The APOE∗3-Leiden Heterozygous Glucokinase Knockout Mouse as Novel Translational Disease Model for Type 2 Diabetes, Dyslipidemia, and Diabetic Atherosclerosis

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Background. There is a lack of predictive preclinical animal models combining atherosclerosis and type 2 diabetes. APOE∗3-Leiden (E3L) mice are a well-established model for diet-induced hyperlipidemia and atherosclerosis, and glucokinase+/− (GK+/−) mice are a translatable disease model for glucose control in type 2 diabetes. The respective mice respond similarly to lipid-lowering and antidiabetic drugs as humans. The objective of this study was to evaluate/characterize the APOE∗3-Leiden.glucokinase+/− (E3L.GK+/−) mouse as a novel disease model to study the metabolic syndrome and diabetic complications. Methods. Female E3L.GK+/−, E3L, and GK+/− mice were fed fat- and cholesterol-containing diets for 37 weeks, and plasma parameters were measured throughout. Development of diabetic macro- and microvascular complications was evaluated. Results. Cholesterol and triglyceride levels were significantly elevated in E3L and E3L.GK+/− mice compared to GK+/− mice, whereas fasting glucose was significantly increased in E3L.GK+/− and GK+/− mice compared to E3L. Atherosclerotic lesion size was increased 2.2-fold in E3L.GK+/− mice as compared to E3L (p = 0.037), which was predicted by glucose exposure (R² = 0.636, p = 0.001). E3L and E3L.GK+/− mice developed NASH with severe inflammation and fibrosis which, however, was not altered by introduction of the defective GK phenotype, whereas mild kidney pathology with tubular vacuolization was present in all three phenotypes. Conclusions. We conclude that the E3L.GK+/− mouse is a promising novel diet-inducible disease model for investigation of the etiology and evaluation of drug treatment on diabetic atherosclerosis.

1. Introduction

The metabolic syndrome consists of a cluster of cardiovascular risk factors, including abdominal obesity, elevated blood pressure, elevated fasting plasma glucose, high serum triglycerides, and low high-density lipoprotein (HDL) levels, and drives the global epidemics of type 2 diabetes (T2D) and cardiovascular disease (CVD). Diabetes increases the CVD risk about twofold [1–3], which is the leading cause of death worldwide, and aggravates nonalcoholic steatohepatitis (NASH) [4] and diabetic nephropathy [5]. These comorbidities emphasize the need for antidiabetic treatments that...
are effective against both T2D and associated cardiovascular complications.

Animal models can be used to learn more about the underlying pathology of diabetic complications and the effect of pharmacological interventions thereon, and a wide range of mouse models combining atherosclerosis and diabetes are described [6]. Most available models are dyslipidemic mice, e.g., ApoE−/− and LDLr−/− mice, with chemically (STZ) or genetically (ob/ob, db/db, and IRS2−/−) induced diabetes [6]. Although these models are widely used in biomedical research and drug development, they do not sufficiently reflect human disease. First, deficiency of the apoe or Idlr gene and STZ treatment result in extreme hyperlipidemia and hyperglycemia, respectively, and may result in overestimation of the contribution of hyperglycemia to diabetic complications. Besides, STZ treatment is difficult to control and creates a type 1 diabetic-like condition. Second, commonly used animal models of T2D (ob/ob and db/db mice) have a wide but unstable hyperglycemic range [7, 8] and are monogenic models of obesity thereby inducing hyperglycemia, which weakens their translatability value as obesity is seldom caused by a monogenic mutation [7, 9]. Last, ApoE−/− and LDLr−/− mice do not respond well to lipid-lowering drugs used in the clinic [10, 11], making these models unsuitable in the development of novel therapeutic strategies against hyperlipidemia and vascular complications.

The objective of this study was to develop a translational mouse model for the metabolic syndrome and diabetic complications by combining diet-induced dyslipidemia and hyperglycemia, with plasma levels translatable to the human complications by combining diet-induced dyslipidemia and hyperlipidemia and vascular complications.

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2. Materials and Methods

2.1. Animals and Breeding. 10–23-week-old female E3L, GK−/−, and E3LGK+−/− mice (n = 6–10/sex/genotype) were used in the study. Both E3L and GK−/− mice are bred on a C57BL/6J background. Since homozygous E3L mice are not viable in utero, these mice are bred heterozygously by breeding E3L X C57BL/6J. GK−/− mice are bred heterozygously (GK+−/− X C57bl/6J) as described previously [26], because the homozygous deletion of GK is postnatally lethal. E3LGK−/− mice were generated by cross-breeding E3L mice with GK−/− mice, thereby generating 27 to 40% offspring of each genotype. Mice were crossed once and were not backcrossed. E3L mice are huApoE3Leiden-huApoC1 double transgenic mice, with both genes located on one genomic DNA construct [12], and therefore, the presence of the E3L phenotype was evaluated by genotyping for ApoC1. The presence of the GK−/− phenotype was evaluated by qPCR as described previously [26]. Females were used because E3L females are more responsive to dietary cholesterol and fat than males. E3L females have a higher VLDL production than males [28] resulting in higher plasma total cholesterol (TC) and triglyceride (TG) levels and development of atherosclerosis [12, 29]. All mice were housed under standard conditions with a 12 h light-dark cycle and had free access to food and water. Body weight was monitored regularly during the study. Animal experiments were approved by the Regional Animal Ethics Committee for Experimental Animals, Göteborg University. All in vivo activities were carried out conforming to the Swedish Animal Welfare Act and regulations SJVFS 2012: 26.

2.2. Experimental Design and Analyses. First, mice were fed a semisynthetic diet, containing saturated fat with 15% (w/w) cacao butter (Western-type diet (WTD); Hope Farms, Woerden, the Netherlands) and 0.15% cholesterol for 7 weeks to study the effect of a mildly lipid-elevating diet on plasma lipid and glucose levels. Subsequently, this diet was supplemented with 10% glucose in the drinking water in weeks 6–7 to investigate whether dietary glucose did modulate these plasma levels. During the following 30 weeks, mice were fed WTD+1.0% cholesterol to induce atherosclerosis [20] (Figure 1). EDTA blood samples were drawn after a 4-hour fast, and plasma parameters were evaluated at different time points throughout the study. The last blood sample was drawn at week 36, and all animals were sacrificed by CO2 inhalation at week 37. Plasma cholesterol, triglycerides, glucose, and insulin were measured throughout the study, and
HbA1c was measured at week 36. Total cholesterol and glucose exposure was calculated by adding up for all intervals the products of the mean cholesterol or glucose level during that interval times the duration of that interval and expressed as mmol/L*weeks. Lipoprotein profiles, alanine transaminase (ALT), and aspartate transaminase (AST) were measured in groupwise-pooled unfasted sacrifice plasma. Urinary albumin/creatinine levels were measured in spot urine collected in week 36. Hepatic lipid content was analyzed in homogenized, snap-frozen liver samples as described previously [30]. Heart and aorta, liver, and kidneys were collected for histopathological analysis of atherosclerosis, NAFLD/NASH and liver fibrosis, and diabetic nephropathy.

2.3. Statistical Analysis. The E3L.GK+/− phenotype was compared to E3L and GK+/+, and significance of differences was calculated parametrically using a one-way ANOVA with Dunnett’s post hoc test. Differences in plasma parameters between the different time points were calculated for each genotype using a one-way ANOVA with a Bonferroni post hoc test. Significance of differences between the E3LGK+/− and E3L mice in atherosclerotic lesion number, severity, and composition was calculated using an independent sample t-test. A multiple regression analysis was performed to predict the effect of variables on lesion size, and linear regression was used to assess correlations between variables. SPSS 22.0 for Windows was used for statistical analysis. Values are presented as means ± SD. All reported p values < 0.05 were considered statistically significant.

For a more detailed description of the applied methods, please see Supplementary Materials online.

3. Results

3.1. Safety Aspects. No clinical signs of deviant behavior were noted in any of the phenotypes. From weeks 0 to 36, all three phenotypes gained 5 ± 2 gram body weight (Suppl. Table I). Plasma pooled per group showed lower AST and ALT values as markers of hepatocellular damage in GK+/− mice when compared to E3LGK+/− and E3L (Suppl. Table I). One mouse was terminated during the study based on human end point criteria.

3.1.1. Plasma Parameters for Metabolic Disease and Response to Diets

(1) E3LGK+/− Mice Are Hyperlipidemic and Hyperglycemic. Plasma cholesterol and triglyceride levels in the E3LGK+/− mice were similar to the E3L mice and increased by 540% (cholesterol) and 450% (triglycerides) when compared to the GK+/− mice (Figures 2(a) and 2(b)), resulting in a significantly increased cholesterol exposure (mmol/L*weeks) (+316%, p < 0.001) (Figure 2(d)). Cholesterol in the E3L and E3LGK+/− mice was mainly contained in VLDL and LDL, and that in GK+/− in HDL (Figure 2(c)). Glucose levels were significantly elevated at all time points except at T = 4 weeks when compared to the E3L mice (Figure 2(e)). Total glucose exposure (mmol/L*weeks) was 429 ± 60, 299 ± 14, and 492 ± 52 mmol/L for E3LGK+/−, E3L, and GK+/+, respectively, and significantly increased in E3LGK+/− when compared to E3L mice (p < 0.001) (Figure 2(h)). Insulin levels did not differ between the strains (Figure 2(f)). HbA1c was increased by 17% when compared to the E3L mice (p = 0.005) (Figure 2(g)). In conclusion, these data show that E3LGK+/− combines both adverse phenotypes with increased lipid levels as in the E3L mice and mildly elevated glucose levels as in the GK+/− mice.

(2) Plasma Cholesterol Levels Are Modulated by the Diet in E3LGK+/− and E3L Mice. Different diets were used in this study to evaluate the response of the mouse model to dietary interventions. Plasma cholesterol, but not triglycerides, increased in both the E3LGK+/− and E3L mice when switched from a chow diet (T = 0 weeks) to a WTD with 0.15% cholesterol added (+143%, p = 0.038; +173%, p = 0.001), whereas plasma lipid levels were not affected in the GK+/− mice (Suppl. Table II). Plasma glucose and insulin levels were not affected by the WTD with 0.15% cholesterol added, except for glucose which increased in the E3L mice (+26%, p = 0.010). Adding 10% glucose to the drinking water further increased plasma cholesterol levels: when compared to T = 0 weeks (chow), cholesterol levels increased by 215% in the E3LGK+/− mice (p = 0.001) and by 224% in the E3L mice (p < 0.001). However, this increase was not significant when compared to T = 4 (WTD with 0.15% cholesterol) (Suppl. Table II). Increasing the amount of cholesterol in the diet to 1.0% further increased plasma cholesterol levels in the E3LGK+/− and E3L mice when compared to T = 0 and when compared to T = 8 (+89%, p = 0.001; +43%, p = 0.013). Insulin levels dropped in the E3L mice at T = 36 weeks when compared to T = 0 weeks (−60%, p = 0.020) and T = 8 weeks (−66%, p = 0.010), whereas this effect was less pronounced in the GK+/− mice (−41%, p = 0.081 compared to T = 8 weeks) and absent in the E3LGK+/− mice. Interestingly, plasma glucose levels in
the E3L.GK+/− and GK+/− mice were not modulated by glucose in the drinking water, indicating that despite reduced glucokinase activity [26], the mice maintain their glucose homeostasis at an increased glucose supply. Altogether, these data show that plasma lipids can be modulated in the E3L.GK+/− mouse model, as in the E3L mice, whereas the elevated glucose levels on chow are not further increased by these dietary interventions.

Figure 2: E3L.GK+/− mice have comparable lipid levels and higher glucose levels as E3L mice. Plasma cholesterol (a) and triglycerides (b) were measured throughout the study. Lipoprotein profiles were assessed by FPLC lipoprotein separation in groupwise-pooled unfasted sacrifice plasma (c). Cholesterol exposure over time was calculated as mmol/L * weeks (d). Plasma glucose (e) and insulin (f) were measured throughout, HbA1c (%) was measured at week 36 (g), and glucose exposure was calculated as mmol/L * weeks (h). Significance of differences was calculated parametrically using a one-way ANOVA with Dunnett’s post hoc test. E3L.GK+/− compared to E3L: *p < 0.05, **p < 0.01, and ***p < 0.001; E3L.GK+/− compared to GK+/−: †p < 0.05, ††p < 0.01, and †††p < 0.001. Data are presented as means ± SD (n = 8–10 per group and for insulin n = 4–8 per group). FPLC: fast protein liquid chromatography; HbA1c: hemoglobine A1c.
3.1.2. Diabetic Macro- and Microvascular Complications in E3L.GK<sup>+/−</sup> Mice

(1) Atherosclerotic Lesion Size and Severity Are Aggravated in E3L.GK<sup>+/−</sup> Mice. One of the most important diabetic complications is increased risk for CVD [1–3], and therefore, we assessed atherosclerotic lesion size, lesion severity, and plaque phenotype, as markers of vulnerability to rupture, in the aortic root. E3L mice developed 0.4 ± 0.5 mild (1–II), 3.6 ± 2.3 moderate (III), and 1.6 ± 1.8 severe (IV–V) lesions per cross-section. The number of severe lesions was significantly increased in the E3L.GK<sup>+/−</sup> mice (2.8-fold; p = 0.038) (Figure 3(a)). When lesion severity was depicted as the percentage of total plaque area that consisted of mild or severe lesions, there was no difference between the E3L and E3L.GK<sup>+/−</sup> mice (Figure 3(b)). However, the total atherosclerotic lesion size was significantly increased by 2.2-fold in the E3L.GK<sup>+/−</sup> mice (68 ± 42 * 1000 μm<sup>2</sup>) as compared to E3L (32 ± 29 * 1000 μm<sup>2</sup>) (p = 0.037) (Figure 3(c)). There were no lesions visible in the GK<sup>+/−</sup> mice (Figure 3(c)). The plaque composition was analyzed in the type III–V lesions, as illustrated by representative images in Figure 4. There were no significant differences between the E3L.GK<sup>+/−</sup> and E3L mice in plaque composition (Figure 3(d)), plaque stability index, or monocyte adherence to the endothelium (data not shown). Collectively, these data show that atherosclerotic lesion size is aggravated in E3L.GK<sup>+/−</sup> as compared to the E3L mice without affecting plaque composition and monocyte adherence.

(2) Elevated Plasma Glucose Levels Contribute to the Increased Development of Atherosclerosis in E3L.GK<sup>+/−</sup> Mice. To explore the contribution of the elevated plasma glucose levels to the increased lesion size, a multiple regression analysis was performed with cholesterol exposure and glucose exposure as covariates after square root transformation of the lesion area. Lesion size was predicted only by glucose exposure as covariates after square root transformation of plasma glucose exposure and lesion area. Lesion size was predicted only by glucose exposure as covariates after square root transformation of plasma glucose exposure and lesion area. Lesion size was predicted only by glucose exposure as covariates after square root transformation of plasma glucose exposure and lesion area.

(3) The GK<sup>+/−</sup> Phenotype Does Not Aggravate Hepatic Steatosis, Inflammation, or Fibrosis. NAFLD/NASH is strongly associated with the metabolic syndrome and type 2 diabetes [31, 32]. To assess whether the GK<sup>+/−</sup> phenotype worsens the development of NASH, liver sections were examined for hepatic steatosis, inflammation, and fibrosis, and liver lipid content was measured. Hepatic macrosteatosis did not differ between the phenotypes (Figure 5(a)), whereas hepatic microsteatosis was significantly elevated by 2.7-fold (p = 0.003) in the E3L.GK<sup>+/−</sup> mice when compared to GK<sup>+/−</sup> (Figure 5(b)), and both E3L.GK<sup>+/−</sup> and E3L had severe liver inflammation which was increased by 6.8-fold (p < 0.001) in E3L.GK<sup>+/−</sup> relative to GK<sup>+/−</sup> (Figure 5(c)). Furthermore, mean fibrosis stage in E3L.GK<sup>+/−</sup> was significantly elevated when compared to GK<sup>+/−</sup> (2.3-fold, p < 0.001) (Figure 5(d)), as well as the percentage Sirius Red-positive area of total liver area (6.1-fold, p = 0.011) (Figure 5(e)). Liver lipids did not differ between the phenotypes (Figures 5(f)–5(h)). Representative images are shown (Figures 5(i)–5(n)). Collectively, these data show that the E3L and E3L.GK<sup>+/−</sup> mice, but not GK<sup>+/−</sup>, develop NASH with severe inflammation and fibrosis, which is not worsened by increased glucose levels. This indicates a dominant role for the combination of the E3L phenotype and dietary cholesterol in the progression of NASH and liver fibrosis.

(4) Mild Kidney Pathology Is Present in All Three Phenotypes. Diabetic nephropathy is becoming an increasingly important cause of morbidity and mortality worldwide and is related to the increasing prevalence of type 2 diabetes. Therefore, kidneys were analyzed for the presence of renal damage focusing on glomerular damage, including mesangial matrix expansion, and tubulointerstitial damage, including interstitial inflammation, fibrosis, and tubular abnormalities, as central causes for loss of kidney function. Nephrin staining was performed to study renal filtration barrier function. There were no differences in inflammation, fibrosis (data not shown), mesangial matrix expansion (Figure 6(a)), or nephrin score (Figure 6(b)) between the phenotypes. Abnormal tubular structures were observed in all three phenotypes but were most pronounced in the GK<sup>+/−</sup> mice, wherein the tubuli showed vacuolization (Figure 6(c)). The pathological changes did not affect permeability in the glomerulus, as measured by the urinary albumin:creatinine ratio (Suppl. Table I). Altogether, we can conclude that mild pathological changes are present, which are not aggravated in the E3L.GK<sup>+/−</sup> mice.

4. Discussion

In the present study, we evaluated the E3L.GK<sup>+/−</sup> mouse as an animal model for diet-induced hyperlipidemia and hyperglycemia and the pathological consequences thereof. We showed that plasma lipids can be titrated to desired and for human relevant levels by adding cholesterol and fat to the diet and that these levels remain stable for a long period (up to 37 weeks). In addition, the E3L.GK<sup>+/−</sup> mice were mildly hyperglycemic and developed more atherosclerosis than the E3L mice, which was related to the higher glucose levels in the E3L.GK<sup>+/−</sup> mice. The E3L and E3L.GK<sup>+/−</sup> mice both developed hepatic steatosis with severe inflammation and fibrosis, which, however, was not altered by introduction of the defective GK phenotype, whereas only mild kidney pathology with tubular vacuolization was present in all three phenotypes.

Translatability of animal models is essential when investigating the pathogenesis of diabetic complications and evaluating drug treatment thereon. Plasma cholesterol and glucose levels in the diet-induced E3L.GK<sup>+/−</sup> mouse model were similar to levels in patients with increased cardiovascular risk [2, 33]. Partial deletion of the Gk gene in the E3L mice did not affect the response of plasma lipids to dietary modulation, and in both the E3L.GK<sup>+/−</sup> and E3L mice, plasma cholesterol levels raised similarly upon feeding a WTD with increasing amounts of cholesterol. Interestingly, glucose
and insulin levels were not affected by the diet but remained stable representing mild hyperglycemia in the E3L.GK+/− and GK+/− mice (10.4 ± 1.4 mmol/L and 14.1 ± 2.6 mmol/L at the end point, respectively). In contrast, glucose levels in male GK+/− mice increase over time on a high-fat diet with plasma levels reaching 18.9 ± 1.0 mmol/L and impaired glucose tolerance [8, 26]. This gender difference may be explained by the C57BL/6J background of the E3L and GK+/− transgenic mice. Upon a high-fat diet, insulin and glucose levels increase over time in C57BL/6J males, consistent with
insulin resistance and glucose intolerance, whereas C57BL/6J females have normal serum insulin concentrations and glucose levels remain constant [34]. Estrogens affect different metabolic pathways in the glucose hemostasis [35], thereby protecting against the risk of developing type 2 diabetes in both premenopausal women [36] and mice [35].

We observed a markedly increased atherosclerotic lesion size in E3L.GK+/− as compared to the E3L mice which was highly significantly correlated with glucose exposure ($R^2 = 0.636, p = 0.001$), suggesting a proatherogenic role of glucose in the development of atherosclerosis. Indeed, it is known that prolonged exposure to hyperglycemia negatively affects the endothelium, vascular smooth muscle cells, and macrophages, and it increases thrombosis while impairing fibrinolysis, leading to formation of atherosclerotic plaques [37]. This may explain the association of diabetes type

![Figure 4: Plaque composition in a severe plaque of an E3L and E3L.GK+/− mouse. Representative images of HPS staining, Movat staining, Sirius Red staining for collagen, immunostaining with α-actin for SMCs, and immunostaining with Mac-3 for macrophages. The arrows depict necrotic areas, including cholesterol clefts. HPS: hematoxylin-phloxine-saffron; SMCs: smooth muscle cells.](image-url)
Figure 5: The GK phenotype does not aggravate hepatic steatosis, inflammation, or fibrosis in E3L mice. Macropalvic steatosis (a) and microvesicular steatosis (b) as percentage of total liver area was determined. The number of inflammatory cell aggregates were counted per mm² (c). Fibrosis grade (1–4) was scored (d) and percentage of area positive for Sirius Red was measured in ImageJ (e). Intrahepatic free cholesterol (f), intrahepatic triglycerides (g), and intrahepatic cholesterol esters (h) were analyzed by HPTLC. Representative images of HE (i–k) and Sirius Red (l–n) staining at a 5x magnification. Significance of differences was calculated parametrically using a one-way ANOVA with Dunnett’s post hoc test. °p < 0.05, ††p < 0.01, and †††p < 0.001 when compared to GK+/−. Data are presented as means ± SD (n = 6–10 per group). HPTLC: high-performance thin-layer chromatography; HE: hematoxylin-eosin.
are no reports in the GK+/− mice, glucokinase deficiency in humans does not aggravate blood pressure [25].

Previously, the GK+/−ApoE−/− mouse model has been developed as a model combining hyperlipidemia and hyperglycemia, which had impaired glucose tolerance and a minimal increase of atherosclerosis relative to ApoE−/− mice [47]. A disadvantage of this model is the ApoE−/− background. ApoE−/− mice are, like LDLr−/− mice, a severe model for hyperlipidemia, and due to the absence of a functional apoE-LDLr-mediated clearance pathway, these mice do not respond well to lipid-lowering drugs (e.g., statins [10], PCSK9 inhibitors [11]) and therefore cannot be used for the evaluation of combination treatment. In contrast, the E3L mice are very suited to study lipoprotein metabolism and lipid modulation [10, 48].

In Figure 7, we give an overview of all registered cholesterol- and glucose-lowering drugs that have been evaluated in the E3L and GK+/− mice, respectively. The E3L mice respond similarly as humans do to lipid-lowering agents, including statins, fibrates, niacin, and PCSK9 inhibitors [11, 14−22], whereas glucose levels are successfully reduced in the GK+/− mice by standard therapeutic agents as insulin, metformin, exendin-4, and GSK9a at doses corresponding to therapeutic drug levels in man [8, 27]. Although these interventions have not been assessed in the E3L.GK+/− mice yet, we carefully speculate about the effects and discuss how the model can be of value for future research. As the E3L.GK+/− mice have similar lipid and glucose levels as their parent models and respond in a similar way to dietary modulations, we propose that both lipid- and glucose-lowering agents will be effective in the combined model. Also, we propose that the E3L.GK+/− mice can be used to examine interactions between glucose and lipid metabolism, e.g., how statin treatment increases the risk of diabetes incidence [49]. Last, atherosclerosis development and cardiovascular safety can be evaluated in the E3L.GK+/− model, which is especially interesting regarding the currently unknown mechanisms by which glucose-lowering agents (e.g., empagliflozin, liraglutide, and semaglutide) improve CV outcome [50−52].

5. Conclusion

Altogether, we conclude that the E3LGK+/− mouse is a promising translatable diet-inducible model, combining
dyslipidemia and hyperglycemia with human-like plasma cholesterol and glucose levels and aggravated atherosclerosis, to study the etiology of diabetic atherosclerosis and for the evaluation of lipid-lowering and antidiabetic drugs and their combination thereon.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Disclosure

An abstract of the presentation of the manuscript has been published in Atherosclerosis Abstracts [53].

Conflicts of Interest

JWJ received research grants from and was speaker on (CME-accredited) meetings sponsored by Amgen, Astellas, AstraZeneca, Daiichi Sankyo, Lilly, Merck-Schering-Plough, Pfizer, Roche, Sanofi-Aventis, the Netherlands Heart Foundation, the Interuniversity Cardiology Institute of the Netherlands, and the European Community Framework KP7 Program. MB, ACA, and ACJR are employees of AstraZeneca and SEH and BL were employees of AstraZeneca during the execution of the study. MGP, AvK, ALM, EJP, AMvdH, and HMGP have nothing to disclose.

Authors’ Contributions

Marianne G. Pouwer and Suvi E. Heinonen contributed equally to this work.
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Supplementary Materials

The Supplementary Materials consist of (1) a detailed description of the applied methods and (2) 2 tables: Table I and Table II. Suppl. Table I: biochemical parameters in E3LGk+/-, E3L, and Gk+/- mice. All depicted parameters are measured at week 36, except for liver weight (week 37). Suppl. Table II: plasma cholesterol levels are modulated by the diet in E3LGk+/- and E3L mice. The response of plasma lipids, glucose, and insulin to the different diets was evaluated. (Supplementary Materials)

References


