

Research Article

Physicochemical Characterization and Cyclodextrin Complexation of the Anticancer Drug Lapatinib

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Lapatinib (LAP), the tyrosine kinase inhibitor drug with moderate bioavailability, was characterized in terms of physicochemical properties: acid-base characteristics, lipophilicity, and solubility. The highly lipophilic nature of the drug and its extremely low water solubility ($S_0 = 0.82 \text{ nM}$) limit the development of a parenteral formulation. In order to enhance solubility and bioavailability, inclusion complex formation with cyclodextrins (CDs) is a promising method of choice. Therefore, LAP-CD interactions were also studied by a multianalytical approach. The stability constants of LAP with native cyclodextrins, determined by UV spectroscopy, identified the seven-membered β -CD as the most suitable host. Continuous variation method (Job's plot) by ^1H NMR showed a 1:1 stoichiometry for the complexes. The geometry of the complex was elucidated by 2D ROESY NMR measurements and molecular modeling, indicating that the partial molecular encapsulation includes the fluorophenyl ring of LAP. Phase-solubility studies with four CDs, β -CD, (2-hydroxypropyl)- β -cyclodextrin (HP- β -CD), randomly methylated- β - (RAMEB-) cyclodextrin, and sulfobutylether- β -cyclodextrin (SBE- β -CD), show an A_L type diagram and highly increased solubility via CD complexation. The results are especially promising with SBE- β -CD, exerting more than 600-fold gain in solubility. The equilibrium and structural information presented herein can offer the molecular basis for an improved drug formulation with enhanced bioavailability.

1. Introduction

Lapatinib (LAP) (Figure 1) is a potent and selective inhibitor of the epidermal growth factor receptor (EGFR) and human epidermal growth factor receptor 2 (HER2) [1].

The drug is marketed as its ditosylate salt, in form of a film-coated tablet (Tyverb/Tykerb®). It is currently used in combination therapy with capecitabine or letrozole for the treatment of HER2-positive advanced or metastatic breast cancer [2, 3]. The pharmacokinetic properties of LAP are not optimal. Its oral bioavailability is moderate at best, it has considerable interpatient variability [4], and it is significantly affected by food intake. Both low-fat and high-fat diets before LAP administration increased its systematic exposure [4, 5]. Besides food intake, gastric pH also influences LAP bioavailability. Higher gastric pH reduces LAP absorption; therefore, concomitant use of proton-pump inhibitors (PPIs)

can decrease its bioavailability [6]. The recommended dose of LAP is 1,250–1,500 mg once daily, which means taking 5–6 tablets at once, reducing patient adherence to therapy. Increasing the bioavailability of LAP could reduce the dose. Thus, a novel formulation with high solubility and bioavailability is currently an unmet medical need.

Enhancement of solubility and the concomitant bioavailability can be achieved by various means, such as solid dispersion, size reduction, cosolvency, polymorphism, formation of water soluble prodrug, or complexes [7]. Cyclodextrin (CD) complexation is one of the most extensively investigated ways to improve drug solubility and bioavailability [8].

CDs are cyclic oligosaccharides comprising six, seven, or eight D-glucose units (α -, β -, and γ -CD, resp.) that have been recognized as pharmaceutical adjuvants for the past 20 years. CDs have a hydrophilic surface responsible for their water solubility and a hydrophobic, inner cavity, where lipophilic

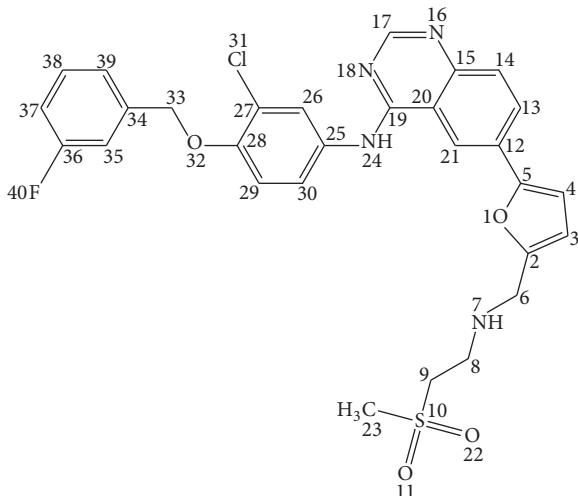


FIGURE 1: Chemical structure and numbering of LAP.

molecules with adequate dimensions can be accommodated forming a noncovalently bonded host-guest complex, conferring an increased aqueous solubility for the encapsulated molecule. Besides native CDs, several modified derivatives are also available, differing in nature, position of substituents, and average degree of substitution, in order to enhance the inclusion capacity and improve the physicochemical properties of the parent, native CDs [9, 10]. Many advantages of inclusion complexation have been reported, such as increased solubility, enhanced bioavailability, improved stability, taste and odor masking, reduced volatility, and modified drug release [11].

So far only a few attempts have been reported to improve the bioavailability of LAP [12–14]. To the best of our knowledge, LAP-CD inclusion complex characterization has yet to be described, despite the fact that CD complexation has been proven to increase the solubility, bioavailability, and anticancer activity of other tyrosine kinase inhibitors erlotinib, gefitinib, imatinib, and so forth [15–19]; moreover, sulfobutylether- β -CD was used for solubilizing LAP in some earlier studies [20, 21].

Here we report the quantified biorelevant physicochemical properties of LAP. On the basis of the protonation constants, lipophilicity, and solubility parameters, we focus on the characterization and optimization of complex formation between various CDs and LAP. The information presented herein can be useful for the development of a new, improved LAP formulation. The methods used in this study include UV and NMR spectroscopy, electrospray ionization-mass spectrometry (ESI-MS), phase-solubility, and molecular modeling.

2. Materials and Methods

2.1. Materials. All CDs (α -CD, β -CD, γ -CD, (2-hydroxypropyl)- β -CD (HP- β -CD), random methylated β - (RAMEB-) CD and sulfobutylether- β -CD (SBE-CD)) were the products of Cyclolab R&D Ltd., Hungary. Degree of substitution (DS)

of HP- β -CD, RAMEB, and SBE-CD was ~4.5, ~12, and 6.5, respectively. LAP was purchased from Eurasia's Ltd., India. Tris(hydroxymethyl)aminomethane (Tris), H₃PO₄, KH₂PO₄, KOH, NaOH, HCl, and KCl were of analytical grade from commercial suppliers. D₂O, DCl, dimethylsulfoxide (DMSO), hexadeuterated dimethylsulfoxide (DMSO-*d*₆), and *n*-octanol were obtained from Sigma-Aldrich, while MeOH was from Merck. Ultrapure, deionized water was prepared by a Milli-Q Direct 8 Millipore system.

2.2. Determination of Protonation Constant by UV-pH Titration. The log K value of LAP was determined by UV-pH titration, using the D-PAS™ ultraviolet spectrophotometer attached to a GLpKa instrument (Sirius Analytical Instruments Ltd., Forest Row, UK). Multiwavelength UV absorbance of the sample solution was monitored throughout the titration using the fiber-optic probe *in situ* [22]. The UV-pH titrations were carried out under nitrogen atmosphere at constant ionic strength ($I = 0.15$ M KCl) and temperature ($t = 25.0 \pm 0.5^\circ\text{C}$). Due to the very low solubility of LAP, the cosolvent method was used to determine its log K value. The cosolvent protonation constants (log K' values) were determined in six different MeOH-water mixtures within 23–43% (w/w). A stock LAP solution of 7 mM concentration was prepared in DMSO. In each experiment, 75 μL of the stock solutions was transferred into sample vials containing 15 mL of 0.15 M KCl solution to achieve the required sample concentration. In each experiment, the acidity of the sample solution was adjusted using 0.5 M HCl to about pH* 2.5 and then titrated with 0.5 M KOH to about pH* 9. Spectral data were recorded in the 200–700 nm region after each pH measurement. About 20 pH readings and absorption spectra were collected from each titration. The log K' values were calculated by RefinementPro™ software.

The measured $\log K'$ values in MeOH-water solutions were then extrapolated to aqueous condition (zero MeOH concentration) according to the Yasuda-Shedlovsky procedure [23]. The Yasuda-Shedlovsky extrapolation method is based on the linear relation between $\log K'$ and the dielectric constant (ϵ) of the cosolvent mixture:

$$\log K' + \log [\text{H}_2\text{O}] = \frac{a}{\varepsilon} + b, \quad (1)$$

where ϵ is the dielectric constant, and $\log[H_2O]$ is the molar water concentration of the given MeOH-water mixture. Three parallel measurements were carried out.

2.3. Determination of the Intrinsic Solubility by Chasing Equilibrium Solubility Method (Cheqsol). The thermodynamic solubility of the nonionized form of the samples was determined by the Chasing Equilibrium Solubility (CheqSol) method [24]. The apparatus used to perform the solubility determinations was a GLpKa titrator and a D-PAS spectrometer controlled by RefinementPro and CheqSol software (Sirius, Forest Row, UK). The titrations were performed under nitrogen atmosphere, at 0.15 M ionic strength and $25.0 \pm 0.5^\circ\text{C}$ using standardized 0.5 M HCl and 0.5 M KOH solutions.

Three parallel measurements were carried out. The implementation of the experiment is described in more detail in our previous articles [19, 25].

2.4. Partition Coefficient Measurement by the Stir-Flask Method. The distribution coefficients were measured from the absorbance of the molecules before and after partitioning using standard stir-flask method as we previously reported [26].

The partition coefficient (P) cannot be measured directly due to the high $\log P$ value of LAP. Therefore, the distribution coefficient (D) was determined at acidic pH (pH = 2), where the molecule is in dicationic form, and at two other pH values at pH 5.83 and 6.11. Three parallel measurements were carried out at each pH. The pH was measured using a Metrohm 6.0204.100 combined pH glass electrode and a Metrohm 780 pH meter. The water/*n*-octanol phase ratio varied between 250 and 600. The $\log P$ values of each macrospecies were calculated based on the following equation:

$$D_{\text{pH}} = \chi_i P_i = \chi_{\text{LAP}^{2+}} P^{\text{LAP}^{2+}} * \chi_{\text{LAP}^+} P^{\text{LAP}^+} * \chi_{\text{LAP}} P^{\text{LAP}}, \quad (2)$$

where χ is the mole fraction of the corresponding macrospecies of the charge indicated.

2.5. Determination of Cyclodextrin Complex Stability by UV Spectroscopy. UV spectroscopic measurements were carried out on a Jasco V-550 diode-array spectrometer (Jasco, Japan) with 10 mm quartz cuvettes at 25°C.

LAP was titrated with a constant amount of native CDs (α -, β -, and γ -CD) at pH = 2 using 0.01 M HCl as solvent. A stock solution of each CD was combined with different volumes of LAP solutions and filled to the same volume. The UV spectra were measured against a blind solution. Three parallel measurements were carried out for each CD. The complex formation equilibrium can be characterized by the following complex stability constant:

$$K_{\text{stab}} = \frac{[\text{CD-LAP}]}{[\text{CD}] [\text{LAP}]}, \quad (3)$$

$$A = \varepsilon_0 [\text{LAP}]_T$$

$$+ (\varepsilon_1 - \varepsilon_0) \frac{(K_{\text{stab}} [\text{CD}]_T + K_{\text{stab}} [\text{LAP}]_T + 1) - \sqrt{(K_{\text{stab}} [\text{CD}]_T + K_{\text{stab}} [\text{LAP}]_T + 1)^2 - 4K_{\text{stab}}^2 [\text{CD}]_T [\text{LAP}]_T}}{2K_{\text{stab}}}. \quad (8)$$

Multiple fitting was performed with Opium program.

2.6. NMR Measurements. All NMR measurements were carried out on an Agilent/Variian VNMR spectrometer (600 MHz for ^1H) with a 5 mm inverse-detection gradient probehead in $\text{D}_2\text{O}/\text{DMSO}-d_6$ 7/3 (v/v) mixtures at 25°C. pD* was set to 2 using DCl solution. Addition of $\text{DMSO}-d_6$ was necessary due to the low solubility of LAP in aqueous solvents, even at acidic pH. The chemical shifts were referenced to the

solvent signal of internal DMSO. Standard pulse sequences and processing routines available in VnmrJ 2.2C/Chempack 4.0 were used.

$$\begin{aligned} [\text{CD}]_T &= [\text{CD}] + [\text{CD-LAP}] \\ [\text{LAP}]_T &= [\text{LAP}] + [\text{CD-LAP}]. \end{aligned} \quad (4)$$

Since absorbance of any of the CDs is negligible and no association other than the CD-LAP formation takes place, it can be safely assumed that only LAP and the complex contribute significantly to the absorbance; therefore, the measured absorbance at any given wavelength is the weighed linear combination of the two absorbances (ε_0 and ε_1 being the molar extinction coefficients of LAP and the complex at a given wavelength, resp.):

$$A = \varepsilon_0 [\text{LAP}] + \varepsilon_1 [\text{CD-LAP}]. \quad (5)$$

Since the equilibrium concentration of LAP can be expressed in terms of the total LAP and the complex concentrations, the above equation takes on the following form:

$$A = \varepsilon_0 [\text{LAP}]_T + (\varepsilon_1 - \varepsilon_0) [\text{CD-LAP}]. \quad (6)$$

The complex concentration can be expressed with the stability constant (K_{stab}) and the total concentrations:

$$K_{\text{stab}} = \frac{[\text{CD-LAP}]}{([\text{CD}]_T - [\text{CD-LAP}]) ([\text{LAP}]_T - [\text{CD-LAP}])}. \quad (7)$$

Solving the above quadratic equation for [CD-LAP] one can derive an equation for absorbance (using (6)), which, if fitted onto the data, provides the complex stability constant:

solvent signal of internal DMSO. Standard pulse sequences and processing routines available in VnmrJ 2.2C/Chempack 4.0 were used.

For determination of complex stoichiometry, Job's continuous variation method was used. Solutions were prepared from LAP and β -CD in complementary amounts, to make up a 1.5 mM summary concentration. The solutions were mixed in different ratios and the ^1H NMR spectra were recorded after 24 h.

The average extent of penetration and the direction of inclusion were investigated by 2D ROESY NMR. Intermolecular NOEs between LAP and CD protons directly involved in the host-guest interaction were detected as cross-peaks. In these experiments, the samples contained 1 mM LAP and 2 mM β -CD. The ROESY spectrum was recorded applying a sweep width of 6 kHz and a spinlock of 3 kHz for a mixing time of 300 ms. 256 increments were collected with 32 repetitions and the measured data matrix was processed as a matrix of 4K (F_2) by 1K (F_1) data points.

2.7. ESI-MS Experiments. High resolution accurate mass of LAP- β -CD inclusion complex was determined with an Agilent 6230 time of flight (TOF) mass spectrometer (Agilent, Germany). Samples were introduced by the Agilent 1260 Infinity LC system, and the mass spectrometer was operated in conjunction with a Jet Stream electrospray ion source in positive ion mode. Reference masses of m/z = 121.050873 and 922.009798 were used to calibrate the mass axis during analysis. The following ion source parameters were employed: flow and temperature of the drying gas: 10 L/min and 320°C; pressure of the nebulizer gas (N_2): 45 psi; capillary voltage: 3000 V; nozzle voltage: 500 V; sheath gas flow and temperature: 10 L/min and 320°C, respectively. TOF-MS parameters were as follows: fragmentor voltage: 300 V; skimmer potential: 170 V; OCT 1 RF Vpp: 750 V. 1 μ L of sample solution, containing 1 mM LAP and 1 mM β -CD, was directly injected into the mass spectrometer. Data were acquired in a 100–3000 m/z mass range and peaks of the hypothesized complex were searched for in the 1000–3000 m/z interval. Mass spectra were processed by Agilent MassHunter B.04.00 software.

2.8. Phase-Solubility Studies. LAP phase-solubility studies were performed according to the method described by Higuchi and Connors [27]. Excess amount of LAP (5–6 mg) was added to 5 mL stock solution with increasing concentrations of different CDs (β -CD, HP- β -CD, RAMEB, and SBE-CD) at pH 2.0 at 0.15 M ionic strength. The obtained suspensions were stirred for 24 hours. After sedimentation, LAP concentrations were measured from the supernatant spectrophotometrically on a Jasco V-550 instrument at 269 nm. Three parallel measurements were carried out.

Assuming 1:1 stoichiometry, the stability constant of the complex (K_{stab}) can be determined using the phase-solubility diagram according to the following equation:

$$K_{\text{stab}} = \frac{\text{slope}}{S_0(1 - \text{slope})}, \quad (9)$$

where S_0 is the intrinsic solubility of LAP in the absence of CDs.

2.9. Molecular Modeling. Molecular modeling study was performed as previously reported [19, 28, 29] by using the semiempirical PM3 method of Hyperchem Professional software (version 8.0). The initial structures of neutral LAP and β -CD were constructed with the help of Hyperchem 8.0 and optimized with PM3. A docking procedure was used to calculate the energy of the complex. LAP was moved into

the CD cavity from the wider rim, step-by-step, performing a geometry optimization after each step until the conformation with the lowest energy was achieved. The docking procedure was repeated using different starting positions, and the lowest achieved energy was considered as the optimal geometry of the complex.

The stability of the complex was estimated from the formation energy:

$$\Delta E = E_{\text{Complex}} - (E_{\text{LAP}} + E_{\text{CD}}). \quad (10)$$

3. Results and Discussion

3.1. Physicochemical Characterization of LAP. Determination of the physicochemical properties of drugs is crucial to understand the pharmacokinetic behavior at the molecular level. Therefore, protonation constants, octanol/water partition coefficient, and the intrinsic solubility of LAP were determined. Due to the very low solubility of LAP, the protonation constants can only be determined by UV-pH titration with mixed-solvent procedure (Figure 2(a)). In our measurements, the MeOH-water cosolvent system was used, where the apparent protonation constants ($\log K'$ values) were determined in six different mixtures; then the aqueous $\log K_s$ were obtained by extrapolation using Yasuda-Shedlovsky method (see (1)) taking into account the dielectric constant of the different cosolvent mixtures (Figure 2(b)). This way $\log K_1 = 7.72 \pm 0.07$ and $\log K_2 = 5.08 \pm 0.05$ values were obtained.

Using the protonation constants, the distribution curves for protonation macropesies of LAP can be depicted as a function of the pH (Figure 3(a)). The diagram shows that, in acidic media, at the pH of the stomach, the dicationic form is overwhelmingly dominant. The compound exists mainly in monocationic form at the pH of the blood (67.4%). Above pH 8, the neutral form dominates, which plays a crucial role in the transport mechanism in general, assuming that orally administrated LAP is absorbed mainly from the intestine.

Lipophilicity of LAP was characterized in terms of partition coefficient, which is the most widely used descriptor of lipophilicity in QSAR studies [30]. The working range of the used stir-flask method is –4 to +4 in $\log D$ values, limited by the accuracy of the phase volume measurement. Based on this fact, the octanol/water partition coefficient at pH values where the nonionized form dominates cannot be measured directly due to the very high $\log P$ value of LAP. Therefore, the $\log D$ values were measured at lower pHs. At pH 2, where the molecule is practically 100% dicationic, the measured $\log D_{\text{pH } 2} = 2.71 \pm 0.06$ characterizes the lipophilicity of the dicationic species. From $\log D$ values at pH 5.83 (3.67 ± 0.07), pH 6.11 (3.90 ± 0.08), and pH 2 (2.71 ± 0.06), the $\log P$ value of the monocationic and neutral species could be calculated using (2). The values are summarized in Table 1, while the pH-dependent lipophilicity profile of LAP is in Figure 3(b).

These values indicate that LAP is an extremely lipophilic molecule and even the dicationic form has the capability to penetrate membranes by passive diffusion. These high $\log P$ values also explain the ability of LAP to cross the blood-brain barrier, which could be a further therapeutic approach in

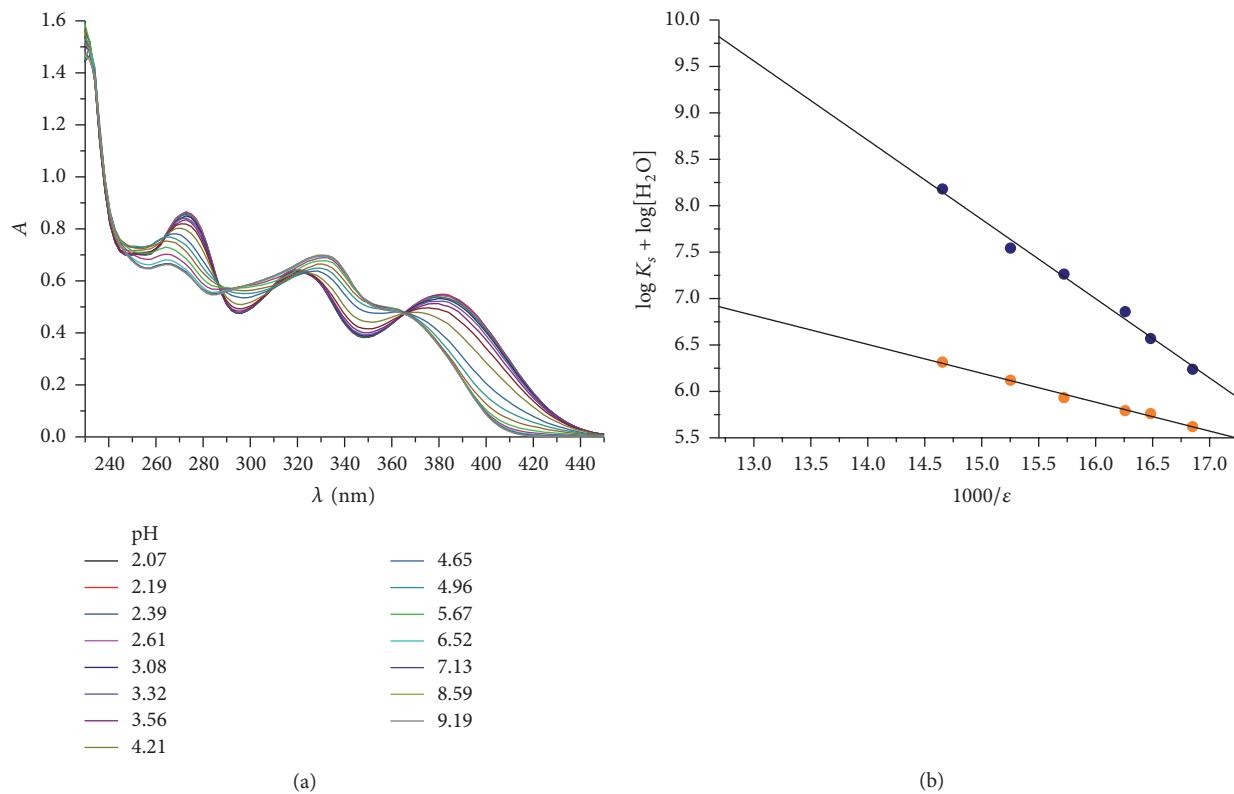


FIGURE 2: Determination of protonation constants of LAP by (a) UV/pH titration of LAP in 43/57 (w/w%) methanol/water mixture and (b) Yasuda-Shedlovsky extrapolation.

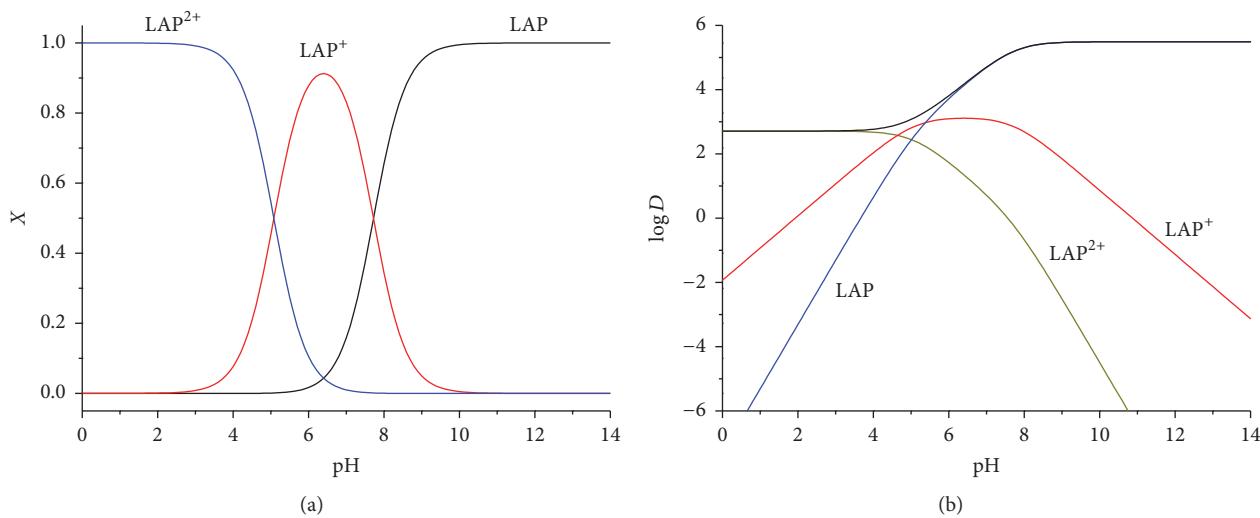


FIGURE 3: (a) Logarithmic distribution diagram for the LAP protonation macrospecies. (b) Contribution of the LAP protonation macrospecies to the lipophilicity profile (in black broad line).

brain metastasis [31]. As a part of our LAP profiling, its intrinsic solubility (the equilibrium solubility of the nonionized form) was measured by the CheqSol method, which resulted in a value of only 0.48 ng/mL. LAP is a class II drug in the Biopharmaceutics Classification System, being of low water solubility and high permeability, which is in good agreement with our determined physicochemical values. These data

quantify that the very poor solubility is the Achilles heel of the LAP administration. It is the root cause for the low bioavailability of the drug the necessity of taking very high doses. These problems could be alleviated by enhancing LAP solubility through CD inclusion complexation. For further pharmaceutical formulation development, optimization of CD-complex formation is crucial; therefore, interactions

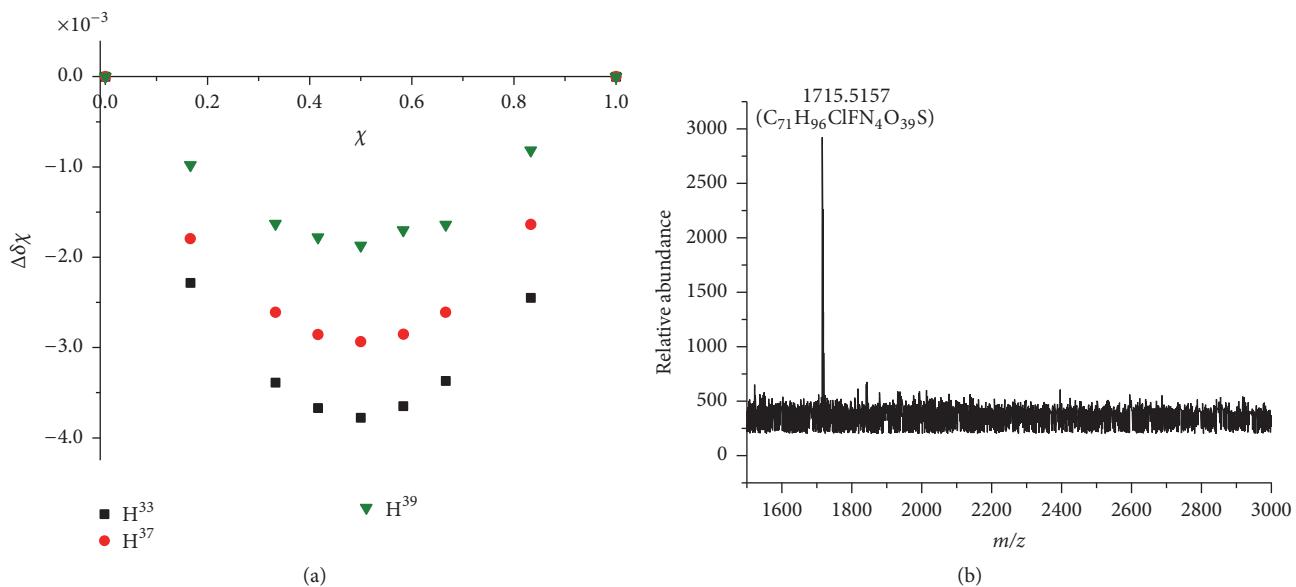


FIGURE 4: Determination of LAP- β -CD stoichiometry. (a) Representative Job plot diagrams of selected LAP protons. (b) Representative mass spectrum of a LAP- β -CD sample.

TABLE 1: Calculated log P values of the different LAP macrospecies.

Macrospecies	log P
Dicationic	2.71 ± 0.07
Monocationic	3.15 ± 0.10
Neutral	5.49 ± 0.10

between LAP and CDs were characterized in solution with a wide variety of complementary analytical techniques and approaches, such as phase-solubility studies, 1H and 2D NMR techniques, UV spectroscopy, and electrospray ionization-time of flight-mass spectrometry (ESI-TOF-MS).

3.2. Determination of LAP- β -CD Stoichiometry: NMR Job Plot and ESI-TOF-MS Studies. In order to draw adequate conclusions on the inclusion process, the knowledge of selector-selectand complexation stoichiometry is a prerequisite. The general method to determine complex stoichiometry is Job's continuous variation method. In this experiment, the 1H NMR chemical shifts (δ) were measured at different LAP concentration/ β -CD concentration (c_{LAP}/c_{CD}) ratios, while the sum of $c_{LAP} + c_{CD}$ was kept constant. The calculated factors ($\Delta\delta\chi_{LAP}$) were plotted as a function of LAP molar ratio (χ_{LAP}). In all cases, the resulting Job plot diagrams showed maxima at 0.5, indicating the typical 1:1 binding stoichiometry. Representative Job plot curves of LAP and β -CD are shown in Figure 4(a).

The complex stoichiometry was also studied by ESI-TOF-MS technique, which provides a powerful means to study noncovalent host-guest inclusion complexes between a CD and the guest molecule [32]. A large number of publications prove that a simple, fast ESI-MS analysis is suitable for the determination of inclusion complex stoichiometry [15, 19, 33];

however, ESI-MS studies imply the question whether the observed peaks represent actual inclusion complexes or rather electrostatic adducts formed during the electrospray process [34], if, however, additional adequate techniques prove that the complex formation ESI-MS is straightforward way to determine complex stoichiometry. Analyzing the MS spectra in the 100–1500 m/z range, the protonated molecular ion of the uncomplexed LAP (m/z 581.1419 [$M+H]^+$), their source fragment ion (m/z 458.1027 [$M+H]^+$), and the characteristic m/z values of uncomplexed β -CD (m/z 1135.3732 [$M+H]^+$, 1157.3531 [$M+Na]^+$) can be observed. Figure 4(b) shows the renormalized upper part (1500–3000 m/z) of the mass spectrum of a sample containing LAP and β -CD in 1:2 molar ratio. The observed signal at m/z 1715.5157 with generated $C_{71}H_{96}ClFN_4O_{39}S$ molecular formula corresponds to the LAP- β -CD 1:1 inclusion complex, which means that ESI-TOF-MS confirms the NMR Job result, verifying the 1:1 stoichiometry.

3.3. Determination of Stability Constant by UV. UV spectroscopy can be a sensitive technique to characterize the CD inclusion complexes, provided that the guest molecule undergoes the significant absorption changes upon complexation. This technique can be used for the determination of CD stability constant of molecules with low solubility as well. Using (8), the stability constants between LAP and native CDs were determined to select the most suitable host. The stability constants determined at pH 2 are listed in Table 2.

Based on these data, we can conclude that, concerning the size, the seven-membered β -CD is the most suitable host molecule for complexation. Thus, further experiments were carried out with β -CD and its derivatives.

3.4. Structure of the LAP- β -CD Complex. 2D ROESY NMR combined with molecular modeling is an appropriate way

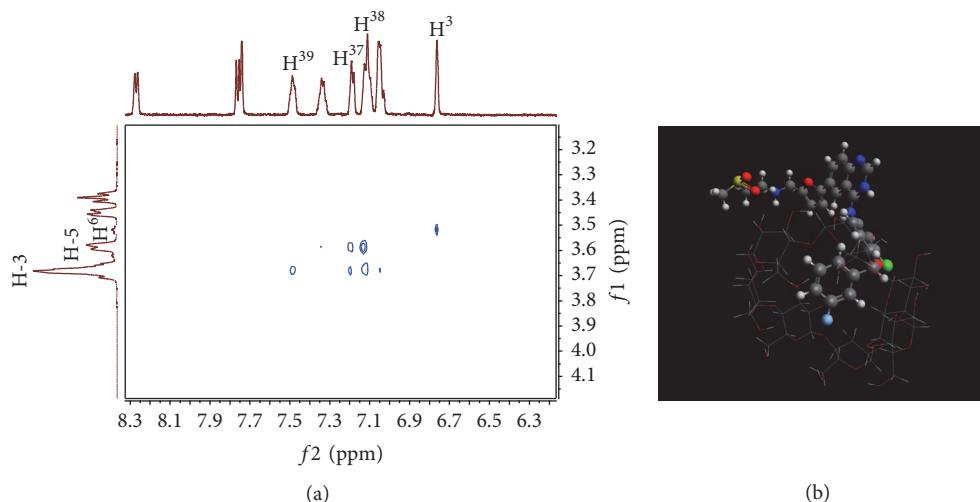


FIGURE 5: Determination of the LAP- β -CD inclusion complex geometry. (a) Expansion of 2D ROESY NMR spectrum. (b) Energy-minimized structure of the complex based on the result of molecular modeling study.

TABLE 2: Determined K_{stab} values at pH 2 for different LAP-CD complexes with UV spectroscopy.

CD	$K_{\text{stab}} (\text{M}^{-1})$
α -CD	46 ± 8
β -CD	121 ± 12
γ -CD	89 ± 6

to characterize the geometry of inclusion complexes. ROESY experiment offers valuable information on spatial proximity between protons of the host and guest molecules by observing intermolecular dipolar cross-correlations, while molecular modeling seeks the energy-minimized structure, which can be visualized at the atomic level [19, 35, 36].

2D NMR ROESY spectra for LAP- β -CD inclusion complex were recorded at $\text{pD}^* 2$, where LAP exhibits an appropriate solubility. The expansion of this 2D spectrum in Figure 5(a) contains an intramolecular cross-peak between H^3 (6.76 ppm) and H^6 (3.52 ppm) nuclei of LAP. From the viewpoint of encapsulation intermolecular cross-peaks of the fluorophenyl protons ($H^{37,38,39}$) with the inner H^3 (3.68 ppm) and H^5 (3.58 ppm) protons of β -CD are crucial. Due to the fact that the H^{39} (7.49 ppm) proton of LAP gives a stronger cross-peak with the H^3 proton of the CD, we can assume that the fluorophenyl moiety is accommodated in the CD cavity from the wider rim. Based on ROESY data, molecular modeling at PM3 level was carried out. The energy-minimization structure is depicted in Figure 5(b). The energy of formation of the complex was found -10.7 kcal/mol , indicating that complex formation is thermodynamically favored.

3.5. Phase-Solubility Studies. Phase-solubility analysis according to Higuchi and Connors is an appropriate technique to demonstrate changes in drug solubility. Moreover, stability constants and stoichiometry can also be determined [9]. Upon addition of increasing amounts of CDs (β -CD,

TABLE 3: K_{stab} values calculated from phase-solubility studies for the CDs employed.

CD	$K_{\text{stab}} (\text{M}^{-1})$
β -CD	131 ± 9
RAMEB	331 ± 15
HP- β -CD	537 ± 20
SBE-CD	31622 ± 120

RAMEB, HP- β -CD, and SBE-CD) at pH 2, linear solubility enhancement was observed for LAP, regardless of the CD employed (Figure 6).

The resulting A_L -type linear positive isotherms reveal 1:1 binding stoichiometry, further supporting Job plot and ESI-TOF-MS results. The K_{stab} values calculated by (9) are summarized in Table 3.

The determined K_{stab} value for β -CD by UV spectroscopy and phase-solubility is in agreement. Moreover similar stability constants were determined for other cyclodextrin complexes of tyrosine kinase inhibitors [15, 16, 19]. The values in Table 3 show that the substitution of the β -CD enhances inclusion complex formation stability as observed also in the case of erlotinib [19]. However, in the case of imatinib, the methylation of the native β -CD reduced the stability of the complex [15], which can be explained by the structural differences of these anticancer drugs. The largest binding constant was observed for SBE- β -CD, which is totally ionized under the applied conditions. Thus, the reason of the more stable complex is the additional electrostatic attractions between the multiply negatively charged hosts and the dicationic LAP. Concerning the solubility enhancing capabilities, it can be stated that CD derivatives enhance the LAP solubility due primarily to the inclusion phenomenon, which can be extended by electrostatic forces. The results are especially promising with SBE- β -CD where more than 600-fold solubility increase could be achieved.

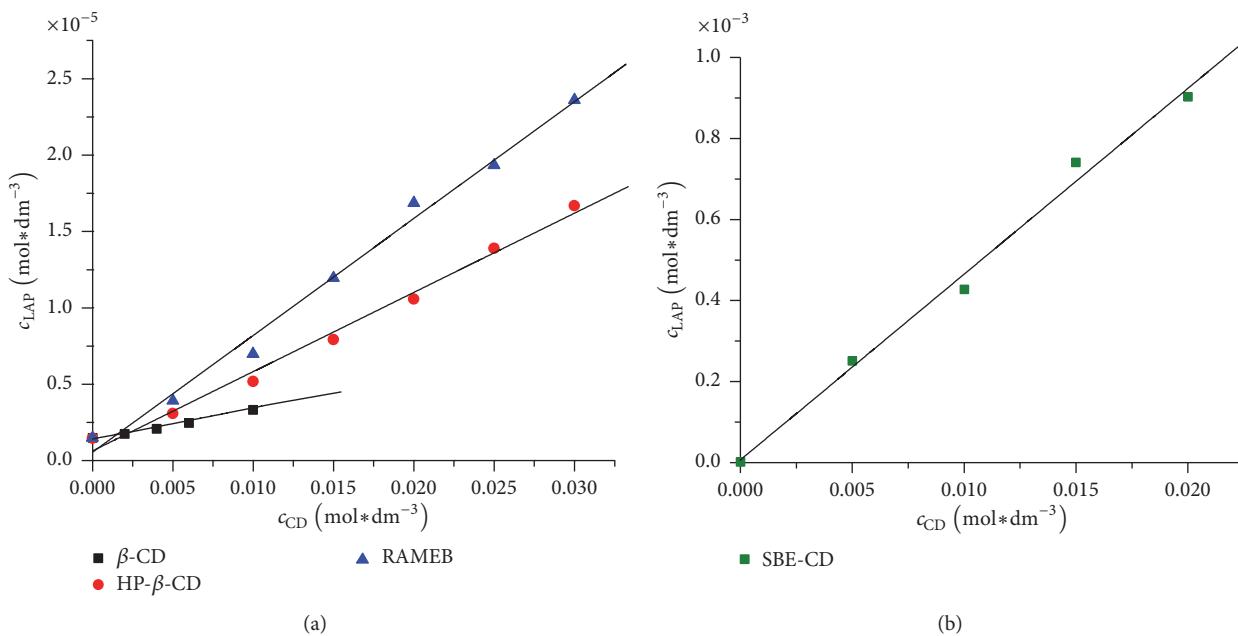


FIGURE 6: Phase-solubility diagrams of LAP-CD complexes.

4. Conclusion

LAP, the anticancer drug, was characterized in terms of lipophilicity, solubility, acid-base, and CD-complex formation properties, using complementary methods, such as UV, NMR, phase-solubility, and molecular modeling. An important practical result of this work is that CDs, especially the substituted derivatives, significantly increase the LAP solubility, which can be a starting point of a new LAP formulation of improved bioavailability.

Competing Interests

The authors declare no conflict of interests.

Authors' Contributions

Gergő Tóth and Ádám Jánoska contributed equally to this work.

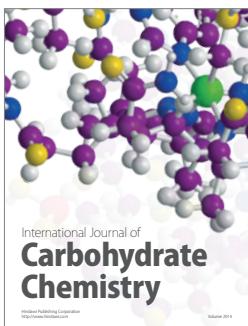
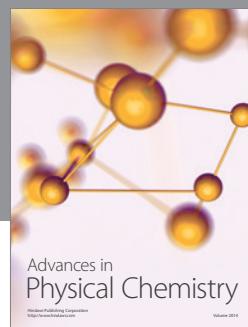
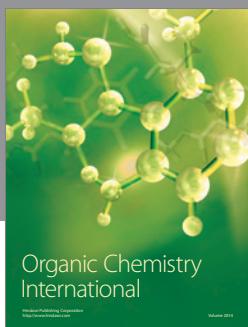
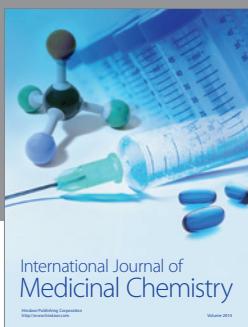
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