Clinical Study

Unexpected High Response Rate to Traditional Therapy after Dendritic Cell-Based Vaccine in Advanced Melanoma: Update of Clinical Outcome and Subgroup Analysis

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We reviewed the clinical results of a dendritic cell-based phase II clinical vaccine trial in stage IV melanoma and analyzed a patient subgroup treated with standard therapies after stopping vaccination. From 2003 to 2009, 24 metastatic melanoma patients were treated with mature dendritic cells pulsed with autologous tumor lysate and keyhole limpet hemocyanin and low-dose interleukin-2. Overall response (OR) to vaccination was 37.5% with a clinical benefit of 54.1%. All 14 responders showed delayed type hypersensitivity positivity. Median overall survival (OS) was 15 months (95% CI, 8–33). Eleven patients underwent other treatments (3 surgery, 2 biotherapy, 2 radiotherapy, 2 chemotherapy, and 4 biochemotherapy) after stopping vaccination. Of these, 2 patients had a complete response and 5 a partial response, with an OR of 63.6%. Median OS was 34 months (range 16–61). Our results suggest that therapeutic DC vaccination could favor clinical response in patients after more than one line of therapy.

1. Introduction

Melanoma accounts for 1%–3% of all malignant tumors and its incidence is increasing in western countries by 6%–7% each year [1–3]. The disease is curable in more than 50% of cases with surgical resection, with an expected 5-year survival of 80%–100% [2]. However, prognosis is poor for patients with advanced disease, with a 5-year life expectancy of <10% and a median survival of 6–8.5 months [4]. Chemotherapy for advanced melanoma continues to be highly unsatisfactory [5, 6]. Dacarbazine (DTIC) is still considered the gold standard of therapy, despite the fact that it has shown “placebo” results in recent phase III trials, obtaining <10% overall response (OR) rates, with an overall survival (OS) of around 6 months [7–9]. Different monochemotherapies and polychemotherapeutic associations, with or without the addition of biological response modifiers, have not proven to be more effective than DTIC [10, 11]. There are no standard second- or third-line therapies.

The relation between the immune system and the tumor is undoubtedly a complex one. The key ability to distinguish between self and non-self is essential for an adequate response to external pathogens and growing tumor cells. Basic research has identified a number of mechanisms underlying spontaneous antitumor immunity and enabled Dunn to formulate the cancer immunoediting hypothesis,
which divides the tumor immune response into three phases: elimination, equilibrium, and escape [13, 14].

Both innate and adaptive immunity are involved in the antitumor immune response [15, 16]. In particular, dendritic cells (DCs) play a crucial role in the interplay between innate and adaptive response towards cancer [17]. As members of the innate immune system, their main function is to present antigens to regulate the activation of the adaptive response. DCs can therefore provide signals of both immunostimulation and tolerance to antigen-specific T lymphocytes, thus determining the T response (Th1/Th2) which depends on the activation status of the DCs at the time of the interaction [18, 19]. There is a convincing rationale for the use of DC-based vaccine. In their review of clinical vaccination trials, Rosenberg et al. observed the highest response rate among studies using dendritic cells (7.1%) [20]. Engell-Noerregaard et al. reviewed 38 publications on clinical trials (from 1996 to 2007) using DC-based vaccination in advanced melanoma. The authors reported an objective response rate of 9% with a clinical benefit of 30% (CR+PR+SD) and 21% stable disease. Although a trend was observed between immunological response and overall survival, no definitive conclusions were drawn [21].

A combination of immunotherapy with standard treatments (chemotherapy, radiotherapy, and surgery) for cancer is an emerging challenge and an emerging paradigm, in contrast to the concept that defines most standard treatments as immunosuppressive. Examination of combined treatments has yielded unexpected results. Antonia et al. reported a very strong objective response to second-line chemotherapy in nonsmall cell lung cancer patients pretreated with vaccination consisting of dendritic cell (DC) transduced with the full-length wild-type p53. This vaccination was started 8 weeks after completion of first-line chemotherapy [22]. A similar observation was made in patients with follicular B-cell lymphoma vaccinated with an anti-idiotype vaccine while in remission. At disease recurrence, patients were treated with second-line chemotherapy (CHOP schedule) obtaining a much higher partial or complete remission than those expected for this disease [23]. Similarly, Gribben et al. reported unexpected high response rates to salvage therapies after vaccination with universal tumor antigen CYP1B1 in solid tumors in a phase 1 trial. Five vaccinated patients who developed immunity to the vaccine had a marked objective response to subsequent therapies [24]. In a review of 3 prostate cancer vaccine trials, researchers underlined that vaccinated patients responded better to subsequent chemotherapy than those who had not been vaccinated [25]. The mechanisms responsible for such results remain unknown, although some data have been published on the effect of gemcitabine on myeloid-derived suppressor cells (MDSC) and on the activity of paclitaxel, which binds toll-like receptors (TLRs) to dendritic cells and induces the production of patterns typical of T-helper type 1 [26].

On the basis of the above, we reviewed and updated the clinical results of a dendritic cell-based clinical vaccine trial in stage IV melanoma patients [27], focusing on a subgroup of 11 patients who underwent other therapies after vaccination.

2. Patients and Methods

2.1. Patient Population. From June 2003 to December 2007, 24 patients with metastatic melanoma were treated with mature-DCs (mDCs) (10 M/vaccine) pulsed with autologous tumor lysate (ATL) and keyhole limpet hemocyanin (KLH). All but 3 of the patients had been heavily pretreated before vaccination. Patients were vaccinated intradermally with mDCs at the base of the thigh about 10 cm from the groin. Interleukin-2 (IL-2) was administered subcutaneously at a dose of 3 MIU/die on days 3, 4, 5, 6, and 7. The procedure was repeated every 2 weeks for four cycles and monthly thereafter until the lysate was finished or evident progression occurred (symptomatic progression or worsening of clinical conditions with a PS > 2 and absence of signs of immunostimulation). Disease evaluation and immunomonitoring in vivo with delayed-type hypersensitivity (DTH) for both ATL and KLH were performed before the first vaccination and every four cycles thereafter. The protocol was approved by the Local Ethics Committee of Forlì Health and Social Services (Azienda USL di Forlì) in 1999.

Prevaccination treatment was as follows: radiotherapy and radical surgery for brain metastases (1 patient), leg perfusion (1), 3 lines of chemotherapy (1), first-line chemotherapy and biochemotherapy after lung metastasis resection (1), high-dose interferon (IFN) for 9 months and bone radiotherapy (1), chemotherapy after low-dose IFN (1) and biochemotherapy (1), or no therapy (2).

Eleven patients underwent other treatments after stopping vaccination due to disease progression (8 patients) or because the ATL was finished (3, of whom 2 had PR and 1 SD). Of these, 3 underwent high-dose IFN, 2 low-dose IFN, and 6 no treatment. Median disease-free survival (DFS) (from exeresis of primary melanoma to first relapse) was 36 months (range 6–108). Two patients had high LDH levels before vaccination which further increased after treatment. Sites of metastasis after vaccination in the subgroup were lymph nodes (6 patients), soft tissue (7), kidney (2), lung (3), and liver (1). Subsequent treatments were as follows: surgical palliative intervention (3 patients, each undergoing >1 nonradical surgical intervention), biotherapy with anti-CTLA-4 monoclonal antibody in a clinical trial (2 patients, one of whom also received hepatic locoregional fotemustine and chemotherapy with dacarbazine [DTIC] and the other, Gamma Knife radiosurgery for brain metastases and fotemustine), chemother-apy (cisplatin [CDDP] plus DTIC-based polychemotherapy) (2), low-dose biochemotherapy (CDDP+DTIC+IL-2) (2), chemotherapy with CDDP+DTIC and high-dose IL-2 (1), and biochemotherapy (CDDP+DTIC+ low-dose IL-2) plus Gamma Knife radiosurgery (1) (Table 1).

2.2. Inclusion Criteria for Vaccine Therapy. Age < 70 years, histologically confirmed diagnosis of melanoma, measurable disease (excluding the presence of brain metastases), previous removal of one or more metastatic lesion from which a sufficient quantity of ATL had been obtained for at least 6 vaccinations, Performance Status (PS) ≤ 2 (according to ECOG criteria), life expectancy > 4 months. Patients
who were in good clinical condition (ECOG ≤ 2) but had stopped vaccination due to disease progression or because the ATL was finished were treated with subsequent standard chemotherapy.

2.3. Autologous Tumor Lysate (ATL) Preparation. Surgically removed tumor samples were mechanically dispersed to create a single-cell suspension. The largest pieces were incubated at 37°C in enzyme mix (collagenase 0.1%, hyaluronidase 0.01%, DNase 0.1%, Sigma, Milan, Italy) in RPMI 1640, (PAA Laboratories GmbH, Pasching, Austria) for 3 hours. At the end of incubation the pellets were washed 3 times with phosphate buffered saline (PBS) and incubated for at least 20 minutes in sterile distilled water. Lysis was monitored by light microscope. Larger particles were removed by centrifugation (10 min at 600 g) and the supernatant was passed through a 0.2-μm filter. Protein contents were determined and aliquots were stored at −80°C until use, after verification of sterility.

### Table 1: Patients who underwent subsequent therapies after vaccine.

<table>
<thead>
<tr>
<th>Patient</th>
<th>Adjuvant therapy</th>
<th>DFS (months)</th>
<th>Prior treatment (Prevaccine)</th>
<th>Prevaccine LDH</th>
<th>Postvaccine LDH</th>
<th>Postvaccine treatment</th>
<th>Best Response</th>
<th>Sites of evaluable metastases</th>
</tr>
</thead>
<tbody>
<tr>
<td>2 P.M.</td>
<td>No 12 BioCT</td>
<td>366</td>
<td>CT</td>
<td>627§</td>
<td></td>
<td>CT</td>
<td>SD</td>
<td>Lung, lymph nodes</td>
</tr>
<tr>
<td>4 G.D.</td>
<td>HD-IFN 37</td>
<td>no</td>
<td>CT and RT + surgery for brain metastases</td>
<td>332</td>
<td>406</td>
<td>CT HD-IL-2</td>
<td>PR</td>
<td>Lymph nodes</td>
</tr>
<tr>
<td>6 T.A.</td>
<td>No 6 BioCT</td>
<td>334</td>
<td>LDH</td>
<td>281</td>
<td></td>
<td>BioCT</td>
<td>PR</td>
<td>Soft tissue</td>
</tr>
<tr>
<td>8 C.P.</td>
<td>No 48</td>
<td>256</td>
<td>CT and RT + surgery for brain metastases</td>
<td>245</td>
<td></td>
<td>BioCT</td>
<td>PR</td>
<td>Soft tissue</td>
</tr>
<tr>
<td>11 M.J.</td>
<td>LD-IFN 108</td>
<td>193</td>
<td>CT and RT + surgery for brain metastases</td>
<td>265</td>
<td></td>
<td>Surgery, Gamma Knife (brain)</td>
<td>PR+*</td>
<td>Lung, kidney, skin, lymph nodes</td>
</tr>
<tr>
<td>15 B.F</td>
<td>No 57 Arm perfusion, BioCT, surgery</td>
<td>603§</td>
<td>Surgery</td>
<td>686§</td>
<td>433</td>
<td>PR+*</td>
<td>Soft tissue, lung, lymph nodes, adrenal gland</td>
<td></td>
</tr>
<tr>
<td>17 B.I</td>
<td>No 9 BioCT</td>
<td>236</td>
<td>Surgery</td>
<td>234</td>
<td></td>
<td>254</td>
<td>PD</td>
<td>Lung, soft tissue</td>
</tr>
<tr>
<td>18 S.L.</td>
<td>HD-INF 36</td>
<td>374</td>
<td>CT (3 different lines)</td>
<td>311</td>
<td></td>
<td>CT (brain), anti-CTLA-4 antibodies</td>
<td>PD</td>
<td>Lung, soft tissue</td>
</tr>
<tr>
<td>22 M.C.</td>
<td>LD-INF 18</td>
<td>No 591§</td>
<td>Hepatic loco-reg CT, DTIC anti-CTLA4ab</td>
<td>988§</td>
<td>493</td>
<td>PR</td>
<td>Liver, soft tissue</td>
<td></td>
</tr>
<tr>
<td>23. R.G</td>
<td>HD-INF 48</td>
<td>Bone RT 299</td>
<td>BioCT, Gamma Knife (brain)</td>
<td>313</td>
<td></td>
<td>na</td>
<td>CR+</td>
<td>Lung, bone</td>
</tr>
<tr>
<td>24. B.R</td>
<td>No 24 CT, LD-INF</td>
<td>236</td>
<td>BioCT</td>
<td>234</td>
<td></td>
<td>na</td>
<td>PD</td>
<td>Lung, soft tissue, lymph nodes</td>
</tr>
</tbody>
</table>

Adj: adjuvant; T: treatment; V: vaccine; HD-IFN: high-dose interferon; BioCT: biochemotherapy; CT: chemotherapy; RT: radiotherapy; LD: low dose; anti-CTLA4ab, anti-CTLA4 monoclonal antibody; §, elevated; *, responses obtained alternating palliative surgery with vaccine.

2.4. DC Generation. DCs were prepared from peripheral blood monocytes (PBMCs) obtained by leukapheresis without previous mobilization. Five to nine liters of blood were processed in each collection. PBMCs were purified on Ficoll–Paque. An aliquot of PBMC was utilized immediately for DC generation and the rest was frozen in bags for use at a later date (4–5 bags/each collection). PBMC were incubated in tissue culture flasks with CellGro DC medium (CellGenix, Freiburg, Germany) at 10 × 106 cells/mL for 2 hours. The nonadherent cells were discarded and the adherent cells were incubated in CellGro DC medium containing 1000 IU/mL rhIL-4 and 1000 IU/mL rhGM-CSF (Cell Genix, Freiburg, Germany) for 7 days to generate a DC-enriched cell population. On day 6, 90% of the DC culture was pulsed with ATL/ATH (100 mg/mL), while the remaining 10% was pulsed with KLH (50 mg/mL). Both cultures were then incubated overnight. On day 7, the cells were defined as immature DCs (iDCs). After eliminating the previous culture medium, pulsed iDCs were cultured for a further 2
days with a cocktail of cytokines (TNFα, IL-1β, IL-6, Cell Genix, Freiburg, Germany; PGE2, Pfizer, Italy). On day 9 they were defined as mature DCs (mDC). iDCs or mDCs were removed, washed, and suspended in sterile saline for therapeutic infusion into the patient.

2.5. Delayed Type Hypersensitivity. ATL (10 μg) and KLH (5 μg) were each suspended in 500 μL of PBS and injected intradermally into the forearm of the patient. PBS alone was used as negative control. Patients received intradermal injections of ATL, KLH, and physiological solution as negative control at separate sites on the forearm. Eight and twenty-four hours later, DTH was assessed by determining the area of erythema and induration using two-dimensional measurements. The DTH response was considered to be positive if the area of erythema was >10 mm.

2.6. Statistical Analysis. Evaluation of response to vaccination was carried out according to modified RECIST criteria, and mixed responses were thus evaluated (decrease in or disappearance of lesions with appearance of new lesions or with modest progression in others). In the event of modest progression, good PS (<2) and positive DTH, vaccination was carried out [28]. Response to postvaccine treatments (11 patients) was carefully evaluated by RECIST criteria, and survival time was calculated as the time between the date of the first vaccination and the date of death from any cause [28] or last followup (June 2009). Overall survival curves were calculated by the Kaplan-Meier method and compared using the Gehan-Wilcoxon test, which tends to weigh the early differences more heavily than other tests belonging to the two-sample rank test family [29].

3. Results

3.1. Patient Characteristics. Twenty-four patients (13 males and 11 females) with a median age of 50 (range 34–75 years) entered the study. Sites of evaluable metastases were viscera (20 patients), bone (1), soft tissue (14), and lymph nodes (13). Prevaccine treatments were biochemotherapy (12 patients), chemotherapy (7), biotherapy (1) radiotherapy (3), or no therapy (3) (Table 2).

3.2. Update of Clinical Vaccine Results. Of the 24 patients treated with vaccine, 2 showed complete response (CR), 2 mixed response (MR), 5 partial response (PR) (Figure 1), and 5 stable disease (SD). The overall response (OR) rate was 37.5% with a clinical benefit of 54.1%. All 14 responders had DTH-positivity to KLH, 10 of whom also to ATL (Table 3). Median overall survival (OS) was 15 months.
3.3. Subgroup (Postvaccine Treatments) Results. The overall response rate to subsequent therapies was 63.6% with 2 CR (1 patient treated with surgery alternating with vaccine and radiosurgery (Gamma Knife) for a small brain metastasis, 1 treated with biochemotherapy plus radiosurgery (Gamma Knife) for a single brain metastasis) and 5 PR (2 patients treated with surgery, 1 patient receiving hepatic locoregional fotemustine+DTIC+anti-CTLA-4 antibody, 1 submitted to DTIC+CDDP and high-dose IL-2, and 1 treated with CDDP+DTIC+ low-dose IL-2), with a median OS of 34 months (median range 16–61). Five of these patients had received one or more treatments before vaccination and experienced an objective response. Of the 11 subgroup patients, 3 had high LDH serum levels after vaccination which normalized during subsequent treatments administered for progressive disease (Table 1). All but one of the patients also had DTH-positivity to at least the KLH test, while 6 were also positive to ATL after vaccination (Figure 5).

3.4. Toxicity. Apart from swelling, redness, and pruritus around the site of inoculation, no noteworthy toxicities or side effects were observed following vaccination. A low fever with mild flu-like symptoms (grade 1-2) was present during administration of IL-2 from days 3 to 7. No autoimmune phenomena were observed apart from the onset of vitiligo in 3 patients, hypothyroidism in 2 and the flaring up of a preexisting vitiligo in one patient (all responders). Toxicity linked to subsequent treatments was coherent with that expected from the different schedules used and no unexpected adverse events were observed.

4. Discussion

Systemic therapy for metastatic melanoma remains disappointing and median survival is not improved significantly by currently available chemotherapy regimens [29]. Clinical response rates to most single agents are lower than 15% [30–34], whereas drug combinations have produced response
Figure 4: PET/CT scans of patient n 19 (MJ) performed in September 2005 (positive PET scan in several metastatic sites), February 2007, and March 2009 (negative PET scan for metasteses). During this period the patient underwent palliative surgery for symptomatic disease (e.g., hematuria due to renal metasteses) or to collect ATL for vaccination that started in February 2005 and terminated in June 2008 because the ATL was finished. Radiosurgery (Gamma Knife) was carried out on a small brain lesion in January 2008. The patient is still in CR (negative PET scan May 2010).

Table 3: Update (June 2009) of results on 24 patients treated with mDC vaccine.

<table>
<thead>
<tr>
<th>Patient ID</th>
<th>n vaccinations</th>
<th>DTH</th>
<th>Best response</th>
<th>after 4 or more vaccinations</th>
<th>Vitiligo</th>
<th>Clinical response</th>
<th>Response duration</th>
<th>OS (Months)</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1) P.M</td>
<td>7</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>−</td>
<td>PR</td>
<td>3</td>
<td>14</td>
</tr>
<tr>
<td>(2) P.M</td>
<td>15</td>
<td>++</td>
<td>++ ++</td>
<td></td>
<td>++</td>
<td>MR</td>
<td>6</td>
<td>22</td>
</tr>
<tr>
<td>(3) R.L.</td>
<td>10</td>
<td>−</td>
<td>++</td>
<td></td>
<td>−</td>
<td>SD</td>
<td>9</td>
<td>14</td>
</tr>
<tr>
<td>(4) G.D.</td>
<td>16</td>
<td>++</td>
<td>++ +</td>
<td>+</td>
<td></td>
<td>CR</td>
<td>8</td>
<td>34</td>
</tr>
<tr>
<td>(5) R.G.</td>
<td>4</td>
<td>−</td>
<td>++ +</td>
<td></td>
<td></td>
<td>PD</td>
<td>—</td>
<td>8</td>
</tr>
<tr>
<td>(6) T.A.</td>
<td>13</td>
<td>−</td>
<td>++</td>
<td></td>
<td></td>
<td>MR</td>
<td>12</td>
<td>41</td>
</tr>
<tr>
<td>(7) B.A.</td>
<td>4</td>
<td>−</td>
<td>−</td>
<td></td>
<td>−</td>
<td>PD</td>
<td>—</td>
<td>7</td>
</tr>
<tr>
<td>(8) C.P.</td>
<td>6</td>
<td>−</td>
<td>++</td>
<td></td>
<td></td>
<td>PD</td>
<td>—</td>
<td>20</td>
</tr>
<tr>
<td>(9) O.M.</td>
<td>4</td>
<td>−</td>
<td>−</td>
<td></td>
<td>−</td>
<td>PD</td>
<td>—</td>
<td>5</td>
</tr>
<tr>
<td>(10) LB.</td>
<td>4</td>
<td>−</td>
<td>−</td>
<td></td>
<td>−</td>
<td>PD</td>
<td>—</td>
<td>3</td>
</tr>
<tr>
<td>(11) M.J.</td>
<td>8+8+8+4</td>
<td>+</td>
<td>++ +</td>
<td>+</td>
<td></td>
<td>CR</td>
<td>15+*</td>
<td>52+</td>
</tr>
<tr>
<td>(12) O.G.</td>
<td>5</td>
<td>−</td>
<td>−</td>
<td></td>
<td>−</td>
<td>PD</td>
<td>—</td>
<td>3</td>
</tr>
<tr>
<td>(13) M.R.</td>
<td>4</td>
<td>+</td>
<td>++</td>
<td></td>
<td></td>
<td>PD</td>
<td>—</td>
<td>6</td>
</tr>
<tr>
<td>(14) D.I.G</td>
<td>10</td>
<td>+</td>
<td>+</td>
<td></td>
<td></td>
<td>CR</td>
<td>36+</td>
<td>39+</td>
</tr>
<tr>
<td>(15) B.F</td>
<td>21</td>
<td>++</td>
<td>++</td>
<td></td>
<td></td>
<td>PR</td>
<td>20+*</td>
<td>40+</td>
</tr>
<tr>
<td>(16) L.I</td>
<td>6</td>
<td>−</td>
<td>−</td>
<td></td>
<td>−</td>
<td>PD</td>
<td>—</td>
<td>7</td>
</tr>
<tr>
<td>(17) B.I</td>
<td>4</td>
<td>++</td>
<td>−/+</td>
<td>+</td>
<td></td>
<td>PR</td>
<td>21+*</td>
<td>31+</td>
</tr>
<tr>
<td>(18) S.L.</td>
<td>9</td>
<td>−</td>
<td>+</td>
<td></td>
<td></td>
<td>SD</td>
<td>7</td>
<td>22</td>
</tr>
<tr>
<td>(19) N.E.</td>
<td>6</td>
<td>++</td>
<td>+</td>
<td></td>
<td></td>
<td>PR</td>
<td>5</td>
<td>8</td>
</tr>
<tr>
<td>(20) B.R</td>
<td>12</td>
<td>+++</td>
<td>+++</td>
<td></td>
<td></td>
<td>SD</td>
<td>8</td>
<td>10</td>
</tr>
<tr>
<td>(21) S.M.</td>
<td>11</td>
<td>−</td>
<td>−</td>
<td></td>
<td>−</td>
<td>PD</td>
<td>—</td>
<td>12</td>
</tr>
<tr>
<td>(22) M.C</td>
<td>6</td>
<td>−</td>
<td>+</td>
<td></td>
<td></td>
<td>PD</td>
<td>—</td>
<td>61+</td>
</tr>
<tr>
<td>(23) R.G.</td>
<td>5</td>
<td>−</td>
<td>−</td>
<td></td>
<td>−</td>
<td>PD</td>
<td>—</td>
<td>60+</td>
</tr>
<tr>
<td>(24) B.R</td>
<td>13</td>
<td>+</td>
<td>++ +</td>
<td></td>
<td></td>
<td>SD</td>
<td>11</td>
<td>16</td>
</tr>
</tbody>
</table>

PR: partial response; MR: mixed response; SD: stable disease; CR: complete response; PD: progressive disease; OS: overall survival; DTH: delayed-type hypersensitivity test; *responses obtained with palliative surgery + vaccine alternating; Grey area, 11 patients given postvaccine therapy.
rates of up to 40% [8, 10, 35]. Although a combination of cytotoxic chemotherapy with biological response modifiers such as IL-2 and IFN has resulted in overall response rates of 40%–60% with about 10%–20% CR, biochemotherapy cannot be offered to a substantial proportion of patients with metastatic melanoma because of its high toxicity [11, 36–40]. Furthermore, recent phase III trials have not demonstrated a clear survival benefit from biochemotherapy compared with that obtained with conventional chemotherapy in patients with metastatic melanoma [30, 41–43]. As demonstrated in most of the phase III studies carried out on combination regimens, the incidence of toxicity increases as more drugs are combined and there is no improvement in median survival. New drug regimens are therefore needed with less morbidity than biochemotherapy but more potent antitumor activity than current standard chemotherapies. In fact, understanding how melanoma overcomes host immunity could be the key to developing strategies targeting components of the antitumor immune response, for example, anti-CTLA-4 agents, which has produced encouraging results. Durable objective response rates have been in the range of 4.6%–15% for patients with metastatic melanoma, with a further increment in long-term SD or progression followed by response [44–46].

Specific tumor vaccines attempt to reverse tumor-induced immune suppression and it is thought that they may prolong survival in immune-responsive patients. Vaccines would seem to trigger immunologic memory and thus subsequent treatments that are capable of upregulating tumor-associated antigen expression or of enhancing cross-presentation in a toll-like receptor 4 dependent manner following chemotherapy- or radiotherapy-induced tumor cell death appear to more successful [47]. A progressive surgical reduction of the tumor mass also seems to intensify the effect of the vaccine.

We observed an OR of 63.6%, which, albeit infrequent in metastatic melanoma after failure of at least one line of therapy, is nevertheless in line with other data published on the treatment of other solid tumors. In our case series, although the 11 patients subjected to postvaccine therapy all had a fairly long initial DFS (median of 36 months) and 5 had also responded to first-line therapy, the response percentage observed after 3-4 lines of therapy was unusually high. It must be underlined that all but one of the 11 patients treated after vaccination had positive DTH to KLH (6/11 also positive to ATL), which seems to support the hypothesis that immunoreactive patients may benefit from being treated even after the failure of vaccine therapy. This highlights the potential usefulness of using vaccine treatment sooner rather than later with the aim of promoting further therapeutic response. In fact, the combination of vaccine with surgery could be effective in reducing the neoplastic mass, facilitating the effect of immunotherapy. Radiotherapy plus vaccination is thought to induce an abscopal effect [48, 49], while chemotherapy, in addition to reducing tumor burden, may induce lymphocytopenia in immunosuppressive cells such as T-regulator lymphocytes [50]. It may even be possible to improve the effect of DC vaccine by combining it with drugs that induce a stronger immunological response (Toll-like receptors) or with agents that inhibit immunosuppression such as antiCTLA-4 antibody. Finally, it would perhaps be useful now to search for predictive factors of immunosuppression, for example, TEM8 expression in dendritic cells. This marker, evaluated at baseline in 4 of the subgroup patients, was low in 3 responders to subsequent therapies and high in one progressive patient, which would seem to confirm previously reported findings by Venanzi et al. [51].

In conclusion, it is clear that the small number of patients and the retrospective setting does not permit any definitive conclusions to be drawn. However, we do feel that our data, together with other findings published in the literature, could form the basis to design new, effective combined treatment strategies. As suggested by other authors [52], it might be useful, for example, to bring forward the time of vaccination with respect to other treatments or to consider the potential of alternating immunotherapy (vaccine alone or vaccine+adjuvant, such as anti-CTLA-4 antibodies) with chemo/radiotherapy or surgery.

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References


