

Supplemental Table

Supplemental Table 1. Toxicity of NPs to different organisms.

NPs	Advertised particle size (nm)	TEM/SEM measured size (nm)	DLS (nm)	Zeta potential (mV)	Test species	Effects	Reference
	25				green algae <i>D. subspicatus</i>	$EC_{50} = 44 \text{ mg/L}$ EC_{50} was beyond the tested range, > 50 mg/L	1
	100	N/A	N/A	N/A			
	25-70	N/A	N/A	N/A	algae <i>P. subcapitata</i>	$EC_{50} = 5.83 \text{ mg Ti/L}$	2
	25-70	N/A	N/A	N/A	<i>S. cerevisiae</i> S288C	not toxic even at 20000 mg/L	3
	N/A	17-64	N/A	N/A	<i>E. coli</i>	$LC_{50} (\text{light}) = 0.16 \text{ mg /L}$ $LC_{50} (\text{dark}) = 4.63 \text{ mg /L}$	4
TiO ₂	27.5	14.2-64.6	N/A	N/A	human lung epithelial cells	not cytotoxic at any concentration up to 100 µg/mL	5
	N/A	91	N/A	+ 48.8	in human peripheral blood lymphocytes4 or BEAS-2B/ IMR-90 cells	no significant induction of DNA damage based on the comet assay	6
	25	N/A	371±323	-18 ± 4.5	<i>S. oneidensis</i>	no significant change in cell survival with increasing concentration	7
	< 25	N/A	185 ± 40	N/A	waste activated sludge	doses up to no inhibitory effect at 150 mg/ g-TSS	8
	50	N/A	N/A	-21	<i>B. subtilis</i> , <i>E. coli</i> and <i>P. fluorescens</i>	did not affect bacterial populations	9
	66	N/A	330	N/A	<i>B. subtilis</i> and	resulting in 75% growth reduction with 1000 ppm	10

				<i>E. coli</i>	resulting in 72% growth reduction with 5000 ppm	
				<i>Vibrio fischeri</i>	$EC_{50} > 20000$ mg/L	
25-70	N/A	N/A	N/A	<i>Daphnia magna</i>	$LC_{50} \sim 20000$ mg/L	11
				<i>Thamnocephalus platyurus</i>	$LC_{50} > 20000$ mg/L	
25-70	N/A	N/A	N/A	<i>Pseudokirchneriella subcapitata</i>	$EC_{50} = 5.83$ mg Ti/L	2
50-70	N/A	N/A	N/A	algae <i>P. subcapitata</i>	$EC_{50} \sim 0.04$ mg Zn/L	2
				<i>E. coli</i>	$EC_{50} = 115.7$ mg/L	
50-70	N/A	N/A	N/A	<i>B. subtilis</i>	$EC_{50} = 85.8$ mg/L	12
				<i>S. aureus</i>	$EC_{50} > 125$ mg/L	
13	N/A	N/A	N/A	<i>E. coli</i>	complete inhibition of <i>E. coli</i> growth at concentrations > 3.4 mM	13
				<i>S. aureus</i>	growth of <i>S. aureus</i> was completely inhibited at concentrations > 1 mM	
ZnO	100	N/A	N/A	Mouse macrophage (Ana-1) cell line	$IC_{50} = 35.26$ mg/L	14
30	N/A	N/A	N/A	Mouse macrophage (Ana-2) cell line	$IC_{50} = 32.43$ mg/L	13
10-30	N/A	N/A	N/A	Mouse macrophage (Ana-3) cell line	$IC_{50} = 24.84$ mg/L	13
50-70	N/A	N/A	N/A	<i>S. cerevisiae</i> S288C	$EC_{50} = 121-134$ mg ZnO/L	12
150	N/A	N/A	N/A	<i>S. agalactiae</i>	0.12 M caused more than 95% inhibition of bacterial growth, 1.2×10^{-3} M, a slight decrease in the number of bacteria colonies was detected	15

					6×10^{-4} M, a slight decrease in the number of bacteria colonies was detected	
N/A	19 ± 7	N/A	-2.9 ± 0.3	<i>E. coli</i>	the bacterial mortality at ZnO NPs concentration of 50 mg/L are 40% in Tap water, 20% in Xixi River and West Lake, and 10% in Qiantang River	16
N/A	47-106	N/A	N/A	<i>E. coli</i>	LC_{50} (light) = 0.048 mg/L LC_{50} (dark) = 0.13 mg/L	4
25	N/A	N/A	N/A	human myeloblastic leukemia cells (HL60)	CC_{50} = 52.80 µg/mL,	17
42	N/A	111±44	- 6.14	normal peripheral blood mononuclear cells (PBMCs)	CC_{50} = 741.82 µg/mL	18
<100	N/A	140 ± 20	N/A	human umbilical vein endothelial cells (HUVECs)	decreased the cellular viability to about 40% of the control at 240 µM	8
20	N/A	N/A	-5	waste activated sludge	inhibitory effect with its dosages increased	9
67	N/A	480	N/A	<i>B. subtilis</i> , <i>E. coli</i> and <i>P. fluorescens</i>	100% mortality	10
				<i>B. subtilis</i>	90% growth reduction at 10 ppm	
				<i>E. coli</i>	48% growth reduction at 1000 ppm	
				<i>Vibrio fischeri</i>	EC_{50} = 1.9 ± 0.2 mg/L	
50-70	N/A	N/A	N/A	<i>Daphnia magna</i>	LC_{50} = 3.2 ± 1.3 mg/L	11
				<i>Thamnocephalus platyurus</i>	LC_{50} = 0.18 ± 0.03 mg/L	
50-70	N/A	N/A	N/A	<i>Pseudokirchneriella subcapitata</i>	EC_{50} ~ 0.04 mg Zn/L	2

20	20.3 ± 1.9	27.0 ± 10.8	- 47.1 ± 1.9	RAW 264.7 L929 cells	EC ₂₀ = 7 mg/L EC ₂₀ = 2.8 mg/L	
80	79.8 ± 5.1	79 ± 25.2	- 49.7 ± 1.8	RAW 264.7 L929 cells	EC ₂₀ = 38 mg/L EC ₂₀ = 17 mg/L	19
110	112.6 ± 7.8	111.6 ± 29.9	- 54.5 ± 1.8	RAW 264.7 L929 cells	EC ₂₀ = 100 mg/L EC ₂₀ > 12 mg/L	
40	46 ± 21	N/A	-13.6	human mesenchymal stem cells (hMSCs)	DNA damage after 1, 3, and 24 h at 0.1 mg/L	20
15						
100	N/A	N/A	N/A	rat liver derived cell line (BRL 3A)	EC ₅₀ = 24 ± 7.25 mg/L EC ₅₀ = 19 ± 5.2 mg/L	21
less than 10 Ag	N/A	N/A	N/A	human hepatoma cells	IC ₅₀ = 3.38 ± 0.55 µg/mL	22
N/A	50 ± 20	85	-30	Human mesenchymal stem cells	57% growth reduction for 20 µg/mL, and 100% growth reduction for 25 µg/mL	23
9-21	N/A	N/A	N/A	Nitrifying Bacteria	EC ₅₀ = 0.14 mg/L	24
7-10	N/A	N/A	N/A	human hepatoma cell line, HepG2	cytotoxicity at higher doses (>1.0 mg/L)	25
10-20	N/A	N/A	N/A	ammonia-oxidizing bacteria	ammoxidation inhibition by nano-Ag was dependant on the level of concentration	26
N/A	57 ± 20	143 ± 9	-44.7 ± 1.6	Natural Estuarine Plankton Community	the growth rates of both phytoplankton and bacterioplankton populations were significantly reduced by Ag NPs at concentrations of ≥ 500 µg/L	27
25	N/A	76.65±5.73	- 4.76±0.19	HEK293T cell	IC ₅₀ = 30 µg/mL	28

N/A	54.3±5.3	101.9±3.3	- 9.9±0.4	the HaCaT cell	$LD_{50} = 54.3 \mu\text{g/mL}$	29
100			-37.3	the human LoVo cell line	the 100 nm nanoparticles exerted indirect effects via serine/threonine protein kinase (PAK), mitogen-activated protein kinase (MAPK), and phosphatase 2A pathways,	30
20			-57.5		the 20 nm nanoparticles induced direct effects on cellular stress, including generation of reactive oxygen species and protein carbonylation.	
10					10 nm AgNPs induced more apoptotic	
50	N/A	N/A	N/A	MC3T3-E1 cells	cells than the larger particles (i.e., 50 and 100 nm).	31
100						
30	N/A	N/A	N/A	algae <i>P. subcapitata</i>	$EC_{50} = 0.71 \text{ mg Cu/L}$	2
20-30	N/A	N/A	N/A	<i>E. coli</i>	$EC_{50} = 28.6 \text{ mg/L}$	
				<i>B. subtilis</i>	$EC_{50} = 61.1 \text{ mg/L}$	12
				<i>S. aureus</i>	$EC_{50} = 65.9 \text{ mg/L}$	
30	N/A	N/A	N/A	<i>S. cerevisiae</i> S288C	$EC_{50} = 13.4 \text{ mg/L}$	3
CuO					the formation of superoxide anions, hydrogen peroxide and single-stranded DNA already at very low sub-toxic levels (0.1 mg Cu/L)	
30	N/A	192.5	N/A	<i>E. coli</i> biosensor		32
N/A	92 ± 12	N/A	N/A	activated sludge	The sludge flocs were unstable after exposure to CuO NPs (50 mg/L)	33
N/A	17-45	N/A	N/A	<i>E. coli</i>	$LC_{50} (\text{light}) = 1.68 \text{ mg/L}$ $LC_{50} (\text{dark}) = 58.3 \text{ mg/L}$	4

					<i>Yeast S. cerevisiae</i> BY4741	$IC_{50} = 4.8 \text{ mg/L}$ in DI $IC_{50} = 643 \text{ mg/L}$ in YPD	
30	N/A	194 ± 16.9	31.9 ± 7.31		<i>Vibrio fischeri</i>	$EC_{50} = 79 \pm 27 \text{ mg/L}$	34
30	N/A	N/A	N/A		<i>Daphnia magna</i> <i>Thamnocephalus platyurus</i>	$LC_{50} = 3.2 \pm 1.6 \text{ mg/L}$ $LC_{50} = 2.1 \pm 0.5 \text{ mg/L}$	11
30	N/A	N/A	N/A		<i>Pseudokirchneriella subcapitata</i>	$EC_{50} = 0.71 \text{ mg Cu/L}$	2
					<i>E. coli</i>	$EC_{50} = 160.2 \text{ mg/L}$	
NiO	10 - 20	N/A	N/A	N/A	<i>B. subtilis</i>	$EC_{50} = 121.9 \text{ mg/L}$	12
					<i>S. aureus</i>	$EC_{50} = 121.1 \text{ mg/L}$	
					<i>E. coli</i> Bacillus subtilis, and Streptococcus aureus	$EC_{50} = 265.5, 144.7, 324 \text{ mg L}^{-1}$	12
Sb ₂ O ₃	90 - 210	N/A	N/A	N/A			
					rat liver derived cell line (BRL 3A)	$EC_{50} = 174.68 \pm 26 \text{ mg/L}$	21
	150	N/A	N/A	N/A		$EC_{50} = 171.58 \pm 25 \text{ mg/L}$	
	30						
MoO ₃		400nm long, nm wide			iMCF-7	$IC_{50} = 275 \mu\text{g/mL}$	35
	N/A	100 - 200	N/A	N/A			
	7	N/A	15	N/A		DNA damage	36
	N/A	20 ± 3	N/A	N/A		cell membrane damage	37
CeO ₂						a reduction of 15 - 19% for the flocculent anaerobic granule sludge at the dosage of 5, 50 and 150 mg CeO ₂ NPs/g-VSS	
less than 25	N/A	N/A	N/A			CeO ₂ NPs/g-VSS	38
					flocculent sludge	a reduction of 35% for the granular sludge at 150 mg CeO ₂ NPs/g-VSS.	

					freshwater alga <i>(P. subcapitata)</i>		
N/A	10 - 20	N/A	N/A			$EC_{50} = 10.3 \pm 1.7 \text{ mg/L}$	39
7	N/A	N/A	N/A		<i>E. coli</i>	lethal	40
33	N/A	125 ± 38	– 6.64		human umbilical vein endothelial cells (HUVECs)	CeO ₂ NPs did not show toxic effects even with doubling of tested dosage to 480 μM	18
Au	10 - 20	N/A	N/A	N/A	ammonia-oxidizing bacteria	no significant effect on ammonoxidation	26
Gold nanorods	N/A	18 × 40	N/A	N/A	in HeLa cells	less toxicity	41
Co ₃ O ₄	N/A	51-132	N/A	N/A	<i>E. coli</i>	$LC_{50} (\text{light}) = 35.06 \text{ mg/L}$ $LC_{50} (\text{dark}) = 55.56 \text{ mg/L}$	4
graphene oxide (GO)	N/A	The lateral length and thickness of GO were approximat ely 0.5–5 μm and 0.8–1.2 nm	295 - 825			did not show significant differences compared with the control at 10 mg/L GO	
carboxyl single-wall ed carbon nanotube s (C-SWCNT)	N/A	The outer diameter, inner diameter and length of C-SWCNT were approximat ely 1–2 nm, 0.8–1.6, and 0.5–3 μm,	396 - 712	N/A	<i>Chlorella vulgaris</i>	with 0.8–28.3% inhibition at 97 h	42

					an average		
SWCNT	N/A	outer diameter of lengths in the range of 10 to 20 μm	N/A	N/A	<i>E. coli</i> cell growth and biofilm formation	biofilms in aquatic systems can potentially recover from the toxic effects of SWNTs after long exposure times (48 h)	43
Silica	N/A	>150 nm long	N/A	N/A	RAW 264.7, a macrophage-like cell line.	single-walled carbon nanotubes, and amorphous silica resulted in a lower toxicity	44
SiO_2	N/A	10	27.2	-37.2	in A549 (a human lung cell line)	no differences in the induction of pulmonary inflammation and lactate dehydrogenase	45
porous SiO_2	N/A	25 \pm 4	N/A	110 \pm 40	waste activated sludge	do not influence mast cell viability	8
nonporous SiO_2	N/A	11 \pm 5	N/A	-16.94	Immune Cells	cell viability drops significantly to 72%	46
SiO_2	20	N/A	N/A	+35	<i>B. subtilis</i>	killed 40% of <i>B. subtilis</i>	9
Fe_3O_4	30	N/A	N/A	205	<i>E. coli</i>	least toxic	10
Al	70	N/A	N/A	N/A	rat liver derived cell line (BRL 3A)	$\text{EC}_{50} > 250 \text{ mg/L}$	21
Al_2O_3	103	N/A	N/A	+ 30	<i>B. subtilis</i>	$\text{EC}_{50} > 250 \text{ mg/L}$	21
					a mortality rate of 57% to <i>B. subtilis</i>		9

					<i>E. coli</i>	a mortality rate of 36% to <i>E. coli</i>	
					<i>P. fluorescens</i>	a mortality rate of 70% to <i>P. fluorescens</i>	
< 50	N/A	130 ± 30	N/A	waste activated sludge		no inhibitory effect	8
		62-65 nm					
N/A	10-70	and 78-88 nm	N/A	<i>B. licheniformis</i>	17% decrease in cell viability at 1µg/mL		48
Fe_2O_3	29	30-60	1580	-17.3	the A549 Cell Line	low toxicity was observed	47
Fe_3O_4	29-30	20-40	< 200	1.8	the A549 Cell Line	no toxicity was observed	47
carbon	< 30	20-40	210	6.9	the A549 Cell Line	no toxicity was observed	47
carbon nanotubes	110-170× (5-9×10 ³)	100-200× (3-7×10 ³)	300× (69×10 ³)	-45.4	the A549 Cell Line	a nonsignificant increase in nonviable cells	47

References

- 1 K. Hund-Rinke and M. Simon, *Environmental Science and Pollution Research*, 2006, **13**, 225-232.
- 2 V. Aruoja, H. C. Dubourguier, K. Kasemets and A. Kahru, *Science of the total environment*, 2009, **407**, 1461-1468.
- 3 K. Kasemets, A. Ivask, H. C. Dubourguier and A. Kahru, *Toxicology in vitro*, 2009, **23**, 1116-1122.
- 4 T. P. Dasari, K. Pathakoti and H.-M. Hwang, *Journal of Environmental Sciences*, 2013, **25**, 882-888.
- 5 R. Y. Prasad, K. Wallace, K. M. Daniel, A. H. Tennant, R. M. Zucker, J. Strickland, K. Dreher, A. D. Kligerman, C. F. Blackman and D. M. DeMarini, *ACS nano*, 2013, **7**, 1929-1942.

- 6 K. Bhattacharya, M. Davoren, J. Boertz, R. P. Schins, E. Hoffmann and E. Dopp, *Particle and Fibre Toxicology*, 2009, **6**, 1.
- 7 M. A. Maurer-Jones, I. L. Gunsolus, B. M. Meyer, C. J. Christenson and C. L. Haynes, *Analytical chemistry*, 2013, **85**, 5810-5818.
- 8 H. Mu, Y. Chen and N. Xiao, *Bioresource technology*, 2011, **102**, 10305-10311.
- 9 W. Jiang, H. Mashayekhi and B. Xing, *Environmental pollution*, 2009, **157**, 1619-1625.
- 10 L. K. Adams, D. Y. Lyon and P. J. Alvarez, *Water research*, 2006, **40**, 3527-3532.
- 11 M. Heinlaan, A. Ivask, I. Blinova, H. C. Dubourguier and A. Kahru, *Chemosphere*, 2008, **71**, 1308-1316.
- 12 Y. W. Baek and Y. J. An, *Science of the Total Environment*, 2011, **409**, 1603-1608.
- 13 K. M. Reddy, K. Feris, J. Bell, D. G. Wingett, C. Hanley and A. Punnoose, *Applied physics letters*, 2007, **90**, 213902.
- 14 W. Song, J. Zhang, J. Guo, J. Zhang, F. Ding, L. Li and Z. Sun, *Toxicology letters*, 2010, **199**, 389-397.
- 15 Z. Huang, X. Zheng, D. Yan, G. Yin, X. Liao, Y. Kang, Y. Yao, D. Huang and B. Hao, *Langmuir*, 2008, **24**, 4140-4144.
- 16 M. Li, D. Lin and L. Zhu, *Environmental pollution*, 2013, **173**, 97-102.
- 17 M. Premanathan, K. Karthikeyan, K. Jeyasubramanian and G. Manivannan, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2011, **7**, 184-192.
- 18 R. Chen, L. Huo, X. Shi, R. Bai, Z. Zhang, Y. Zhao, Y. Chang and C. Chen, *ACS nano*, 2014, **8**, 2562-2574.
- 19 M. V. Park, A. M. Neigh, J. P. Vermeulen, L. J. de la Fonteyne, H. W. Verharen, J. J. Briedé, H. van Loveren and W. H. de Jong, *Biomaterials*, 2011, **32**, 9810-9817.
- 20 S. Hackenberg, A. Scherzed, M. Kessler, S. Hummel, A. Technau, K. Froelich, C. Ginzkey, C. Koehler, R. Hagen and N. Kleinsasser, *Toxicology letters*, 2011, **201**, 27-33.
- 21 S. Hussain, K. Hess, J. Gearhart, K. Geiss and J. Schlager, *Toxicology in vitro*, 2005, **19**, 975-983.
- 22 S. Kim, J. E. Choi, J. Choi, K. H. Chung, K. Park, J. Yi and D. Y. Ryu, *Toxicology in vitro*, 2009, **23**, 1076-1084.

- 23 S. Kittler, C. Greulich, J. Diendorf, M. Koller and M. Epple, *Chemistry of Materials*, 2010, 22, 4548-4554.O. Choi and Z. Hu, *Environmental science & technology*, 2008, **42**, 4583-4588.
- 24 O. Choi and Z. Hu, *Environmental science & technology*, 2008, **42**, 4583-4588.
- 25 K. Kawata, M. Osawa and S. Okabe, *Environmental science & technology*, 2009, **43**, 6046-6051.
- 26 Z. Luo, Z. Chen, Z. Qiu, Y. Li, G. Du Laing, A. Liu and C. Yan, *Chemosphere*, 2015, **120**, 737-742.
- 27 M. S. Baptista, R. J. Miller, E. R. Halewood, S. K. Hanna, C. M. R. Almeida, V. M. Vasconcelos, A. A. Keller and H. S. Lenihan, *Environmental science & technology*, 2015, **49**, 12968-12974.
- 28 Y. Chen, Z. Wang, M. Xu, X. Wang, R. Liu, Q. Liu, Z. Zhang, T. Xia, J. Zhao and G. Jiang, *ACS nano*, 2014, **8**, 5813-5825.
- 29 K. K. Comfort, L. K. Braydich-Stolle, E. I. Maurer and S. M. Hussain, *ACS nano*, 2014, **8**, 3260-3271.
- 30 T. Verano-Braga, R. Miethling-Graff, K. Wojdyla, A. Rogowska-Wrzesinska, J. R. Brewer, H. Erdmann and F. Kjeldsen, *ACS nano*, 2014, **8**, 2161-2175.
- 31 T. H. Kim, M. Kim, H. S. Park, U. S. Shin, M. S. Gong and H. W. Kim, *Journal of Biomedical Materials Research Part A*, 2012, **100**, 1033-1043.
- 32 O. Bondarenko, A. Ivask, A. Käkinen and A. Kahru, *Environmental pollution*, 2012, **169**, 81-89.
- 33 J. Hou, L. Miao, C. Wang, P. Wang, Y. Ao and B. Lv, *Bioresource technology*, 2015, **176**, 65-70.
- 34 K. Kasemets, S. Suppi, K. Künnis-Beres and A. Kahru, *Chemical research in toxicology*, 2013, **26**, 356-367.
- 35 T. Anh Tran, K. Krishnamoorthy, Y. W. Song, S. K. Cho and S. J. Kim, *ACS applied materials & interfaces*, 2014, **6**, 2980-2986.
- 36 M. Auffan, J. Rose, T. Orsiere, M. De Meo, A. Thill, O. Zeyons, O. Proux, A. Masion, P. Chaurand and O. Spalla, *Nanotoxicology*, 2009, **3**, 161-171.
- 37 W. Lin, Y. w. Huang, X. D. Zhou and Y. Ma, *International journal of toxicology*, 2006, **25**, 451-457.

- 38 J. Ma, X. Quan, X. Si and Y. Wu, *Bioresource technology*, 2013, **149**, 346-352.
- 39 N. J. Rogers, N. M. Franklin, S. C. Apte, G. E. Batley, B. M. Angel, J. R. Lead and M. Baalousha, *Environmental Chemistry*, 2010, **7**, 50-60.
- 40 A. Thill, O. Zeyons, O. Spalla, F. Chauvat, J. Rose, M. Auffan and A. M. Flank, *Environmental science & technology*, 2006, **40**, 6151-6156.
- 41 T. S. Hauck, A. A. Ghazani and W. C. Chan, *Small*, 2008, **4**, 153-159.
- 42 X. Hu, S. Ouyang, L. Mu, J. An and Q. Zhou, *Environmental science & technology*, 2015, **49**, 10825-10833.
- 43 D. F. Rodrigues and M. Elimelech, *Environmental science & technology*, 2010, **44**, 4583-4589.
- 44 D. Dutta, S. K. Sundaram, J. G. Teeguarden, B. J. Riley, L. S. Fifield, J. M. Jacobs, S. R. Addleman, G. A. Kaysen, B. M. Moudgil and T. J. Weber, *Toxicological Sciences*, 2007, **100**, 303-315.
- 45 T. M. Sager, D. W. Porter, V. A. Robinson, W. G. Lindsley, D. E. Schwegler-Berry and V. Castranova, *Nanotoxicology*, 2007, **1**, 118-129.
- 46 M. A. Maurer-Jones, Y. S. Lin and C. L. Haynes, *ACS nano*, 2010, **4**, 3363-3373.
- 47 H. L. Karlsson, P. Cronholm, J. Gustafsson and L. Moller, *Chemical research in toxicology*, 2008, **21**, 1726-1732.
- 48 S. Pakrashi, S. Dalai, D. Sabat, S. Singh, N. Chandrasekaran and A. Mukherjee, *Chemical research in toxicology*, 2011, **24**, 1899-1904.