Review Article

Adipokines and Hepatic Insulin Resistance

Yu Li,¹ Lin Ding,¹ Waseem Hassan,^{1,2} Daoud Abdelkader,¹ and Jing Shang¹

¹ National Center for Drug Screening and State Key Laboratory of Natural Medicines, China Pharmaceutical University, Jiangsu Province 210009, China

² Department of Pharmacy, Bahauddin Zakariya University, Multan 60800, Pakistan

Correspondence should be addressed to Jing Shang; shangjing21cn@163.com

Received 6 March 2013; Accepted 24 April 2013

Academic Editor: Ji Li

Copyright © 2013 Yu Li et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Obesity is a major risk factor for insulin resistance and type 2 diabetes. Adipose tissue is now considered to be an active endocrine organ that secretes various adipokines such as adiponectin, leptin, resistin, tumour necrosis factor- α , and interleukin-6. Recent studies have shown that these factors might provide a molecular link between increased adiposity and impaired insulin sensitivity. Since hepatic insulin resistance plays the key role in the whole body insulin resistance, clarification of the regulatory processes about hepatic insulin resistance by adipokines in rodents and human would seem essential in order to understand the mechanism of type 2 diabetes and for developing novel therapeutic strategies to treat it.

1. Introduction

In many developed and developing countries, obesity has reached epidemic proportions, resulting in an increasing prevalence of type 2 diabetes characterized by insulin resistance of peripheral tissues such as liver, muscle, and fat which cannot be overcome by hypersecretion of pancreatic beta cells [1]. One survey conducted in 2000 revealed that more than 150 million people in the world suffered from type 2 diabetes [2] and 80% of these cases were related to obesity. Because various studies have demonstrated that hepatic insulin resistance plays a central role in the development of type 2 diabetes and obesity is centrally involved in increasing the clinical risk of diabetes, visceral adipose tissue is now thought to provide a link between obesity and hepatic insulin resistance.

Adipose tissue was traditionally regarded as a passive energy reservoir. However, since the discovery of leptin and subsequent identification of other adipose tissue-derived cytokines (e.g., adiponectin and resistin) in the last two decades [3], it became clear that adipose tissue is an active endocrine organ. Obese adipose tissue also secretes various inflammatory cytokines, such as interleukin-6 (IL-6) and tumour necrosis factor- α (TNF- α) [4]. All of these cytokines, termed adipokines, act in an autocrine, paracrine, or endocrine fashion to control various metabolic functions.

Some of these adipokines have been implicated in the development of hepatic insulin resistance. Indeed, they may act locally or distally to alter insulin sensitivity in insulin-targeted organs such as liver which is also discussed in detail by Marra and Bertolani [5] previously or may act through neuroendocrine, autonomic, or immune pathways. For example, activation of proinflammatory pathways in adipose tissue is known to interfere with insulin signaling and induce hepatic insulin resistance [6]. Although, the role of adipocytokines on insulin resistance has been comprehensively elucidated elsewhere [7, 8], but recently specialized review on their actions in hepatic insulin resistance is missing. In this paper, we focus on the role of a series of adipokines and discuss how they influence hepatic insulin sensitivity.

2. Adipokines and Hepatic Insulin Resistance

2.1. Adiponectin. Since the identification of adiponectin as a protein exclusively produced from adipocytes exclusively by Scherer et al. in 1995 [9], it has grabbed much attention by scientific communities, primarily due to its inverse relationship with hepatic insulin resistance. It was first named as adipocytes complement related protein of 30 kDa (ACRP30) on the basis of structural similarity with the complement C1q fraction [9]. At the same time other research groups

also termed it as AdipoQ, adipose most abundant gene transcript 1 (apM1) or (gelatin-binding protein 28) GBP28, in both mice and humans [10]. Following the characterization of its genomic organization, comprising three exons and its 3q27 localization in 1999 [11] the protein was named adiponectin [12]. In 2004, the gene was named adipocytes C1q and collagen-domain-containing (ACDC), but actual nomenclature given by the Human Genome Organization (HUGO) nomenclature is now ADIPOQ [13].

The adipokine adiponectin is greatly expressed in human serum where it accounts for 0.01% of total plasma protein and is predominantly secreted by adipose tissue [14] (reports of secretion in differentiating preadipocytes [15] and placenta [16]) in inverse proportion to the body mass index [17]. A unique feature of the structure of adiponectin is its ability to assemble into several characteristic oligomeric isoforms, including trimers (low molecular weight (LMW)), hexamers (middle molecular weight (MMW)), and the oligomeric complexes comprising 18 protomers or above (high molecular weight (HMW)) [18]. Adiponectin presents predominantly in the circulation in the three oligomeric complexes cited previously [19]. Trimeric adiponectin is the basic unit block of adiponectin. Two LMW adiponectin molecules linked by disulfide bonds to form hexameric adiponectin [18]. Adiponectin may also form hetero-oligomers with additional members of the C1q/TNF-related protein (CTRP) family such as the CTRP9 [20].

The role of adiponectin in hepatic insulin resistance has evolved into a major factor and seen by many as therapeutic option now. The generalized effects of adiponectin on insulin resistance can be cited elsewhere [21, 22]. In this portion we will review the major studies describing the role of adiponectin and its receptors [23] in hepatic insulin resistance.

Hepatic insulin resistance is a hall mark for the spectrum of many diseases, and it is an independent predictor for metabolic disorders. Recent evidence suggests that visceral adipose tissue is a metabolic and inflammatory organ that signals and modulates the action and metabolism of the brain, liver, muscle, and cardiovascular system [24, 25]. The hormone plays a role in the attenuation of the hepatic metabolic disorders that may result in type 2 diabetes [26], insulin resistance, obesity [27], and nonalcoholic fatty liver disease (NAFLD) [28].

Recent data has exhibited more specification as most of the studies have implicated HMW oligomers in attenuation of hepatic insulin resistance. For instance, the synthesis of the HMW oligomers is necessary to mediate the insulin sensitizing effects of adiponectin on the suppression of hepatic gluconeogenesis in primary rat hepatocytes [29]. Moreover, HMW oligomers of adiponectin potentially blunted hyperglycemia in diabetic mice through the inhibition of hepatic glucose production [30]. Further, acute injection of recombinant adiponectin enriched with the HMW oligomers resulted in a marked activation of AMP-activated kinase (AMPK) in the liver, while chronic infusion with this protein leads to prolonged alleviation of hyperglycemia and insulin resistance in *db/db* diabetic mice [31]. Animal studies are matched to the clinical data exhibiting that the ratio

of HMW/total adiponectin correlates closely with hepatic insulin sensitivity [32]. The role of the HMW oligomer as a predominant active form of adiponectin mediating its hepatic actions is also supported by two recent independent reports demonstrating that the insulin-sensitizing effects of the peroxisome proliferator-activated receptor gamma (PPAR- γ) agonist thiazolidinediones were diminished in *ob/ob* obese mice with the targeted mutation of the adiponectin gene [33]. Notably, treatment with thiazolidinediones, which is an insulin sensitizer caused a selective increase of the HMW oligomeric adiponectin [32]. Pioglitazone, another insulin sensitizer, has remarkably increased the HMW adiponectin, which correlated greatly with the increased hepatic insulin action [34].

Plethora of animal data has been published describing the role of adiponectin in hepatic insulin resistance. Different animal models have supported this supposition widely. For example, a high scale study of adiponectin function in animal models of obesity, insulin resistance, and type 2 diabetes provided strong evidence that adiponectin promotes insulin sensitivity in muscle and in the liver [35]. Further, most of the adiponectin knockout mice were more insulin resistant than controls, although to different degrees, and this factor has been associated with hepatic insulin resistance [36].

Moreover, adiponectin-deficient mice showed mild insulin resistance in the liver with administration of standard diet [36]. Similar evidences were extracted on adiponectin receptor (AdipoR) studies, as *adipoR1* and *adipoR2* knockout mice exhibit mild insulin resistance [37]. Further, *AdipoR2* knockout mice showed reduced diet-induced insulin resistance but promoted type 2 diabetes [38]. In another study, secretion of wild type adiponectin has been employed [39] and caused insulin sensitization when expressed in either liver [39] or adipose tissue [40]. Interestingly, infusion of adiponectin inhibited both the expression of hepatic gluconeogenic enzymes and the rate of endogenous glucose production by the liver. The latter was dependent on insulin, because adiponectin alone had no significant effect on glucose output in cultured hepatocytes. Obviously, both wild-type and type 2 diabetic mice were shown to reduce the expression of gluconeogenic enzymes and elevated phosphorylation of hepatic AMP-activated protein kinase (AMPK), and the effects were attributed to adiponectin due to its circulating levels [41]. Indeed AMPK has currently established itself as a target for adiponectin, and its phosphorylation has been linked with the adiponectin level [42]. It is worth mentioning here that most of the adiponectin effects on liver has been attributed to either AMPK activation or nuclear receptor involvement, and this activation is mediated by *AdipoR2*, but a recent ground breaking study refuted this as an only view and put forward new dimensions in future adiponectin research. Motoharu and his group have shown that adiponectin effects on hepatic system can be independent of *AdipoR2* and can also be mediated by inflammatory cells [43].

Clinical studies have described the similar evidences regarding the role of adiponectin in liver insulin resistance. Instantly, adiponectin levels are associated, in healthy humans, with plasma concentrations of various liver function

indices [44]. Further, a clinical study showed that plasma level of adiponectin was closely linked with hepatic lipids and insulin resistance in patients administered with pioglitazone [45]. Another clinical study supported the similar view and showed that increased adiponectin level could help restore the hepatic insulin resistance in severely obese women [46].

In conclusion, adiponectin is a promising target for hepatic insulin resistance and resultantly metabolic disorder, but further clinical research is required to translate similar impacts in humans keeping in mind the pleiotropic nature of this hormone.

2.2. Leptin. In 1994, *ob* gene was first cloned on mice chromosome 6 [47]. A year later, Halaas found that leptin was the gene product of *ob* in *colon bacillus* [48]. Leptin is a 16-KDa hormone secreted by adipocytes which plays a key role in the regulation of food intake, energy metabolism, saccharide, lipid metabolism, and so forth. The levels of leptin in adipose tissue and plasma are dependent on the amount of energy stored like fat as well as the status of energy balance. Therefore, leptin levels are higher in obese individuals and increase with overfeeding [49, 50].

Lots of studies reported that a strong positive correlation exists between the concentration of leptin and insulin sensitivity and obesity [51–54], but the anti-insulin resistance effect of leptin, independently of its activity on weight control, was also reported [55, 56]. The mechanisms that explain the direct role of leptin in hepatic insulin resistance are not well understood. Studies in animal model showed that exogenous leptin administration leads to a dramatic improvement in insulin resistance that is independent of decreased caloric intake [57]. Clinical studies showed that chronic leptin treatment ameliorates insulin-stimulated hepatic and peripheral glucose metabolism in severely insulin-resistant lipodystrophic patients [58]. A study showed that in leptin receptor-deficient Koletsky rats, adenovirally induced expression of leptin receptors in the area of the hypothalamic arcuate nucleus improved peripheral insulin sensitivity via enhancement of suppression of hepatic glucose production, with no change of insulin-stimulated glucose uptake or disposal, and leptin regulated hepatic insulin sensitivity via phosphatidylinositol-3-OH kinase (PI3K) signaling in the liver of this animal model [59].

In summary, leptin acts as an important target to improve the hepatic sensitive to insulin, and the precise mechanisms about leptin underlying hepatic insulin resistance should be investigated by experiments *in vivo* and *in vitro*.

2.3. Resistin. Resistin is a member of a family of cysteine-rich proteins referred to as resistin-like molecules, termed RELMs, which include RELM- α , RELM- β , and the recently discovered RELM- γ [60]. It was discovered in 2001 by Dr Mitchell A. Lazar [61] and was called resistin, since mice injected with resistin exhibited insulin resistance. Crystallographic studies of resistin have determined its complex hexameric structure. Resistin is proved to circulate in two distinct assembly states, likely corresponding to hexamers and trimers. Infusion of an intertrimer disulfide bonds-lacking resistin showed more potent effects on hepatic insulin

sensitivity than those observed with wild-type resistin [62]. This result suggested an activation effect of different inter-trimer disulfide bonds and further suggested a potential target site for the receptor interaction.

In rodents, resistin primarily expressed in and secreted from mature adipocytes, with some expression in pancreatic islets and portions of the pituitary and hypothalamus. However, resistin is expressed primarily by macrophages and seems to be involved in the recruitment of other immune cells and the secretion of proinflammatory factors in humans [63]. The diverse expression and regulatory patterns may be explained by the evidence that human and mouse resistins have diverse genomic organizations [64]. Moreover, at the protein level, human resistin is only 55% identical to its murine counterpart.

A study in rodents showed that the infusion of resistin rapidly induced severe hepatic insulin resistance, resulting in a reduced insulin-mediated suppression of gluconeogenesis and increased glycogenolysis [65]. Consistent with these reports, resistin induced insulin resistance with a robust decrease in insulin-stimulated phosphorylation of Akt and glycogen synthase kinase3 (GSK3) human liver cell line HepG2 cells [66], pointing to a specific role of resistin in the initiation of hepatic insulin resistance. Furthermore, mice treated with an antiresistin IgG [67] or expressing a dominant negative resistin [68] exhibit low fasted blood glucose levels due to reduced hepatic glucose production.

In a word, resistin could induce hepatic insulin resistance by inhibiting the phosphorylation of Akt and GSK3, and its effect on insulin sensitivity is opposite to those reported for the adipocyte-secreted hormone adiponectin, which increases insulin sensitivity of the same liver-specific functions.

2.4. IL-6. IL-6 is secreted by many cell types, including immune cells, fibroblasts, endothelial cells, myocytes, and a variety of endocrine cells [69]. Adipose tissue contributes to 10–35% of circulating IL-6 [70] in resting, healthy humans, and the production is greater in obese subjects [71]. It is a multifunctional cytokine that has been well known for its anti-inflammatory and proinflammatory effect in immune responses. IL-6 signals through a cell-surface type I cytokine receptor complex consisting of the specific receptor subunit IL-6 receptor (IL-6R) and the signal-transducing component glycoprotein 130 (gp130), which is the common signal transducer for several cytokines including leukemia inhibitory factor (LIF), ciliary neurotrophic factor, oncostatin M, IL-11, and cardiotrophin-1 [72]. In contrast, the expression of IL-6R is restricted to certain tissues. IL-6 interacts with its receptor IL-6R and gp130 protein to form a complex that activates the receptor IL-6R.

The inflammatory regulator IL-6 has also emerged as a factor that is implicated in hepatic insulin resistance. However, the role of IL-6 in the etiology of insulin resistance is not fully understood. Impaired insulin receptor signaling and insulin-dependent glycogen synthesis were found in HepG2 cells and primary mouse hepatocytes acutely pretreated with IL-6 [73]. IL-6 caused reduced insulin signal transduction in the liver of mice [74]. These results seemed to be consistent

with the hypothesis that IL-6 may have a negative effect on insulin resistance. But in healthy humans under the basal condition, treatment by recombinant human IL-6 (rhIL-6) at a physiological concentration neither impaired the whole body glucose disposal nor increased endogenous glucose production [75]. In patients with type 2 diabetes, splanchnic glucose output did not increase with acute infusion of rhIL-6 [76], while glucose disposal was not impaired, suggesting that IL-6 might have favorable action on insulin action.

IL-6 mediates insulin sensitivity through many distinct mechanisms. Of note, there is a significant correlation between the level of IL-6 in adipose tissue and several metabolic parameters such as fasting plasma glucose, basal and insulin-stimulated glucose transport, and whole-body glucose disposal [77]. Thus, IL-6 may play an important role in insulin action in these subjects. Moreover, IL-6 mediated suppressor of cytokine signaling (SOCS-3) pathway in liver, leading to impairment of insulin actions [78].

To sum up, the effect of IL-6 on hepatic insulin sensitivity is uncertain; it appeared to be determined by whether it is present acutely or chronically; the latter is the setting associated with insulin resistance.

2.5. TNF- α . TNF- α is a proinflammatory cytokine produced by various types of cells, mainly but not only inflammatory cells like macrophages and lymphocytes. The release of TNF- α in noninflammatory cells has also been reported, at lesser degrees. Since the first recognized sequencing of TNF- α in 1984, it has established itself as a prime proinflammatory cytokine [79], making it the subject of continuous pondering by researchers. It is now clear that TNF- α is involved in array of pathological conditions like obesity, congestive heart failure, inflammations, and insulin resistance [80], nevertheless the exact role and degree of its effects are still not clearly understood.

TNF- α has been proposed as a link between adiposity and the development of insulin resistance [81], because the majority of type 2 diabetic subjects are obese [82]. Elevated levels of TNF- α have been observed in obese and insulin resistance humans and animals [83]. TNF- α -mediated pathway interference has shown protection in diet induced animal models of obesity and metabolic syndrome [84]. A clinical study has observed improvement in the insulin sensitivity under anti-TNF- α therapy [85]. Anti-TNF- α therapy has also shown to attenuate in insulin resistance in animal studies involving fructose-fed rats [86]. Further, TNF- α expression is increased in adipose tissue in obese rodents and humans [87], and reducing TNF- α signaling either by knocking TNF- α out or by infusing blocking antibodies can reduce insulin resistance in obese rodents [88]. More recent clinical evidence confirms the proportional rise of TNF- α and IR with parallel increase in body mass index (BMI) factor [89]. A recent cohort study on nondiabetic patients with insulin resistance has shown concomitant decrease in TNF- α level accompanied by an improvement in insulin resistance after administration of rosiglitazone [90]. Another reported study has observed the direct dose-dependant effects of TNF- α and higher doses linked directly to insulin sensitivity [91]. Moreover, insulin stimulating effects on glycemic levels and glucose uptake

have been reversed by the administration of TNF- α [92]. A clinical trial which consists of 15 women with normal glucose tolerance and developed gestational diabetes mellitus has proved TNF- α as the sole biomarker and predictor for insulin resistance during pregnancy [93]. In one rodent model, obese mice lacking either TNF- α or its receptors showed protection against developing insulin resistance [94]. Liver resident macrophages (Kupffer cells) in human body are the primary source of TNF- α ; diet induced macrophages which subsequently release the TNF- α are found to be involved in insulin resistance in paracrine signaling.

TNF- α is reported to demonstrate wide range of effects in pathogenesis of insulin resistance. The precise signaling pathway is, however, not clear yet even after scores of studies and scientific data put forward on this particular subject. However, the most attention grabbing signaling pathway in liver supported by numerous evidences is through the phosphorylation of the insulin receptor substrate-1 (IRS-1) protein on serine residues. This could prevent its interaction with the insulin receptor beta subunit and stop the insulin signaling pathway [95–97]. However, the precise role of TNF- α in human insulin signaling requires extensive scientific investigation in future to clear its signaling pathway.

Conversely, numerous clinical studies have not been able to support similar findings and failed to prove its role in the relationship between type 2 diabetes coexisting with insulin resistance [98, 99]. TNF- α antibody infusion in humans was shown not to alter insulin sensitivity, fueling lingering uncertainty about the biological relevance of this pathway in human insulin resistance states [100].

Conclusively, TNF- α role in modulating insulin resistance with no doubt varies from species to species and depends on animal models adopted for particular study. Further, due to its release by various types of cell types involving both inflammatory and noninflammatory cells which then can be involved in insulin resistance pathogenesis both by autocrine or paracrine manner, TNF- α is the subject of uncertainty regarding its precise role in insulin resistance. Without doubt, further scientific work is required in order to unravel its exactness in insulin resistance.

3. Summary

During the last couple of years it has been shown that adipokines play an important role in physiology and pathophysiology of insulin sensitivity. Our review summarizes recent findings regarding the relationship between adipocytokines and hepatic insulin resistance (Figure 1). The mechanisms by which adipocytokines promote or relieve insulin resistance are complex, and our understanding is yet incomplete. According to most of the recent studies, it seems that excessive adipose tissue may be detrimental partially through secretion of the following cytokines: TNF- α , IL-6, and resistin. In contrast, the presence of adipose tissue is vital in the prevention of hepatic insulin resistance, at least in part, via secretion of the following cytokines: leptin and adiponectin. Indeed, adipocytokines are major regulators of hepatic insulin sensitivity potentially linking

FIGURE 1: The effects of adipokines on hepatic glucose metabolism and insulin signalling. Both of adiponectin (APN) and leptin can decrease hepatic gluconeogenesis. Resistin (REN) can increase hepatic gluconeogenesis and glycogenolysis. Moreover, interleukin-6 (IL-6) can decrease glycogen synthesis, and tumor necrosis factor α (TNF- α) can decrease glucose uptake in liver. Both of them can block hepatic insulin signalling by interfection of insulin receptor signalling and insulin signal transduction.

insulin resistance and obesity. Further work is needed to clearly determine how they regulate the insulin signaling in hepatocytes and influence insulin sensitivity in other tissues (e.g., muscle and adipose tissue) in rodents and human and their contribution to glucose homeostasis in diabetes.

Authors' Contribution

Yu Li and Lin Ding equally contributed to the project.

Acknowledgments

The study supported by "National Science and Technology Infrastructure Program of China" (2012BAI30B001), "Mega-projects of Science Research for the 12th Five-Year Plan of China" (2011ZX09401-007), "Project Program of State Key Laboratory of Natural Medicines, China Pharmaceutical University (JKGZ201108), and "Jiangsu Yanjiushengchuangxin Plan" (CXLX11-0782).

References

- [1] A. R. Saltiel, "The molecular and physiological basis of insulin resistance: emerging implications for metabolic and cardiovascular diseases," *Journal of Clinical Investigation*, vol. 106, no. 2, pp. 163–164, 2000.
- [2] S. Matthaei, M. Stumvoll, M. Kellerer, and H. U. Haring, "Pathophysiology and pharmacological treatment of insulin resistance," *Endocrine Reviews*, vol. 21, no. 6, pp. 585–561, 2000.
- [3] T. Kadowaki, T. Yamauchi, N. Kubota, K. Hara, K. Ueki, and K. Tobe, "Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome," *Journal of Clinical Investigation*, vol. 116, no. 7, pp. 1784–1792, 2006.
- [4] H. Tilg and A. R. Moschen, "Inflammatory mechanisms in the regulation of insulin resistance," *Molecular Medicine*, vol. 14, no. 3-4, pp. 222–231, 2008.
- [5] F. Marra and C. Bertolani, "Adipokines in liver diseases," *Hepatology*, vol. 50, no. 3, pp. 957–969, 2009.
- [6] G. S. Hotamisligil, "Inflammation and metabolic disorders," *Nature*, vol. 444, no. 7121, pp. 860–867, 2006.
- [7] K. Rabe, M. Lehrke, K. G. Parhofer et al., "Adipokines and insulin resistance," *Molecular Medicine*, vol. 14, no. 11, pp. 741–751, 2008.
- [8] B. Antuna-Puente, B. Feve, S. Fellahi, and J. P. Bastard, "Adipokines: the missing link between insulin resistance and obesity," *Diabetes and Metabolism*, vol. 34, no. 1, pp. 2–11, 2008.
- [9] P. E. Scherer, S. Williams, M. Fogliano, G. Baldini, and H. F. Lodish, "A novel serum protein similar to C1q, produced exclusively in adipocytes," *Journal of Biological Chemistry*, vol. 270, no. 45, pp. 26746–26749, 1995.
- [10] J. J. Díez and P. Iglesias, "The role of the novel adipocyte-derived hormone adiponectin in human disease," *European Journal of Endocrinology*, vol. 148, no. 3, pp. 293–300, 2003.
- [11] K. Saito, T. Tobe, S. Minoshima et al., "Organization of the gene for gelatin-binding protein (GBP28)," *Gene*, vol. 229, no. 1-2, pp. 67–73, 1999.
- [12] N. Ouchi, S. Kihara, Y. Arita et al., "Novel modulator for endothelial adhesion molecules: adipocyte-derived plasma protein adiponectin," *Circulation*, vol. 100, no. 25, pp. 2473–2476, 1999.
- [13] F. Vasseur, D. Meyre, and P. Froguel, "Adiponectin, type 2 diabetes and the metabolic syndrome: lessons from human genetic studies," *Expert Reviews in Molecular Medicine*, vol. 8, no. 27, pp. 1–12, 2006.
- [14] K. Maeda, K. Okubo, I. Shimomura, T. Funahashi, Y. Matsuzawa, and K. Matsubara, "cDNA cloning and expression of a novel adipose specific collagen-like factor, apM1 (adipose most abundant gene transcript 1)," *Biochemical and Biophysical Research Communications*, vol. 221, no. 2, pp. 286–289, 1996.
- [15] C. Lara-Castro, Y. Fu, B. H. Chung, and W. T. Garvey, "Adiponectin and the metabolic syndrome: mechanisms mediating risk for metabolic and cardiovascular disease," *Current Opinion in Lipidology*, vol. 18, no. 3, pp. 263–270, 2007.
- [16] J. Chen, B. Tan, E. Karteris et al., "Secretion of adiponectin by human placenta: differential modulation of adiponectin and its receptors by cytokines," *Diabetologia*, vol. 49, no. 6, pp. 1292–1302, 2006.
- [17] Y. Arita, S. Kihara, N. Ouchi et al., "Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity," *Biochemical and Biophysical Research Communications*, vol. 257, no. 1, pp. 79–83, 1999.
- [18] Y. C. Woo, A. W. Tso, and A. Xu, "Combined use of serum adiponectin and tumor necrosis factor- α receptor 2 levels was comparable to 2-hour post-load glucose in diabetes prediction," *PLoS One*, vol. 7, no. 5, pp. 1–8, 2012.
- [19] F. Simpson and J. P. Whitehead, "Adiponectin-It's all about the modifications," *International Journal of Biochemistry and Cell Biology*, vol. 42, no. 6, pp. 785–788, 2010.
- [20] G. W. Wong, S. A. Krawczyk, C. Kitidis-Mitrokostas et al., "Identification and characterization of CTRP9, a novel secreted glycoprotein, from adipose tissue that reduces serum glucose in mice and forms heterotrimer with adiponectin," *FASEB Journal*, vol. 23, no. 1, pp. 241–258, 2009.

- [21] T. Yamauchi, J. Kamon, Y. Ito et al., "Cloning of adiponectin receptors that mediate antidiabetic metabolic effects," *Nature*, vol. 423, no. 6941, pp. 762–769, 2003.
- [22] B. Antuna-Puente, B. Feve, S. Fellahi, and J. P. Bastard, "Adipokines: the missing link between insulin resistance and obesity," *Diabetes and Metabolism*, vol. 34, no. 1, pp. 2–11, 2008.
- [23] Y. Matsuzawa, "Adiponectin: a key player in obesity related disorders," *Current Pharmaceutical Design*, vol. 16, no. 17, pp. 1896–1901, 2010.
- [24] O. Ukkola and M. Santaniemi, "Adiponectin: a link between excess adiposity and associated comorbidities?" *Journal of Molecular Medicine*, vol. 80, no. 11, pp. 696–702, 2002.
- [25] J. J. Díez and P. Iglesias, "The role of the novel adipocyte-derived hormone adiponectin in human disease," *European Journal of Endocrinology*, vol. 148, no. 3, pp. 293–300, 2003.
- [26] O. Renaldi, B. Pramono, H. Sinorita, L. B. Purnomo, R. H. Asdie, and A. H. Asdie, "Hypoadiponectinemia: a risk factor for metabolic syndrome," *Acta medica Indonesiana*, vol. 41, no. 1, pp. 20–24, 2009.
- [27] K. Dadson, Y. Liu, and G. Sweeney, "Adiponectin action: a combination of endocrine and autocrine/paracrine effects," *Frontiers in Endocrinology*, vol. 2, no. 62, pp. 1–14, 2011.
- [28] A. H. Berg, T. P. Combs, X. Du, M. Brownlee, and P. E. Scherer, "The adipocyte-secreted protein Acrp30 enhances hepatic insulin action," *Nature Medicine*, vol. 7, no. 8, pp. 947–953, 2001.
- [29] L. Xie, D. Boyle, D. Sanford, P. E. Scherer, J. E. Pessin, and S. Mora, "Intracellular trafficking and secretion of adiponectin is dependent on GGA-coated vesicles," *Journal of Biological Chemistry*, vol. 281, no. 11, pp. 7253–7259, 2006.
- [30] N. Vrachnis, P. Belitsos, and S. Sifakis, "Role of adipokines and other inflammatory mediators in gestational diabetes mellitus and previous gestational diabetes mellitus," *International Journal of Endocrinology*, vol. 2012, Article ID 549748, 12 pages, 2012.
- [31] A. Gastaldelli, S. A. Harrison, R. Belfort-Aguilar et al., "Importance of changes in adipose tissue insulin resistance to histological response during thiazolidinedione treatment of patients with nonalcoholic steatohepatitis," *Hepatology*, vol. 50, no. 4, pp. 1087–1093, 2009.
- [32] A. Baranova, S. J. Gowder, K. Schlauch et al., "Gene expression of leptin, resistin, and adiponectin in the white adipose tissue of obese patients with non-alcoholic fatty liver disease and insulin resistance," *Obesity Surgery*, vol. 16, no. 9, pp. 1118–1125, 2006.
- [33] T. Yamauchi, J. Kamon, Y. Minokoshi et al., "Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase," *Nature Medicine*, vol. 8, no. 11, pp. 1288–1295, 2002.
- [34] A. R. Nawrocki, M. W. Rajala, E. Tomas et al., "Mice lacking adiponectin show decreased hepatic insulin sensitivity and reduced responsiveness to peroxisome proliferator-activated receptor γ agonists," *Journal of Biological Chemistry*, vol. 281, no. 5, pp. 2654–2660, 2006.
- [35] T. Yamauchi, Y. Nio, T. Maki et al., "Targeted disruption of AdipoR1 and AdipoR2 causes abrogation of adiponectin binding and metabolic actions," *Nature Medicine*, vol. 13, no. 3, pp. 332–339, 2007.
- [36] Y. Liu, M. D. Michael, S. Kash et al., "Deficiency of adiponectin receptor 2 reduces diet-induced insulin resistance but promotes type 2 diabetes," *Endocrinology*, vol. 148, no. 2, pp. 683–692, 2007.
- [37] T. P. Combs, U. B. Pajvani, A. H. Berg et al., "A transgenic mouse with a deletion in the collagenous domain of adiponectin displays elevated circulating adiponectin and improved insulin sensitivity," *Endocrinology*, vol. 145, no. 1, pp. 367–383, 2004.
- [38] J. Y. Kim, E. van de Wall, M. Laplante et al., "Obesity associated improvements in metabolic profile through expansion of adipose tissue," *Journal of Clinical Investigation*, vol. 117, no. 9, pp. 2621–2637, 2007.
- [39] A. H. Berg, T. P. Combs, X. Du, M. Brownlee, and P. E. Scherer, "The adipocyte-secreted protein Acrp30 enhances hepatic insulin action," *Nature Medicine*, vol. 7, no. 8, pp. 947–953, 2001.
- [40] T. Yamauchi, Y. Nio, T. Maki et al., "Targeted disruption of AdipoR1 and AdipoR2 causes abrogation of adiponectin binding and metabolic actions," *Nature Medicine*, vol. 13, no. 3, pp. 332–339, 2007.
- [41] M. Awazawa, K. Ueki, K. Inabe et al., "Adiponectin enhances insulin sensitivity by increasing hepatic IRS-2 expression via a macrophage-derived IL-6-dependent pathway," *Cell Metabolism*, vol. 13, no. 4, pp. 401–412, 2011.
- [42] A. L. Bermejo, P. Botas, T. Funahashi et al., "Adiponectin, hepatocellular dysfunction and insulin sensitivity," *Clinical Endocrinology*, vol. 60, no. 2, pp. 256–263, 2004.
- [43] M. Bajaj, S. Suraamornkul, P. Piper et al., "Decreased plasma adiponectin concentrations are closely related to hepatic fat content and hepatic insulin resistance in pioglitazone-treated type 2 diabetic patients," *Journal of Clinical Endocrinology and Metabolism*, vol. 89, no. 1, pp. 200–206, 2004.
- [44] E. Lin, L. S. Phillips, T. R. Ziegler et al., "Increases in adiponectin predict improved liver, but not peripheral, insulin sensitivity in severely obese women during weight loss," *Diabetes*, vol. 56, no. 3, pp. 735–742, 2007.
- [45] Y. Zhang, R. Proenca, M. Maffei, M. Barone, L. Leopold, and J. M. Friedman, "Positional cloning of the mouse obese gene and its human homologue," *Nature*, vol. 372, no. 6505, pp. 425–432, 1994.
- [46] J. L. Halaas, K. S. Gajiwala, M. Maffei et al., "Weight-reducing effects of the plasma protein encoded by the obese gene," *Science*, vol. 269, no. 5223, pp. 543–546, 1995.
- [47] J. V. Silha, M. Krsek, J. V. Skrha, P. Sucharda, B. L. G. Nyomba, and L. J. Murphy, "Plasma resistin, adiponectin and leptin levels in lean and obese subjects: correlations with insulin resistance," *European Journal of Endocrinology*, vol. 149, no. 4, pp. 331–335, 2003.
- [48] J. Wang, S. Obici, K. Morgan, N. Barzilay, Z. Feng, and L. Rossetti, "Overfeeding rapidly induces leptin and insulin resistance," *Diabetes*, vol. 50, no. 12, pp. 2786–2791, 2001.
- [49] P. Zimmet, A. Hodge, M. Nicolson et al., "Serum leptin concentration, obesity, and insulin resistance in Western Samoans: Cross Sectional Study," *British Medical Journal*, vol. 313, no. 7063, pp. 965–969, 1996.
- [50] J. W. Kolaczynski, M. R. Nyce, R. V. Considine et al., "Acute and chronic effect of insulin on leptin production in humans: studies in vivo and in vitro," *Diabetes*, vol. 45, no. 5, pp. 699–701, 1996.
- [51] M. Gabriel, S. Jinagouda, R. Boyadjian et al., "Is leptin the link between obesity and insulin resistance?" *Diabetes & Metabolism*, vol. 45, no. 3, pp. 16–24, 1997.
- [52] S. Dagogo-Jack, C. Fanelli, D. Paramore, J. Brothers, and M. Landt, "Plasma leptin and insulin relationships in obese and nonobese humans," *Diabetes*, vol. 45, no. 5, pp. 695–698, 1996.

- [53] R. Lichnovská, S. Gwozdziwiczová, R. Chlup et al., "Serum leptin in the development of insulin resistance and other disorders in the metabolic syndrome," *Biomedical Papers of the Medical Faculty of the University Palacký, Olomouc, Czechoslovakia*, vol. 149, no. 1, pp. 119–126, 2005.
- [54] K. R. Segal, M. Landt, and S. Klein, "Relationship between insulin sensitivity and plasma leptin concentration in lean and obese men," *Diabetes*, vol. 45, no. 3, pp. 988–991, 1996.
- [55] P. Z. Zimmet, V. R. Collins, M. P. De Courten et al., "Is there a relationship between leptin and insulin sensitivity independent of obesity? A population-based study in the Indian Ocean nation of Mauritius," *International Journal of Obesity*, vol. 22, no. 2, pp. 171–177, 1998.
- [56] I. Shimomura, R. E. Hammer, S. Ikemoto, M. S. Brown, and J. L. Goldstein, "Leptin reverses insulin resistance and diabetes mellitus in mice with congenital lipodystrophy," *Nature*, vol. 401, no. 6748, pp. 73–76, 1999.
- [57] J. German, F. Kim, G. J. Schwartz et al., "Hypothalamic leptin signaling regulates hepatic insulin sensitivity via a neurocircuit involving the vagus nerve," *Endocrinology*, vol. 150, no. 10, pp. 4502–4511, 2009.
- [58] C. M. Steppan, S. T. Bailey, S. Bhat et al., "The hormone resistin links obesity to diabetes," *Nature*, vol. 409, no. 6818, pp. 307–312, 2001.
- [59] C. M. Steppan, E. J. Brown, C. M. Wright et al., "A family of tissue-specific resistin-like molecules," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 98, no. 2, pp. 502–506, 2001.
- [60] S. D. Patel, M. W. Rajala, L. Rossetti, P. E. Scherer, and L. Shapiro, "Disulfide-dependent multimeric assembly of Resistin family hormones," *Science*, vol. 304, no. 5674, pp. 1154–1158, 2004.
- [61] K. M. Barnes and J. L. Miner, "Role of resistin in insulin sensitivity in rodents and humans," *Current Protein and Peptide Science*, vol. 10, no. 1, pp. 96–107, 2009.
- [62] S. Ghosh, A. K. Singh, B. Aruna, S. Mukhopadhyay, and N. Z. Ehteshami, "The genomic organization of mouse resistin reveals major differences from the human resistin: functional implications," *Gene*, vol. 305, no. 1, pp. 27–34, 2003.
- [63] E. D. Muse, S. Obici, S. Bhanot et al., "Role of resistin in diet-induced hepatic insulin resistance," *Journal of Clinical Investigation*, vol. 114, no. 2, pp. 232–239, 2004.
- [64] L. Zhou, H. Sell, K. Eckardt, Z. Yang, and J. Eckel, "Conditioned medium obtained from in vitro differentiated adipocytes and resistin induce insulin resistance in human hepatocytes," *FEBS Letters*, vol. 581, no. 22, pp. 4303–4308, 2007.
- [65] C. M. Steppan, S. T. Bailey, S. Bhat et al., "The hormone resistin links obesity to diabetes," *Nature*, vol. 409, no. 6818, pp. 307–312, 2001.
- [66] K. H. Kim, L. Zhao, Y. Moon, C. Kang, and H. S. Sul, "Dominant inhibitory adipocyte-specific secretory factor (ADSF)/resistin enhances adipogenesis and improves insulin sensitivity," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 17, pp. 6780–6785, 2004.
- [67] J. P. Bastard, M. Maachi, J. T. Van Nhieu et al., "Adipose tissue IL-6 content correlates with resistance to insulin activation of glucose uptake both in vivo and in vitro," *Journal of Clinical Endocrinology and Metabolism*, vol. 87, no. 5, pp. 2084–2089, 2002.
- [68] S. J. Van, "Interleukin-6: an overview," *Annual Review of Immunology*, vol. 8, pp. 253–278, 1990.
- [69] S. K. Fried, D. A. Bunkin, and A. S. Greenberg, "Omental and subcutaneous adipose tissues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid," *Journal of Clinical Endocrinology and Metabolism*, vol. 83, no. 3, pp. 847–850, 1998.
- [70] V. Mohamed-Ali, S. Goodrick, A. Rawesh et al., "Subcutaneous adipose tissue releases interleukin-6, but not tumor necrosis factor- α , in vivo," *Journal of Clinical Endocrinology and Metabolism*, vol. 82, no. 12, pp. 4196–4200, 1997.
- [71] V. Vicennati, A. Vottero, C. Friedman, and D. A. Papanicolaou, "Hormonal regulation of interleukin-6 production in human adipocytes," *International Journal of Obesity*, vol. 26, no. 7, pp. 905–911, 2002.
- [72] J. J. Senn, P. J. Klover, I. A. Nowak, and R. A. Mooney, "Interleukin-6 induces cellular insulin resistance in hepatocytes," *Diabetes*, vol. 51, no. 12, pp. 3391–3399, 2002.
- [73] P. J. Klover, T. A. Zimmers, L. G. Koniaris, and R. A. Mooney, "Chronic exposure to interleukin-6 causes hepatic insulin resistance in mice," *Diabetes*, vol. 52, no. 11, pp. 2784–2789, 2003.
- [74] A. L. Carey, G. R. Steinberg, S. L. Macaulay et al., "Interleukin-6 increases insulin-stimulated glucose disposal in humans and glucose uptake and fatty acid oxidation in vitro via AMP-activated protein kinase," *Diabetes*, vol. 55, no. 10, pp. 2688–2697, 2006.
- [75] A. Steensberg, C. P. Fischer, M. Sacchetti et al., "Acute interleukin-6 administration does not impair muscle glucose uptake or whole-body glucose disposal in healthy humans," *Journal of Physiology*, vol. 548, no. 2, pp. 631–638, 2003.
- [76] J. J. Senn, P. J. Klover, I. A. Nowak et al., "Suppressor of cytokine signaling-3 (SOCS-3), a potential mediator of interleukin-6-dependent insulin resistance in hepatocytes," *Journal of Biological Chemistry*, vol. 278, no. 16, pp. 13740–13746, 2003.
- [77] S. E. Borst, "The role of TNF- α in insulin resistance," *Endocrine*, vol. 23, no. 2-3, pp. 177–182, 2004.
- [78] S. Gwozdziwiczova, R. Lichnovska, R. B. Yahia et al., "TNF α in the development of insulin resistance and other disorders in metabolic syndrome," *Biomedical Papers of the Medical Faculty of the University Palacký, Olomouc, Czechoslovakia*, vol. 149, no. 1, pp. 109–117, 2005.
- [79] E. P. Araújo, C. T. De Souza, M. Ueno et al., "Infliximab restores glucose homeostasis in an animal model of diet induced obesity and diabetes," *Endocrinology*, vol. 148, no. 12, pp. 5991–5997, 2007.
- [80] F. C. Huvers, C. Popa, M. G. Netea, F. H. J. Van Den Hoogen, and C. J. Tack, "Improved insulin sensitivity by anti-TNF α antibody treatment in patients with rheumatic diseases," *Annals of the Rheumatic Diseases*, vol. 66, no. 4, pp. 558–559, 2007.
- [81] N. Togashi, N. Ura, K. Higashiura, H. Murakami, and K. Shimamoto, "Effect of TNF- α -converting enzyme inhibitor on insulin resistance in fructose-fed rats," *Hypertension*, vol. 39, no. 2, pp. 578–580, 2002.
- [82] B. M. De Taeye, T. Novitskaya, O. P. McGuinness et al., "Macrophage TNF- α contributes to insulin resistance and hepatic steatosis in diet-induced obesity," *American Journal of Physiology—Endocrinology and Metabolism*, vol. 293, no. 3, pp. E713–E725, 2007.
- [83] K. T. Uysal, S. M. Wiesbrock, M. W. Marino, and G. S. Hotamisligil, "Protection from obesity-induced insulin resistance in mice lacking TNF- α function," *Nature*, vol. 389, no. 6651, pp. 610–614, 1997.

- [84] J. J. Swaroop, D. Rajarajeswari, and J. N. Naidu, "Association of TNF- α , with insulin resistance in type 2 diabetes mellitus," *Indian Journal of Medical Research*, vol. 135, pp. 127–130, 2012.
- [85] T. Bastuk, A. Unsal, T. Ulas et al., "Effects of rosiglitazone treatment on insulin resistance and TNF- α levels in patients with chronic kidney disease: A Prospective Study," *European Review for Medical and Pharmacological Sciences*, vol. 16, no. 11, pp. 1519–1524, 2012.
- [86] P. D. G. Miles, O. M. Romeo, K. Higo, A. Cohen, K. Rafaat, and J. M. Olefsky, "TNF- α -induced insulin resistance in vivo and its prevention by troglitazone," *Diabetes*, vol. 46, no. 11, pp. 1678–1683, 1997.
- [87] C. Rask-Madsen, H. Domínguez, N. Ihlemann, T. Hermann, L. Køber, and C. Torp-Pedersen, "Tumor necrosis factor- α inhibits insulin's stimulating effect on glucose uptake and endothelium-dependent vasodilation in humans," *Circulation*, vol. 108, no. 15, pp. 1815–1821, 2003.
- [88] J. P. Kirwan, S. Hauguel-De Mouzon, J. Lepercq et al., "TNF- α is a predictor of insulin resistance in human pregnancy," *Diabetes*, vol. 51, no. 7, pp. 2207–2213, 2002.
- [89] K. T. Uysal, S. M. Wiesbrock, M. W. Marino, and G. S. Hotamisligil, "Protection from obesity-induced insulin resistance in mice lacking TNF- α function," *Nature*, vol. 389, no. 6651, pp. 610–614, 1997.
- [90] L. E. Bernstein, J. Berry, S. Kim et al., "Effects of etanercept in patients with the metabolic syndrome," *Archives of Internal Medicine*, vol. 166, no. 8, pp. 902–908, 2006.
- [91] H. Dominguez, H. Storgaard, M. C. Rask et al., "Metabolic and vascular effects of tumor necrosis factor- α blockade with etanercept in obese patients with type 2 diabetes," *Journal of Value Resonance*, vol. 42, no. 6, pp. 517–525, 2005.
- [92] F. Ofei, S. Hurel, J. Newkirk, M. Sopwith, and R. Taylor, "Effects of an engineered human anti-TNF- α antibody (CDP571) on insulin sensitivity and glycemic control in patients with NIDDM," *Diabetes*, vol. 45, no. 3, pp. 881–885, 1996.
- [93] S. Fernández-Veledo, I. Nieto-Vazquez, C. M. Rondinone, and M. Lorenzo, "Liver X receptor agonists ameliorate TNF α -induced insulin resistance in murine brown adipocytes by downregulating protein tyrosine phosphatase-1B gene expression," *Diabetologia*, vol. 49, no. 12, pp. 3038–3048, 2006.
- [94] G. S. Hotamisligil, P. Arner, J. F. Caro, R. L. Atkinson, and B. M. Spiegelman, "Increased adipose tissue expression of tumor necrosis factor- α in human obesity and insulin resistance," *Journal of Clinical Investigation*, vol. 95, no. 5, pp. 2409–2415, 1995.
- [95] T. C. Wascher, J. H. N. Lindeman, H. Sourij, T. Kooistra, G. Pacini, and M. Roden, "Chronic TNF- α neutralization does not improve insulin resistance or endothelial function in "healthy" men with metabolic syndrome," *Molecular Medicine*, vol. 17, no. 3–4, pp. 189–193, 2011.
- [96] J. P. Bastard, M. Maachi, C. Lagathu et al., "Recent advances in the relationship between obesity, inflammation, and insulin resistance," *European Cytokine Network*, vol. 17, no. 1, pp. 4–12, 2006.
- [97] P. Gual, Y. Le Marchand-Brustel, and J. F. Tanti, "Positive and negative regulation of insulin signaling through IRS-1 phosphorylation," *Biochimie*, vol. 87, no. 1, pp. 99–109, 2005.
- [98] J. A. Olivares-Reyes, A. Arellano-Plancarte, and J. R. Castillo-Hernandez, "Angiotensin II and the development of insulin resistance: implications for diabetes," *Molecular and Cellular Endocrinology*, vol. 302, no. 2, pp. 128–139, 2009.
- [99] J. M. Tishinsky, L. E. Robinson, and D. J. Dyck, "Insulin-sensitizing properties of adiponectin," *Biochimie*, vol. 94, no. 10, pp. 2131–2136, 2012.
- [100] C. Buechler, J. Wanninger, and M. Neumeier, "Adiponectin, a key adipokine in obesity related liver diseases," *World Journal of Gastroenterology*, vol. 17, no. 23, pp. 2801–2811, 2011.