevated prevalence in this age group is mainly due to poor hand hygiene [32].

Table 5: Sensitivity and resistance of Gram-negative isolates.

<table>
<thead>
<tr>
<th>Antibiotics</th>
<th>Sensitivity (%)</th>
<th>Resistance (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ampicillin</td>
<td>—</td>
<td>100</td>
</tr>
<tr>
<td>Amikacin</td>
<td>63.6</td>
<td>36.4</td>
</tr>
<tr>
<td>Amoxicillin-clavulanic acid</td>
<td>18.2</td>
<td>36.4</td>
</tr>
<tr>
<td>Cefepime</td>
<td>36.4</td>
<td>63.6</td>
</tr>
<tr>
<td>Cefotaxime</td>
<td>72.7</td>
<td>27.3</td>
</tr>
<tr>
<td>Ceftazidime</td>
<td>72.7</td>
<td>27.3</td>
</tr>
<tr>
<td>Cefuroxime</td>
<td>9.1</td>
<td>90.9</td>
</tr>
<tr>
<td>Ciprofloxacin</td>
<td>72.7</td>
<td>27.3</td>
</tr>
<tr>
<td>Colistin</td>
<td>81.8</td>
<td>18.2</td>
</tr>
<tr>
<td>Ertapenem</td>
<td>72.7</td>
<td>27.3</td>
</tr>
<tr>
<td>Fosfomycin</td>
<td>54.5</td>
<td>45.5</td>
</tr>
<tr>
<td>Gentamycin</td>
<td>72.7</td>
<td>27.3</td>
</tr>
<tr>
<td>Imipenem</td>
<td>72.7</td>
<td>27.3</td>
</tr>
<tr>
<td>Meropenem</td>
<td>72.7</td>
<td>27.3</td>
</tr>
<tr>
<td>Levofloxacin</td>
<td>72.7</td>
<td>27.3</td>
</tr>
<tr>
<td>Piperacillin</td>
<td>54.5</td>
<td>45.5</td>
</tr>
<tr>
<td>Piperacillin-tazobactam</td>
<td>63.6</td>
<td>36.4</td>
</tr>
<tr>
<td>Tigecycline</td>
<td>36.4</td>
<td>63.3</td>
</tr>
<tr>
<td>Tobramycin</td>
<td>54.5</td>
<td>45.5</td>
</tr>
<tr>
<td>Trimethoprim-sulfamethoxazole</td>
<td>72.7</td>
<td>27.3</td>
</tr>
</tbody>
</table>

Table 6: Resistance rates of Gram-positive and Gram-negative isolates for common tested antibiotics.

<table>
<thead>
<tr>
<th>Antibiotics (%)</th>
<th>Resistance profile for Gram-positive isolates</th>
<th>Resistance profile for Gram-negative isolates</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td><em>S. aureus</em></td>
<td>CoNS</td>
</tr>
<tr>
<td>Ampicillin</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>Ciprofloxacin</td>
<td>14.3</td>
<td>24.4</td>
</tr>
<tr>
<td>Amoxicillin-clavulanic acid</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>Gentamycin</td>
<td>64.3</td>
<td>81.8</td>
</tr>
<tr>
<td>Fosfomycin</td>
<td>7.1</td>
<td>24.4</td>
</tr>
<tr>
<td>Trimethoprim-sulphamethoxazole</td>
<td>28.6</td>
<td>17.1</td>
</tr>
</tbody>
</table>
Gram-negative bacteria only partially contribute to ocular infections, according to the study of Mohammad [37]. The most isolated strains were Enterobacter cloacae (4.1%) and Serratia marcescens (4.1%). Different results were observed in Ethiopia, where the most isolated Gram-negative strain is Pseudomonas aeruginosa (11.3%). Among Gram-positive bacteria, ampicillin, amoxicillin-clavulanic acid, cefoxitin, erythromycin, and oxacillin showed a lower performance, recording resistance rates greater than 63.8%. daptomycin, teicoplanin, tigecycline, moxifloxacin, rifampicin, mupirocin, rifampicin, teicoplanin, tigecycline, vancomycin, and nitrofurantoin had exhibited a higher efficacy against Gram-positive strains (S > 81%). Methylcellulose resistance was detected in 72% of the isolated CoNS. A lower incidence of these strains was observed in the United States (47.4%), in Uganda (27.6%), and in Ethiopia (45.2%). Concerning Gram-negative bacteria, all strains had shown a high rate of resistance to ampicillin and cefuroxime (R > 90.9%). They had exhibited high efficacy of cefotaxime, ceftazidime, colistin, ertapenem, meropenem, imipenem, gentamycin, levofloxacin, and trimethoprim-sulfamethoxazole (R > 72.7%). However, fluoroquinolones represent the most used antibiotics in ophthalmic practice, and they remain effective against the strains responsible for ocular infections [38]. The Gram-negative and positive isolated bacteria in our University Hospital had shown a low rate of resistance to the fluoroquinolones tested (ciprofloxacin, moxifloxacin, and levofloxacin). Antibiotics, belonging to this class, inhibit DNA gyrase and topoisomerase IV, enzymes that are involved in DNA replication [39]. These drugs are broad-spectrum antibiotics, providing excellent coverage against most ocular pathogens [40]. It is well tolerated on the ocular surface, and the topical use reduces the development of bacterial resistance [41].

These data can be the starting point for outlining the guideline in the treatment of the pediatric patient’s ocular infection. In conclusion, the main goal of our study was to report the bacterial profile and antibiotic susceptibility pattern of ocular infection in pediatric patients in order to know the epidemiology of our hospital, reducing the antibiotic resistance and improving the empirical treatment with factual and statistical information.

Data Availability

Epidemiological data used to support the results of this study are included in the article.

Conflicts of Interest

None of the authors have any conflicts of interest related to this submission.

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