Research Article

Recyclable Xanthan/TiO\textsubscript{2} Composite Cryogels towards the Photodegradation of Cr(VI) Ions and Methylene Blue Dye

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1. Introduction

Porous 3D polymer structures have a high surface area and low density, making them applicable as filters, catalysts, and insulators. A reliable way to produce 3D polymer structures is removing the solvent of a precursor gel of interest causing minimal damage to the original structure. When the solvent is exchanged by supercritical CO\textsubscript{2} and then CO\textsubscript{2} is eliminated by pressure decrease, the resulting monoliths are classified as aerogels [1]. When the precursor gel is frozen and the solvent is removed by freeze-drying, the monoliths are classified as cryogels [2]. Aerogels and cryogels can be tailored by choosing the material and synthesis that best fit the desired application [3, 4].

Composite cryogels are prepared by combining polymer and reinforcing particles to improve the mechanical, thermal, electric, magnetic, and catalytic properties. For instance, poly(N-isopropylacrylamide) cryogels reinforced with silica nanoparticles presented superior mechanical and thermal behavior in comparison to bare poly(N-isopropylacrylamide) cryogels [5]. Polysaccharide/clay aerogels and cryogel compositions are interesting because they are biodegradable systems. Some examples of successful biopolymer/clay porous structures are cellulose/clay [6, 7], casein/clay [8], alginate/clay [9], and xanthan gum/agar/clay [10].

Xanthan gum (XG) is produced at large scale by Xanthomonas campestris during the fermentation of monosaccharides [11]. It is a polysaccharide composed by D-glucosyl, D-mannosyl, and D-glucuronyl acid residues in a 2:2:1 molar ratio and variable proportions of O-acetyl and pyruvyl residues. Side chains consist of a trisaccharide composed of mannose (\(\beta\)-1,4) gluconic acid (\(\beta\)-1,2) mannose attached to alternate glucose residues in the backbone by \(\alpha\)-1,3 linkages [12]. Above pH 4.5, the D-gluconic acid and pyruvyl residues are deprotonated, and XG chains behave as polyanions. Under these conditions, XG chains can form physical
or chemical networks either by interacting with polyvalent cations [13, 14] or by crosslinking with multifunctional molecules, respectively [12]. Citric acid is an efficient nontoxic crosslinker for polysaccharides; the esterification reaction among the carboxylic acid groups and polysaccharide hydroxyl groups takes place upon heating at 165°C for seven min [15–17]. XG is biodegradable and biocompatible; for this reason, it is widely used in food and drug formulation [12, 18] and as scaffold for tissue engineering [19–23].

The combination of XG with inorganic particles improves the bioaffinity of XG scaffold, physical and chemical properties towards bare XG. For instance, XG/hydroxyapatite nanocomposites presented improved mechanical behavior [24, 25] and served as scaffolds for the proliferation of osteoblasts [24]. XG/montmorillonite in combination with chitosan [26] or agar [10] formed foams with superior mechanical properties. XG/bioglass hybrid scaffolds reinforced with cellulose nanocrystals presented improved mechanical stability in dry and wet states and good compatibility with osteoblasts [27]. XG/SiO2 composites presented excellent mechanical properties and high capacity for the adsorption of methylene blue (MB) and Bismarck brown dyes [28]. Favorable interaction among XG chains and TiO2 particles led to improved rheological behavior and interior wall coatings [29].

In the present study, composite cryogels of XG were prepared with different contents of TiO2 P25 nanoparticles. The characterization of composite cryogels comprised compression tests, swelling degree determination, Fourier transform infrared vibrational spectroscopy in the attenuated total reflectance mode (FTIR-ATR), scanning electron microscopy (SEM), and X-ray microtomography (CT) analyses. The catalytic properties of XG/TiO2 composite cryogels were tested upon immersion in aqueous solutions containing methylene blue (MB) or Cr(VI) ions under exposition of UV radiation. The photoinduced reaction showed better correlation with first-order kinetics model. The possibility of composite cryogels recycling was also evaluated. MB molecules and Cr(VI) ions were chosen because they can be found as pollutants in natural waters.

2. Materials and Methods

2.1. Materials. Xanthan gum, XG (Kelzan®, CP Kelco, Brazil, Mw ~ 1.2 × 10⁶ g mol⁻¹), commercial titanium dioxide particles TiO2 P25 (AEROSIL®), Evonik, Brazil), citric acid (Labsynth, Brazil, 192.13 g mol⁻¹), and sodium hypophosphite (Labsynth, Brazil, 87.98 g mol⁻¹) were used as received. The chemical structures of XG and citric acid are provided in Figures 1(a) and 1(b), respectively. Hydrochloric acid (Labsynth, Brazil, 1.17 g mL⁻¹, 37%, 36.46 g mol⁻¹) and sodium hydroxide (Labsynth, Brazil, 40.00 g mol⁻¹) were used to adjust solutions pH during synthesis and long-term stability tests. Solutions of potassium dichromate (Labsynth, Brazil, 294.18 g mol⁻¹) and methylene blue (M9140, Sigma-Aldrich, 319.85 g mol⁻¹) were used to test the catalysis effectiveness of hybrid cryogels under UVC light.

2.2. Synthesis of Composite Cryogels. Figure 1(c) shows schematically the preparation of composite cryogels. First, TiO2 particles were dispersed in Milli-Q water at 1.0 g L⁻¹, 2.0 g L⁻¹, or 4 g L⁻¹ under magnetic stirring for 30 min. Then, XG, citric acid (crosslinker), and sodium hypophosphite (catalyst) were added to the TiO2 dispersions, so that their final concentrations amounted to 20 g L⁻¹, 1.0 g L⁻¹, and 0.5 g L⁻¹, respectively. The contents of TiO2 particles in relation to the XG mass amounted to 5%, 10%, or 20%. The pH was adjusted to 2, 4, or 7 by adding droplets of HCl 0.1 mol L⁻¹ or NaOH 0.1 mol L⁻¹. After 8 h under magnetic stirring, the precursor composite gels were kept in the refrigerator at 7°C overnight. As control, XG cryogels were prepared in the absence of TiO2 particles.

The precursor gels were poured either into rectangular polypropylene molds (25 mm × 17 mm × 13 mm) for mechanical tests, cylindrical acrylic molds of 10 mm diameter and 6 mm high for morphological analyses, or polystyrene Petri dish 35 mm diameter and 2 mm high for the catalytic assays. The photographs of molds and resulting cryogels are presented as Supplementary Material SM1. The samples were frozen during 2 hours in a standard freezer at −25°C, followed by 24 h of freeze-drying. After that, the cryogels were withdrawn from the molds and heated for 7 min at 165°C to promote the esterification among citric acid and hydroxyl groups from XG chains [17]. The resulting cryogels were rinsed with Milli-Q water in order to remove unreacted molecules and freeze-dried again; they were coded as XG, XG/TiO2 5%, XG/TiO2 10%, and XG/TiO2 20%.

2.3. Characterization. The X-ray diffractograms (XRD) of TiO2 particles were recorded with a Rigaku Miniflex diffractometer (Tokyo) operating at 30 kV, 15 mA, and λ (CuKα) = 0.154 nm, 0.5 min⁻¹, sampling width of 0.01°, at room temperature in the 2θ range between 10° and 90°. Dynamic light scattering (DLS) measurements were performed in a Zetasizer Nano-ZS90 Malvern equipment with a He-Ne laser operating at 632.8 nm at 25°C for dispersions of TiO2 at 0.01 wt%, pH 7.0, pH 4.0, and pH 2.0 after five min equilibration. The average particle diameter (z-average, Dz) and polydispersity (PDI) of each sample were calculated using the cumulant method for the corresponding autocorrelation function. The Dz and PDI values represent the mean values for triplicates. Scanning electron microscopy (SEM) analyses were performed in a Jeol microscope FEG7401F equipped with a field-emission gun operating at voltage of 3 kV. Droplets of diluted dispersion of TiO2 were deposited on clean Si wafers and allowed to dry at 40°C overnight.

The apparent density (ρapp) of composite cryogels was determined by weighing them in an analytical balance and measuring their dimensions with a pachometer over ten samples at 22 ± 1°C and relative humidity of 60 ± 10%. The swelling degree (SD) was determined with a precision tensiometer Krüss K100 at 22 ± 1°C and relative humidity of 65 ± 5%. The SD was calculated as the mass of sorbed water after 10 min divided by the mass of dried adsorbent, which was in average 12 ± 1 mg. Fourier transform infrared vibrational spectroscopy in the attenuated total reflectance mode (FTIR-ATR) spectra were obtained with a PerkinElmer Frontier with Zn/Se crystal equipment with resolution of 4 cm⁻¹ and in
the range of 600 cm\(^{-1}\) to 4000 cm\(^{-1}\). Compression tests were performed with a digital dynamometer IP 90DI-10 with a load cell of 10.0 N, limit of applied force of 0.1 N, accuracy of 0.5\% at 22 ± 1 °C, and relative humidity of 60 ± 10\%.

SEM analyses were conducted with a JEOL Neoscope JCM 5000 equipment, operating at voltage of 10 kV, on samples coated by sputtering with 5 nm of gold. X-ray microtomography (CT) analyses were performed with a Skyscan 1272 Bruker equipment, operating at 20 kV and 175 μA. The sample was rotated stepwise (0.6° per step) through 360°, and images were recorded at each step with an exposure time of 5 s, yielding spatial resolution of 10 μm. In average, 600 X-ray images were taken in 60 min, scanning along the whole sample, yielding images with spatial resolution of 10 μm. The thresholding of the calculated gray scale images (from 13 to 255) and the transformation of the CT data into microstructural parameters were performed for all samples in the same way, using the CTAn Bruker software.

2.4. Photoreduction of Cr(VI) and Methylene Blue (MB). Discs of cryogels (2 mm high, 35 mm diameter, ~0.050 g) were inserted in Petri dishes (55 mm diameter) containing 5 mL of 120 mg L\(^{-1}\) (4 × 10\(^{-4}\) mol L\(^{-1}\)) K\(_2\)Cr\(_2\)O\(_7\) solution at pH 1.0 or 2.0. As a control experiment, pieces of bare XG cryogels were tested under the same conditions. The systems were arranged equidistant from an UVC lamp (Osram 36 W, 265 nm at 10 cm of the bulb), which was mounted in the center of a closed box (Supplementary Material SM2). The systems were irradiated during 15 min, then an aliquot of 1 mL was withdrawn for the quantification of Cr(VI) in the solution by means of spectrophotometry at 434 nm (Beckman-Coulter DU650 spectrophotometer), which is a wavelength for the maximal absorbance. After absorbance reading, which took less than one minute, the aliquot was returned to the Petri dish. The procedure was repeated 6 times, totaling 90 minutes of irradiation. The reduction of Cr(VI) to Cr(III) was monitored using as reference the absorbance of the initial 4 × 10\(^{-4}\) mol L\(^{-1}\) K\(_2\)Cr\(_2\)O\(_7\) solution, which was not irradiated and was kept in the absence of UV irradiation.

Discs of cryogels (2 mm high, 35 mm diameter, ~0.050 g) were inserted in Petri dishes (55 mm diameter) containing 5 mL of 5.0 mg L\(^{-1}\) (15.6 μmol L\(^{-1}\)) MB solution prepared in 50 mM Tris-HCl, pH 7.0 [30]. At this, pH MB molecules are positively charged [31]. As a control experiment, pieces of bare XG cryogels were tested under the same conditions. The systems were irradiated during 15 min, and then an aliquot of 1 mL was withdrawn for the quantification of MB.
in the solution by means of spectrophotometry at 664 nm, which is a wavelength for the maximal absorbance. After absorbance reading, the aliquot was returned to the Petri dish. The procedure was repeated 8 times, totaling 120 minutes of irradiation. The photobleaching of MB was monitored using the initial MB solution (~17 μmol L⁻¹), which was not irradiated and was kept in the absence of UV irradiation.

3. Results and Discussion

3.1. Characterization of TiO₂ Particles. Figure 2(a) shows the X-ray diffractogram obtained for TiO₂ P25 particles. It presents typical diffraction peaks of anatase and rutile iso-morphic forms. The weight percentages of anatase and rutile, \( X_A \) and \( X_R \), respectively, were estimated using the Spurr-Myers equations [32].

\[
\begin{align*}
X_A &= \frac{100}{(1 + 1.265( I_R / I_A ))}, \\
X_R &= \frac{100}{(1 + 0.8( I_A / I_R ))}, \\
\end{align*}
\]

where \( I_A \) and \( I_R \) are the intensity of anatase peak at \( 2θ = 25.25° \) (101) and the intensity of rutile peak at \( 2θ = 27.42° \) (110). The contents of anatase and rutile amounted to 77% and 23%, respectively, which are in agreement with literature values [33].

Figure 2(b) shows the size distribution determined for TiO₂ P25 particles dispersed at pH 7.0, pH 4.0, and pH 2.0, the mean \( D_z \) values amounted to (344 ± 15) nm, (289 ± 8) nm, and (281 ± 4) nm, respectively, and the mean PDI values amounted to 0.45 ± 0.06, 0.28 ± 0.03, and 0.31 ± 0.04, respectively. The average sizes of the anatase and rutile elementary particles are 85 nm and 25 nm, respectively [34]. Thus, the observed \( D_z \) values probably correspond to the aggregates of TiO₂ P25. Moreover, the experimental \( D_z \) data indicated that at pH 7.0, the particles tend to form larger aggregates because it is close to the isoelectric point (pI) of TiO₂ P25, which is reported as 6.2 [35], whereas at pH 4.0 or 2.0, they presented similar mean \( D_z \) values. SEM images obtained for TiO₂ P25 particles dispersed at pH 2.0 were provided as Supplementary Material SM3, they revealed the predominance of particles with mean size of (30 ± 5) nm and aggregates ranging from 100 nm to 500 nm in agreement with the \( D_z \) values.

3.2. Characterization of Composite Cryogels. In order to evaluate the effect of TiO₂ content in the cryogels on the compressive modulus, all cryogels were prepared at pH 4.0 because at this pH no tendency of particle aggregation was observed (Figure 2(b)). Typical compressive stress-strain curves determined for bare XG cryogels and XG/TiO₂ cryogels with 5%, 10%, and 20% TiO₂ prepared at pH 4.0 are presented in the Supplementary Material Figure SM4; the corresponding Young compressive modulus (\( E \)) values were determined from the slopes of linear regions. Figure 3 shows that the mean \( E \) values increased with the increase of TiO₂ content in the cryogels up to 10%, where the maximum \( E \) value achieved 100 ± 7 kPa. Similar behavior was observed for TiO₂ in epoxy composites [36] and cellulose nanocrystals in cellulose acetate butyrate composites [37]. A possible explanation for this effect is that for TiO₂ contents larger than 10%, the dispersion of the filler in the matrix is no longer effective, reducing the compressive strength.

In order to evaluate the effect of the precursor pH on the physicochemical properties of resulting cryogels,
Composite cryogels were prepared with 10% TiO$_2$ at pH 2.0, 4.0, and 7.0. Table 1 shows the compression modulus ($E$) and apparent density ($\rho_{app}$) values determined for bare XG and XG/TiO$_2$ 10% cryogels prepared at pH 2.0, 4.0, and 7.0. The data represent mean values and standard deviations of triplicate.

The $E$ values determined for the composite cryogels prepared at pH 2.0 or pH 4.0 were approximately fourfold and threefold that determined for bare XG cryogels, respectively. This interesting finding is in agreement with the observation that the addition of clay increased the $\rho_{app}$ and $E$ values of XG aerogel composites [10]. In general, for cellular materials, the $E$ values are expected to increase if the $\rho_{app}$ increase [38]. Thus, if the inorganic particles are well distributed in the polymeric matrix, increasing the composite density, the improvement of mechanical properties is expected [37].

The preparation of composite cryogels at pH 7.0 led to a drastic decrease of compressive modulus, indicating that at pH 7.0 the interactions among TiO$_2$ particles and XG chains were weakened. At pH 7.0, the particles probably tend to aggregate due to the proximity to the pI of TiO$_2$ (6.2) [35], impairing the mechanical properties. The mean $E$ value determined for XG/TiO$_2$ 10% composite cryogels prepared at pH 7 was almost twofold that obtained for bare XG cryogels. It was expected because the mean $\rho_{app}$ value of composite cryogels was 1.3-fold larger than that of bare XG cryogels.

Figure 4 shows the SEM images obtained for bare XG and XG/TiO$_2$ 10% cryogels prepared at pH 2.0, 4.0, and 7.0. All cryogels presented isotropic open cells, regardless of the pH or composition. Bare XG hydrogels prepared from precursors containing acetic acid or hydrochloric acid presented similar morphological features [39]. The composite cryogels presented in average larger pores than the bare XG cryogels. In order to gain insight about the morphometric parameters, microtomography (CT) analyses were performed for XG and XG/TiO$_2$ 10% cryogels prepared at pH 4.0. Figure 5 and Table 2 show the typical reconstructed CT images and the corresponding morphometric parameters. Table 2 shows that the $\rho_{app}$ values calculated from CT analyses were similar to those calculated by dividing the mass by the volume presented in Table 1. XG and XG/TiO$_2$ 10% cryogels presented similar $\rho_{app}$, surface area ($S$), and porosity ($P$) values. However, the values of mean pore size ($d$) calculated for XG/TiO$_2$ 10% cryogels were larger than that for XG cryogels, in agreement with the SEM images in Figure 4. In average, the walls of XG/TiO$_2$ 10% cryogels were thicker ($h = 61$ $\mu$m) than those of XG cryogels ($h = 47$ $\mu$m), evidencing the integration of TiO$_2$ particles to the cryogels walls.

One potential application for TiO$_2$ hybrid composites is the photocatalytic reduction of pollutants present in aqueous media [40]. For this reason, the long-term stability in water was also investigated. Figure 6 shows photographs taken for XG and XG/TiO$_2$ 10% cryogels prepared at pH 2.0, 4.0, and

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**Table 1:** Compression modulus ($E$) and density ($\rho_{app}$) values determined for bare XG and XG/TiO$_2$ 10% cryogels prepared at pH 2.0, 4.0, and 7.0. The data represent mean values and standard deviations of triplicate.

<table>
<thead>
<tr>
<th>pH</th>
<th>XG</th>
<th>XG/TiO$_2$ 10%</th>
<th>XG</th>
<th>XG/TiO$_2$ 10%</th>
</tr>
</thead>
<tbody>
<tr>
<td>2.0</td>
<td>30 ± 3</td>
<td>120 ± 8</td>
<td>24.0 ± 0.4</td>
<td>28 ± 1</td>
</tr>
<tr>
<td>4.0</td>
<td>29 ± 3</td>
<td>100 ± 7</td>
<td>23 ± 1</td>
<td>25 ± 1</td>
</tr>
<tr>
<td>7.0</td>
<td>29 ± 3</td>
<td>49 ± 3</td>
<td>23.8 ± 0.7</td>
<td>30 ± 1</td>
</tr>
</tbody>
</table>

---

**Figure 3:** Dependence of compression modulus ($E$) and apparent density on the TiO$_2$ content in the composites from precursors prepared at pH 4.0. The data represent mean values and standard deviations of triplicate.

---

**Figure 4:** SEM images obtained for bare XG and XG/TiO$_2$ 10% cryogels prepared at pH 2.0, 4.0, and 7.0. All cryogels presented isotropic open cells, regardless of the pH or composition. Bare XG hydrogels prepared from precursors containing acetic acid or hydrochloric acid presented similar morphological features [39]. The composite cryogels presented in average larger pores than the bare XG cryogels. In order to gain insight about the morphometric parameters, microtomography (CT) analyses were performed for XG and XG/TiO$_2$ 10% cryogels prepared at pH 4.0. Figure 5 and Table 2 show the typical reconstructed CT images and the corresponding morphometric parameters. Table 2 shows that the $\rho_{app}$ values calculated from CT analyses were similar to those calculated by dividing the mass by the volume presented in Table 1. XG and XG/TiO$_2$ 10% cryogels presented similar $\rho_{app}$, surface area ($S$), and porosity ($P$) values. However, the values of mean pore size ($d$) calculated for XG/TiO$_2$ 10% cryogels were larger than that for XG cryogels, in agreement with the SEM images in Figure 4. In average, the walls of XG/TiO$_2$ 10% cryogels were thicker ($h = 61$ $\mu$m) than those of XG cryogels ($h = 47$ $\mu$m), evidencing the integration of TiO$_2$ particles to the cryogels walls.

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Figure 4: SEM images obtained for XG and XG/TiO$_2$ 10% cryogels prepared at pH 2.0, 4.0, and 7.0.

Figure 5: X-ray computer tomography images for (a) XG/TiO$_2$ 10% and (b) XG cryogels, which were prepared at pH 4.0 in molds of 7 mm and 5 mm diameter, respectively. Typical slices, from a total of 600 slices for each sample, are presented in the upper images. The reconstruction of 3D boxes (2 mm $\times$ 2 mm $\times$ 1 mm) was based on the slices and served for the morphometric analyses. The colors resulted from the transformation of the gray scale (red from 25% to 35% max attenuation, yellow from 35% to 45% attenuation, and white from 45% to 100%).
Table 2: Morphometric parameters from CT analyses: specific area (S), mean pore size (d), wall thickness (h), and integrated porosity (P) were determined for XG and XG/TiO2 10% cryogels.

<table>
<thead>
<tr>
<th>Cryogel</th>
<th>Φ (mm)</th>
<th>ρ_app (kg m⁻³)</th>
<th>S (m² g⁻¹)</th>
<th>d (μm)</th>
<th>h (μm)</th>
<th>P (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>XG</td>
<td>5</td>
<td>24 ± 2</td>
<td>2.5 ± 0.1</td>
<td>39 ± 2</td>
<td>47 ± 1</td>
<td>93.2</td>
</tr>
<tr>
<td>XG/TiO2 10%</td>
<td>7</td>
<td>26 ± 2</td>
<td>2.8 ± 0.4</td>
<td>45 ± 2</td>
<td>61 ± 1</td>
<td>93.8</td>
</tr>
</tbody>
</table>

7.0 just after immersion in Milli-Q water and after 7, 14, and 21 days in Milli-Q water. Just after immersion, only XG and XG/TiO2 10% cryogels prepared at pH 2.0 or pH 4.0 kept their original form; those prepared at pH 7.0 swelled and dissolved. This behavior corroborates with the low compressive strength observed for the XG/TiO2 10% cryogels prepared at pH 7.0 (Table 1), which was attributed to weak interaction among XG chains and TiO2 particles. If the TiO2 particles were intimately attached to the XG chains, the cryogels would have higher E value and better stability in water.

After seven days in contact with Milli-Q water, the cryogels prepared at pH 2.0 or 4.0 swollen but did not dissolve, whereas those synthesized at pH 7.0 dissolved completely. After 14 or 21 days in contact with water, the XG cryogels prepared at pH 2.0 or 4.0 remained the same. On the other hand, after 14 or 21 days, the XG/TiO2 10% cryogels prepared at pH 2.0 presented fungi on the surface, whereas those prepared at pH 4.0 were completely free of fungi and the supernatant turned turbid, indicating that TiO2 particles were partially released to the medium, avoiding fungi proliferation. The antimicrobial properties of TiO2 particles are well described in the literature [41]; one possible mechanism is the peroxidation and decomposition of fatty acids present in the bacteria membrane cell by the photoactivity of TiO2 particles [42].

The esterification among citric acid molecules and XG chains were favored at pH 2.0 or 4.0 because the carboxylic acid groups are protonated [16, 17]. At pH 7.0, the esterification was not possible due to the deprotonation of XG and citric acid groups. Considering the pI of TiO2 P25 at pH 6.2 [35], at pH 2.0 or 4.0 the particles surface is enriched by the Ti-O-H groups, which can undergo esterification with the carboxylic acid groups from citric acid and XG chains. The photographs showed that composite cryogels prepared at pH 4.0 remained stable in water for a long time, but the supernatant became slightly turbid, indicating that some TiO2 particles were not completely free of fungi and the supernatant turned turbid, indicating that TiO2 particles were partially released to the medium, avoiding fungi proliferation. The antimicrobial properties of TiO2 particles are well described in the literature [41]; one possible mechanism is the peroxidation and decomposition of fatty acids present in the bacteria membrane cell by the photoactivity of TiO2 particles [42].

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The photocatalytic properties of XG and XG/TiO2 10% cryogels prepared at pH 4.0 were evaluated in the reduction of Cr(VI) to Cr(III) ions and in the photobleaching of methylene blue (MB), under exposition of UV radiation. Control experiments were conducted in the dark. The irradiation of TiO2 with UV light results in photon absorption and excitation of an electron (e⁻_VB) from valence band (VB) to the conduction band (CB), thereby generating a positive electron hole in the valence band (h⁺_VB) [43]:

\[
\text{TiO}_2 + h\nu_{(UV)} \rightarrow h^+_\text{VB} + e^-_\text{VB} \tag{2}
\]

The e⁻_VB and h⁺_VB species are present on the TiO2 particles surface. The e⁻_VB can react with O₂ to form superoxide radicals or hydroperoxide radicals, and the h⁺_VB can react with water, generating hydroxyl radicals:

\[
e^-_\text{VB} + O_2 \rightarrow O_2^- \tag{3}
\]
\[
h^+_\text{VB} + H_2O \rightarrow OH + H^+ \tag{4}
\]

Under acid conditions, the reduction of Cr(VI) to Cr(III) follows:

\[
\text{Cr}_2\text{O}_7^{2-} + 14H^+ + 6e^-_\text{VB} \rightarrow 2\text{Cr}^{3+} + 7H_2O \tag{5}
\]

Figure 7(a) shows the FTIR-ATR spectra obtained for TiO2 P25 (powder), XG (powder), and XG and XG/TiO2 10% cryogels prepared at pH 4.0. The complete band assignment is presented as Supplementary Material Table SM1. Briefly, a broad band from 3500 to 3000 cm⁻¹ appeared in all spectra, it was attributed to O-H stretching. Except for the TiO2 P25 (powder) spectrum, the C-H stretching band at 2910 cm⁻¹ was present in all spectra. The spectral region from 1100 to 1800 cm⁻¹ was shown in Figure 7(b). The most important feature is that the ester bonds resulting from the esterification among citric acid carboxylic acid groups and/or XG hydroxyl groups were evidenced by the appearance of the band at 1725 cm⁻¹ in the FTIR-ATR spectra determined for XG and XG/TiO2 10% cryogels which was attributed to C=O axial deformation; noteworthy, this band was absent in the XG (powder) [16].

3.3. Photocatalytic Properties of Cryogels. The photocatalytic properties of XG and XG/TiO2 10% cryogels prepared at pH 4.0 were evaluated in the reduction of Cr(VI) to Cr(III) ions and in the photobleaching of methylene blue (MB), under exposition of UV radiation. Control experiments were conducted in the dark. The irradiation of TiO2 with UV light results in photon absorption and excitation of an electron (e⁻_VB) from valence band (VB) to the conduction band (CB), thereby generating a positive electron hole in the valence band (h⁺_VB) [43]:

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h^+_\text{VB} + H_2O \rightarrow OH + H^+ \tag{4}
\]

Under acid conditions, the reduction of Cr(VI) to Cr(III) follows:

\[
\text{Cr}_2\text{O}_7^{2-} + 14H^+ + 6e^-_\text{VB} \rightarrow 2\text{Cr}^{3+} + 7H_2O \tag{5}
\]

Figure 8 shows the decrease of Cr(VI) concentration as a function of time at pH 1.0 and 2.0. In order to increase the contact area of the cryogels with the Cr(VI) ions in solution, the cryogels were prepared as discs (about 2 mm thick, 35 mm diameter) and immersed in the solution. The reduction of Cr(VI) to Cr(III) was observed in solutions, which were in contact with XG and XG/TiO2 10% cryogels. In both situations, the experimental data were better fitted with the first-order (Figures 8(b) and 8(d)) model than with the second-order (Supplementary Material SM6) model, in agreement with the photoreduction of Cr(VI) by pure TiO2 particles [44]. Table 3 presents the fitting parameters. At pH 2.0, the rate constants were similar in both cases, whereas at pH 1.0 the reduction rate was slightly larger for XG/TiO2...
10% than for XG cryogels. The rate constants in Table 3 are large in comparison to those reported for pure TiO₂, for instance, at pH 2 the \( k \) value is reported as 0.00373 min⁻¹ [44], five times smaller than those determined for XG/TiO₂ 10% cryogels. The efficiency of bare XG cryogels is due the reducing ends of XG chains. Efficient reduction of Cr(VI)
ions to Cr(III) ions was also observed for alginate [45] and chitosan [46] matrices.

The mechanism for the photodegradation of methylene blue (MB) molecules mediated by TiO₂ under UV radiation is well reported in the literature [47]. The MB molecules react with the reactive oxygen species (ROS) in equations (3) and (4), resulting in CO₂, H₂SO₄, and HNO₃ and as final products [47]. Figure 9(a) shows the decrease of MB concentration as a function of time at pH 7.0 (Tris-HCl buffer), the cryogel discs were immersed in the MB solutions while the systems were irradiated. The experimental data fitted better the first-order model (Figure 9(b)) than the second-order model (Supplementary Material SM7), as evidenced by the \( R^2 \) values in Table 3. The \( k^1 \) value determined for the photodegradation of MB with XG/TiO₂ 10% cryogels was twice the \( k^1 \) value determined for bare XG cryogels and 10 times \( k^1 \) value determined for the control, evidencing the catalytic efficiency of the composite cryogels. The \( k^1 \) values determined for the photobleaching of MB in contact with TiO₂ P25 and under laser pulse with 50, 100, 150, and 200 mJ energy amounted to 0.00433, 0.00519, 0.00741, and 0.00577 min⁻¹, respectively [48]; the \( k^1 \) values were similar to that
determined for bare XG cryogels. The control experiment was done in the absence of cryogels.

The initial concentration of MB was reduced by 46% and 95% upon contact with XG/TiO$_2$ 10% cryogels under UV radiation after 60 min and 5.2 h, respectively. For comparison, under optimum conditions, hydrogels of poly(acrylic acid) crosslinked with N,N-methylene bis-acrylamide (MBA), and TiO$_2$ reduced 95% of the initial MB concentration after 5 h under sunlight radiation [49]. In order to illustrate the photocatalytic effect of TiO$_2$ in the composites cryogels, 1 mL of MB solution at 7.8 $10^{-4}$ mol L$^{-1}$ was added to bare XG and XG/TiO$_2$ 10% cryogels, irradiated by UV for 2 h. Figure 10 shows the photographs of XG and XG/TiO$_2$ 10% before the addition of MB solution, just after the addition of MB solution, after 1 h and 2 h of UV radiation; the photographs clearly showed the catalytic effect of TiO$_2$ on the photodegradation of MB.

The possibility of recycling the XG/TiO$_2$ 10% cryogels was evaluated. After the photoreduction of Cr(VI) or photodegradation of MB, they were rinsed with the corresponding solvents (HCl or Tris-HCL bufer) and reused without losing the original shape or efficiency for five times.

### 4. Conclusions

The results presented in this study evidenced that the TiO$_2$ P25 particles enhanced the mechanical and catalytic properties of XG cryogels. The pH of the precursor gel played a crucial role on the chemical stability and mechanical properties of composite cryogels. Close to the pI of TiO$_2$ particles...
(pH 7), the interactions among XG chains and particles were not favored, yielding weak cryogels. On the other hand, the precursor preparation at pH 2 or pH 4 favored the interactions among XG hydroxyl and carboxylic acid groups and TiO$_2$ particles by H bonding. The outstanding mechanical properties and stability in water enabled the application of XG/TiO$_2$ 10% as adsorbents for pollutants present in aqueous media. Their efficiency in the reduction of Cr(VI) to Cr(III) ions and in the degradation of MB, upon UV radiation, could be attested in repeated cycles, without losing the original shape. Thus, the XG/TiO$_2$ 10% cryogels displayed mechanical and catalytic properties, which evidenced their potential as environmentally friendly adsorbents.

**Data Availability**

The data used to support the findings of this study are included within the supplementary information file.

**Conflicts of Interest**

The authors declare that there is no conflict of interest regarding the publication of this paper.

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**Supplementary Materials**

Figure S1: photographs of molds used and the resulting cryogels. Figure S2: experimental setup for the photocatalytic experiments. Figure S3: typical SEM images of TiO$_2$ P25 particles. The particles were not coated with gold prior to the analyses because they are semiconductor. The mean size of TiO$_2$ particles amounted to (30 ± 5) nm, as an average of 15 measurements (red lines). The aggregates ranged from 100 nm to 500 nm, in agreement with the $D_z$ values. Figure S4: typical compressive stress- (σ-) strain curves determined for bare XG cryogels (0%) and XG/TiO$_2$ cryogels with 5%, 10%, and 20% TiO$_2$ prepared at pH 4. Figure S5: typical Milli-Q water sorption curves determined for XG and XG/TiO$_2$ 10% cryogels prepared at pH 4. Figure S6: fittings to second-order kinetic model for the experimental data of reduction of Cr(VI) at (a) pH 1.0 and (b) pH 2.0 (see Figure 8). Figure S7: fittings to second-order kinetic model for the experimental data of degradation of MB at pH 7.0 (see Figure 9). Table S1: bands assignment regarding the FTIR spectra obtained for XG and TiO$_2$ powder and XG and XG/TiO$_2$ 10% cryogels. (Supplementary Materials)

**References**


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**Figure 10:** Photographs of XG and XG/TiO$_2$ 10% cryogels before the addition of MB solution, just after the addition of MB solution (0 h), after 1 h and 2 h of UV radiation.


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