Review Article

The Role of T Helper (T_H)17 Cells as a Double-Edged Sword in the Interplay of Infection and Autoimmunity with a Focus on Xenobiotic-Induced Immunomodulation

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Extensive research in recent years suggests that exposure to xenobiotic stimuli plays a critical role in autoimmunity induction and severity and that the resulting response would be exacerbated in individuals with an infection-aroused immune system. In this context, heavy metals constitute a prominent category of xenobiotic substances, known to alter divergent immune cell responses in accidentally and occupationally exposed individuals, thereby increasing the susceptibility to autoimmunity and cancer, especially when accompanied by inflammation-triggered persistent sensitization. This perception is learned from experimental models of infection and epidemiologic studies and clearly underscores the interplay of exposure to such immunomodulatory elements with pre- or postexposure infectious events. Further, the T_H 17 cell subset, known to be associated with a growing list of autoimmune manifestations, may be the “superstar” at the interface of xenobiotic exposure and autoimmunity. In this review, the most recently established links to this nomination are short-listed to create a framework to better understand new insights into T_H 17’s contributions to autoimmunity.

1. Introduction

Long-term exposure to xenobiotic substances induces hyperactivity of the immune system, thereby increasing the incidence of autoimmune diseases (AD), especially in infection-aroused systems. Circumstances dating back to earlier exposure, as in case of heavy-metal industry workers or current exposure as in individuals harboring amalgam teeth filling, favor incidence of inflammatory processes and most likely AD [1–4]. Exposure to infectious agents leads to the induction of various cellular pathways essential to the microbe’s infectivity, survival, and virulence, thus making it difficult for the microbe to go undetected by the host’s immune system [5]. Upon pathogen recognition, production of a proinflammatory response, primarily by macrophages, NK and NKT cells, is the subsequent event in the early phase of the infection

[5, 6]. Further, the coordination between innate and adaptive immune defense systems ensures a successful eradication of pathogens, and such developed cytokine milieu determines the induction of a specific T-cell-mediated response that is critical for an effective and complete pathogen clearance. However, whether the induction of a strong host inflammation constitutes an adaptive advantage to the host or pathogen remains debated. Indeed, many disorders, including AD [7] and cancer [8], are associated with and maintained by chronic inflammation; for review, see [9, 10]. The association of cancer incidence with exposure to heavy metals, such as cadmium [11], or following attainment of chronic inflammation, as in case of colitis-associated cancer, has been widely anticipated.

Research on T_H 17 cells has suggested a crucial role in autoimmunity. Despite developing autoimmune signs in the absence of detectable IL-17 levels, as in case of

choriomeningitis-virus-induced model of type 1 diabetes [12], a key role of T_H17 cells and their related molecules was underscored in many previously assigned “ T_H1 -mediated” AD including rheumatoid arthritis (RA), psoriasis, systemic lupus erythematosus (SLE), and multiple sclerosis (MS), as well as, the experimental autoimmune encephalomyelitis—EAE [7, 9, 13–15]. Variations in disease susceptibility or outcome may be a result of co-exposure to one or multiple xenobiotic substances or infectious pathogens, so that a xenobiotic-induced polarized immune response triggers the development of AD in genetically predisposed individuals [1, 2, 4, 16–19]. The IL-17 response, while constituting a protective arm defending the body against various infections, also functions as a double-edged sword constituting a risk factor that mediates the development and/or induction of AD, mostly manifested following pathogenic and xenobiotic-induced chronic inflammation; it then acts as a double-edged sword, constituting a risk factor that mediates the development and/or induction of AD, mostly manifested following pathogenic and xenobiotic-induced chronic inflammation. In the next sections, we revisit our view on the T_H17 cells’ role in autoimmunity [9] and provide a brief description of the double-sided role of T_H17 cells and their related molecules IL-17, IL-21, and IL-22 and their participation at the initiation/induction of autoimmunity as a consequence of xenobiotic exposure.

2. T_H17 Cells and Their Associated Molecules Link Infection to Autoimmunity

T cells differentiate and expand into distinct lineages including T_H1 , T_H2 , iT_{Reg} , and T_H17 cells [9], whereas iT_{Reg} cells differentiate under subimmunogenic antigen presentation both during chronic inflammation and under normal homeostatic conditions of the gut and function to control severe chronic allergic inflammation and as a barrier to the eradication of tumors [20, 21]. T_H17 cells derive from $CD161^+$ precursors in umbilical cord blood and newborn thymus [22] and likely constitute the most prominent T cell subset at the crossroads of infection and autoimmunity. The contributions of T_H17 cells have prompted and were the results of intensive scientific research, which is reflected by a growing list of publications in this field (Figure 1), and have in turn led to identification of T_H17 cells’ markers, as well as, their differentiation and commitment program [23]. Figure 2 demonstrates the major T cell subsets, their interaction with T_H17 cells and the main contributions of the latter.

Recently, several groups delivered compelling evidence of the effects of T_H17 -associated cytokines, namely IL-17, IL-21, IL-22, and IL-23, on inflammatory responses elicited by extracellular, as well as, facultative and obligate intracellular pathogens including bacteria and fungi. Exemplified contributions of IL-17 response to some infectious diseases are summarized in Table 1.

2.1. T_H17 Cells-Associated Molecules and Their Contributions to Anti-Infectious Responses. In comparison to the frequent appearance of T_H1 , the relative rarity of T_H17 in inflamed

FIGURE 1: Frequencies of IL-17-producing T helper (T_H)17 cells and their related molecules as appeared in title or title/abstract of PubMed publications. The “keywords” { T_H17 } OR {IL-17*} OR {IL-21} OR {IL-22} OR {IL-23*} OR {CTLA-8} OR {CCL20} OR {ROR*} and their synonyms combined with “publication date” {1993} through {2011} were given as search parameters.

tissues was attributed to their *RorC*-dependent expression of the oxidase IL4I1, which impairs CD3 signaling and hence constrains IL-2 production and cell proliferation [24]. As we recently reviewed, the recruitment of T_H17 cells to inflammatory tissues accompanies the expression of the chemokine receptor CCR6, in addition to CCR4, IL-23R (involved in the survival/maturation program of T_H17 cells) [9], and CD161 [22]. T_H17 cells are considered as potent inflammation inducers that, in addition to production of IL-17, differentially produce IL-6, IL-2, IL-8, IL-9, TNF- α , IL-17F, IL-21, IL-22, IL-26, IFN- γ , and the chemokine CCL20 and induce activation and recruitment of other cells including neutrophils that are pivotal in inflammation and AD [9, 25]. Through their cytokine/chemokine production, T_H17 cells act on a broad range of cell types initiating the expression of antibodies, metalloproteinases, prostaglandin E2 (PGE₂), and antimicrobial peptides and inducing cyclooxygenase 2 activity [9, 26], constituting, thereby, a link between innate and adaptive immune responses. In addition to its role as an arm of adaptive immunity, the current perception categorizes IL-17 also as an innate cytokine, produced mainly by NK cells [27] as well as by $\gamma\delta$ T cells [28–31], $CD8^+$ T cells, and mast cells [9, 32]. Indeed, detecting functional T_H17 cells and production of protective IL-17 during the early phase of the immune response [33–35] and the activation of T_H17 that

FIGURE 2: Differentiation and commitment of IL-17-producing T helper (T_H)17 cells in the midpoint of other coacting cells in favor of and against the harboring individual. Upon antigen recognition, presentation and cosignaling, naive (T_H 0) cells differentiate in the presence of distinct cytokine milieu into effector (T_H 1, T_H 2, T_H 9, T_H 17, T_H 22, and TFH) and T regulatory (T_{Reg}) cells. Signaling cytokine and other molecules activate lineage-unique transcription factors that ultimately mediate cell differentiation and maturation. Whereas activation of STAT1 induces T-bet expression, STAT6 signaling upregulates GATA3 expression; both cell lineages reciprocally regulate each other and regulate the generation of T_H 17 cells through their hallmark effector cytokines, IFN- γ and IL-4, respectively, though IFN- γ is produced by T_H 17 cells in some disease settings and in response to certain infections. Differentiation of T_H 17 necessitates costimulatory signals of CD28 and ICOS (the last is not mandatory) and the absence of T_H 1 and T_H 2 cytokines (IL-12/IFN- and IL-4) and their master transcription factors; a task taken over by TGF- β is to constrain T_H 1 and T_H 2 during T_H 17 differentiation program. T_H 17 and T_{Reg} are likely descendants of the same ancestor cell lineage, which differentiates in presence of low levels of TGF- β and IL-6/IL-21 or other proinflammatory cytokines (IL-1, TNF- α , and IL-18) into T_H 17 cells; TGF- β signals through Smad2 protein pathway and is indispensable for induction of expression ROR γ t. High levels of TGF- β alone induce Foxp3 expression and hence T_{Reg} cell differentiation. Following their final commitment and upregulation of IL-23R, T_H 17 cells require IL-23 signaling that is crucial for their survival and effector functions including production of IL-22, as well as, cell plasticity including later production of IFN- γ . T_H 22 may differentiate from T_H 0 cells or through a local commitment of T_H 17 cells homed in the epidermis. Contributions of T_H 17 cells entail activations of aryl hydrocarbon receptor (AhR) signaling and production of IL-22 upon exposure to xenobiotic substances. IL-17 and IL-17F increase production of IL-6, IL-8, prostaglandin E2 (PGE2), monocyte chemoattractant protein-1 (MCP-1), and the granulocyte colony-stimulating factor (G-CSF) by various cells including macrophages, fibroblasts, keratinocytes, and epithelial and endothelial cells and ultimately promote inflammatory diseases, AD, and/or cancer. These cytokines together with IL-21 and IL-22 are also implicated in mediating protective as well as pathogenic processes in various disease settings.

even precedes the differentiation of T_H 1 cells [33], together with the later contribution of T_H 17 [36], highlight the crucial importance of IL-17 and other T_H 17-related cytokines in the early, as well as, the late phase of infection. The upregulation of TLR1 and TLR2 and dectin 1 by IL-17-producing $\gamma\delta$ T cells [37] supports this belief. Moreover, studies on nucleotide oligomerization domain knockout mice (Nod1 $^{-/-}$ and Nod2 $^{-/-}$) demonstrated that this “early” T_H 17 response was Nod1- and Nod2-dependent, and hence they have been given the name innate (i) T_H 17 cells [35]. Therefore, the new look of IL-17-producing cells comprises their contribution in building the first line of host defense, besides mediating and shaping adaptive responses required for ultimate clearance.

Based on a wealth of experimental data, the contribution of T_H 17 cells to infection is manifold. As in the case of oral infection, the importance of IL-17 in protection against infection seems to be crucial to attain a mucosal barrier in the intestine as in case of *Salmonella* [30, 38], in mediating protection against oral brucellosis [39], promoting granulopoiesis through induction of granulocyte colony-stimulating factor G-CSF [40], and neutrophil influx through inducing neutrophil chemotactic CXCL8 (IL-8), macrophage chemotactic protein (MCP)-1, and macrophage inflammatory proteins- (MIP-) 1 and MIP-2 [9, 40, 41]. Additionally, we and others found that IL-17 activates phagocytosis and neutrophil cytotoxic activity [42]. Therefore, IL-17R $^{-/-}$ mice

TABLE 1: Mechanistic investigations on the role of T_H17 cells and their related molecules in various infectious diseases.

Diagnosis	Role	Observations on T _H 17-associated molecules	Citations
(1) Bacterial infections			
<i>Bacillus subtilis</i>	Pathogenic/protective	Increased lung inflammation and collagen deposition; delay in bacterial clearance in IL-17R ^{-/-} compared with WT counterparts	[118]
<i>Francisella tularensis</i>	Pathogenic	Intranasal inoculation induces T _H 17 response and PGE ₂ production in the lung; inhibition of PGE ₂ production increased IFN- γ and decreased bacteremia	[119]
<i>Saccharopolyspora rectivirgula</i>	Pathogenic	Induction of IL-17-mediated hypersensitivity pneumonitis in mice; reduced lung inflammation and fibrosis in IL-17R ^{-/-} mice	[120, 121]
<i>Klebsiella pneumoniae</i> , <i>Bordetella pertussis</i> , & <i>S. pneumoniae</i>	Protective	Mounting of an IL-17 and IL-22 response; defects in T _H 17 response increased susceptibility	[12, 122]
<i>Staphylococcus aureus</i>	Protective	High infection incidence correlated with defect in T _H 17 response	[59]
<i>Listeria monocytogenes</i>	Protective	IL-17-mediated cross-protection following immunization with <i>M. pulmonis</i> ; blockade of bacterial growth following transfer of IL-17-producing $\gamma\delta$ and double negative $\alpha\beta$ T cells into RAG2 ^{-/-} mice	[30, 60]
<i>Shigella flexneri</i>	Protective	Restriction of bacterial growth mediated by T _H 17 response	[28]
<i>Citrobacter</i> and <i>Salmonella</i> sp.	Protective	Innate T _H 17 response-dependent protection; protective effect of IL-17 and IL-22; decrease in phagocytic activity and increase in bacterial burden upon IL-17 neutralization and its correlation with T _H 17 response in Hg-exposed mice	[29, 35, 38] and Hemdan and Abul El-Saad, unpublished
<i>Mycobacterium tuberculosis</i> and <i>M. bovis</i>	Protective	IL-17 ^{-/-} mice reveal a reduced IFN- γ production by CD4 ⁺ T cells, impaired granuloma formation, and chemokine expression	[61]
<i>Mycobacterium tuberculosis</i>	Protective	Correlation of reduced T _H 17 responses in patients with active tuberculosis with decreased expression of IL-6R on CD4 ⁺ T cells	[123]
<i>Chlamydia</i> sp.	Protective	Enhanced bacterial growth and decreased mouse survival upon applying anti-IL-17 mAb	[34]
	Pathogenic	Applying IL-17RA antagonist reversed the susceptible phenotype of C3H/HeN mice	[58]
Inflammatory bowel disease—IBD (Crohn's disease and ulcerative colitis)	Pathogenic	Enhanced differentiation T _H 17 and IL-17 expression levels and NK activities in IBD	[124, 125]
	Protective	IL-22 mediated protection against IBD	[54]
(2) Protozoal infection			
<i>Toxoplasma gondii</i>	Pathogenic	IL-23-mediated IL-22 and MMP-2 upregulation in the ileum of infected mice; MMP-2 deficiency offered protection	[47]
	Protective	Increased mortality in IL-17 ^{-/-} mice	[126]
(3) Fungal infections			
<i>Candida</i> sp.	Protective	Involvement of IL-17, IL-17F, IL-22, and IL-23 in mediating natural defense against candidiasis	[45]
<i>Aspergillus fumigatus</i>		Induced IL-17 response mediates pathogen clearance	[127]
(4) Viral infection			
Theiler's murine encephalomyelitis virus infection	Pathogenic	Induction of antiapoptotic molecules by IL-17 and thereby promoting persistent infection; boosting lytic function of CTLs and ameliorating disease upon neutralizing IL-17; association of lower T _H 17 with higher virus-specific CD8 ⁺ T cell responses in resistant mouse than in susceptible strain	[128]

TABLE 1: Continued.

Diagnosis	Role	Observations on T _H 17-associated molecules	Citations
Respiratory syncytial virus (RSV)	Pathogenic	Elevated IL-6 and IL-17 levels in tracheal aspirate samples from severely ill infants and in infected mice; IL-17 blockade decreased the exacerbated disease via increasing RSV-specific CD8 ⁺ T cells, T-bet, IFN- γ , eomesodermin, and granzyme B	[129]
HBV	Pathogenic/protective	Distinct effects associated with heterogeneous T _H 17 populations: IL-17 with inflammation and ALT levels, IL-22 with protection of hepatocytes, and IL-21 with virus clearance	[130]
HCV	Pathogenic	Hepatitis-C-virus-infected patient revealed upregulated T _H 17 cell cytokines that became downregulated by combined treatment with pegylated IFN and ribavirin	[131]
Simian immunodeficiency virus (SIV)/HIV	Pathogenic	Induction of TGF- β and IL-18 during the acute phase in SIV-infected rhesus macaques proposed to be associated with induction of IL-17-producing NKT cells	[132]
	Protective	Association of disease progression with loss of T _H 17 and induction of T _{Reg} cells; T _H 17 cell frequency correlated negatively with viral load	[63, 133, 134]
Herpes simplex virus (HSV-1)		Infiltration of T _H 1 preceded T _H 17 cells, the latter showed lower responsiveness ability to HSV-1; diminished stromal keratitis severity in IL-17R ^{-/-} infected mice and upon IL-17 neutralization in WT mice	[135]
(5) Nematode infection			
<i>Trichinella spiralis</i>	Pathogenic	Correlation of T _H 17 response with increase of smooth muscle contraction probably causing gut dysfunction; association of IL-17/IL-23 axis induction with increased mortality in mice coinfecting with malaria and nematode	[136]

revealed increased systemic dissemination of *S. typhimurium* from the gut [29]. The same strategy seems to be attained to combat extracellular pathogens such as *Klebsiella pneumoniae* [12] and fungal infections (e.g., *Candida* sp.), mainly due to defective IL-17 immunity [43, 44], mediated by eliciting production of autoantibodies (AAs) against IL-17, IL-17F, and IL-22 that contribute to chronic mucocutaneous candidiasis [45, 46]. A third arm of T_H17 cells is built through IL-22, which is also produced by other cell types including NK22 and lymphoid tissue inducer cells [47], as well as by skin homing T_H22 cells [48, 49]. Besides its accepted role against infection, it induces tissue repair offering protection against injury [49]. Although both IL-22 and IL-17 or IL-17F synergize to stimulate expression of human beta-defensin (HBD)-2, S100 calcium binding protein A9 (S100A9) and enhanced the expression of S100A7 and S100A8 [9], IL-22, rather than IL-17, seems to contribute more to the epidermal and mucosal immunity [47, 49]. It synergizes with TNF- α to induce secretion of initial complement factors C1r and C1s, antimicrobial peptides S100A7 and HBD-2, and antimicrobial chemokines CXCL-9/-10/-11 in primary human keratinocytes [50]. In a three-dimensional skin infection model, stimulation of keratinocytes with T_H22 supernatants or by adding IL-22 plus TNF- α effectively inhibited *C. albicans* growth and maintained epithelial survival, and the combinatorial stimulation of keratinocytes with IL-22 plus TNF- α most effectively conserved the integrity of the epidermal barrier as compared with IFN- γ , IL-17, IL-22, or TNF- α alone [50]. IL-22 also functions to induce an acute phase systemic response that extends beyond IL-22R-expressing

cells and revealed diverse significant impact on coagulation and cellular constituents of blood, in addition to induction of thymic atrophy, body weight loss, and renal proximal tubule metabolic activity and biochemical changes in the liver, including induction of fibrinogen, CXCL1, and serum amyloid A [51]. Besides its contribution to protection against bacterial infection [50, 52], IL-22 plays an important role in protection against viral infection, for example, hepatitis B virus [53]. On the other side, IL-22 is implicated in the induction of IBD [54] and AD such as experimental autoimmune myocarditis [9] and psoriatic disease through the induction of keratinocyte proliferation and cytokine and chemokine release [7]. This reflects the dark side of the T_H17 story; that is, a promoted T_H17 response may reflect a current or predict incidence of AD.

2.2. T_H1-T_H17 Cells Interaction during Infection. Although a protective role against intracellular bacteria such as *Listeria monocytogenes* [55] or *S. typhimurium* (N. Y. A. Hemdan and A. M. Abu El-Saad, unpublished data) may be attributed to T_H17 response, this may be rather compensatory to a defective IL-12/IFN- γ axis as previously demonstrated by N. N. Orgun et al. [56], or a complementary function to indirectly induce type I response mediated by APCs endowing thereby a protection against infection, as in case of the obligate intracellular bacteria *Chlamydia muridarum* [57]. In case of infection with *S. typhimurium* and *C. muridarum*, neutralizing IL-17 significantly reduced pathogen-specific T_H1 but promoted higher T_H2 responses. DCs isolated from IL-17-neutralized

mice demonstrated lower expression of CD40, MHC II, and IL-12 production, but higher level of IL-10 compared with control mice [57]. Furthermore, neutralizing IL-17 in case of *S. typhimurium* significantly reduced phagocytosis as well as T_H1 cytokine production (N. Y. A. Hemdan and A. M. Abu El-Saad, unpublished data). Moreover, delivery of an IL-17R antagonist that resulted in a 50% reduction in the neutrophilic infiltration in lungs following *Chlamydia* infection reversed the susceptible phenotype of C3H/HeN mice [58], indicating a key role of IL-17 in induction of neutrophil infiltration. The compromised IL-17 response in HIS (Job's syndrome) patients that contributed to higher susceptibility to *Staphylococcus aureus* infection [59] is evidenced by a recent finding that coinfection with influenza A abrogated host defense, which was rescued by overexpression of IL-23 and markedly improved bacterial clearance [52]. Influenza A was found to inhibit T_H17 differentiation and substantially decreased IL-17, IL-22, and IL-23 production after *S. aureus* infection. Interestingly, IL-17-mediated cross-protection against secondary *L. monocytogenes* infection has been demonstrated following immunization with *Mycoplasma pulmonis* [60].

In addition to the function of T_H17 cells as a substitute for a defective T_H1 response, synergism between T_H17 and T_H1 cells is proposed following infection or postvaccination challenge with *Mycobacterium* sp., based on the observation that IL-17^{-/-} mice revealed a reduced IFN- γ production by CD4⁺ T cells and impaired granuloma formation and expression of chemokines CXCL9, CXCL10, and CXCL11 [61]. Also, an enhanced T_H1 memory response in the lungs of vaccinated mice infected with *M. tuberculosis* was dependent upon IL-23/IL-17 axis [61]. In a model of TCR $\alpha\beta$ ^{-/-} mouse [62], where adoptive transfer of either T_H1 or T_H17 cells restored bacterial burdens and innate immune cell infiltrates to wild-type animals level, T_H17 transferred cells revealed plasticity within the CNS compartment with an ultimate T_H1 -like cytokine profile, and this might be the reason for restoration of a strong innate immune response against infection with pyogenic bacteria; for review on various forms of T_H17 cell plasticity, refer to Hemdan [10]. Furthermore, the importance of T_H17 cells in combating HIV-associated bacterial infections has been recently elucidated [63].

2.3. Interaction of T_H17 Cells with Commensal Bacteria. Of a crucial importance is the recent clue linking the induction of T_H17 response with gut commensal bacteria. Colonization of the small intestine of mice with a single commensal microbe, segmented filamentous bacterium (SFB), was sufficient to induce IL-17- and IL-22-producing T_H17 cell responses in the lamina propria, and this was correlated with enhanced expression of inflammation- and antimicrobial-associated genes and increased resistance to the intestinal pathogens *Citrobacter* and *Salmonella* [35, 64]. Induction of T_H17 cells mediated autoimmune arthritis in K/BxN mice [65], whereas when the same mice were held under germ-free conditions, autoimmune arthritis was strongly attenuated and mice revealed reductions in serum AAs titers, splenic AAs-secreting cells, germinal centers, and splenic T_H17 cells

as well as the lack of T_H17 cells in the small intestinal lamina propria [65]. These findings suggest the role of T_H17 cells not only in defending the gastrointestinal tract against pathogens, but also in mediating AD (Table 1). How does the immune system monitor the resident intestinal microbes and coordinate between host defense and tolerance and how do dysregulated host-microbe interactions lead to intestinal inflammation were recently discussed [66]. The increased production of IL-17 and IL-23 by PBMCs derived from patients of primary Sjogren's syndrome upon TLR2, TLR4, and TLR6 stimulation [67] highlights the link between TLR ligation and autoimmune induction in such disease settings, where the participation of T_H17 cytokines in their pathogenesis is evident [68, 69].

2.4. T_H17 Cells in the Bone Disease. One of the most important contributions of T_H17 cells involves bone metabolism and bone disease. The coincidence of chronic inflammation and osteoporosis (OP) or osteoarthritis (OA) is quite anticipated and raised a debate about IL-17's contribution. Several hallmark inflammatory mediators including TNF- α , IL-1, IL-6, IFN- γ , receptor activator of NF- κ B (RANK), and RANK ligand (RANKL) are of crucial importance not only at the primary inflammation site, but also in bone metabolism [70, 71]. Although a protective role of IL-17 against bone loss has been described [72], induction of osteoclastogenesis by T_H17 cells has been suggested in various inflammatory models [73]. Proinflammatory cytokines correlated with osteoclastogenic or antiosteoclastogenic manifestations in human OP and OA, for example, negative correlations of hip bone mineral density (BMD) with TNF- α in OA and with RANKL/RANK in OP [70]. In a mouse model of type II diabetes, whereas osteocalcin and osteoprotegerin (osteoblast-specific bone forming markers) were decreased, osteoclast-driven bone resorption markers such as IL-6 and RANK were elevated and coincided with enhanced RANKL and IL-17 expression by CD4⁺ cells; IL-17 induction was directly promoted upon leptin treatment [74]. The authors proposed that leptin and IL-6 stimulate IL-17 production and, thereby, induce RANKL-mediated osteoclastogenesis. A direct link of IL-17 to osteoclast induction was proved in cultures of PBMCs drained from patients with Crohn's disease [71]. Altogether, IL-17 may be a valuable target for controlling bone diseases, at least those accompanying chronic inflammations as in Crohn's disease or inflammatory arthritis.

Overall, in addition to expecting counterprotective impacts of T_H17 -associated cytokines, the induction of T_H17 response seems to be an intrinsic feature originally evolved to fight bacterial, viral, and fungal infections. However, what drives such an immune arm to react against the body's own elements, that is, the loss of tolerance, remains elusive. Upon infection, it seems to be a failure to eliminate the invader, whereby an inflammation-potent cell response is amplified, whose army calls for other inflammation competent cells that might have lost the ability to recognize the body's own MHC molecules and therefore attack the self and/or induce production of AAs. Such a modified response attained through a persistent infection constitutes an additional load against the

system's strategy of pathogen clearance and the culmination of the immune response to its steady state thereafter. In other words, boosting such a potent inflammatory cell type as T_H17 through an initial inflammation, for example, through inflammatory cytokine-mediated induction of NF- κ B, see next, should normally be accompanied by induction of a regulation program; otherwise autoimmunity occurs. On the basis of current understanding, we propose that the T_H17 -driven autoimmune response is manifold, attaining its incidence through (i) activating T_H1 responses and the later conversion of T_H17 themselves into T_H1 -like cells or double T_H1/T_H17 cytokine producers having the inflammatory potency of both subsets; (ii) activating B cells and their production of AAs, especially through IL-21-dominated responses; (iii) inducing inflammatory cells like macrophages and neutrophils and their recruitment through induction of chemokines, facilitating thereby tissue destruction and release of intrinsic cellular factors, which, in turn, leads to local or systemic induction of the autoimmune traits; and (iv) promoting cytotoxicity of NK and CD8⁺ cells and the conversion of the latter cells into IL-17 producers that further magnify the whole response. On the basis of current knowledge, introducing the T_H17 efficacy as a potent inflammatory lineage, targeting IL-23/ T_H17 axis, may be a promising approach that paves the way for additive and alternative treatment of chronic inflammation and AD [75].

3. T_H17 Cells Are Key Players in Heavy-Metal-Elicited Autoimmunity

Whereas some heavy metals (e.g., copper, selenium, iron, and zinc) are essential to maintain our metabolism, the majority of heavy metals are non-essential, for example, arsenic (As^{3+} , As^{4+}), cadmium (Cd^{2+}), chromium (Cr^{3+} , Cr^{4+}), mercury (Hg^{2+}), and lead (Pb^{2+}), and are ranked among the most highly toxic substances. Great evidence exists that various heavy metals elicit immunomodulation increasing thereby the incidence of human AD and cancer [3, 18, 76]. It has been recently found that patients with autoimmune thyroiditis (AT) and other AD, including MS, psoriasis, SLE, and atopic eczema, showed increased lymphocyte reactivity to inorganic Hg^{2+} , Ni^{3+} , and other metals and that replacement of amalgam in Hg^{2+} -allergic subjects resulted in improvement of health in about 70% of AT patients [3]. Furthermore, recent data implied that exposure of mice to low micromolar concentrations of Cd^{2+} and Hg^{2+} induces a robust T_H17 response, that was also inferred by mild but significant increase of IL-17 profile in serum of individuals occupationally exposed to the same metals, as well as a robust *ex vivo* T_H17 response (N. Y. A. Hemdan et al., unpublished data).

3.1. Ligation of Metal Ion with the Aryl Hydrocarbon Receptor. Like other xenobiotic stimuli [77], one mechanism so far delineated is the ligation of metal ion with the aryl hydrocarbon receptor (AhR) as in the case of Cd^{2+} , As^{3+} , Cr^{6+} [78, 79], and Pb^{2+} [80]. Such ligand-specific activation of the AhR was found to regulate the balance between T_H17 and T_{Reg} cell responses [81]. Whereas AhR activation by TCDD

(dioxin) induces functional T_{Reg} cells that suppress EAE, activation by 6-formylindolo[3,2-b]carbazole induces T_H17 cells and ultimately the disease severity. AhR is expressed by T_H17 cells, $\gamma\delta$ T cells, and DCs [82, 83] and is indispensable for IL-22 production as evidenced by AhR^{-/-} mouse studies [82] and by downregulation of the AhR on RNA-mediated interference, as well as, by applying AhR agonists [48], where it substantially altered the balance of IL-22- versus IL-17-producing cells. In DCs, activation of AhR induces expression of IDO1 and IDO2 that mediate induction of T_{Reg} cells [83]. Therefore, we hypothesized that exposure to heavy metals may mediate autoimmune initiation/induction through metal ligation of AhR. Recent works of other groups and our unpublished data indicate the association of IL-22 with the appearance of autoimmune signs, as inferred in the pathogenesis of psoriasis [7]. These data raise the AhR as a sequential segment linking such potent inflammatory T_H17 cells with the heavy-metal-induced autoimmune induction and reveal a mechanism for further differentiation of T_H17 into T_H22 under organ- or pathogen-specific conditions, including AhR ligation pathways. Therefore, targeting AhR may offer a possibility for differential regulation of T_H17 cytokines and thereby reduction of autoimmune susceptibility in heavy-metal occupationally exposed individuals; however, the paradoxical effect of various AhR ligands should be considered.

AhR-mediated immunomodulation by heavy metals may involve several mechanisms; one of which is the regulation of CYP1A1 expression, for example, in the case of Pb^{2+} [80] that coincided with increase of heme-oxygenase- (HO-) 1 mRNA level and production of reactive oxygen species (ROS). Induction of CYP1A1 expression by AhR ligation is well documented; its downstream signal mediates cellular responses probably through modifying cytokine secretion, including IL-6 [84], IL-17, and IL-22 [37, 82, 85]. Induction of oxidative stress has been reported in case of Pb^{2+} [86], Cd^{2+} [87–89], and Hg^{2+} exposure [90], inferred by reduced activity of antioxidant enzymes catalase (CAT), superoxide dismutase (SOD), and glutathione peroxidase (GPx) and reduction of glutathione (GSH). Smoking, a major source of Cd [91], also initiates ROS production accompanied by augmented cell signaling pathways implicated in the pathogenesis of AD such as psoriasis that implicates mitogen-activated protein kinase (MAPK), NF- κ B and Janus kinase JAK/STAT [92], and reduced antioxidants malondialdehyde and SOD [93]. Oxidative stress comprising production of free radicals such as reactive oxygen and nitrogen species is closely related to inflammation and discussed as an underlying mechanism of inflammatory diseases, accompanying activation of the leucocytes and generation of peroxynitrite, at the early stages of induction, or appearing before the incidence of various AD such as IBD [94, 95], systemic sclerosis [96], SLE [97], psoriasis [93, 98], and in cardiovascular inflammation [99]. Therefore, depletion of endogenous antioxidants such as GSH, Cu, and ZnSOD was manifested in experimental models of IBD [100], accompanied, however, by induction of HO-1. The latter catalyzes CO production, and, although it

is considered as a prooxidant due to iron released from HO activity [101], it may be a compensatory response to oxidative stress and chronic inflammation [102], whose protective function has been elucidated in various AD including MS [103].

The protective role of antioxidants in combating inflammation has been clarified in various models by applying exogenous antioxidants or by manipulating expression of endogenous antioxidants. Depletion of NF-E2-related factor 2 (Nrf2), for instance, markedly enhanced susceptibility of experimental IBD [104]. Moreover, by disrupting GSH metabolism through targeting GSH peroxidase (GPx) 1 and GPx2 derived development of colitis [105], or by depleting GSH through curcumin-elicited glyoxalase 1 activity inhibition that enhanced the anti-inflammatory response as well as anticarcinogenic potency [106], it became clear how close the metabolic stress is related to cell response and cell survival, an assumption that is confirmed by many studies [107]. Unfortunately, although the role of T_H17 cells-related products such as IL-17 in the previous experimental settings has not been addressed, T_H17 together with T_H1 cells are drawn as being major players in the IBD as well as in other inflammatory concerts [108]. Therefore, the elevation of ROS levels links heavy metal exposure to induction of inflammation and cancer. Indeed, induction of T_H17 cell response by heavy metals is likely a downstream event of hydrogen-peroxide- (H_2O_2 -) mediated IL-6 induction, which is found to protect resident lung cells from ROS-induced injury [109]. Ye et al. [110] found that fumarates induced type II DCs as a result of initial GSH depletion followed by induction of HO-1, which interacts with AP-1 and NF- κ B sites of *IL23p19* promoter and inactivates STAT1 and thereby improves T_H1 - and T_H17 -mediated AD including MS and psoriasis.

3.2. Modification of NF- κ B Signaling. A key pathway through which heavy metals, amongst other xenobiotic substances, exert impacts on the immune response occurs via modifying NF- κ B signaling. Several distinct NF- κ B activation pathways are identified, including responses to various cell stresses and stimuli such as proinflammatory cytokines TNF- α and IL-1, bacterial products, genotoxic stimuli such as ionizing radiation and some chemotherapeutic drugs [88], in addition to exposure of various cell types to heavy metals such as Cd^{2+} [88, 111]. This may highlight NF- κ B activation as a trait of carcinogenicity assigned to heavy metals including Cd^{2+} [112]. The increased production of cytokines, for example, IL-6 and IL-17, in murine models and in heavy-metal-exposed individuals (Hemdan & Abul El-Saad, unpublished data), might ultimately lead to excessive induction of NF- κ B-mediated chronic inflammation [113]. This is consistent with the involvement of IL-17 in the differentiation of plasma cells mediated by NF- κ B-regulated TF Twist-1 [114]; we recall the correlation of higher IL-17 levels with the severity of various AD and the appearance of autoimmune signs accompanying exposure to heavy metals. Therefore, delicate intervention to regulate NF- κ B activation may help prevent chronic inflammation, simultaneous tissue cell damage, and reduce

incidence of AD in individuals occupationally exposed to heavy metals or those with an accidental exposure history.

3.3. Disruption of Ca^{2+} Homeostasis. A third event by which heavy metals like Cd^{2+} modify cell survival and function is modifying Ca^{2+} displacement and ultimately adherence/tight junctions, mediated by disrupting expression and translocation of E-cadherin/ β -catenin, in a way that mimics Wnt-signaling [88, 115], providing a clue for the carcinogenicity of heavy metals. Previous studies [116] as well as our unpublished data identified some protein kinases including ROCK-II as a target of Cd^{2+} - and Hg^{2+} -mediated induction of IL-17, which activates NF- κ B through CIKS/Act1 adaptor proteins, inducing thereby chronic inflammation and cancer.

3.4. Modification of Zinc Metabolism. A further important element in the interplay of heavy metals such as Cd^{2+} with immunomodulation is represented by its interaction with zinc. Zinc was found to inhibit cancer through regulating various oncogenic pathways including NF- κ B, AP-1, Notch-1, and PI3K/Akt, apoptosis, cytotoxicity, regulating tumor suppressors such as p53 and macrophage phagocytic activities and increased production of ROS and inflammatory cytokines TNF- α , IL-1 β , IL-8, VCAM, and MCP-1 [112]. Similarly, Zn probably inhibits STAT3 activation and thereby T_H17 -mediated collagen-induced arthritis [117]. Via targeting such oncogenic pathways, Zn supplements may participate in attaining promising antitumor approaches, at least in cases where Cd^{2+} is considered to have a carcinogenic potential.

4. Concluding Remarks

Ongoing research provides a preponderance of evidence that T_H17 cells and related molecules act as double agents both in favor of but also against the harboring individual. They elicit various antimicrobial mechanisms on one hand, but, on the other hand, when dysregulated, likely triggered by xenobiotic agents including pollutants and infectious agents, initiate/promote chronic inflammatory/autoimmune manifestations. A delineation of the underlying mechanisms that culminate into hyperactivation of T_H17 cells and the resultant production of related mediators would facilitate the development of potential therapeutic approaches to combat their deteriorating effects but simultaneously allow their benefits to act. Therefore, various research directions gave more attention in the last decade to T_H17 -cells-related molecules to help attain and evaluate valuable therapeutic strategies. Manipulating T_H17 differentiation and function by targeting differentiation/promoting cytokines, transcription factors, or commensal-bacteria-elicited immune induction may be valuable for treating AD; however, the risk of increasing the vulnerability of attacking infections should remain in focus. Therefore, it may be of worth to apply prophylactic antibacterial and antifungal therapy in case of treating patients of AD with IL-17/-22/-23 inhibitors.

Take-Home Messages

- (i) T_H17 cells and their related products act as double agents both to mediate various antimicrobial mechanisms and to initiate/promote chronic inflammatory/autoimmune manifestations.
- (ii) Simultaneous or successive exposure to xenobiotic substances and infectious agents renders the genetically susceptible individual vulnerable to autoimmune incidence via induction of inflammatory mediators.
- (iii) A compromise should be met to facilitate development of potential therapeutics aiming at targeting AD via inhibiting differentiation of T_H17 cells and/or commitment factors for the benefit of attaining the antimicrobial response intact, for example, by applying prophylactic therapy combined with IL-17/-22/-23 inhibitors.

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References

- [1] M. Schiraldi and M. Monestier, "How can a chemical element elicit complex immunopathology? Lessons from mercury-induced autoimmunity," *Trends in Immunology*, vol. 30, no. 10, pp. 502–509, 2009.
- [2] P. D. Pigatto and G. Guzzi, "Linking mercury amalgam to autoimmunity," *Trends in Immunology*, vol. 31, no. 2, pp. 48–49, 2010.
- [3] M. Hybenova, P. Hrdá, J. Prochazkova, V. Stejskal, and I. Sterzl, "The role of environmental factors in autoimmune thyroiditis," *Neuroendocrinology Letters*, vol. 31, no. 3, pp. 283–289, 2010.
- [4] S. M. Al-Mogairen, "Induction of autoimmunity in brown Norway rats by oral and parenteral administration of nickel chloride," *Lupus*, vol. 19, no. 3, pp. 262–267, 2010.
- [5] A. N. Layton and E. E. Galyov, "Salmonella-induced enteritis: molecular pathogenesis and therapeutic implications," *Expert Reviews in Molecular Medicine*, vol. 9, no. 18, pp. 1–17, 2007.
- [6] A. Haraga, M. B. Ohlson, and S. I. Miller, "Salmonellae interplay with host cells," *Nature Reviews Microbiology*, vol. 6, no. 1, pp. 53–66, 2008.
- [7] S. P. Raychaudhuri, "Role of IL-17 in psoriasis and psoriatic arthritis," *Clinical Reviews in Allergy and Immunology*, vol. 44, no. 2, pp. 183–193, 2013.
- [8] I. Kryczek, K. Wu, E. Zhao et al., "IL-17+ regulatory T Cells in the microenvironments of chronic inflammation and cancer," *Journal of Immunology*, vol. 186, no. 7, pp. 4388–4395, 2011.
- [9] N. Y. Hemdan, G. Birkenmeier, G. Wichmann et al., "Interleukin-17-producing T helper cells in autoimmunity," *Autoimmunity Reviews*, vol. 9, no. 11, pp. 785–792, 2010.
- [10] N. Y. Hemdan, "Anti-cancer versus cancer-promoting effects of the interleukin-17-producing T helper cells," *Immunology Letters*, vol. 149, pp. 123–133, 2013.
- [11] N. Sawada, M. Iwasaki, M. Inoue et al., "Long-term dietary cadmium intake and cancer incidence," *Epidemiology*, vol. 23, no. 3, pp. 368–376, 2012.
- [12] T. L. Van Belle, E. Esplugues, J. Liao, T. Juntti, R. A. Flavell, and M. G. von Herrath, "Development of autoimmune diabetes in the absence of detectable IL-17A in a CD8-driven virally induced model," *Journal of Immunology*, vol. 187, no. 6, pp. 2915–2922, 2011.
- [13] G. M. Delgoffe, K. N. Pollizzi, A. T. Waickman et al., "The kinase mTOR regulates the differentiation of helper T cells through the selective activation of signaling by mTORC1 and mTORC2," *Nature Immunology*, vol. 12, no. 4, pp. 295–304, 2011.
- [14] J. Ma, R. Wang, X. Fang, Y. Ding, and Z. Sun, "Critical role of TCF-1 in repression of the IL-17 gene," *PLoS ONE*, vol. 6, no. 9, Article ID e24768, 2011.
- [15] K. Hirota, J. H. Duarte, M. Veldhoen et al., "Fate mapping of IL-17-producing T cells in inflammatory responses," *Nature Immunology*, vol. 12, no. 3, pp. 255–263, 2011.
- [16] N. Y. A. Hemdan, F. Emmrich, K. Adham et al., "Dose-dependent modulation of the in vitro cytokine production of human immune competent cells by lead salts," *Toxicological Sciences*, vol. 86, no. 1, pp. 75–83, 2005.
- [17] N. Y. A. Hemdan, F. Emmrich, U. Sack et al., "The in vitro immune modulation by cadmium depends on the way of cell activation," *Toxicology*, vol. 222, no. 1–2, pp. 37–45, 2006.
- [18] N. Y. A. Hemdan, F. Emmrich, S. Faber, J. Lehmann, and U. Sack, "Alterations of Th1/Th2 reactivity by heavy metals: possible consequences include induction of autoimmune diseases," *Annals of the New York Academy of Sciences*, vol. 1109, pp. 129–137, 2007.
- [19] N. Y. Hemdan, "The role of interleukin-12 in the heavy metal-elicited immunomodulation: relevance of various evaluation methods," *Journal of Occupational Medicine and Toxicology*, vol. 3, no. 1, article 25, 2008.
- [20] M. A. Curotto de Lafaille and J. J. Lafaille, "Natural and adaptive Foxp3+ regulatory T cells: more of the same or a division of labor?" *Immunity*, vol. 30, no. 5, pp. 626–635, 2009.
- [21] S. Issazadeh-Navikas, R. Teimer, and R. Bockermann, "Influence of dietary components on regulatory T cells," *Molecular Medicine*, vol. 18, no. 1, pp. 95–110, 2012.
- [22] L. Maggi, V. Santarlasci, M. Capone et al., "CD161 is a marker of all human IL-17-producing T-cell subsets and is induced by RORC," *European Journal of Immunology*, vol. 40, no. 8, pp. 2174–2181, 2010.
- [23] N. Y. Hemdan, G. Birkenmeier, and G. Wichmann, "Key molecules in the differentiation and commitment program of T helper 17 (Th17) cells up-to-date," *Immunology Letters*, vol. 148, pp. 97–109, 2012.
- [24] V. Santarlasci, L. Maggi, M. Capone et al., "Rarity of human T helper 17 cells is due to retinoic acid orphan receptor-dependent mechanisms that limit their expansion," *Immunity*, vol. 36, no. 2, pp. 201–214, 2012.
- [25] M. Ankathatti Munegowda, Y. Deng, S. J. Mulligan, and J. Xiang, "Th17 and Th17-stimulated CD8+ T cells play a distinct role in Th17-induced preventive and therapeutic antitumor immunity," *Cancer Immunology, Immunotherapy*, vol. 60, no. 10, pp. 1473–1484, 2011.
- [26] L. G. Pinto, T. M. Cunha, S. M. Vieira et al., "IL-17 mediates articular hypernociception in antigen-induced arthritis in mice," *Pain*, vol. 148, no. 2, pp. 247–256, 2010.

- [27] S. T. Passos, J. S. Silver, A. C. O'Hara, D. Sehly, J. S. Stumhofer, and C. A. Hunter, "IL-6 promotes NK cell production of IL-17 during toxoplasmosis," *Journal of Immunology*, vol. 184, no. 4, pp. 1776–1783, 2010.
- [28] G. Sellge, J. G. Magalhaes, C. Konradt et al., "Th17 cells are the dominant T cell subtype primed by *Shigella flexneri* mediating protective immunity," *Journal of Immunology*, vol. 184, no. 4, pp. 2076–2085, 2010.
- [29] M. Raffatellu, R. L. Santos, D. E. Verhoeven et al., "Simian immunodeficiency virus-induced mucosal interleukin-17 deficiency promotes *Salmonella* dissemination from the gut," *Nature Medicine*, vol. 14, no. 4, pp. 421–428, 2008.
- [30] L. Rioll-Blanco, V. Lazarevic, A. Awasthi et al., "IL-23 receptor regulates unconventional IL-17-producing T cells that control bacterial infections," *Journal of Immunology*, vol. 184, no. 4, pp. 1710–1720, 2010.
- [31] T. Dejima, K. Shibata, H. Yamada et al., "Protective role of naturally occurring interleukin-17A-producing $\gamma\delta$ cells in the lung at the early stage of systemic candidiasis in mice," *Infection and Immunity*, vol. 79, no. 11, pp. 4503–4510, 2011.
- [32] A. J. Hueber, D. L. Asquith, A. M. Miller et al., "Cutting edge: mast cells express IL-17A in rheumatoid arthritis synovium," *Journal of Immunology*, vol. 184, no. 7, pp. 3336–3340, 2010.
- [33] J. Pötl, C. Botteron, E. Tausch et al., "Tracing functional antigen-specific CCR6⁺ Th17 cells after vaccination," *PLoS ONE*, vol. 3, no. 8, Article ID e2951, 2008.
- [34] X. Zhang, L. Gao, L. Lei et al., "A MyD88-dependent early IL-17 production protects mice against airway infection with the obligate intracellular pathogen *Chlamydia muridarum*," *Journal of Immunology*, vol. 183, no. 2, pp. 1291–1300, 2009.
- [35] K. Geddes, S. J. Rubino, J. G. Magalhaes et al., "Identification of an innate T helper type 17 response to intestinal bacterial pathogens," *Nature Medicine*, vol. 17, no. 7, pp. 837–844, 2011.
- [36] M. B. Torchinsky, J. Garaude, A. P. Martin, and J. M. Blander, "Innate immune recognition of infected apoptotic cells directs TH17 cell differentiation," *Nature*, vol. 458, no. 7234, pp. 78–82, 2009.
- [37] B. Martin, K. Hirota, D. J. Cua, B. Stockinger, and M. Veldhoen, "Interleukin-17-producing $\gamma\delta$ T cells selectively expand in response to pathogen products and environmental signals," *Immunity*, vol. 31, no. 2, pp. 321–330, 2009.
- [38] I. Godinez, M. Raffatellu, H. Chu et al., "Interleukin-23 orchestrates mucosal responses to *Salmonella enterica* serotype typhimurium in the intestine," *Infection and Immunity*, vol. 77, no. 1, pp. 387–398, 2009.
- [39] B. Clapp, J. A. Skyberg, X. Yang, T. Thornburg, N. Walters, and W. P. Pascual, "Protective live oral brucellosis vaccines stimulate Th1 and Th17 cell responses," *Infection and Immunity*, vol. 79, no. 10, pp. 4165–4174, 2011.
- [40] P. Ye, F. H. Rodriguez, S. Kanaly et al., "Requirement of interleukin 17 receptor signaling for lung CXC chemokine and granulocyte colony-stimulating factor expression, neutrophil recruitment, and host defense," *Journal of Experimental Medicine*, vol. 194, no. 4, pp. 519–527, 2001.
- [41] M. Miyamoto, O. Prause, M. Sjöstrand, M. Laan, J. Lötval, and A. Lindén, "Endogenous IL-17 as a mediator of neutrophil recruitment caused by endotoxin exposure in mouse airways," *Journal of Immunology*, vol. 170, no. 9, pp. 4665–4672, 2003.
- [42] H. Hoshino, M. Laan, M. Sjöstrand, J. Lötval, B. E. Skoogh, and A. Lindén, "Increased elastase and myeloperoxidase activity associated with neutrophil recruitment by IL-17 in airways in vivo," *Journal of Allergy and Clinical Immunology*, vol. 105, no. 1, pp. 143–149, 2000.
- [43] A. Puel, S. Cypowyj, J. Bustamante, J. F. Wright, L. Liu, H. K. Lim et al., "Chronic mucocutaneous candidiasis in humans with inborn errors of interleukin-17 immunity," *Science*, vol. 332, no. 6025, pp. 65–68, 2011.
- [44] L. Liu, S. Okada, X.-F. Kong et al., "Gain-of-function human STAT1 mutations impair IL-17 immunity and underlie chronic mucocutaneous candidiasis," *Journal of Experimental Medicine*, vol. 208, no. 18, pp. 1635–1648, 2011.
- [45] K. Kisand, A. S. Bøe Wolff, K. T. Podkrajsek et al., "Chronic mucocutaneous candidiasis in APECED or thymoma patients correlates with autoimmunity to Th17-associated cytokines," *Journal of Experimental Medicine*, vol. 207, no. 2, pp. 299–308, 2010.
- [46] A. Puel, R. Döflinger, A. Natividad et al., "Autoantibodies against IL-17A, IL-17F, and IL-22 in patients with chronic mucocutaneous candidiasis and autoimmune polyendocrine syndrome type I," *Journal of Experimental Medicine*, vol. 207, no. 2, pp. 291–297, 2010.
- [47] M. Muñoz, M. M. Heimesaat, K. Danker et al., "Interleukin (IL)-23 mediates *Toxoplasma gondii*-induced immunopathology in the gut via matrix metalloproteinase-2 and IL-22 but independent of IL-17," *Journal of Experimental Medicine*, vol. 206, no. 13, pp. 3047–3059, 2009.
- [48] S. Trifari, C. D. Kaplan, E. H. Tran, N. K. Crellin, and H. Spits, "Identification of a human helper T cell population that has abundant production of interleukin 22 and is distinct from TH-17, TH1 and TH2 cells," *Nature Immunology*, vol. 10, no. 8, pp. 864–871, 2009.
- [49] S. Eyerich, K. Eyerich, D. Pennino et al., "Th22 cells represent a distinct human T cell subset involved in epidermal immunity and remodeling," *Journal of Clinical Investigation*, vol. 119, no. 12, pp. 3573–3585, 2009.
- [50] S. Eyerich, J. Wagener, V. Wenzel et al., "IL-22 and TNF- α represent a key cytokine combination for epidermal integrity during infection with *Candida albicans*," *European Journal of Immunology*, vol. 41, no. 7, pp. 1894–1901, 2011.
- [51] S. C. Liang, C. Nickerson-Nutter, D. D. Pittman et al., "IL-22 induces an acute-phase response," *Journal of Immunology*, vol. 185, no. 9, pp. 5531–5538, 2010.
- [52] A. Kudva, E. V. Scheller, K. M. Robinson et al., "Influenza A inhibits Th17-mediated host defense against bacterial pneumonia in mice," *Journal of Immunology*, vol. 186, no. 3, pp. 1666–1674, 2011.
- [53] Y. Zhang, M. A. Cobleigh, J.-Q. Lian et al., "A proinflammatory role for interleukin-22 in the immune response to hepatitis B virus," *Gastroenterology*, vol. 141, no. 5, pp. 1897–1906, 2011.
- [54] L. A. Zenewicz, G. D. Yancopoulos, D. M. Valenzuela, A. J. Murphy, S. Stevens, and R. A. Flavell, "Innate and adaptive interleukin-22 protects mice from inflammatory bowel disease," *Immunity*, vol. 29, no. 6, pp. 947–957, 2008.
- [55] M. Miyamoto, M. Emoto, Y. Emoto et al., "Neutrophilia in LFA-1-deficient mice confers resistance to listeriosis: possible contribution of granulocyte-colony-stimulating factor and IL-17," *Journal of Immunology*, vol. 170, no. 10, pp. 5228–5234, 2003.
- [56] N. N. Orgun, M. A. Mathis, C. B. Wilson, and S. W. Sing, "Deviation from a strong Th1-dominated to a modest Th17-dominated CD4 T cell response in the absence of IL-12p40 and type I IFNs sustains protective CD8 T cells," *Journal of Immunology*, vol. 180, no. 6, pp. 4109–4115, 2008.

- [57] H. Bai, J. Cheng, X. Gao et al., "IL-17/Th17 promotes type 1 T cell immunity against pulmonary intracellular bacterial infection through modulating dendritic cell function," *Journal of Immunology*, vol. 183, no. 9, pp. 5886–5895, 2009.
- [58] X. Zhou, Q. Chen, J. Moore, J. K. Kolls, S. Halperin, and J. Wang, "Critical role of the interleukin-17/interleukin-17 receptor axis in regulating host susceptibility to respiratory infection with *Chlamydia* species," *Infection and Immunity*, vol. 77, no. 11, pp. 5059–5070, 2009.
- [59] Y. Minegishi, M. Saito, M. Nagasawa et al., "Molecular explanation for the contradiction between systemic Th17 defect and localized bacterial infection in hyper-IgE syndrome," *Journal of Experimental Medicine*, vol. 206, no. 6, pp. 1291–1301, 2009.
- [60] A. N. Sieve, K. D. Meeks, S. Bodhankar et al., "A novel IL-17-dependent mechanism of cross protection: respiratory infection with mycoplasma protects against a secondary listeria infection," *European Journal of Immunology*, vol. 39, no. 2, pp. 426–438, 2009.
- [61] S. A. Khader, G. K. Bell, J. E. Pearl et al., "IL-23 and IL-17 in the establishment of protective pulmonary CD4⁺ T cell responses after vaccination and during *Mycobacterium tuberculosis* challenge," *Nature Immunology*, vol. 8, no. 4, pp. 369–377, 2007.
- [62] M. M. Holley and T. Kielian, "Th1 and Th17 cells regulate innate immune responses and bacterial clearance during central nervous system infection," *Journal of Immunology*, vol. 188, no. 3, pp. 1360–1370, 2012.
- [63] M. Salgado, N. I. Rallón, B. Rodés, M. López, V. Soriano, and J. M. Benito, "Long-term non-progressors display a greater number of Th17 cells than HIV-infected typical progressors," *Clinical Immunology*, vol. 139, no. 2, pp. 110–114, 2011.
- [64] I. I. Ivanov, K. Atarashi, N. Manel et al., "Induction of intestinal Th17 cells by segmented filamentous bacteria," *Cell*, vol. 139, no. 3, pp. 485–498, 2009.
- [65] H.-J. Wu, I. I. Ivanov, J. Darce et al., "Gut-residing segmented filamentous bacteria drive autoimmune arthritis via T helper 17 cells," *Immunity*, vol. 32, no. 6, pp. 815–827, 2010.
- [66] C. Abraham and R. Medzhitov, "Interactions between the host innate immune system and microbes in inflammatory bowel disease," *Gastroenterology*, vol. 140, no. 6, pp. 1729–1737, 2011.
- [67] S.-K. Kwok, M.-L. Cho, Y.-M. Her et al., "TLR2 ligation induces the production of IL-23/IL-17 via IL-6, STAT3 and NF- κ B pathway in patients with primary Sjögren's syndrome," *Arthritis Research and Therapy*, vol. 14, no. 2, article R64, 2012.
- [68] F. Ciccia, G. Guggino, A. Rizzo et al., "Potential involvement of IL-22 and IL-22-producing cells in the inflamed salivary glands of patients with Sjögren's syndrome," *Annals of the Rheumatic Diseases*, vol. 71, no. 2, pp. 295–301, 2012.
- [69] W. H. Abdulahad, F. G. M. Kroese, A. Vissink, and H. Bootsma, "Immune regulation and B-cell depletion therapy in patients with primary Sjögren's syndrome," *Journal of Autoimmunity*, vol. 39, pp. 103–111, 2012.
- [70] J. Zupan, R. Komadina, and J. Marc, "The relationship between osteoclastogenic and anti-osteoclastogenic pro-inflammatory cytokines differs in human osteoporotic and osteoarthritic bone tissues," *Journal of Biomedical Science*, vol. 19, no. 1, article 28, 2012.
- [71] A. E. Oostlander, V. Everts, T. Schoenmaker et al., "T cell-mediated increased osteoclast formation from peripheral blood as a mechanism for crohn's disease-associated bone loss," *Journal of Cellular Biochemistry*, vol. 113, no. 1, pp. 260–268, 2012.
- [72] J. Goswami, N. Hernández-Santos, L. A. Zuniga, and S. L. Gaffen, "A bone-protective role for IL-17 receptor signaling in ovariectomy-induced bone loss," *European Journal of Immunology*, vol. 39, no. 10, pp. 2831–2839, 2009.
- [73] F.-L. Yuan, X. Li, W.-G. Lu et al., "Type 17 T-helper cells might be a promising therapeutic target for osteoporosis," *Molecular Biology Reports*, vol. 39, no. 1, pp. 771–774, 2012.
- [74] H. Y. Won, J.-A. Lee, Z. S. Park et al., "Prominent bone loss mediated by RANKL and IL-17 produced by CD4⁺ T cells in tallyho/JngJ mice," *PLoS ONE*, vol. 6, no. 3, Article ID e18168, 2011.
- [75] É. Toussiot, "The IL23/Th17 pathway as a therapeutic target in chronic inflammatory diseases," *Inflammation and Allergy*, vol. 11, no. 2, pp. 159–168, 2012.
- [76] J. J. Putila and N. L. Guo, "Association of arsenic exposure with lung cancer incidence rates in the United States," *PLoS ONE*, vol. 6, no. 10, Article ID e25886, 2011.
- [77] K. W. Bock and C. Köhle, "Ah receptor: dioxin-mediated toxic responses as hints to deregulated physiologic functions," *Biochemical Pharmacology*, vol. 72, no. 4, pp. 393–404, 2006.
- [78] R. H. Elbekai and A. O. S. El-Kadi, "Modulation of aryl hydrocarbon receptor-regulated gene expression by arsenite, cadmium, and chromium," *Toxicology*, vol. 202, no. 3, pp. 249–269, 2004.
- [79] F. M. Kluxen, N. Höfer, G. Kretzschmar, G. H. Degen, and P. Diel, "Cadmium modulates expression of aryl hydrocarbon receptor-associated genes in rat uterus by interaction with the estrogen receptor," *Archives of Toxicology*, vol. 86, pp. 591–601, 2012.
- [80] H. M. Korashy and A. O. S. El-Kadi, "Transcriptional and posttranslational mechanisms modulating the expression of the cytochrome P450 1A1 gene by lead in HepG2 cells: a role of heme oxygenase," *Toxicology*, vol. 291, no. 1–3, pp. 113–121, 2012.
- [81] F. J. Quintana, A. S. Basso, A. H. Iglesias et al., "Control of Treg and TH17 cell differentiation by the aryl hydrocarbon receptor," *Nature*, vol. 453, no. 7191, pp. 65–71, 2008.
- [82] M. Veldhoen, K. Hirota, A. M. Westendorf et al., "The aryl hydrocarbon receptor links TH17-cell-mediated autoimmunity to environmental toxins," *Nature*, vol. 453, no. 7191, pp. 106–109, 2008.
- [83] C. F. A. Vogel, S. R. Goth, B. Dong, I. N. Pessah, and F. Matsumura, "Aryl hydrocarbon receptor signaling mediates expression of indoleamine 2,3-dioxygenase," *Biochemical and Biophysical Research Communications*, vol. 375, no. 3, pp. 331–335, 2008.
- [84] B. C. DiNatale, I. A. Murray, J. C. Schroeder et al., "Kynurenine acid is a potent endogenous aryl hydrocarbon receptor ligand that synergistically induces interleukin-6 in the presence of inflammatory signaling," *Toxicological Sciences*, vol. 115, no. 1, pp. 89–97, 2010.
- [85] J.-M. Ramirez, N. C. Bremilla, O. Sorg et al., "Activation of the aryl hydrocarbon receptor reveals distinct requirements for IL-22 and IL-17 production by human T helper cells," *European Journal of Immunology*, vol. 40, no. 9, pp. 2450–2459, 2010.
- [86] I. Baranowska-Bosiacka, I. Gutowska, M. Marchlewicz et al., "Disrupted pro- and antioxidative balance as a mechanism of neurotoxicity induced by perinatal exposure to lead," *Brain Research*, vol. 1435, pp. 56–71, 2012.
- [87] H. Oguzturk, O. Ciftci, M. Aydin, N. Timurkaan, A. Beytur, and F. Yilmaz, "Ameliorative effects of curcumin against acute cadmium toxicity on male reproductive system in rats," *Andrologia*, vol. 44, pp. 243–249, 2012.

- [88] F. Thévenod, "Cadmium and cellular signaling cascades: to be or not to be?" *Toxicology and Applied Pharmacology*, vol. 238, no. 3, pp. 221–239, 2009.
- [89] M. Galazyn-Sidorczuk, M. M. Brzóska, J. Rogalska, A. Roszczenko, and M. Jurczuk, "Effect of zinc supplementation on glutathione peroxidase activity and selenium concentration in the serum, liver and kidney of rats chronically exposed to cadmium," *Journal of Trace Elements in Medicine and Biology*, vol. 26, no. 1, pp. 46–52, 2012.
- [90] M. Pal and M. Ghosh, "Studies on comparative efficacy of α -linolenic acid and α -eleostearic acid on prevention of organic mercury-induced oxidative stress in kidney and liver of rat," *Food and Chemical Toxicology*, vol. 50, no. 3–4, pp. 1066–1072, 2012.
- [91] M. Tellez-Plaza, A. Navas-Acien, K. L. Caldwell, A. Menke, P. Muntner, and E. Guallar, "Reduction in cadmium exposure in the United States population, 1988–2008: the contribution of declining smoking rates," *Environmental Health Perspectives*, vol. 120, no. 2, pp. 204–209, 2012.
- [92] A. W. Armstrong, E. J. Armstrong, E. N. Fuller, M. E. Sockolov, and S. V. Voyles, "Smoking and pathogenesis of psoriasis: a review of oxidative, inflammatory and genetic mechanisms," *British Journal of Dermatology*, vol. 165, no. 6, pp. 1162–1168, 2011.
- [93] E. Attwa and E. Swelam, "Relationship between smoking-induced oxidative stress and the clinical severity of psoriasis," *Journal of the European Academy of Dermatology and Venereology*, vol. 25, no. 7, pp. 782–787, 2011.
- [94] B. Khor, A. Gardet, and R. J. Xavier, "Genetics and pathogenesis of inflammatory bowel disease," *Nature*, vol. 474, no. 7351, pp. 307–317, 2011.
- [95] H. Zhu and Y. R. Li, "Oxidative stress and redox signaling mechanisms of inflammatory bowel disease: updated experimental and clinical evidence," *Experimental Biology and Medicine*, vol. 237, no. 5, pp. 474–480, 2012.
- [96] T. Yamamoto, "Autoimmune mechanisms of scleroderma and a role of oxidative stress," *Self/Nonself*, vol. 2, no. 1, pp. 4–10, 2011.
- [97] K. W. Mathis, M. Venegas-Pont, C. W. Masterson, N. J. Stewart, K. L. Wasson, and M. J. Ryan, "Oxidative stress promotes hypertension and albuminuria during the autoimmune disease systemic lupus erythematosus," *Hypertension*, vol. 59, no. 3, pp. 673–679, 2012.
- [98] S. A. Gabr and A. H. Al-Ghadir, "Role of cellular oxidative stress and cytochrome c in the pathogenesis of psoriasis," *Archives of Dermatological Research*, vol. 304, no. 6, pp. 451–457, 2012.
- [99] E. Profumo, B. Buttari, and R. Rigano, "Oxidative stress in cardiovascular inflammation: its involvement in autoimmune responses," *International Journal of Inflammation*, vol. 2011, Article ID 295705, 6 pages, 2011.
- [100] Ç. A. İşman, B. Ç. Yeğen, and İ. Alican, "Methimazole-induced hypothyroidism in rats ameliorates oxidative injury in experimental colitis," *Journal of Endocrinology*, vol. 177, no. 3, pp. 471–476, 2003.
- [101] R. J. Castellani, P. I. Moreira, G. Perry, and X. Zhu, "The role of iron as a mediator of oxidative stress in Alzheimer disease," *BioFactors*, vol. 38, no. 2, pp. 133–138, 2012.
- [102] N. G. Abraham and A. Kappas, "Pharmacological and clinical aspects of heme oxygenase," *Pharmacological Reviews*, vol. 60, no. 1, pp. 79–127, 2008.
- [103] P. Fagone, K. Mangano, M. Coco et al., "Therapeutic potential of carbon monoxide in multiple sclerosis," *Clinical and Experimental Immunology*, vol. 167, no. 2, pp. 179–187, 2012.
- [104] T. O. Khor, M.-T. Huang, K. H. Kwon, J. Y. Chan, B. S. Reddy, and A.-N. Kong, "Nrf2-deficient mice have an increased susceptibility to dextran sulfate sodium-induced colitis," *Cancer Research*, vol. 66, no. 24, pp. 11580–11584, 2006.
- [105] R. S. Esworthy, R. Aranda, M. G. Martín, J. H. Binder, S. W. Doroshov, and F.-F. Chu, "Mice with combined disruption of gpx1 and gpx2 genes have colitis," *The American Journal of Physiology*, vol. 281, no. 3, pp. G848–G855, 2001.
- [106] T. Santel, G. Pflug, N. Y. A. Hemdan et al., "Curcumin inhibits glyoxalase 1: a possible link to its anti-inflammatory and anti-tumor activity," *PLoS ONE*, vol. 3, no. 10, Article ID e3508, 2008.
- [107] C.-C. Chen, H.-L. Chen, C.-W. Hsieh, Y.-L. Yang, and B.-S. Wung, "Upregulation of NF-E2-related factor-2-dependent glutathione by carnosol provokes a cytoprotective response and enhances cell survival," *Acta Pharmacologica Sinica*, vol. 32, no. 1, pp. 62–69, 2011.
- [108] T. Olsen, R. Rismo, G. Cui, R. Goll, I. Christiansen, and J. Florholmen, "TH1 and TH17 interactions in untreated inflamed mucosa of inflammatory bowel disease, and their potential to mediate the inflammation," *Cytokine*, vol. 56, no. 3, pp. 633–640, 2011.
- [109] H. Kida, M. Yoshida, S. Hoshino et al., "Protective effect of IL-6 on alveolar epithelial cell death induced by hydrogen peroxide," *The American Journal of Physiology*, vol. 288, no. 2, pp. L342–L349, 2005.
- [110] K. Ghoreschi, J. Brück, C. Kellerer et al., "Fumarates improve psoriasis and multiple sclerosis by inducing type II dendritic cells," *Journal of Experimental Medicine*, vol. 208, no. 11, pp. 2291–2303, 2011.
- [111] J. R. Napolitano, M.-J. Liu, S. Bao et al., "Cadmium-mediated toxicity of lung epithelia is enhanced through NF- κ B-mediated transcriptional activation of the human zinc transporter ZIP8," *The American Journal of Physiology*, vol. 302, no. 9, pp. L909–L918, 2012.
- [112] B. Bao, A. Thakur, Y. Li et al., "The immunological contribution of NF- κ B within the tumor microenvironment: a potential protective role of zinc as an anti-tumor agent," *Biochimica et Biophysica Acta*, vol. 1825, no. 2, pp. 160–172, 2012.
- [113] L. Li, L. Huang, A. L. Vergis et al., "IL-17 produced by neutrophils regulates IFN- γ -mediated neutrophil migration in mouse kidney ischemia-reperfusion injury," *Journal of Clinical Investigation*, vol. 120, no. 1, pp. 331–342, 2010.
- [114] A. Doreau, A. Belot, J. Bastid et al., "Interleukin 17 acts in synergy with B cell-activating factor to influence B cell biology and the pathophysiology of systemic lupus erythematosus," *Nature Immunology*, vol. 10, no. 7, pp. 778–785, 2009.
- [115] I. Lindner, N. Y. A. Hemdan, and M. Buchold, " α 2-macroglobulin inhibits the malignant properties of astrocytoma cells by impeding β -catenin signaling," *Cancer Research*, vol. 70, no. 4, pp. 277–287, 2010.
- [116] S. U. Sønder, S. Saret, W. Tang, D. E. Sturdevant, S. F. Porcella, and U. Siebenlist, "IL-17-induced NF- κ B activation via CIKS/Act1 physiologic significance and signaling mechanisms," *Journal of Biological Chemistry*, vol. 286, no. 15, pp. 12881–12890, 2011.
- [117] C. Kitabayashi, T. Fukada, M. Kanamoto et al., "Zinc suppresses Th17 development via inhibition of STAT3 activation," *International Immunology*, vol. 22, no. 5, pp. 375–386, 2010.
- [118] P. L. Simonian, C. L. Roark, F. Wehrmann et al., "IL-17A-expressing T cells are essential for bacterial clearance in a murine model of hypersensitivity pneumonitis," *Journal of Immunology*, vol. 182, no. 10, pp. 6540–6549, 2009.

- [119] M. D. Woolard, L. L. Hensley, T. H. Kawula, and J. A. Frelinger, "Respiratory *Francisella tularensis* live vaccine strain infection induces Th17 cells and prostaglandin E2, which inhibits generation of gamma interferon-positive T cells," *Infection and Immunity*, vol. 76, no. 6, pp. 2651–2659, 2008.
- [120] A. D. Joshi, D. J. Fong, S. R. Oak et al., "Interleukin-17-mediated immunopathogenesis in experimental hypersensitivity pneumonitis," *The American Journal of Respiratory and Critical Care Medicine*, vol. 179, no. 8, pp. 705–716, 2009.
- [121] P. L. Simonian, C. L. Roark, F. Wehrmann et al., "Th17-polarized immune response in a murine model of hypersensitivity pneumonitis and lung fibrosis," *Journal of Immunology*, vol. 182, no. 1, pp. 657–665, 2009.
- [122] F. Wang, J. Xu, Y. Liao et al., "Tim-3 ligand galectin-9 reduces IL-17 level and accelerates *Klebsiella pneumoniae* infection," *Cellular Immunology*, vol. 269, no. 1, pp. 22–28, 2011.
- [123] X. Chen, M. Zhang, M. Liao et al., "Reduced Th17 response in patients with tuberculosis correlates with IL-6R expression on CD4+ T cells," *The American Journal of Respiratory and Critical Care Medicine*, vol. 181, no. 7, pp. 734–742, 2010.
- [124] Z. Liu, L. Yang, Y. Cui et al., "IL-21 enhances NK cell activation and cytolytic activity and induces Th17 cell differentiation in inflammatory bowel disease," *Inflammatory Bowel Diseases*, vol. 15, no. 8, pp. 1133–1144, 2009.
- [125] T. Sugihara, A. Kobori, H. Imaeda et al., "The increased mucosal mRNA expressions of complement C3 and interleukin-17 in inflammatory bowel disease," *Clinical and Experimental Immunology*, vol. 160, no. 3, pp. 386–393, 2010.
- [126] M. N. Kelly, J. K. Kolls, K. Happel et al., "Interleukin-17/interleukin-17 receptor-mediated signaling is important for generation of an optimal polymorphonuclear response against *Toxoplasma gondii* infection," *Infection and Immunity*, vol. 73, no. 1, pp. 617–621, 2005.
- [127] J. L. Werner, A. E. Metz, D. Horn et al., "Requisite role for the dectin-1 β -glucan receptor in pulmonary defense against *aspergillus fumigatus*," *Journal of Immunology*, vol. 182, no. 8, pp. 4938–4946, 2009.
- [128] Y.-H. Jin, H. S. Kang, M. Mohindru, and B. S. Kim, "Preferential induction of protective T cell responses to Theiler's virus in resistant (C57BL/6 X SJL)F1 mice," *Journal of Virology*, vol. 85, no. 6, pp. 3033–3040, 2011.
- [129] S. Mukherjee, D. M. Lindell, A. A. Berlin et al., "IL-17-Induced pulmonary pathogenesis during respiratory viral infection and exacerbation of allergic disease," *The American Journal of Pathology*, vol. 179, no. 1, pp. 248–258, 2011.
- [130] Z. Huang, J. C. van Velkinburgh, B. Ni, and Y. Wu, "Pivotal roles of the interleukin-23/T helper 17 cell axis in hepatitis B," *Liver International*, vol. 32, pp. 894–901, 2012.
- [131] M. A. Jimenez-Sousa, R. Almansa, C. de la Fuente et al., "Increased Th1, Th17 and pro-fibrotic responses in hepatitis C-infected patients are down-regulated after 12 weeks of treatment with pegylated interferon plus ribavirin," *European Cytokine Network*, vol. 21, no. 2, pp. 84–91, 2010.
- [132] L. Campillo-Gimenez, M.-C. Cumont, M. Fay et al., "AIDS progression is associated with the emergence of IL-17-producing cells early after simian immunodeficiency virus infection," *Journal of Immunology*, vol. 184, no. 2, pp. 984–992, 2010.
- [133] A. Elhed and D. Unutmaz, "Th17 cells and HIV infection," *Current Opinion in HIV and AIDS*, vol. 5, no. 2, pp. 146–150, 2010.
- [134] D. Chege, P. M. Sheth, T. Kain et al., "Sigmoid Th17 populations, the HIV latent reservoir, and microbial translocation in men on long-term antiretroviral therapy," *AIDS*, vol. 25, no. 6, pp. 741–749, 2011.
- [135] A. Suryawanshi, T. Veiga-Parga, N. K. Rajasagi et al., "Role of il-17 and th17 cells in herpes simplex virus-induced corneal immunopathology," *Journal of Immunology*, vol. 187, no. 4, pp. 1919–1930, 2011.
- [136] Y. Fu, W. Wang, J. Tong et al., "Th17: a new participant in gut dysfunction in mice infected with *trichinella spiralis*," *Mediators of Inflammation*, vol. 2009, Article ID 517052, 7 pages, 2009.