Role of Mast Cells and Type 2 Innate Lymphoid (ILC2) Cells in Lung Transplantation

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The multifunctional role of mast cells (MCs) in the immune system is complex and has not fully been explored. MCs reside in tissues and mucous membranes such as the lung, digestive tract, and skin which are strategically located at interfaces with the external environment. These cells, therefore, will encounter external stimuli and pathogens. MCs modulate both the innate and the adaptive immune response in inflammatory disorders including transplantation. MCs can have pro- and anti-inflammatory functions, thereby regulating the outcome of lung transplantation through secretion of mediators that allow interaction with other cell types, particularly innate lymphoid cells (ILC2). ILC2 cells are a unique population of hematopoietic cells that coordinate the innate immune response against a variety of threats including infection, tissue damage, and homeostatic disruption. In addition, MCs can modulate alloreactive T cell responses or assist in T regulatory (Treg) cell activity. This paper outlines the current understanding of the role of MCs in lung transplantation, with a specific focus on their interaction with ILC2 cells within the engrafted organ.

1. Introduction

1.1. Mast Cells. The multifunctional role of mast cells (MCs) within the immune system has been clarified since their discovery by Paul Ehrlich in 1878 [1–3]. CD34+ progenitor cells circulate in the blood and migrate into peripheral tissues where they further differentiate into mature MCs under the influence of various tissue-specific factors such as extracellular matrix proteins, adhesion molecules, cytokines, and chemokines [4]. MCs act as key immune and inflammatory sentinels by initiating and shaping the inflammatory response through the rapid activation of IgE-dependent and -independent innate immune pathways [5–8]. The most well-known MC activation pathway involves IgE/FceRI signaling, but MCs are also triggered via pattern recognition receptors such as Toll-like receptors (TLRs), complement, neuropeptides, cytokines, and many other stimuli [9].

MCs are present in all tissues and are particularly abundant in tissues and mucous membranes, such as the lung and digestive tract. MCs have this strategic later location in order to respond to external inflammatory stimuli and pathogens [4, 10, 11]. MCs can produce growth factors, costimulatory
molecules, and numerous pro- and anti-inflammatory mediators. These cells are heterogeneous in nature [12], and the response to an external stressor may be altered by the local microenvironment. MCs release >200 mediators including prestored factors, such as histamine [2] and tryptase [13], as well as de novo synthesized such as chemokines and cytokines in response to allergic or nonimmune triggers [14, 15].

The important role of MCs in both the innate and adaptive immune responses [16–19] has led to speculation that MCs may play a crucial role in organ allograft rejection [17, 20–23]. In contrast to other immune-competent organs, the transplanted lung is constantly exposed to airborne antigens that may activate the local immune response and thereby modulate MC activity [10, 24–29]. For example, activated MCs release IL-2 [30], IL-7 [31], IL-3, IL-6, IL-9, IL-10, IFN-γ, and TNF-α and chemokines (CXCL8, CCL2, and CCL5) which have all been implicated in organ transplant and rejection [10, 14, 15, 25, 32]. In addition, MCs may enhance chronic rejection by the induction of fibrotic pathways [33] in the lung [29], kidney [34–36], and heart [37, 38].

Regulatory T cells (Tregs) are essential in maintaining tolerance to self-antigens, preventing excessive immune responses and in abrogating autoimmunity during graft rejection [39–41]. The use of MC-deficient mice has emphasized the important role of MCs in the activation of Treg-mediated immunoregulatory activities during transplant rejection [42]. In agreement with this, the absence of MCs is associated with significantly reduced cardiac allograft survival after heterotopic heart transplantation in rats [43]. Mechanistically, this may involve the ability of MCs to act as antigen-presenting cells and to mediate allograft reactions [12, 44].

Activated MCs influence the activity of many other cell types [45]. In turn, the function of MCs is controlled by factors such as proteases, complement [46], TLR ligands [47], and stem cell factor (SCF) released by other immune cells and by structural cells such as fibroblasts and smooth muscle cells. These factors either prime MCs for mediator release or directly induce MC degranulation [48].

MCs are histologically categorized into two phenotypes based on their protease content termed MC-tryptase (MCT) and MC-tryptase/chymase (MCTC) [24]. However, it remains unclear which MC phenotype is involved in regulating transplant rejection. The phenotype of MCs varies over time following transplantation with the MCTC being the main phenotype implicated in chronic rejection after fibrosis in the transplanted kidney [49]. Indeed, the phenotypic shift from MCT to MCTC cells may be associated with a progressive and potentially irreversible decline in allograft function [50].

These data together indicate that MCs are important immune effector cells during lung allograft rejection, but the role of these cells in organ transplant rejection is still not completely clear. Type 2 innate lymphoid cells (ILC2) cells are found in the vicinity of MCs in lung tissue, and both cell types can communicate with each other [51]. In addition, ILC2s are involved in epithelial and lung tissue repair [52, 53] and ILC2 are found in the lung parenchyma and bronchoalveolar lavage (BAL) fluid of subjects undergoing lung transplant [54]. In this review, we discuss how MCs and ILC2 can modulate transplant rejection of the lung.

2.1. Innate Lymphoid Cells (ILCs). ILCs are a novel population of hematopoietic cells [55] that develop from common lymphoid progenitors in fetal liver and bone marrow [56, 57]. These cells are multifunctional and found throughout the body but are more prominent at barrier surfaces such as the lung and mucosal membranes [54, 58, 59]. Three types of ILCs exist (ILC1, 2, and 3), and these are functionally analogous to T-helper (Th) 1, Th2, and Th17 cell subsets [54, 60]. ILCs have a lymphoid morphology and release similar profiles of cytokines and eicosanoids as their respective Th cells but lack the T cell antigen receptor [60, 61]. Exposure of ILC progenitors (ILCP) to cytokines such as IL-25 and IL-33 induces ILC2 cells which are able to release cytokines IL-5, IL-9, and IL-13 [32, 54].

In the lung, ILC2s are mainly localized to the epithelium and perform a variety of protective immune functions [62, 63]. For example, ILC2s and their cytokines play critical roles in the protection of airway epithelial cells (ECs) against pathogens and regulate the repair of damaged cells [52, 64]. Since ILC2s have a protective role by organizing the innate immune response against infection and tissue damage, it is likely that they are involved in regulating transplant rejection [54, 65, 66].

Expansion of ILC2s is driven by exposure to numerous immune factors including the cytokines IL-2 [67], IL-4 [68], IL-25, IL-33 [69–71], thymic stromal lymphopoietin (TSLP) [72], IL-9 [52, 73], IL-1β [69], and TNF-like ligand 1A [55]. In addition, eicosanoids such as prostaglandin D2 (PGD2) and leukotriene D4 (LTD4) [69] can drive the development of ILC2s. In contrast, inflammatory or immune suppressors such as montelukast, corticosteroids, prostaglandin L2, IL-27, IFN-γ, and lipoxin A4 suppress ILC2 proliferation and cytokine production [70, 74, 75] (Figure 1).

2. Role of MCs and ILC2s in Rejection/ Survival of the Transplanted Lung

Since the first successful lung transplant in 1983 [76], the number of operations has grown substantially [77]. A lung transplant is generally the final treatment option for patients with end-stage lung disease. Various types of injury can damage a grafted organ. Some processes are due to the surgical procedure itself, for example, the sectioning of vessels and nerves or, in the transplanted lung, of the conducting airways. Other processes are inflammatory in nature, due to reperfusion of the graft or the onset of early allogeneic reactions. The lack of efficient tissue repair mechanisms could severely impair graft functioning, and the events involved in restoration of the transplanted airways utilize a variety of cell types [78].

2.1. The Role of IL-33 and IL-13. After lung transplant, IL-33 is released into the extracellular space which results in the activation of immune and inflammatory cells such as ECs,
dendritic cells (DCs), MCs, ILC2s, and CD4+ T cells [79–82]. IL-33 is an alarm signal that triggers ECs in the lung and other cells present at the mucosal barrier, to reverse or prevent cell damage [20, 83, 84]. In addition, IL-33 recruits and activates cytokine production by ILC2s and by MCs [80, 82]. For example, ATP, released from the damaged epithelium and acting via the P2X7 receptor [66, 69, 83], in combination with IL-33, triggers the production of IL-13 by ILC2s. MCs can also release IL-33 following IgE cross-linking [51, 85]. IL-33, therefore, acts as a “sensor of cell injury” via MCs [86], and, in a feed-forward manner, activates MCs to produce IL-2. MC-derived IL-2 enhances the activation of Th9 cells and further releases IL-9 in a feed-forward manner. In addition, IL-2 released from MCs induces tolerance in Treg cells and regulates transplantation survival. Activated ILC2s, MCs, and Th2 cells release Areg which is important in promoting EC repair from injury. Damaged epithelial cells, seen during transplant rejection, release IL-33 and ATP, which together can act on ILC2 and MCs to enhance their activity. A number of inhibitors including montelukast, IL-27, corticosteroid, PGI2, and lipoxin A4, can suppress the activation and/or proliferation of ILC2s and their release of inflammatory mediators. Abbreviations: Areg: amphiregulin; ECs: epithelial cells; IF: interleukin; ILC2: type 2 innate lymphoid cell; MCs: mast cells; PG12: prostaglandin I2; Th9: T-helper type 9; Treg: T regulatory cells.

Figure 1: The role of Th9 cells and IL-9 in lung transplantation. IL-9 released by Th9 cells act on MCs to produce IL-2. MC-derived IL-2 leads to the expansion of CD45+ ILC2, which enhances the activation of Th9 cells and the further release of IL-9 in a feed-forward manner. In addition, IL-2 released from MCs induces tolerance in Treg cells and regulates transplantation survival. Activated ILC2s, MCs, and Th2 cells release Areg which is important in promoting EC repair from injury. Damaged epithelial cells, seen during transplant rejection, release IL-33 and ATP, which together can act on ILC2 and MCs to enhance their activity. A number of inhibitors including montelukast, IL-27, corticosteroid, PGI2, and lipoxin A4, can suppress the activation and/or proliferation of ILC2s and their release of inflammatory mediators. Abbreviations: Areg: amphiregulin; ECs: epithelial cells; IF: interleukin; ILC2: type 2 innate lymphoid cell; MCs: mast cells; PG12: prostaglandin I2; Th9: T-helper type 9; Treg: T regulatory cells.
Human MCs produce lipid mediators such as PGD2 and LTD4 following FcεRI activation [31, 94–96]. PGD2 stimulates ILC2 migration into the lung and drives the production of type 2 cytokines via its receptor CRTH2 [97] which is a distinctive marker of human ILC2s [31, 98]. In contrast, IL-2 produced by ILC2s assists Treg survival [30, 99] and thus supports survival of the transplanted organ. In addition, activated lung ECs release PGD2 to enable recruitment of ILC2s, basophils, MCs, and Th2 cells into the inflamed airway [100].

ILC2s produce amphiregulin (Areg) which promotes epithelial cell repair [34, 101, 102]. Areg is also produced by MCs and Th2 cells which further indicates a critical feed-forward interaction within the MC-ILC2 nexus [91, 101, 103] (Figure 1). Conversely, ECs release factors such as IL-25, IL-33, and TSLP which drive ILC [91, 104]. IL-25 and IL-33 induce different types of ILCs. IL-33 induces ILC2s whereas IL-25 preferentially elicits multipotent progenitor (MPP-) type 2 cells, a new population of innate cells which also promote type 2 immunity [105]. IL-7 has also been described as a crucial factor in the development of ILC2s [31]. The in vivo sources of IL-7 required for ILC development are unknown, but IL-7 is critical for the generation and maintenance of all lymphocytes and is expressed by stromal cells [106].

These data highlight the interplay between MCs and ILC2s in maintaining the integrity of the respiratory epithelium and restoring the lung during infection of the transplanted lung [78, 107, 108] (Figure 2).

### 3. Possible Role of MCs and ILC2s in Lung Allograft Rejection

Due to the lung’s distinctive anatomic position, long-term graft survival is comparatively lower than with other solid organs such as the heart, liver, and kidney [76]. According to the 2016 report, a half-life of heart, kidney, and liver transplants endures around 12 years but the median survival after lung transplantation in the same condition is around 5.8 years [109]. This may result from injury occurring during the lung transplant and the lack of organized tissue repair mechanisms which together damage the grafted organ [78] or an immunological reaction to the foreign organ leading to graft dysfunction and failure [110].

#### 3.1. The Role of Fibrosis

Lung allograft rejection occurs due to both acute (AR) and chronic (CR) rejection processes [111–113]. Lung MCs play a dual role in the transplanted lung being implicated in both the induction of organ rejection and the induction of immune tolerance [10, 14, 50]. In contrast to the role of MC in AR, various studies have correlated CR with the fibrosis-inducing activity of MCs [10]. The proinflammatory cytokines TGF-β, IL-13, IL-1β, IL-17A, and IL-37 all contribute to the fibrotic [20]. IL-25 is also believed to be important as it induces a dramatic increase in both IL-13 and TGF-β expression in the lungs [70, 114].

Obliterative bronchiolitis (OB) occurs when graft survival is compromised after transplant of the donor lung [27, 113]. OB is the major cause of allograft rejection affecting at least 60% of recipients within 5 years of transplant [27, 115]. The release of profibrotic mediators such as TGF-β1 and basic fibroblast growth factor (bFGF) by MCs led to the examination of MCs in OB [116]. In addition to TGF-β1 and bFGF, other MC mediators including IL-4, TNF-α, histamine, heparin, chymase, and cathepsin G stimulate fibroblast proliferation and/or induce collagen synthesis [11, 31, 115, 117].

MCs and other immune cells accumulate around the vessels and airways during AR [118, 119]. Furthermore, MC hyperplasia occurs in areas of luminal fibrosis in both AR and CR of human lung allografts and is associated with the release of bFGF and histamine [29]. MC stabilization using cromolyn prevented the development of chronic lung allograft rejection in rats, again emphasizing the critical role of MCs in this process [95, 96]. Not only has bFGF been implicated in driving fibrosis-induced chronic lung allograft rejection but enhanced expression of bFGF may be a biomarker of rejection [37, 120]. Finally, TGF-β1 acts cooperatively with IL17 in fibrosis [115] and clinical observations indicate that TGF-β1 expression predicts the failure/success of the organ transplant [115, 121].

#### 3.2. The Role of Th2 Cytokines

The cytokines IL-4 [10] and IL-13 [98], produced directly or indirectly by MCs and ILC2, are important during the development of chronic lung rejection [115]. IL-13 can drive transplant rejection due to fibrosis [31, 98]. ILC2-derived IL-13 promotes the migration of activated DCs into the local lymph nodes and thereby induce naïve T cells to differentiate into Th2 cells and increase IL-13 and IL-4 production [31, 122]. MC-derived IL-4 is an inducer of fibroblast activation during the development of chronic rejection [123].

In AR of lung allografts, MCs may increase allo-specific T cell responses which are unfavorable for the engrafted organ [22]. MCs activate T cells by presenting the antigen either directly in the context of MHC II [27, 124] or indirectly through the release of cytokines [10]. Numerous mediators produced by MCs and/or ILC2s [10, 31, 55, 60] have been implicated in either the survival or the rejection of transplanted lungs [125]. Many of these mediators act on T cells; for example, IL-4 and TNF-α can augment MHC-II on antigen-presenting cells (APC) and in the presence of IFNα induce T cell proliferation [116]. In addition to TNF-α inducing the recruitment of T cells and enhancing their interaction with antigen-presenting cells [10], MC-derived TNF-α can drive donor-derived DCs toward a tolerogenic phenotype [126]. Conversely, a reduction in the release of IL-10 and TGF-β1 from T cells supports the development of acute rejection [10] in part by effects on Tregs [127].

In summary, donor T cells are continually primed and activated to react against the host causing graft-versus-host disease (GvHD) that leads to tissue damage and death [40]. Finally, MC-derived mediators can upregulate the expression of adhesion molecules such as VCAM-1 and ICAM-1 on endothelial and granulocytes and enhance the transendothelial migration of T cells. This effect of MC products on adhesion molecule expression is amplified by crosstalk with ILC2s [52, 116].
4. Conclusion

ILC2s are juxtaposed with MCs in the lung and directly communicate with MCs to induce the release of numerous mediators from both cell types that are implicated in either the survival or the rejection of transplanted lungs. MCs affect ILC2 activities directly by the release of PGD2 or indirectly via the release of the proteases chymase and tryptase to promote IL-13 production and that of the other Th2 cytokines. IL-13 acts in a synergistic manner with IL-33 released from airway ECs to reverse or prevent tissue damage during the transplant. Moreover, IL-13 can induce epithelial cell hyperproliferation and collagen deposition leading to pulmonary fibrosis. IL-4 released by MC also triggers fibroblast activation leading to lung fibrosis. MC-derived TGF-β1 further enhances fibrotic activity. Abbreviations: Areg: amphiregulin; DCs: dendritic cells; ECs: epithelial cell; IL: interleukin; LTD4: leukotriene D4; PGD2: prostaglandin D2; TGF-β1: transforming growth factor beta; Th2: T helper type 2 cell; TSLP: thymic stromal lymphopoietin.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References


