High Performace Liquid Chromatographic Determination of Nicardipine Hydrochloride in Human Plasma

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Abstract. A sensitive high-performance liquid chromatographic method was developed for the estimation of nicardipine hydrochloride in human plasma. Varying amount of nicardipine hydrochloride (2.5 to 150 ng/0.5 mL) and fixed quantity (100 ng/0.5 mL) of nifedipine (internal standard) was added to blank human plasma, and a single step extraction was carried out with ethyl acetate. The mixture was centrifuged, ethyl acetate layer separated, dried and reconstituted with 100µL of acetonitrile. Twenty microliters of this solution was injected into a reverse phase C-18 column using a mobile phase consisting of acetonitrile: 0.02M potassium dihydrogen phosphate (pH 4.0) in the ratio of 60:40v/v and the eluents were monitored at 239 nm. The method was validated for its linearity, precision and accuracy. The calibration curve was linear in the range of 5-150 ng/0.5 mL of plasma and the lower detection limit was 2.5 ng/0.5mL of plasma. The intra- and inter-day variation was found to be less than 2.5% indicating that the method is highly precise. The mean recovery of nicardipine hydrochloride from plasma samples was 89.6±2.60%. The proposed HPLC method was applied for the estimation of nicardipine hydrochloride in human plasma after oral administration of an immediate release nicardipine hydrochloride capsule (dose 30 mg) to 6 adult male volunteers. There was no interference of either the drug metabolites or other plasma components with the proposed HPLC method for the estimation of nicardipine hydrochloride in human plasma. Due to its simplicity, sensitivity, high precision and accuracy, the proposed HPLC method may be used for biopharmaceutical and pharmacokinetic evaluation of nicardipine hydrochloride and its formulations in humans

Keywords: Nicardipine hydrochloride, HPLC, Human plasma, Pharmacokinetic
Introduction
Nicardipine hydrochloride, a potent calcium channel antagonist, is used in the treatment of angina pectoris and hypertension\textsuperscript{1,2}. The terminal half-life of nicardipine hydrochloride after single oral administration in human subjects varies from 2 to 4 h. After oral administration, nicardipine hydrochloride undergoes extensive first-pass elimination, and inter-and intra-subject variability of plasma concentration is observed\textsuperscript{2}. Because of the first-pass elimination, oral bioavailability\textsuperscript{2} of nicardipine hydrochloride in human subjects has been reported to be as low as 30-35\%. Several analytical methods have been reported for the estimation of nicardipine hydrochloride in plasma by HPLC\textsuperscript{3-11}, HPLC with gas chromatography-mass spectrometry\textsuperscript{12} and GC-MS combined with chiral stationary phase HPLC\textsuperscript{13}. The HPLC methods are simple, sensitive, precise and highly accurate, and require small quantity of sample. But some of the reported HPLC methods\textsuperscript{8,10,11} utilized special columns for the estimation of nicardipine hydrochloride in biological samples. The HPLC with gas chromatography-mass spectrometry\textsuperscript{12} and GC-MS combined with chiral stationary phase HPLC\textsuperscript{13} are time consuming and the process is expensive and tedious. The present study describes a rapid and simple HPLC method for the determination of nicardipine hydrochloride in human plasma by using RP C-18 column.

Experimental
Nicardipine hydrochloride was obtained from M/s. ICN Biomedicals, USA. Nifedipine was a gift sample from M/s Torrent Laboratories, Ahmedabad, India. Potassium dihydrogen phosphate was of analytical grade and supplied by M/s. S.D Fine-Chem limited, Mumbai, India. Acetonitrile and water used were of HPLC grade (E-Merck). All other reagents used in the study were of AR quality (Qualigens). A commercial (M/s. Zenith Goldline Pharmaceuticals, USA) capsules dosage form containing 30 mg of nicardipine hydrochloride was used in the study.

Instrumentation
A gradient HPLC (Shimadzu HPLC Class VP series) with two LC-10AT VP pumps, variable wave length programmable UV/VIS Detector SPD-10A VP, CTO-10AS VP Column oven (Shimadzu), SCL-10A VP system controller (Shimadzu), a disposable guard column LC-18 (Pelliguard\textsuperscript{TM}, LC-18, 2 cm, Supelco, Inc., Bellefonte, PA.) and RP C-18 column (250 mm X 4.6 mm I.D., particle size 5 \(\mu\)m; YMC Inc., USA) was used. The HPLC system was equipped with the software "Class-VP series version 5.03 (Shimadzu)".

HPLC conditions
Both acetonitrile and 0.02 M potassium dihydrogen phosphate solution were filtered before use through 0.45-\(\mu\)m membrane filter, and were pumped from the respective solvent reservoirs in the ratio of 60:40v/v to the column at a flow rate of 1 mL/min which yielded a column back-pressure of 74-75 kg/cm\textsuperscript{2}. The run time was set at 10 min and the column temperature was maintained at 40\(^\circ\)C. The volume of each injection was 20 \(\mu\)l. Prior to injecting solutions, the column was equilibrated for at least 30 min with the mobile phase flowing through the systems. The detector sensitivity was set at 0.0001 a.u.f.s and eluents were monitored at 239 nm and the data was acquired, stored and analyzed with the software “Class-VP series version 5.03 (Shimadzu)”.

Procedure
Solutions were prepared on a weight basis and volumetric flasks were used to minimize solvent evaporation. The quantification of the drug was accomplished by an internal standard method. Stock solutions of nicardipine hydrochloride and nifedipine were prepared
by dissolving 100 mg of drug in 100 mL volumetric flask containing 70 mL of methanol, vortexed for about 15 min and then made up to volume with methanol. Daily working standard solutions of nicardipine hydrochloride was prepared by suitable dilution of the stock solution with mobile phase.

**Calibration solutions**

Five sets of plasma samples with varying drug concentrations were prepared by spiking drug-free plasma with an appropriate volume (100 µL) of a known amount nicardipine hydrochloride at a concentration range of 2.5 to 150 ng/0.5 mL of plasma along with 100 µL of 0.1 µg/0.5 mL of nifedipine (internal standard) solution.

**Plasma extraction procedure**

An aliquot of plasma (0.5 mL) was accurately measured into a 10-mL glass tube with a teflon-lined cap, followed by the addition of 100 µL of 0.1 µg/0.5 mL of nifedipine solution as internal standard. To the above plasma samples 5 mL of ethyl acetate was added as extracting solvent, vortexed for 5 min, centrifuged at 3,000 rpm for 10 min, organic layer was separated and dried under vacuum evaporation. The residue was reconstituted with 0.1 mL of acetonitrile. Twenty microliters of the resultant solution was injected into the column (RP-C18).

**Accuracy and precision**

Aliquots of blank plasma (0.5 mL) were spiked with fixed quantity of internal standard (0.1µg/0.5 mL) and of nicardipine hydrochloride solutions (100 µL) with various strengths to yield concentrations of 5, 20, 50, 100 and 150 ng. Each of these samples was extracted as described above and the resulting reconstituted solutions were injected into the HPLC column (n=5). Each sample was prepared in five sets on three consecutive days and were assayed by the proposed HPLC method to find the precision and accuracy of the method. Accuracy was expressed as the mean percentage error [(mean measured concentration)/(expected concentration)] X 100. Precision was calculated as intra- and inter-day coefficient of variation (CV) [%C.V=(SD/mean) X 100]. Least square regression method was used to determine the regression equation and correlation coefficient.

**Results and Discussion**

Nicardipine hydrochloride and nifedipine appeared on the chromatograph in approximately 6.490 and 4.978 min respectively with no interfering peaks (Figure 1). The run time of the method was set at 10 minutes. When the same drug solution was injected 5 times, the retention time of both the drug and internal standard were same. This indicates that the proposed HPLC method is rapid, which in turn shows that the method consumes lesser amounts of expensive HPLC solvents. Table 1 shows the mean peak area ratios of nicardipine hydrochloride to nifedipine (internal standard) solutions for 5 such determinations. When the concentration of nicardipine hydrochloride and its respective peak area ratios (peak area nicardipine hydrochloride/ peak area of nifedipine) were subjected to regression analysis by least squares method, a high correlation coefficient was observed (r=0.9987) in the range of 5 to 150 ng/0.5 mL. However, the lower detection limit was found to be 2.5 ng/0.5 mL of human plasma. The regression of nicardipine hydrochloride concentration over its peak area ratio was found to be Y=0.02864+0.01161X where ‘Y’ is the peak area ratio and ‘X’ is the concentration of nicardipine hydrochloride in human plasma. This regression equation is used to estimate the amount of nicardipine hydrochloride in plasma or in validation study (precision and accuracy).
When the proposed HPLC method was validated for intra- and inter-day variation, it showed less than 2.5% of CV (Table 2). To assess the assay recovery from plasma after extraction with ethyl acetate, known quantity of the drug (10, 20, 50 or 100ng) along with internal standard was added to drug-free plasma and the resulting reconstituted solutions were repeatedly injected on the same day. The coefficient of variation (CV) in the peak area ratio for five replicate injections was found to be less than 3.5% and there was a high recovery (Table 3) of the drug (95.04±3.09%). Thus, the results show that the proposed HPLC method is highly reproducible and accurate.

![Figure 1](image)

**Table 1.** Calibration and precision of the HPLC assay

<table>
<thead>
<tr>
<th>Amount of nicardipine hydrochloride added to 0.5 mL of human plasma (ng)</th>
<th>Peak-area ratio*</th>
<th>%C.V.</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>5</td>
<td>0.06182</td>
<td>1.06</td>
</tr>
<tr>
<td>10</td>
<td>0.15048</td>
<td>1.64</td>
</tr>
<tr>
<td>20</td>
<td>0.29358</td>
<td>0.56</td>
</tr>
<tr>
<td>50</td>
<td>0.62888</td>
<td>1.65</td>
</tr>
<tr>
<td>100</td>
<td>1.20797</td>
<td>2.01</td>
</tr>
<tr>
<td>150</td>
<td>1.74818</td>
<td>0.98</td>
</tr>
</tbody>
</table>

*: Mean of five determinations Regression Equation: \( Y=0.02864+0.01161X \) (r=0.9987)

**Table 2.** Inter- and intra-day precision for nicardipine hydrochloride assay in human plasma of the proposed HPLC method

<table>
<thead>
<tr>
<th>Nicardipine hydrochloride concentration* ng/mL</th>
<th>Concentration of nicardipine hydrochloride (ng/mL) found</th>
<th>Intra-day Precision</th>
<th>Concentration of nicardipine hydrochloride (ng/mL) found</th>
<th>Inter-day Precision</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>5.01</td>
<td>2.10</td>
<td>100.20±2.2</td>
<td>4.99</td>
</tr>
<tr>
<td>20</td>
<td>20.02</td>
<td>19.99</td>
<td>100.01±0.60</td>
<td>19.99</td>
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<tr>
<td>50</td>
<td>49.98</td>
<td>1.02</td>
<td>99.96±1.02</td>
<td>49.98</td>
</tr>
<tr>
<td>100</td>
<td>100.0</td>
<td>1.65</td>
<td>100.10±1.65</td>
<td>99.97</td>
</tr>
<tr>
<td>150</td>
<td>150.0</td>
<td>1.98</td>
<td>99.99±1.98</td>
<td>149.97</td>
</tr>
</tbody>
</table>

*: Mean of five determinations
Table 3. Experimental values obtained in the recovery of Nicardipine hydrochloride from plasma

<table>
<thead>
<tr>
<th>Amount of standard drug, ng</th>
<th>Added</th>
<th>Recovered*</th>
<th>%CV</th>
<th>% Recovery</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>8.96±0.26</td>
<td>2.9</td>
<td>89.6±2.60</td>
<td></td>
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<tr>
<td>20</td>
<td>17.72±0.68</td>
<td>3.8</td>
<td>88.60±3.4</td>
<td></td>
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<tr>
<td>50</td>
<td>43.25±1.51</td>
<td>3.49</td>
<td>86.60±3.62</td>
<td></td>
</tr>
<tr>
<td>100</td>
<td>95.04±2.69</td>
<td>2.83</td>
<td>95.04±3.09</td>
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</tr>
</tbody>
</table>

*: Mean of five determinations

In order to find the utility of this HPLC method for the estimation of nicardipine hydrochloride in plasma in need-based situations, an immediate release capsule dosage form (dose 30 mg) was administered to six volunteers. The plasma concentration of the drug was estimated by the proposed HPLC method. There was no interference of any other plasma component with the drug and internal standard peaks. The plasma concentration of nicardipine hydrochloride in plasma samples is shown in Table 4. The sensitivity of the assay is sufficient for the determination of nicardipine hydrochloride in human plasma at 0.5 h and 12 h after administration of an immediate release capsule. The results of the study show that the proposed HPLC method is simple, rapid, sensitive, precise and accurate for the estimation of nicardipine hydrochloride in human plasma and could be used for biopharmaceutical and pharmacokinetic evaluation of drug and its formulations.

Acknowledgements

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References

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