Spectroscopic and Conductometric Analysis of Gabapentin

SARA M. ANIS, MERVAT M. HOSNY*, HISHAM E. ABDELLATEF and MOHAMED N. EL- BALKINY

Analytical Chemistry Department, Faculty of Pharmacy
Zagazig University, Zagazig, 44519, Egypt
mervat2200@yahoo.com

Received 11 December 2010; Revised 12 February 2011; Accepted 28 February 2011

Abstract: Four simple, sensitive and reproducible methods were developed for the determination of gabapentin (GPT) in pure form and in pharmaceutical preparations. Methods A and B are based on the reaction of cupric chloride with gabapentin to form stable complex, which could be measured spectrophotometrically at $\lambda_{\text{max}}$ 246 nm (method A) or by using conductometric technique (method B). While method C and D depends on the formation of ion pair complex between the studied drug and bromothymol blue, bromocresol green respectively this was extractable with methylene chloride. The concentration ranges were 40-95 $\mu$g mL$^{-1}$, 1-15 mg, 100-800 and 10-150 $\mu$g mL$^{-1}$ for methods A, B, C and D respectively .The optimization of various experimental conditions were described .The results obtained showed good recoveries, Ringbom optimum concentration ranges were calculated, in addition to molar absorptivity and sandell’s sensitivity, detection and quantification limits. The methods were successfully applied to the determination of GPT in bulk and pharmaceutical preparations. The results were favorably comparable with the official method. The molar combining ratio for methods (A-B) was found to be (2:1) (drug: reagent) while for method (C-D) it was found to be (1:1).

Keywords: Cupric chloride, Bromothymol blue, Bromocresol green, Spectrophotometrically; Conductometric technique

Introduction

Gabapentin (1- (aminomethyl)cyclo-hexaneacetic acid) is a structural analogue of g-aminobutyric acid (GABA) (Scheme 1) and its action is attributed to the irreversible inhibition of the enzyme GABA-transaminase, thus preventing the physiological degradation of GABA in the brain; a secondary mechanism of a blockade for GABA uptake is also
suggested\(^1\), it is an antiepileptic effective in the treatment of partial seizures with or without secondary generalization and is used as adjunctive therapy in patients unresponsive to or intolerant of standard antiepileptic drugs\(^2\). A survey of the literature reveals that there were few reported methods for the determination of gabapentin using spectrophotometric technique\(^3-5\), spectrofluorimetry\(^6,7\), capillary electrophoresis\(^8\), LC-MS\(^9\) and HPLC\(^10,11\).

![Scheme 1. Chemical structure of gabapentin](image)

An inspection of both available methods for the cited drug reveals that only few spectrophotometric work were done, although conductometry is a rapid method and requires simple procedures, it has not been yet applied to the determination of gabapentin. The USP 30 described a liquid chromatographic method for its determination\(^12\).

The aim of this study was to apply simple, accurate, sensitive and reproducible reactions to analyze GPT in pure form and in pharmaceutical preparations, this study described methods that can be used in laboratories where modern and expensive equipment, such as that required for GC or HPLC is not available.

**Experimental**

Absorption spectra for all measurements were carried out using Shimadzu UV-260 double beam recording spectrometer with a 1 cm cell holder. All conductometric measurements were recorded using conductometer model CM-1K, Tokyo TOA electronics Ltd Japan. The pH values of solutions were measured using a Chemocadet pH meter.

**Materials and reagents**

Analytical grade reagents and double distilled water were used to prepare all solutions. GPT pure drug was obtained from Godecke AG, Germany under license of Park-Davis. Aqueous solutions of 1 and 4 mg mL\(^{-1}\) of GPT was prepared by dissolving 100 and 400 mg of pure drug in 100 mL double distilled water respectively.

Stock solution of cupric chloride (Aldrich Chemical Co. Ltd) was prepared as 0.1% solution in double distilled water also 10\(^{-2}\) M solution was prepared by dissolving 0.171 g in 100 mL double distilled water. Bromothymol blue sodium salt (BDH Chemicals Ltd., Poole, England) was prepared as 0.05% solution in double distilled water. Bromocresol green (Aldrich Chemical Co. Ltd) 0.025% solution was prepared by dissolving the weighed amount in 2.5 mL 0.1 M NaOH then completed to 100 mL using double distilled water\(^13\).

Borate buffer pH 7.5 was used by dissolving 2.5 g of sodium chloride, 2.85 g of sodium tetraborate and 10.5 g of boric acid in sufficient water to produce 1000 mL. Adjust the pH if necessary\(^14\). Also acetate buffer of pH 3.7 was used by dissolving 10 g of anhydrous sodium acetate in 300 mL water, adjust to pH 3.7 with glacial acetic acid and dilute to 1000 mL with water. If necessary, readjust to pH 3.7 with glacial acetic acid or anhydrous sodium acetate as required, before use\(^14\).

Gaptin capsules (labeled to contain 100 mg gabapentin per capsule) were obtained from Delta Pharma, Egypt.
Spectroscopic and Conductometric Analysis of Gabapentin

**Standard Drug Solutions**

The contents of ten capsules were emptied, pulverized. An accurately weighed amount equivalent to 100 and 400 mg GPT were extracted by shaking with 50 mL distilled water, filtered, transferred to a 100 mL volumetric flask, completed to the mark using double distilled water. The general procedures were then followed using standard addition method.

**General procedures**

*Spectrophotometric procedure using cupric chloride (method A)*

Aliquots containing 0.4-0.95 mg of gabapentin drug solution were transferred into 10 mL volumetric flasks, 1 mL of borate buffer pH 7.5 was added, mixed then 2 mL of cupric chloride 0.1% solution was added, the volume was made up to 10 mL using distilled water, then the absorbance was measured at 246 nm, against a reagent blank prepared according to the same treatment.

*Conductometric procedure (method B)*

Aliquot of sample solution containing 1-15 mg of drug were transferred to a 50 mL calibrated flask, volume was made up to the mark using bi-distilled water. The contents of the calibrated flask were transferred to a beaker and the conductivity cell was immersed. 10-2 M cupric chloride was used for titration, the conductance was measured subsequent to each addition of reagent solution and after thorough stirring for two min, corrected for dilution effect, by means of the following equation, assuming that conductivity is a linear function of dilution.

\[
\Omega_{\text{correct}}^{-1} = \Omega_{\text{obs}}^{-1} \left( \frac{v_1 + v_2}{v_1} \right)
\]

Where \(\Omega_{\text{obs}}^{-1}\) is the observed electrolytic conductivity, \(v_1\) is the initial volume and \(v_2\) is the volume of reagent added. A graph of corrected conductivity versus the volume of added titrant is constructed and the end-point is determined.

*Ion pair procedure using bromothymol blue (method C)*

Aliquots containing 1-8 mg of gabapentin drug solution were transferred into 125 mL separating funnels, 1.5 mL acetate buffer pH 3.7 and 1.5 mL 0.05% bromothymol blue were added in order, mixed then the aqueous solution was extracted with an equal volume of methylene chloride and shaken for 30 sec, the mixture was allowed to separate into two phases. The organic layer was collected and dried over anhydrous sodium sulfate, completed to 10 mL with methylene chloride, the absorbance of the extract was measured at 411 nm, against a reagent blank prepared according to the same treatment.

*Ion pair procedures using bromocresol green (method D)*

Aliquots containing 0.1-1.5 mg of gabapentin drug solution were transferred into 125 mL separating funnels and then treated with about 1.5 mL of acetate buffer pH 3.7, 2.5 mL of 0.025% bromocresol green were then added and mixed, then the aqueous solution was extracted with an equal volume of methylene chloride and shaken for 30 sec, the mixture was allowed to separate into two phases. The organic layer was collected and dried with anhydrous sodium sulfate, completed to 10 mL with methylene chloride, the absorbance of the extract was measured at 411 nm, against a reagent blank prepared according to the same treatment.

**Determination of the stoichiometry of the reaction**

In order to ascertain the stoichiometry of reaction Job's method of continuous variation was carried out using the same molarity of drug and reagent.
Results and Discussion

Spectrophotometric procedures using cupric chloride (method A)

Binary complexes have been widely used in spectrophotometric analysis of many pharmaceutical compounds\textsuperscript{17-19}. In this paper, the formed binary complex consists of the studied drug gabapentin and the metal ions, copper(II). This complex is water soluble with absorption maximum at 246 nm, (Figure 1). The effects of the reagent concentrations, pH and temperature with respect to maximum sensitivity, adherence to beer’s law and stability, have been studied through control experiments. The optimum conditions were established by varying one variable at a time and observing its effect on the absorbance of colored species\textsuperscript{20}:

- 2 mL of 0.1% cupric chloride solution was found to be satisfactory for maximum absorbance and was used throughout this investigation (Figure 2).
- 1 mL of borate buffer pH 7.5 was needed to achieve best results (Figure 3).
- The reaction proceeded maximally at room temperature, no heating was required.

\begin{figure}[h]
\centering
\includegraphics[width=\textwidth]{absorption_spectra.png}
\caption{Absorption spectra of the complex formed through reaction of: 75 µg mL\textsuperscript{-1} gabapentin with cupric chloride in presence of borate buffer pH 7.5, 75 µg mL\textsuperscript{-1} gabapentin with borate buffer pH 7.5, cupric chloride with borate buffer pH 7.5.}
\end{figure}

\begin{figure}[h]
\centering
\includegraphics[width=\textwidth]{reagent_volume_effect.png}
\caption{Effect of 0.1% cupric chloride volume on the reaction between cupric chloride and 75 µg mL\textsuperscript{-1} gabapentin.}
\end{figure}
Figure 3. Effect of borate buffer pH 7.5 volume on the reaction between cupric chloride and 75 µg mL\(^{-1}\) gabapentin

Conductometric procedures using cupric chloride (method B)

Conductometric analysis can be used in many titration procedures when ionic solutions are involved. As the conductance of a solution is related to the total ionic content, it can be applied to follow reactions that results in a change in this quantity. Conductance measurements are used successfully in quantitative titration of systems in which the conductance of the solution varies before and after the equivalence point. In these cases, the titration curve can be represented by two lines intersecting at the end point\(^{21-22}\).

Investigations were carried out to establish the most favorable conditions for the ion associates formation of gabapentin with cupric chloride to achieve sharp end point. The optimum conditions for performing the titration in a quantitative manner were elucidated as described below.

Effect of solvent

Seven different titrations were attempted:
(i) Aqueous drug solution with aqueous reagent solution,
(ii) Ethanol drug solution with ethanol reagent solution,
(iii) Drug solution with reagent solution, both in ethanol–water (50%, v/v) mixture
(iv) Methanol drug solution with methanol reagent solution,
(v) Drug solution with reagent solution, both in methanol–water (50 v/v) mixture
(vi) Acetone drug solution with acetone reagent solution and
(vii) Drug solution with reagent solution, both in acetone–water (50% v/v) mixture.

Preliminary experiments showed that procedure in aqueous media was the most suitable for successful results, because in other procedures turbid solution was formed which caused some errors.

Reagent’s concentration

Different concentrations of cupric chloride solution were tried ranging from 2.5×10\(^{-3}\) to 2×10\(^{-2}\) molar solution. The optimum concentration of the reagent was 10\(^{-2}\) M in titration of the studied drug to achieve a constant and highly stable conductance reading within 1-2 min of mixing. Concentrations less or more than these limits showed only poor inflection at the end point.

Representative titration curve is shown in (Figure 4). Two straight lines are obtained, intersecting at the end-point, the first branch ascending the second one, conductance values slightly increase after the equivalence point.
The conductance measured before the addition of the titrant (volume of Cu$^{2+}$ equal zero) is related to the dissociation of the proton of the carboxylic group present in the gabapentin molecule. Up to the equivalence point, the titration involves the gradual substitution of the protons of the gabapentin molecule by cations of Cu$^{2+}$. This increase of the conductance is because the mobility of the ion H$^+$ is larger than that of ion Cu$^{2+}$, causing an increase in the slope of the conductometric curve (first branch of the curve). After the equivalence point, the measured conductance is the sum of the Cu$^{2+}$ and Cl$^-$ present in the solution. As the sum of the mobilities of those ions is smaller than that of the H$^+$ cation, there is a decrease in the slope of the second section of the titration curve. The equivalence point is defined as the point of intersection of the two straight segments.

The conductometric titrations of different volumes of 10$^{-2}$ M cupric chloride solution were performed. The results show an obvious maximum in the conductance curve at drug-reagent molar ratio of (2:1). The optimum concentration ranges for determination of gabapentin was in the range of 1-15 mg. At such range, distinct inflections and stable conductance reading were obtained.

**Ion pair procedures using bromothymol blue (method C)**

The utility of bromothymol blue as ion-pairing reagent in assay of gabapentin was investigated here. The spectra of the reaction products show characteristic $\lambda_{max}$ at 411 nm (Figure 5). The experimental conditions were established by varying one variable and observing its effect on the absorbance of the colored species as discussed below:

- 1.5 mL of 0.05% bromothymol blue was found to be sufficient for giving best results (Figure 6).
- Using different buffers of different pH in the range from (2-11), the intensity of the color of the formed complex increased when 1.5 mL of acetate buffer of pH 3.7 was used (Figure 7).
- It was found that a single extraction of the ternary complex for 30 seconds was sufficient for complete extraction.
- Methylene chloride was found to be the most convenient solvent for the studied drug.
Figure 5. Absorption spectra of the ion pair formed through reaction of: 800 µg mL\(^{-1}\) gabapentin with 0.05% bromothymol blue blank solution

Figure 6. Effect of volume of 0.05% bromothymol blue on the reaction of bromothymol blue with 800 µg mL\(^{-1}\) gabapentin

Figure 7. Effect of volume of acetate buffer pH 3.7 on the reaction of bromothymol blue with 800 µg mL\(^{-1}\) gabapentin
Ion pair procedure using bromocresol green (method D)

The utility of bromocresol green as ion-pairing reagent in assay of gabapentin was investigated here. The spectra of the reaction products show characteristic \( \lambda_{\text{max}} \) at 411 nm (Figure 8). The experimental conditions for the reaction between bromocresol green and gabapentin are discussed below:

- 2.5 mL of 0.025% bromocresol green was found to be sufficient to give maximum absorbance (Figure 9).
- Trying different buffer systems of different pH values, the intensity of the color of the formed complex increased when 1.5 mL of acetate buffer of pH 3.7 was used (Figure 10).
- It was found that a single extraction of the ternary complex for 30 seconds was sufficient for complete extraction of the formed complex.
- Methylene chloride was found to be the most convenient solvent for the studied drug.

![Figure 8](image1)

**Figure 8.** Absorption spectra of the ion pair formed through reaction of 100 µg mL\(^{-1}\) gabapentin with 0.025% bromocresol green blank solution

![Figure 9](image2)

**Figure 9.** Effect of volume of 0.025% bromocresol green on the reaction of bromocresol green with 50 µg mL\(^{-1}\) gabapentin
**Figure 10.** Effect of acetate buffer pH 3.7 volume on the reaction of bromocresol green with 50 µg mL\(^{-1}\) gabapentin

**Stoichiometric relationship**

Using Job’s method of continuous variation, the molar ratio of gabapentin to cupric chloride was found to be 2:1, while for bromothymol blue and bromocresol green it was found to be 1:1 (Figure 11-13). The mechanisms of the methods are suggested in Scheme 2.

**Figure 11.** Determination of the stoichiometry of the reaction of: Gabapentin (5\(\times\)10\(^{-3}\) M) and cupric chloride (5\(\times\)10\(^{-3}\) M)

**Figure 12.** Determination of the stoichiometry of the reaction of: Gabapentin (1.25\(\times\)10\(^{-3}\) M) and bromothymol blue (1.25\(\times\)10\(^{-3}\) M)
Figure 13. Determination of the stoichiometry of the reaction of gabapentin (1.25x10^{-3} M) and bromocresol green (1.25x10^{-3} M)

Scheme 2. Proposed reactions of gabapentin with A) Cupric chloride B) Bromothymol blue C) Bromocresol Green

Methods of validation
The developed analytical methods were validated as per ICH (The International Conference on Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use), guideline and USP (United States Pharmacopeia) requirement^{23-24}, applying a pharmaceutical preparation analysis. Under the described experimental conditions, calibration graphs were constructed for all of the studied methods, as can be seen from (Table 1), linear relationship was found between the absorbance at λ_{max} and the concentration
Spectrscopic and Conductometric Analysis of Gabapentin

of the drug in the ranges of 40-95 µg mL⁻¹, 100-800 and 10-150 µg mL⁻¹ for methods A, C and D respectively. It was observed that method D is the most sensitive one. The values of beer's law limits, ringbom concentrations ranges, regression equations, correlation coefficients, molar absorptivity, sandell's sensitivity, limit of detection (LOD) and limit of quantification (LOQ) for each method were all summarized in (Table 1). The optimum concentration ranges of gabapentin that can be measured accurately as evaluated from the Ringbom plot were 56.2-85.11, 199.5-707.9, 19.95-85.11 µg mL⁻¹ for methods A, C and D respectively. The standard deviations, relative standard deviations, standard errors, variances were all listed in (Table 2), the average percent recoveries as can be seen from the same table indicates good accuracy of the methods.

**Table 1.** Spectral characteristics and statistical data of the regression equations for the product formed through reactions (A, C and D)

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Method A</th>
<th>Method C</th>
<th>Method D</th>
</tr>
</thead>
<tbody>
<tr>
<td>Linearity range, µg mL⁻¹</td>
<td>40-95</td>
<td>100-800</td>
<td>10-150</td>
</tr>
<tr>
<td>Ringbom range, µg mL⁻¹</td>
<td>56.2-85.11</td>
<td>199.5-707.9</td>
<td>19.95-85.11</td>
</tr>
<tr>
<td>Molar absorptivity, mol⁻¹ cm⁻³</td>
<td>1.33x10⁴</td>
<td>1.99x10²</td>
<td>1.54x10³</td>
</tr>
<tr>
<td>Sandell's sensitivity, µg mL⁻¹ per 0.001 A</td>
<td>7.76x10⁻³</td>
<td>1.17x10⁻⁴</td>
<td>8.98 x 10⁻⁴</td>
</tr>
<tr>
<td>Regression equation:</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Intercept (a)</td>
<td>-0.549</td>
<td>0.076</td>
<td>0.175</td>
</tr>
<tr>
<td>Slope (b)</td>
<td>0.016</td>
<td>0.0092</td>
<td>0.0053</td>
</tr>
<tr>
<td>Correlation coefficient (r)</td>
<td>0.9995</td>
<td>0.9998</td>
<td>0.9999</td>
</tr>
<tr>
<td>Variance</td>
<td>1.09</td>
<td>1.38</td>
<td>1.71</td>
</tr>
<tr>
<td>Limit of detection LOD, µg mL⁻¹</td>
<td>1.179</td>
<td>1.44</td>
<td>1.61</td>
</tr>
<tr>
<td>Limit of quantitation LOQ, µg mL⁻¹</td>
<td>3.89</td>
<td>4.75</td>
<td>5.30</td>
</tr>
</tbody>
</table>

*calculated on the basis of the molecular weight of the drug

**Table 2.** Determination of gabapentin by using methods (A-D)

<table>
<thead>
<tr>
<th>Method</th>
<th>Taken, µg mL⁻¹</th>
<th>Found, µg mL⁻¹</th>
<th>Recovery %</th>
<th>Taken, mg</th>
<th>Found, mg</th>
<th>Recovery %</th>
<th>Taken, µg mL⁻¹</th>
<th>Found, µg mL⁻¹</th>
<th>Recovery %</th>
<th>Taken, µg mL⁻¹</th>
<th>Found, µg mL⁻¹</th>
<th>Recovery %</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>40</td>
<td>40.25</td>
<td>100.6</td>
<td>1</td>
<td>0.99</td>
<td>99.31</td>
<td>100</td>
<td>98.91</td>
<td>98.91</td>
<td>10</td>
<td>9.81</td>
<td>98.11</td>
</tr>
<tr>
<td>B</td>
<td>50</td>
<td>49.00</td>
<td>3</td>
<td>3</td>
<td>3.01</td>
<td>100.47</td>
<td>200</td>
<td>201.09</td>
<td>100.54</td>
<td>25</td>
<td>24.91</td>
<td>99.62</td>
</tr>
<tr>
<td>C</td>
<td>60</td>
<td>58.88</td>
<td>3</td>
<td>5</td>
<td>5.00</td>
<td>100.00</td>
<td>300</td>
<td>303.26</td>
<td>101.09</td>
<td>50</td>
<td>50.00</td>
<td>100.00</td>
</tr>
<tr>
<td>D</td>
<td>75</td>
<td>74.69</td>
<td>7</td>
<td>7</td>
<td>7.02</td>
<td>100.29</td>
<td>400</td>
<td>405.43</td>
<td>101.36</td>
<td>75</td>
<td>76.42</td>
<td>101.89</td>
</tr>
<tr>
<td></td>
<td>85</td>
<td>83.13</td>
<td>9</td>
<td>8.99</td>
<td>99.89</td>
<td>98.48</td>
<td>500</td>
<td>492.39</td>
<td>98.48</td>
<td>100</td>
<td>101.13</td>
<td>101.13</td>
</tr>
<tr>
<td></td>
<td>90</td>
<td>89.25</td>
<td>11</td>
<td>10.96</td>
<td>99.63</td>
<td>100.41</td>
<td>800</td>
<td>803.26</td>
<td>101.41</td>
<td>150</td>
<td>149.62</td>
<td>99.75</td>
</tr>
<tr>
<td></td>
<td>95</td>
<td>94.56</td>
<td>13</td>
<td>13.01</td>
<td>100.08</td>
<td>15.07</td>
<td>99.54</td>
<td>100.47</td>
<td>98.98</td>
<td>15.07</td>
<td>15.07</td>
<td>100.02</td>
</tr>
</tbody>
</table>

Mean* 98.98 100.02 100.13 100.08

\( p = 0.05 \)

N   7          8          6          6
S.D. 1.04       0.405      1.173      1.309
R.S.D. 1.05      0.404      1.172      1.308
V   1.09       0.164      1.38       1.71
S.E. 0.394     0.143      0.479      0.534

*Mean of three different experiments
The proposed methods were applied for determination of the selected drug in its capsule form (Table 3). Experiments showed that there was no interference from the additives e.g. lactose, fructose, magnesium stearate and starch. The methods performance was assessed using the t-test (for accuracy) and a variance ratio F-values, both values did not exceed the theoretical values (95% confidence limit), so we concluded that the proposed methods do not differ significantly from the official one, (Table 4).

**Table 3.** Determination of gaptin capsule by using methods (A-D)

<table>
<thead>
<tr>
<th></th>
<th>Method A</th>
<th>Method B</th>
<th>Method C</th>
<th>Method D</th>
</tr>
</thead>
<tbody>
<tr>
<td>Taken,</td>
<td>µg mL⁻¹</td>
<td>Found,</td>
<td>Recovery</td>
<td>Taken,</td>
</tr>
<tr>
<td>µg mL⁻¹ Added</td>
<td></td>
<td>µg mL⁻¹</td>
<td>%</td>
<td>µg mL⁻¹</td>
</tr>
<tr>
<td>40</td>
<td>40.56</td>
<td>101.41</td>
<td>99.30</td>
<td>100</td>
</tr>
<tr>
<td>60</td>
<td>59.44</td>
<td>99.06</td>
<td>8</td>
<td>80</td>
</tr>
<tr>
<td>80</td>
<td>66.19</td>
<td>98.72</td>
<td>100.13</td>
<td>600</td>
</tr>
<tr>
<td>65</td>
<td>70.56</td>
<td>100.18</td>
<td>9.99</td>
<td>600</td>
</tr>
<tr>
<td>Mean</td>
<td>100.16</td>
<td>98.72</td>
<td>99.86</td>
<td>100.75</td>
</tr>
<tr>
<td>(p = 0.05)</td>
<td>N</td>
<td>5</td>
<td>5</td>
<td></td>
</tr>
<tr>
<td></td>
<td>S.D.</td>
<td>1.18</td>
<td>0.706</td>
<td></td>
</tr>
<tr>
<td></td>
<td>V</td>
<td>1.39</td>
<td>0.499</td>
<td></td>
</tr>
<tr>
<td></td>
<td>S.E.</td>
<td>0.529</td>
<td>0.353</td>
<td></td>
</tr>
</tbody>
</table>

*Mean of three different experiments

**Table 4.** Statistical data for the determination gabapentin using method (A-D) compared with reference method.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Reference method</th>
<th>Method A</th>
<th>Method B</th>
<th>Method C</th>
<th>Method D</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean (p = 0.05)</td>
<td>99.59</td>
<td>98.98</td>
<td>100.02</td>
<td>100.13</td>
<td>100.08</td>
</tr>
<tr>
<td>N</td>
<td>5</td>
<td>7</td>
<td>8</td>
<td>6</td>
<td>6</td>
</tr>
<tr>
<td>S.D.</td>
<td>0.650</td>
<td>1.04</td>
<td>0.405</td>
<td>1.173</td>
<td>1.309</td>
</tr>
<tr>
<td>R.S.D</td>
<td>0.653</td>
<td>1.05</td>
<td>0.404</td>
<td>1.172</td>
<td>1.308</td>
</tr>
<tr>
<td>V</td>
<td>0.423</td>
<td>1.09</td>
<td>0.164</td>
<td>1.38</td>
<td>1.71</td>
</tr>
<tr>
<td>t</td>
<td>-</td>
<td>1.15 (2.228)</td>
<td>1.48 (2.201)</td>
<td>0.913 (2.262)</td>
<td>0.758 (2.262)</td>
</tr>
<tr>
<td>F</td>
<td>-</td>
<td>2.58 (4.53)</td>
<td>2.58 (4.12)</td>
<td>3.26 (5.19)</td>
<td>4.04 (5.19)</td>
</tr>
</tbody>
</table>

*Theoretical values of t and F at p = 0.05

**Conclusion**

The data given above reveals that the proposed methods introduce new techniques for the determination of gabapentin. The studied methods showed the advantage of being simple, accurate and sensitive with good precision and accuracy. Using of bromocresol green allowed the detection of gabapentin in small amount of 10 µg mL⁻¹. With these methods, one can do the analysis in a short time at low cost without losing accuracy. The proposed methods can be used as alternative methods to reported ones for the routine determination of gabapentin in the pure form and in pharmaceutical formulations.
References
