

Research Article

A Comprehensive Prognostic and Immune Analysis of Ferroptosis-Related Genes Identifies SLC7A11 as a Novel Prognostic Biomarker in Lung Adenocarcinoma

Li Qian,¹ Fu Wang,² Shu-min Lu,³ Hua-jie Miao,¹ Xin He,¹ Jia Feng,¹ Hua Huang,¹ Rong-feng Shi ,⁴ and Jian-guo Zhang ¹

¹Department of Pathology, Affiliated Hospital of Nantong University, Nantong, Jiangsu, China

²Department of Traditional Chinese Medicine, Nantong Maternity and Child Health Care Hospital Affiliated to Nantong University, Nantong, Jiangsu, China

³Department of Oncology, Xinhua Hospital Affiliated to Shanghai Jiao Tong University School of Medicine, Shanghai, China

⁴Department of Interventional Radiology, Affiliated Hospital of Nantong University, Nantong, Jiangsu, China

Correspondence should be addressed to Rong-feng Shi; shirongfeng140@hotmail.com and Jian-guo Zhang; jgz_edu@163.com

Received 28 January 2022; Revised 2 April 2022; Accepted 4 April 2022; Published 25 April 2022

Academic Editor: Dawei Cui

Copyright © 2022 Li Qian et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Lung adenocarcinoma (LUAD) is still one of the illnesses with the greatest mortality and morbidity. As a recently identified mode of cellular death, the activation of ferroptosis may promote the effectiveness of antitumor therapies in several types of tumors. However, the expression and clinical significance of Ferroptosis-associated genes in LUAD are still elusive. The RNA sequencing data of LUAD and relevant clinical data were downloaded from The Cancer Genome Atlas (TCGA) datasets. Subsequently, potential prognostic biomarkers were determined by the use of biological information technology. The R software package “ggalluvial” was applied to structure Sanguini diagram. Herein, our team screened 14 dysregulated ferroptosis-associated genes in LUAD. Among them, only four genes were associated with clinical outcome of LUAD patients, including ATP5MC3, FANCD2, GLS2, and SLC7A11. In addition, we found that high SLC7A11 expression predicted an advanced clinical progression in LUAD patients. Additionally, 8 immune checkpoint genes and 7 immune cells for LUAD were recognized to be related to the expression of SLC7A11. KEGG assays indicated that high expression of SLC7A11 might participate in the modulation of intestinal immune network for IgA generation and Staphylococcus aureus infection. Overall, our findings revealed that SLC7A11 might become a potentially diagnostic biomarker and SLC7A11 might serve as an independent prognosis indicator for LUAD.

1. Introduction

Pulmonary carcinoma is the leading cause of tumor-related death across the globe [1]. Lung adenocarcinoma (LUAD) predominantly arises from small airway epitheliums and type II alveolar cells, accounting for 45% of lung cancer [2, 3]. The progresses in the therapy of NSCLC sufferers, like surgery, radiotherapy, chemical therapy, targeted treatment, and immune checkpoint suppressor treatment, have been reported; nevertheless, the 5-year survival is still below 15%, mainly due to cancer metastasis [4, 5]. In addition, up to now, the causal links as to the occurrence and progression

of pancreas carcinoma remain elusive [6]. It is imperative to reveal potential molecule-level causal links and identify valid treatment targets and new prognosis markers for LUAD.

Recently, certain researches have explored cancer ferroptosis [7]. Ferroptosis, an iron-dependent pathway of cellular death, is different from novel types of apoptotic cellular death like apoptosis, pyrolysis, and autophagy [8, 9]. Ferroptosis is induced via the cumulation of lipoperoxidation products and cytotoxic ROS originated from Fe metabolic activity [10, 11]. Fe metabolic activity and lipoperoxidation signal transmission are identified as the predominant mediating

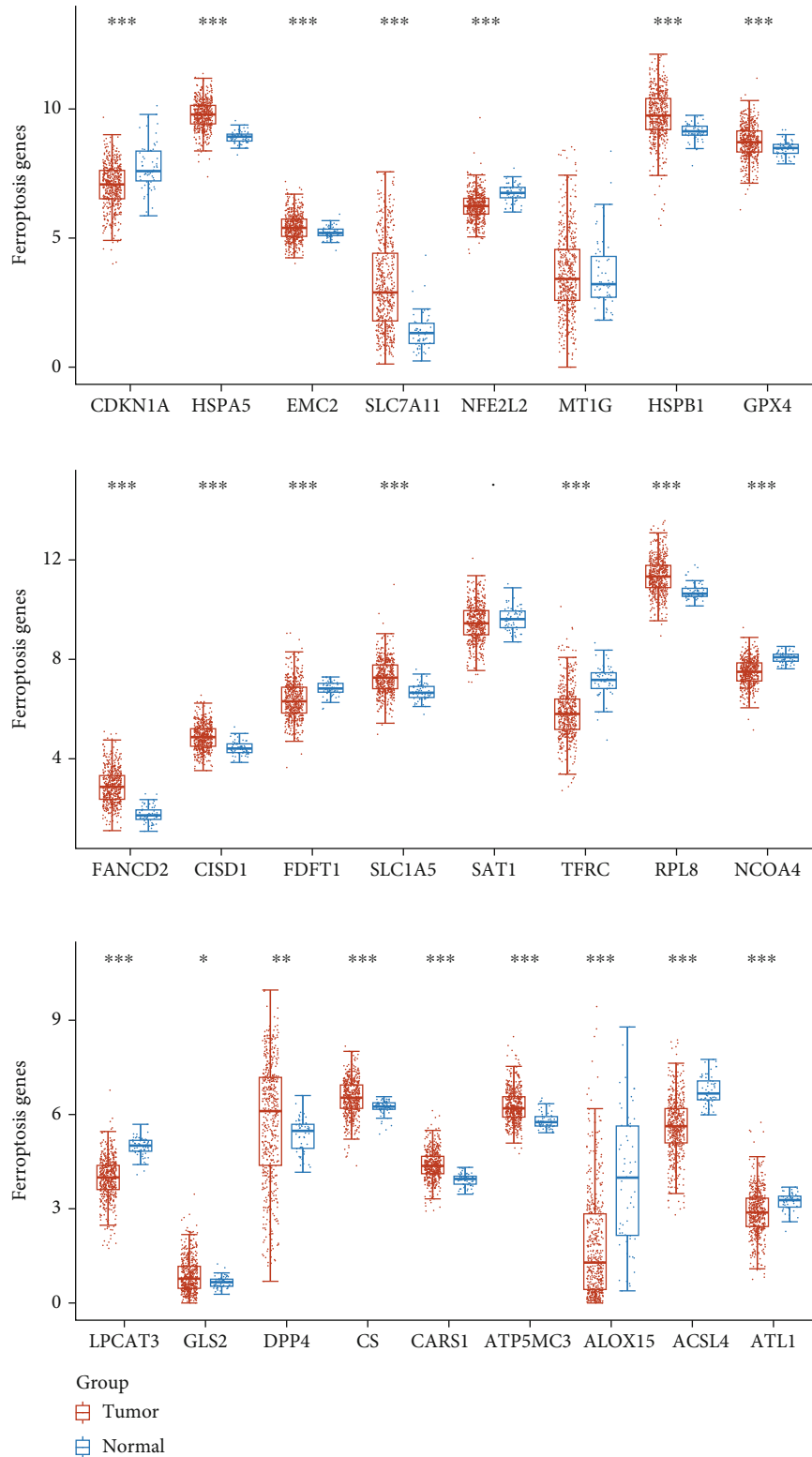


FIGURE 1: The expressing pattern of ferroptosis-associated genes in LUAD using TCGA datasets. *** $p < 0.001$, ** $p < 0.01$, and* $p < 0.05$.

factors of ferroptosis [12, 13]. Ferroptosis induces mitochondria function disorder and cytotoxic lipoperoxidation, which are pivotal for the inhibition of tumor progression [14, 15]. Several ferroptosis-related genes have been functionally

identified [16, 17]. For instance, NRF2 modulates the susceptibility of mankind pulmonary carcinoma cells to cystine deprivation-triggered ferroptosis through the FOCAD-FAK signal path [18]. Therefore, investigating the ferroptosis-

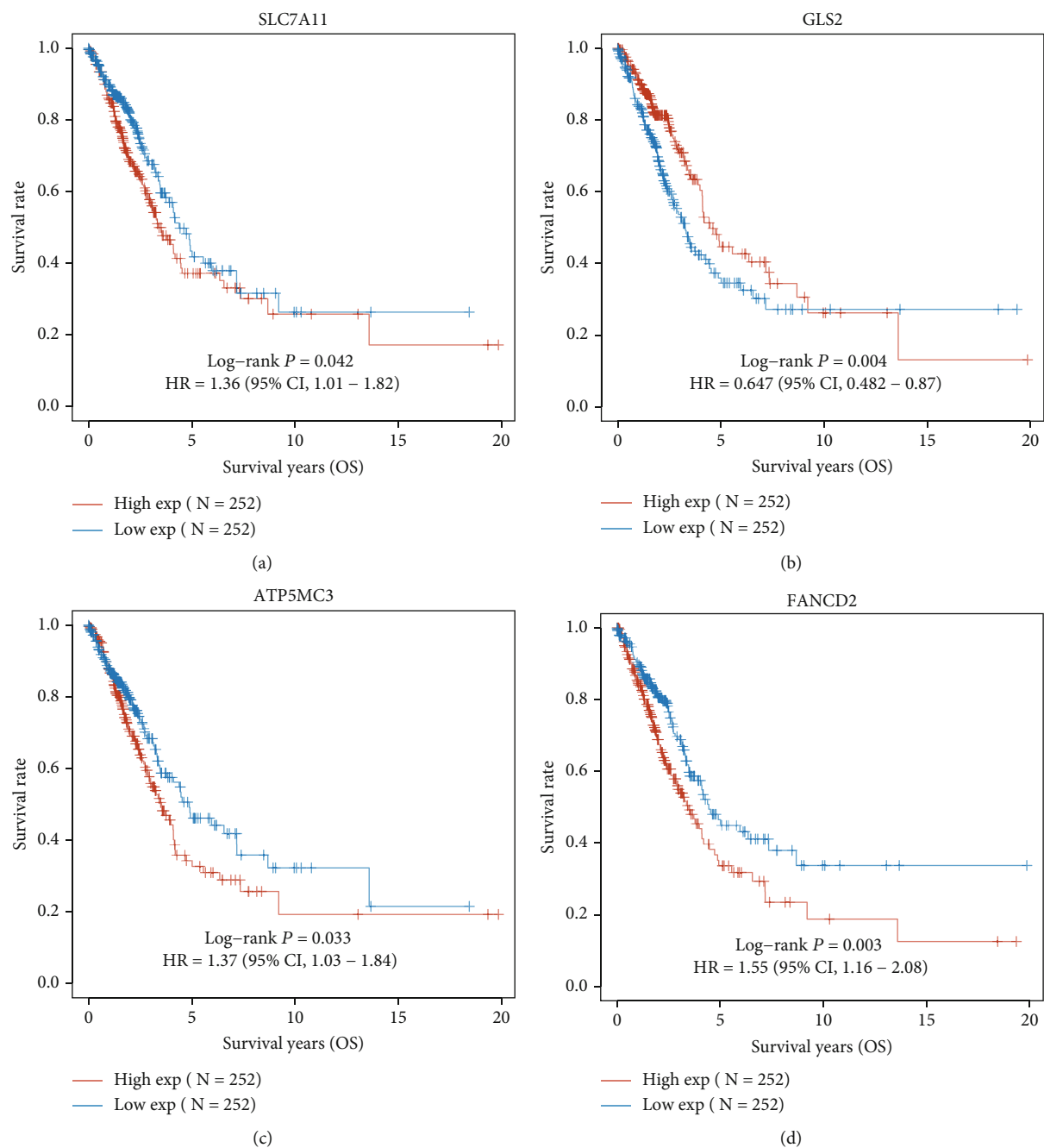


FIGURE 2: The survival-associated genes in LUAD were identified using Kaplan-Meier methods, including (a) SLC7A11, (b) GLS2, (c) ATP5MC3, and (d) FANCD2.

related gene expression profile and its prognostic value in LUAD may develop new strategies for the treatment of LUAD.

Herein, our team analyzed the expressing pattern of ferroptosis-associated genes using TCGA datasets and identified 24 dysregulated genes in LUAD. Then, we further explore their prognostic values. By now, there still have been few reports concerning the expression and function of SLC7A11 in human LUAD. Our attention focused on SLC7A11, and we aimed to explore the possible association between SLC7A11 expression and immune activity.

2. Materials and Methods

2.1. Data Collection. Level 3 raw counts of the RNA sequencing data, cancer mutation burden, aneuploidy scores, and relevant clinic information from an overall 513 LUAD sufferers were obtained from TCGA. Clinical variables, like sex, age, and pathology phase, were assessed as well. Ferroptosis-related genes are derived from GSEA (<http://www.gsea-msigdb.org>).

2.2. Screening of the Abnormally Expressed Ferroptosis-Related Genes in LUAD. For the sake of identifying the

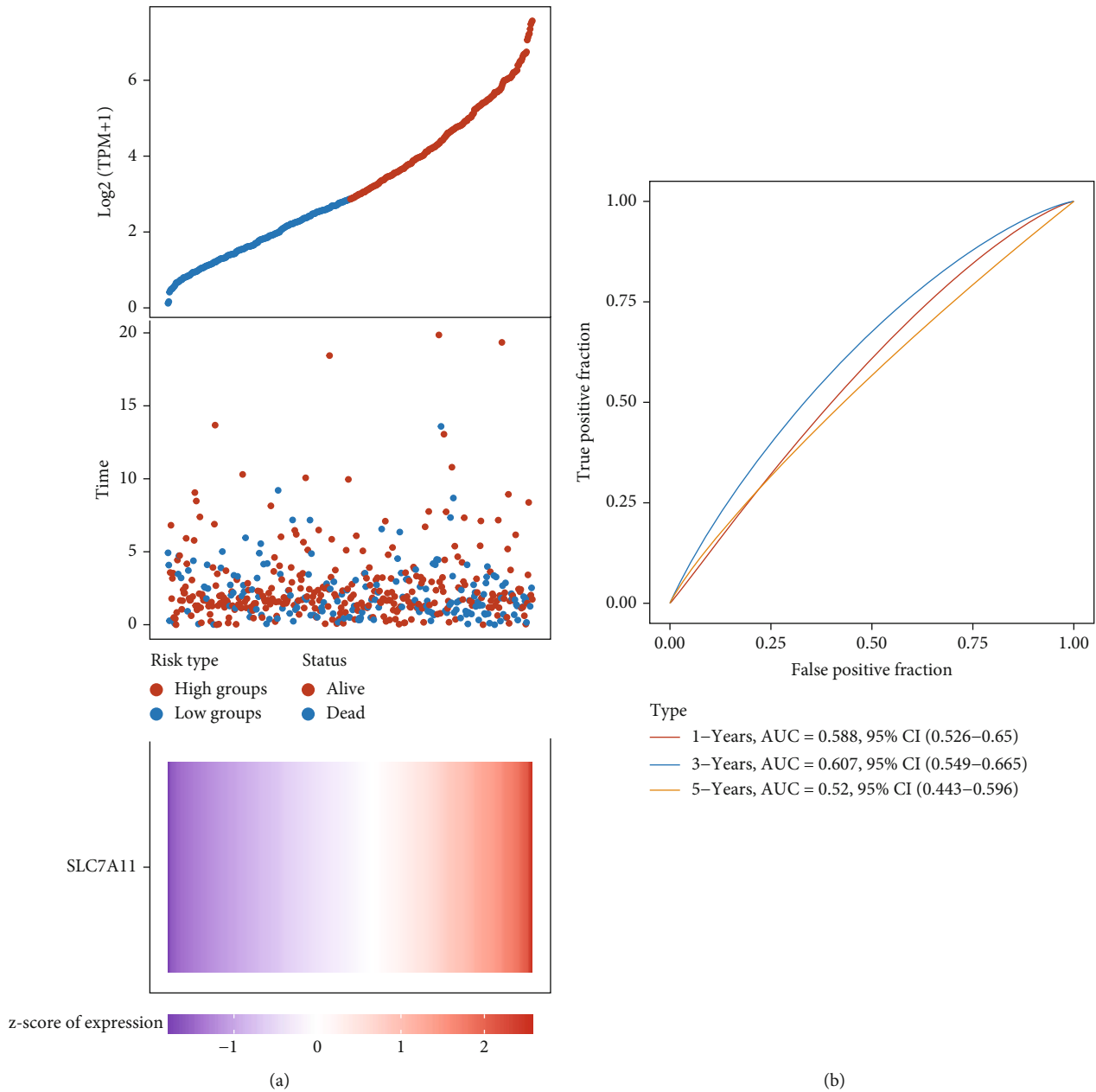


FIGURE 3: (a) The curve of risk scoring. Survival status of the patients. (b) Time-reliant ROC analyses for SLC7A11.

aberrantly expressed ferroptosis-related genes, R program 4.0.5 was leveraged to contrast the expression profiles of LUAD samples to healthy samples. DEG analyses of individual genes were completed via edgeR Bioconductor package [14]. The $|\log_2(\text{fold change [FC]})| > 2$, p value < 0.01 , and $\text{FDR} < 0.01$ were the liminal values for the determination of the aberrantly expressed ferroptosis-related genes.

2.3. Screening of Ferroptosis-Associated and Survival-Associated Genes. The association between the expressions of the aberrantly expressed ferroptosis-associated genes and survival time of LUAD was investigated. The survival function was plotted with the Kaplan-Meier plots. The entire analysis approaches aforesaid were completed via the “sur-

vival” and “survminer” R package [19]. $p < 0.05$ had significance on statistics.

2.4. The Association between SCL7A11 and Clinicopathological Parameters. The association between the expression of SCL7A11 and clinicopathological variables was analyzed through box plots and Sankey diagrams via independent specimen nonparametric assays in the TCGA cohort. Sankey diagrams were drawn via “ggforce” package in R.

2.5. TIMER Database Analysis. TIMER is a database for the analysis of immunocyte infiltrates in several tumors. Such database uses pathology test-verified statistic approach to speculate cancer immunity infiltration via neutrophilic cells, macrophages, dendritic cells, B cells, and CD4/CD8 T cells.

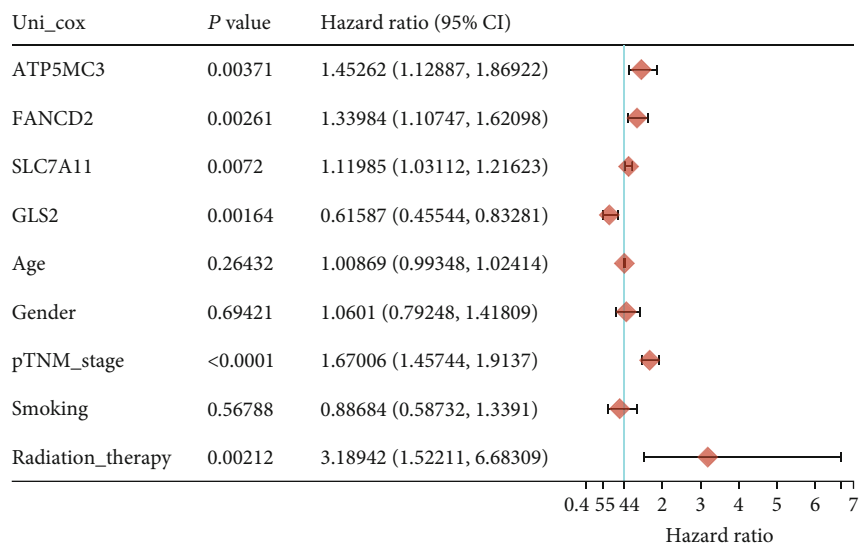


FIGURE 4: Univariable Cox regressive analyses were completed on all clinical covariates to identify its impact on survival.

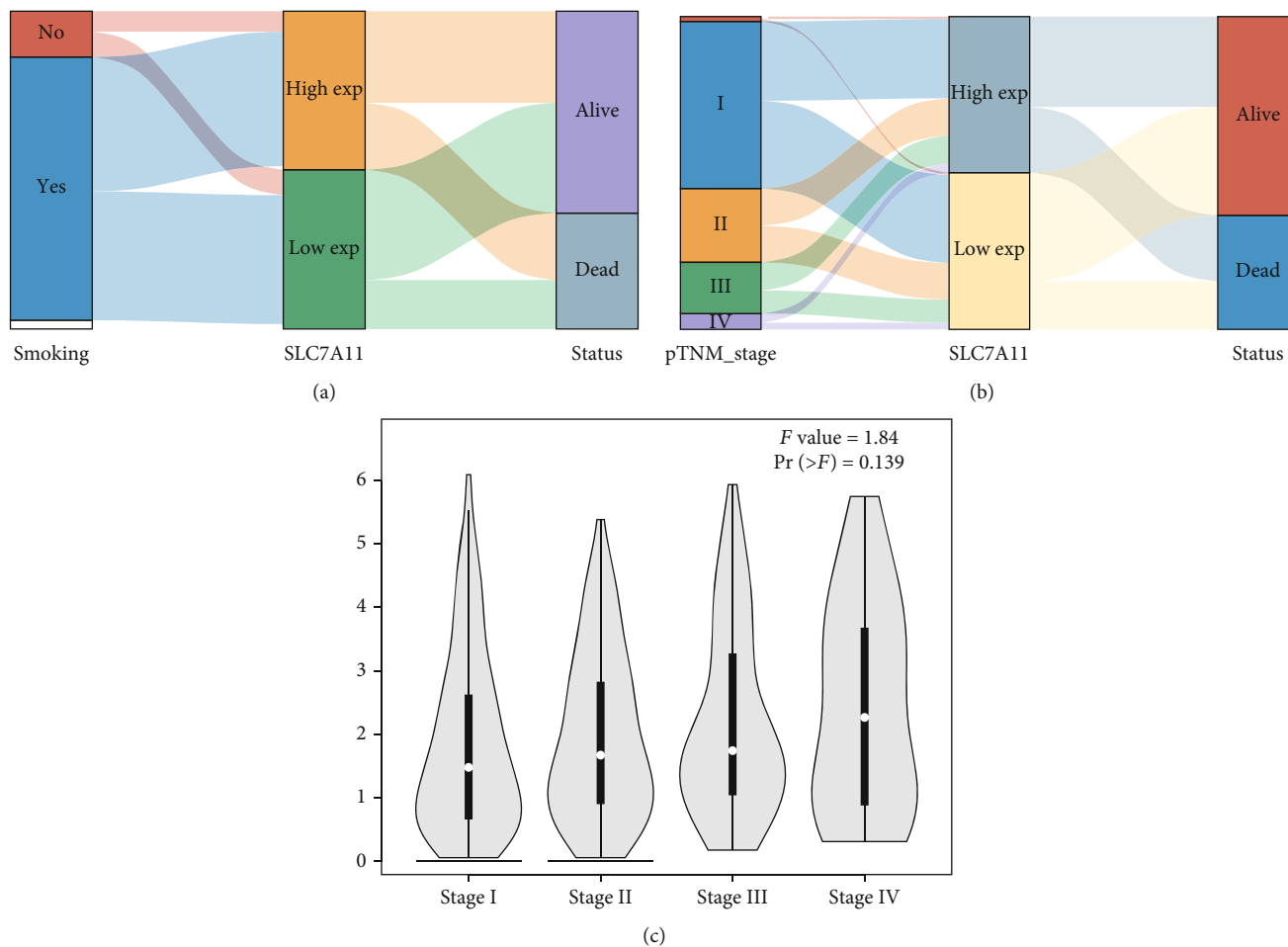


FIGURE 5: The correlation between SLC7A11 and clinicopathological parameters in LUAD. Sankey diagrams between SLC7A11 and (a) Smoking and (b) pTNM_stage. (c) High expression of SCL7A11 was identified in LUAD samples with phases III-IV in contrast to phases I-II.

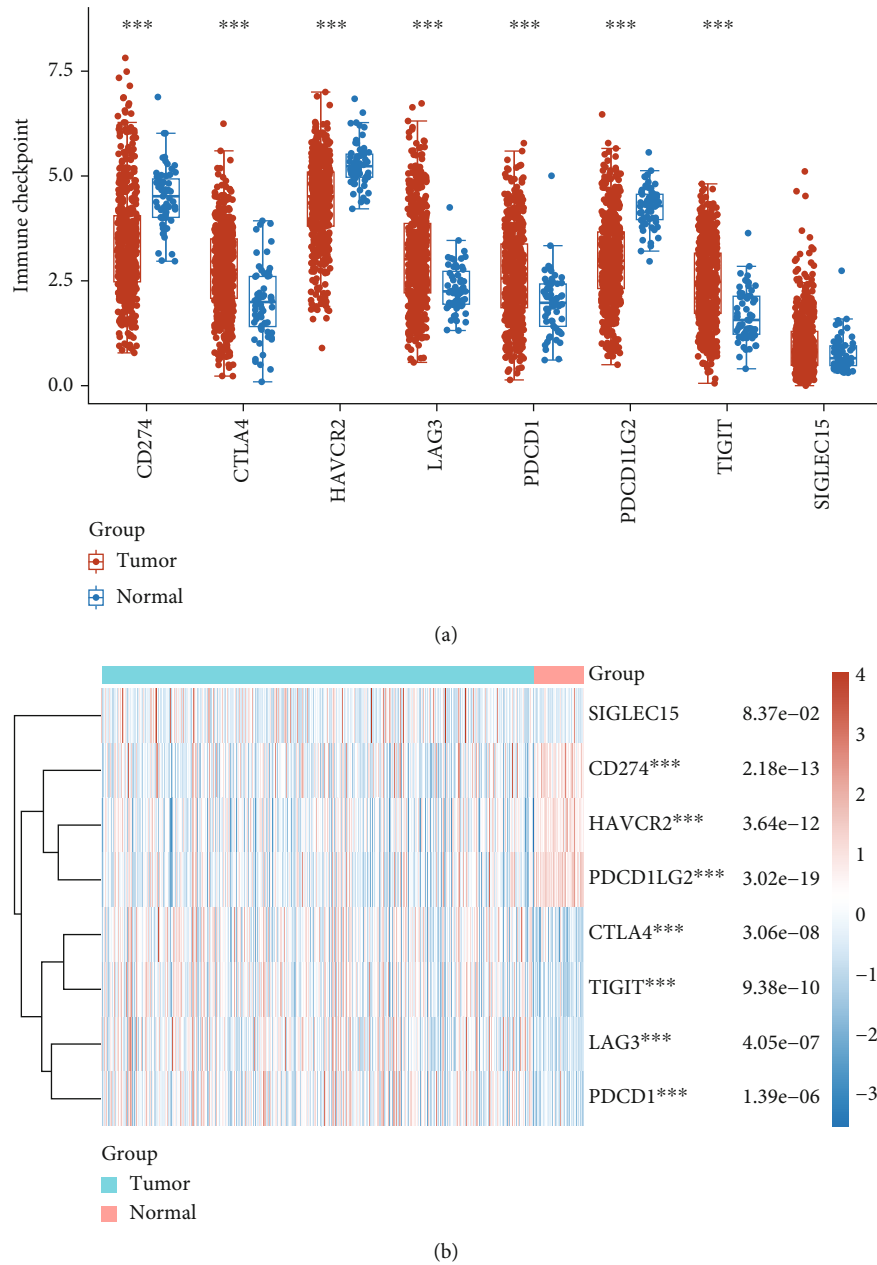


FIGURE 6: (a, b) The dysregulated expression of immune checkpoints in LUAD samples in contrast to nontumor specimens. *** $p < 0.001$.

Our team first utilized such database to evaluate diversities in SCL7A11 expressing levels in specific cancer types via the TIMER database, and our team investigated the relationship between the expression of SCL7A11 and infiltrative level via specific immunocyte subsets.

2.6. Gene Set Enrichment Analysis (GSEA). GSEA was completed to analyze the biology pathway in LUAD layered via the midvalue of SCL7A11 expression. The assay was finished as per the protocol recommendation from the Broad Institute Gene Set Enrichment Analysis website. The GSEA was completed via the GSEA 4.0.3 program. NOM ($p < 0.05$) and FDR $q < 0.05$ ($p < 0.05$) had significance on statistics.

2.7. Statistical Analysis. The entire analysis was completed via R 4.0.5 and the proper packages. A two-sided t -test was employed to examine the association between the expression of SCL7A11 and clinic characteristics. Survival curves were drawn via the Kaplan-Meier approach and evaluated via log-rank tests. ROC curve analyses were completed for risk scoring computed on the foundation of screened genetic hallmark via survivalROC R package. Statistic assays were bilateral, with $p \leq 0.05$ having significance on statistics.

3. Results

3.1. Twenty-Three Aberrantly Expressed Ferroptosis-Related Genes in LUAD Were Identified. To explore the possible

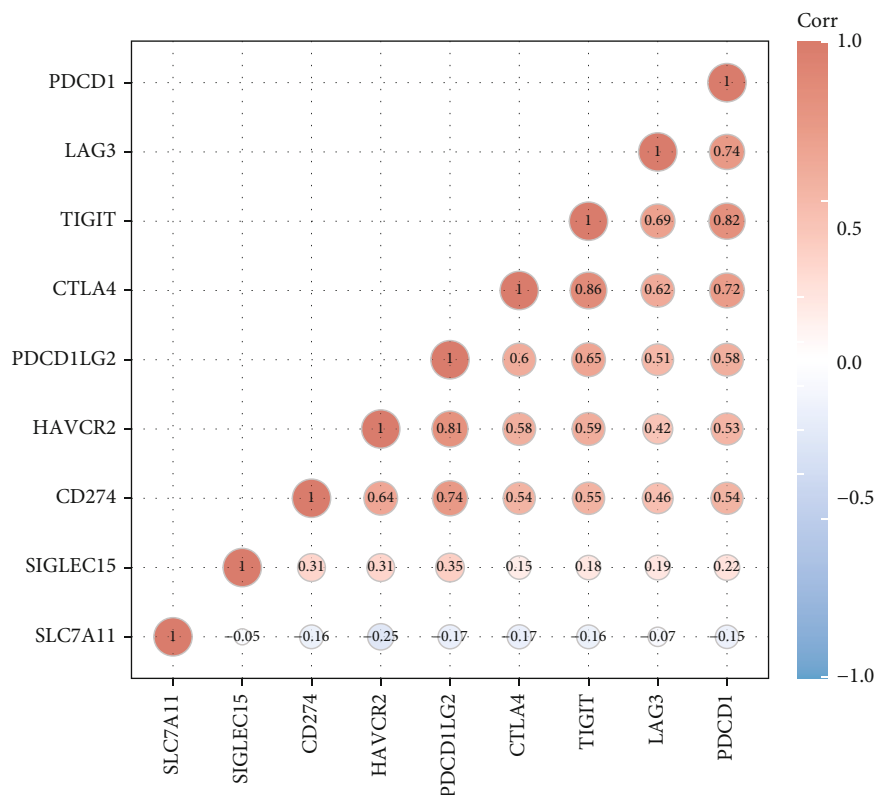


FIGURE 7: The association between SCL7A11 expression and several immune checkpoints in LUAD.

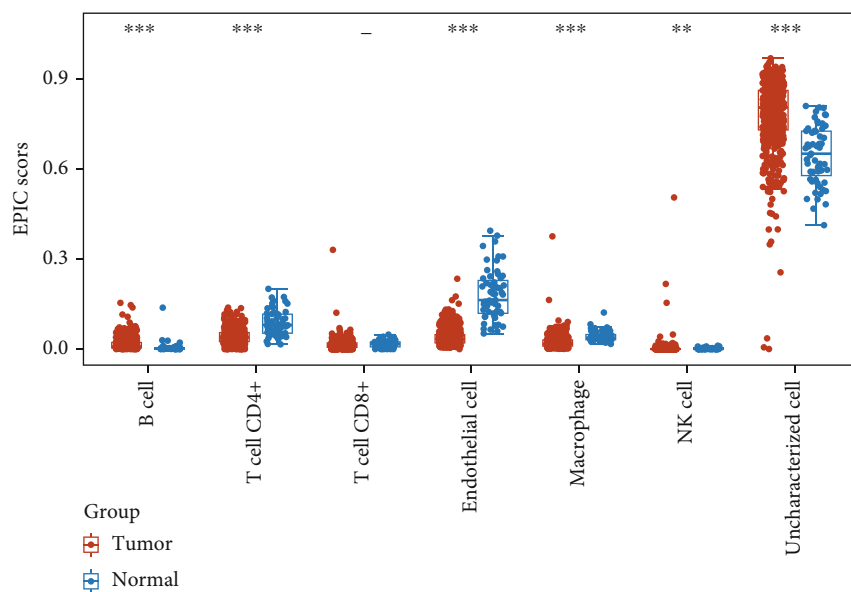
function of aberrantly expressed ferroptosis-related genes in LUAD, our team analyzed the expressing pattern of ferroptosis-related genes via TCGA datasets. We observed that 23 ferroptosis-related genes which exhibited a dysregulated level in LUAD, such as CDKN1A, HSPA5, EMC2, SLC7A11, NFE2L2, HSPB1, GPX4, FANCD2, C1SD1, FDFT1, SLC1A5, TFRC, RPL8, NCOA4, LPCAT3, DPP4, CS, CARS1, ATP5MC3, ALOX15, ACSL4, GLS2, and ACSL4 (Figure 1).

3.2. Identification of Survival-Related and Ferroptosis-Related Genes in LUAD. Then, we performed Kaplan-Meier methods to screen survival-related and ferroptosis-related genes in LUAD. Among the above 23 aberrantly expressed ferroptosis-associated genes, we observed four genes were related to the OS of LUAD sufferers, including SCL7A11 (Figure 2(a)), GLS2 (Figure 2(b)), ATP5MC3 (Figure 2(c)), and FANCD2 (Figure 2(d)). Our attention focused on SCL7A11. The survival status of the entire LUAD sufferers is presented by Figure 3(a). Time-reliant ROC analyses revealed the prognosis accuracy was 0.588 at 1 year, 0.607 at 3 years, and 0.52 at 5 years, separately (Figure 3(b)). Moreover, univariate assays suggested the above four genes, pTNM_stage, and Radiation_therapy were risk factors for the clinical outcome of LUAD patients (Figure 4). Moreover, we performed Sankey diagrams to investigate the relationship between the expression of SCL7A11 and clinicopathological parameters, finding that high SCL7A11 predicted an advanced pTNM_stage (Figures 5(a) and 5(b)). High expres-

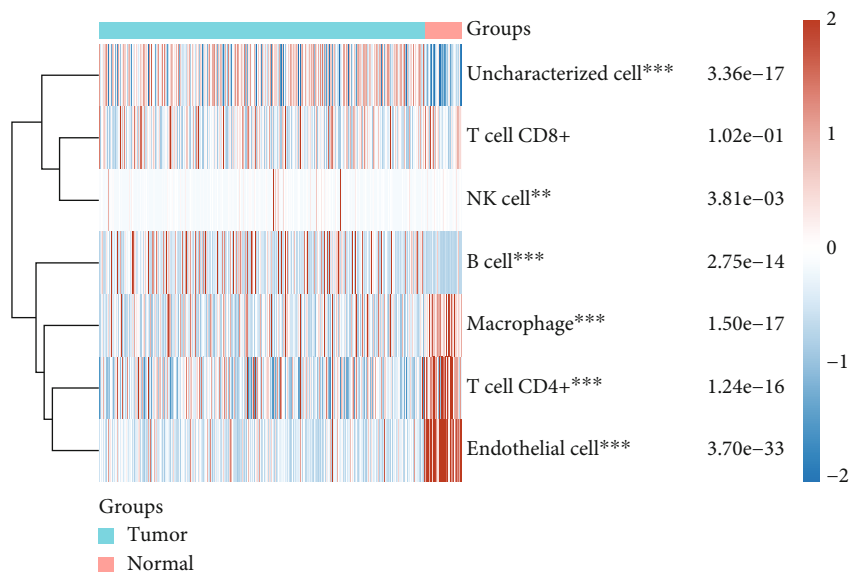
sion of SCL7A11 was identified in LUAD samples with phases III-IV in contrast to the phases I-II (Figure 5(c)).

3.3. The Relationship between the Expression of SCL7A11 and Immune Checkpoint in LUAD. For the sake of exploring the underlying function of SCL7A11 in immunoactivity, we analyzed the expressing pattern of several immune checkpoint in LUAD. As presented by Figures 6(a) and 6(b), our team discovered that the expression of CTLA4, LAG3, PDCD1, TIGIT, and SIGLEC15 was distinctly increased in LUAD specimens in contrast to nontumor specimens, while the expression of HAVCR2, CD274, and PDCD1LG2 was distinctly decreased in LUAD samples in contrast to nontumor specimens. The outcomes of correlation analyses revealed that the expression of SCL7A11 was associated with the expression of most immune checkpoints (Figure 7).

3.4. A Significant Correlation between SCL7A11 and Cancer-Infiltrating Immunocytes in LUAD. Then, we analyzed the expression of cancer-infiltrating immunocytes in LUAD, and observed that B cell, T cell CD4+, endothelial cell, macrophages, NK cell, and uncharacterized cell exhibited a dysregulated level in LUAD samples in contrast to nontumor specimens (Figures 8(a) and 8(b)). For the sake of knowing the association between the expression of SCL7A11 and cancer-infiltrating immunocytes, the kit of TIMER was employed. A remarkable association was observed between the expression of SCL7A11 and diverse kinds of immunocytes, like B cell and macrophage (Figure 9).



(a)



(b)

FIGURE 8: Continued.

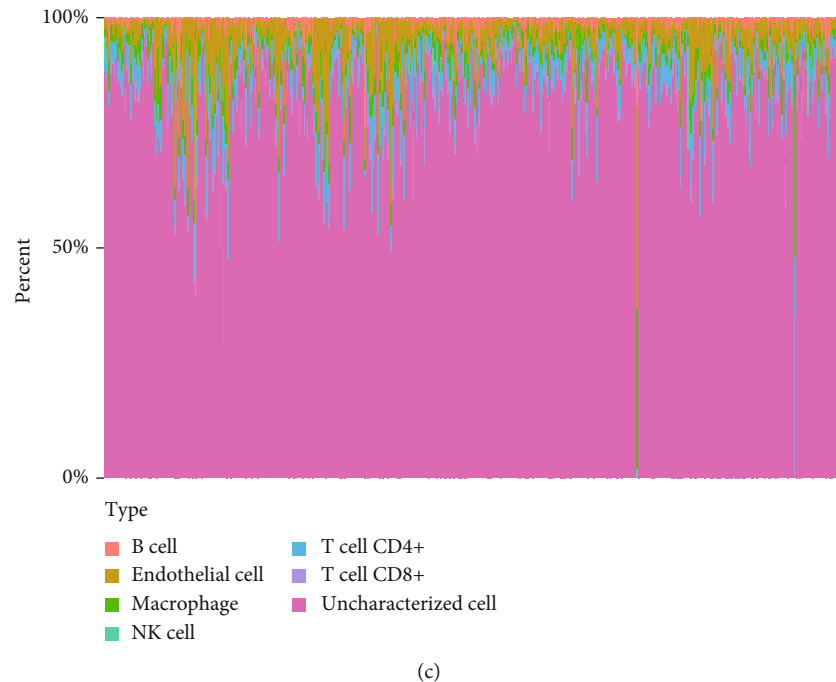


FIGURE 8: (a, b) The expressing feature of immune cells in LUAD specimens and nontumor specimens. *** $p < 0.001$ and ** $p < 0.01$.

3.5. Gene Ontology (GO) and the Kyoto Encyclopedia of Genes and Genomes (KEGG) Enrichment Analysis of DEGs. To explore the function of SCL7A11 in LUAD, our team selected DEGs between LUAD specimens with high SCL7A11 expression and LUAD specimens with low SCL7A11 expression. The GO analyses revealed that the GO annotations of differentially expressed genes were separated into 3 parts: biological process (BP), cell composition (CC), and molecular function (MF). Terms were in an ascending order as per the FDR results. Posterior to the selection process, we discovered that (FDR<0.05) along with DEGs were concentrated in BP, like response to metal ion, cellular ketone metabolic process, cell projection membrane, clathrin-coated endocytic vesicle, carboxylic acid binding, and monocarboxylic acid binding (Figure 10(a)). The KEGG analyses revealed that DEGs were predominantly enriched in Staphylococcus aureus infection, intestine immunity network for IgA generation, glutathione metabolism, viral myocarditis, steroid hormone biosynthesis, and arachidonic acid metabolism (Figure 10(b)).

4. Discussion

Based on the Chinese statistics as to LUAD development, it is evident that LUAD prognosis factors preventing and curing LUAD require further exploration [20]. Investigating novel molecule-level prognosis and prediction biomarkers is a hotspot in medical researches [21, 22]. Recently, more and more researches have reported the underlying effects of ferroptosis-related genes utilized as new diagnosis and prognosis markers for LUAD [23–25]. Herein, our team identified 23 dysregulated ferroptosis-associated genes in LUAD. Among them, SLC7A11, GLS2, ATP5MC3, and

FANCD2 were associated with five-year survivals of LUAD patients, suggesting their potential use as novel biomarkers.

Previously, several studies have reported the function of the above four genes in several types of tumors, including LUAD. For instance, nuclear and cytoplasmic localization of FANCD2 was identified in ovarian carcinoma from both healthy and ovarian carcinoma sufferers [26]. Sufferers with cytoplasm localization of FANCD2 had remarkably better median survival duration (50 months), in contrast to sufferers in the absence of cytoplasm localization of FANCD2. In addition, cytoplasm FANCD2 was discovered to be capable of binding protein participating in the native immune system, cell reaction to thermal stress, amyloid fiber formation, and estrogen-mediated signal transduction [27]. High ATP5MC3 expression predicted a poor prognosis in prostate cancer, colon cancer, and endometrial cancer [28–30]. TGF- β 1-mediated inhibition of SLC7A11 drives vulnerability to GPX4 suppression in hepatic cell cancer [31]. In this study, we focused on SLC7A11 and found that high SLC7A11 expression predicted an advanced clinical pTNM_stage in LUAD patients. In clinical practice, pTNM_stage has been considered an important factor for the prediction of clinical outcome of LUAD patients, and its degree was strongly associated with distant metastasis [32, 33]. Thus, our findings suggested SLC7A11 may play a functional role in tumor metastasis. In addition, a survival study confirmed that high SLC7A11 expression predicted a poor prognosis in LUAD patients, which may be due to its potential function on tumor stage and metastasis.

With the development of cancers, our immunosystems are stimulated to inhibit tumorous progression [34]. It is unfortunate that oncocytes leverage a variety of methods to postpone or even prevent our immunosystems from

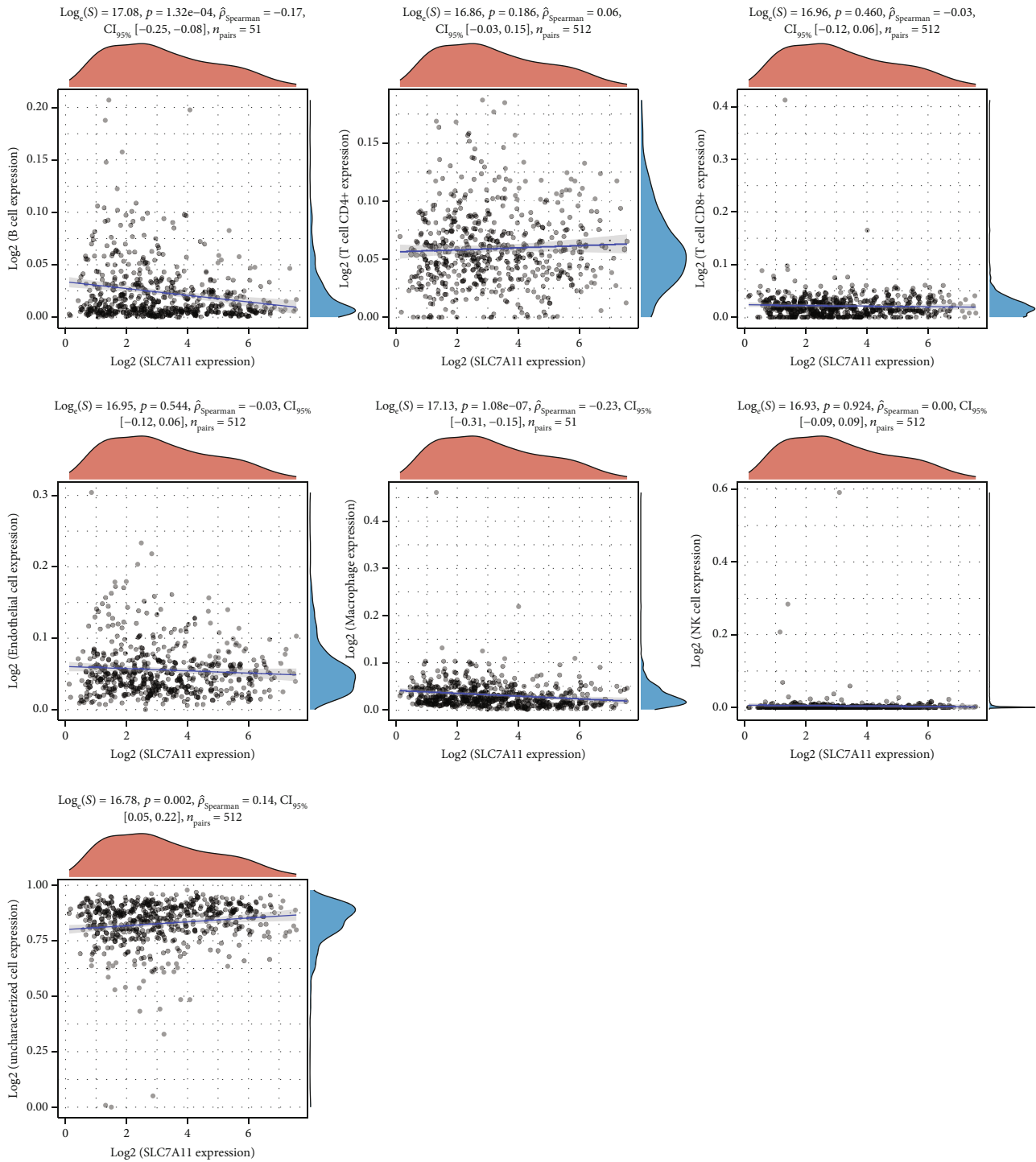
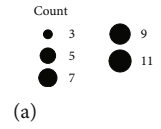
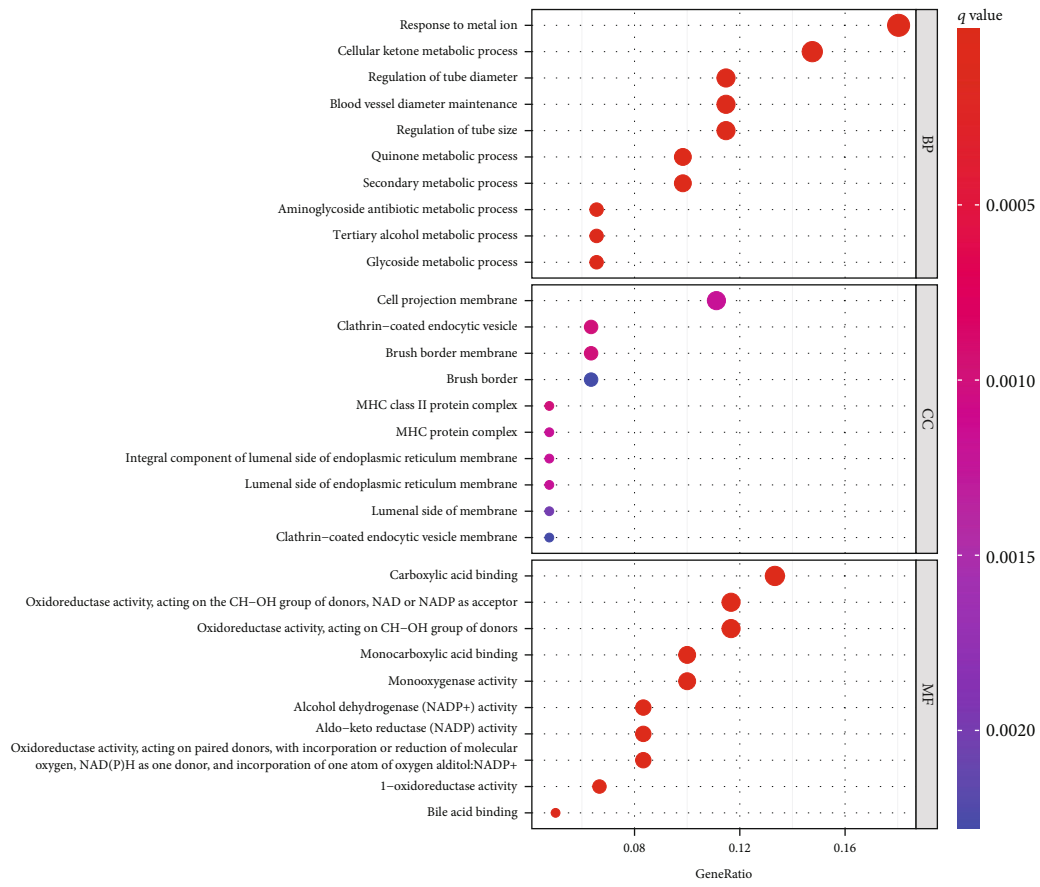


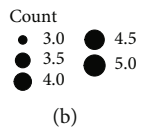
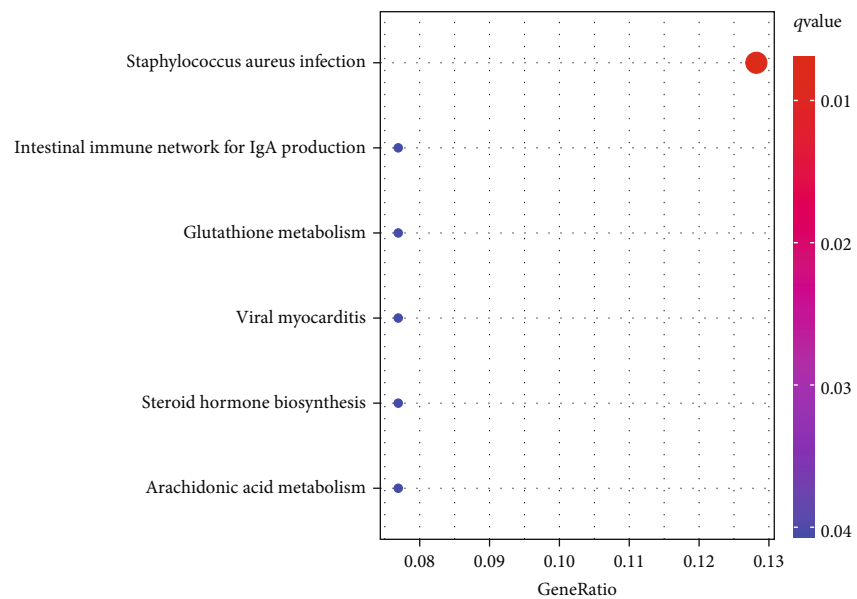
FIGURE 9: The association analysis between the expression of SCL7A11 and immunocytes.

repressing cancers, a phenomenon called immunoescape [35, 36]. The onset of immunoescape normally induces the development of malignancies, metastases, unsatisfactory prognoses, and immune therapy failures [37, 38]. Immune checkpoints, like programmed cell death protein-1 (PD-1), PD ligand-1 (PD-L1), and cytotoxic T-lymphocyte-associated protein 4 (CTLA4), account for immunoescape [39, 40]. Herein, our team discovered that eight immune

checkpoints exhibited a dysregulated level in LUAD. Moreover, we found SCL7A11 expression was associated with several Immune checkpoints, indicating NLRP3 promoted immune escape in LUAD. Moreover, our team discovered that the expression of SCL7A11 was negatively associated with the expression of B cells and macrophage while being positively associated with that of T cell CD4+. Those associations imply an underlying causal link in which SCL7A11



(a)



(b)

FIGURE 10: (a) GO analyses and (b) biological pathway enrichment analyses of the dysregulated genes.

modulates T cell functions in LUAD. Collectively, those discoveries reveal that the SLC7A11 is vital for recruiting and regulating immune infiltration cells in LUAD.

Nonetheless, there are certain deficiencies herein. Firstly, the present paper was a research finished retrospectively, although our team strove to involve as many data sets as possible to more strictly validate such hallmark. Further prospective researches were required to evidence the prognosis value of SLC7A11. Secondly, this report was restricted by the lack of experiment proofs. More researches exploring lineage cells and encompassing animal assays are warranted to substantiate the discoveries herein and unravel the molecule-level causal links.

5. Conclusion

In summary, our study suggested that four ferroptosis-related genes could be used as a prognostic indicator for LUAD with which to predict patient risk. In addition, SLC7A11 was related to immune therapy-associated biological markers, revealing its application significance in forecasting the potency of immune therapy.

Data Availability

The datasets utilized and/or evaluated during this study are accessible upon valid request from the corresponding authors.

Conflicts of Interest

The authors hereby state that they do not have any conflicts of interest.

Authors' Contributions

Li Qian and Fu Wang contributed equally to this work.

Acknowledgments

This work was supported by the National Natural Science Foundation of China (grant number 82102167) and China Postdoctoral Science Foundation (grant number 2019M651928).

References

- [1] C. Gridelli, A. Rossi, D. P. Carbone et al., "Non-small-cell lung cancer," *Nature Reviews Disease Primers*, vol. 1, no. 1, article 15009, 2015.
- [2] L. T. Tanoue, N. T. Tanner, M. K. Gould, and G. A. Silvestri, "Lung cancer screening," *American Journal of Respiratory and Critical Care Medicine*, vol. 191, no. 1, pp. 19–33, 2015.
- [3] M. B. Schabath and M. L. Cote, "Cancer progress and priorities: lung cancer," *Cancer Epidemiology, Biomarkers & Prevention*, vol. 28, no. 10, pp. 1563–1579, 2019.
- [4] K. Wadowska, I. Bil-Lula, Ł. Trembecki, and M. Śliwińska-Mossoń, "Genetic markers in lung cancer diagnosis: a review," *International Journal of Molecular Sciences*, vol. 21, no. 13, p. 4569, 2020.
- [5] M. Alexander, S. Y. Kim, and H. Cheng, "Update 2020: management of non-small cell lung cancer," *Lung*, vol. 198, no. 6, pp. 897–907, 2020.
- [6] C. Stapelfeld, C. Dammann, and E. Maser, "Sex-specificity in lung cancer risk," *International Journal of Cancer*, vol. 146, no. 9, pp. 2376–2382, 2020.
- [7] R. Tang, J. Xu, B. Zhang et al., "Ferroptosis, necroptosis, and pyroptosis in anticancer immunity," *Journal of Hematology & Oncology*, vol. 13, no. 1, p. 110, 2020.
- [8] C. Liang, X. Zhang, M. Yang, and X. Dong, "Recent progress in ferroptosis inducers for cancer therapy," *Advanced Materials*, vol. 31, no. 51, article e1904197, 2019.
- [9] T. Hirschhorn and B. R. Stockwell, "The development of the concept of ferroptosis," *Free Radical Biology & Medicine*, vol. 133, pp. 130–143, 2019.
- [10] J. Zheng and M. Conrad, "The metabolic underpinnings of ferroptosis," *Cell Metabolism*, vol. 32, no. 6, pp. 920–937, 2020.
- [11] P. Lei, T. Bai, and Y. Sun, "Mechanisms of ferroptosis and relations with regulated cell death: a review," *Frontiers in Physiology*, vol. 10, p. 139, 2019.
- [12] B. R. Stockwell, J. P. Friedmann Angeli, H. Bayir et al., "Ferroptosis: a regulated cell death nexus linking metabolism, redox biology, and disease," *Cell*, vol. 171, no. 2, pp. 273–285, 2017.
- [13] H. Imai, M. Matsuoka, T. Kumagai, T. Sakamoto, and T. Koumura, "Lipid peroxidation-dependent cell death regulated by GPx4 and ferroptosis," *Current Topics in Microbiology and Immunology*, vol. 403, pp. 143–170, 2017.
- [14] T. Xu, W. Ding, X. Ji et al., "Molecular mechanisms of ferroptosis and its role in cancer therapy," *Journal of Cellular and Molecular Medicine*, vol. 23, no. 8, pp. 4900–4912, 2019.
- [15] X. Chen, C. Yu, R. Kang, G. Kroemer, and D. Tang, "Cellular degradation systems in ferroptosis," *Cell Death and Differentiation*, vol. 28, no. 4, pp. 1135–1148, 2021.
- [16] Y. Tang, C. Li, Y. J. Zhang, and Z. H. Wu, "Ferroptosis-related long non-coding RNA signature predicts the prognosis of head and neck squamous cell carcinoma," *International Journal of Biological Sciences*, vol. 17, no. 3, pp. 702–711, 2021.
- [17] B. Tang, J. Zhu, J. Li et al., "The ferroptosis and iron-metabolism signature robustly predicts clinical diagnosis, prognosis and immune microenvironment for hepatocellular carcinoma," *Cell Communication and Signaling: CCS*, vol. 18, no. 1, p. 174, 2020.
- [18] P. Liu, D. Wu, J. Duan et al., "NRF2 regulates the sensitivity of human NSCLC cells to cystine deprivation-induced ferroptosis via FOCAD-FAK signaling pathway," *Redox Biology*, vol. 37, article 101702, 2020.
- [19] K. Ito and D. Murphy, "Application of ggplot2 to pharmacometric graphics," *CPT: Pharmacometrics & Systems Pharmacology*, vol. 2, no. 10, article e79, 2013.
- [20] X. J. Zhou, R. Li, X. Liu, and Y. Q. Qu, "Advances in deubiquitinating enzymes in lung adenocarcinoma," *Journal of Cancer*, vol. 12, no. 18, pp. 5573–5582, 2021.
- [21] V. C. Kok and C. C. Yu, "Cancer-derived exosomes: their role in cancer biology and biomarker development," *International Journal of Nanomedicine*, vol. Volume 15, pp. 8019–8036, 2020.
- [22] V. A. Hristova and D. W. Chan, "Cancer biomarker discovery and translation: proteomics and beyond," *Expert Review of Proteomics*, vol. 16, no. 2, pp. 93–103, 2019.

- [23] X. Gao, M. Tang, S. Tian, J. Li, and W. Liu, "A ferroptosis-related gene signature predicts overall survival in patients with lung adenocarcinoma," *Future Oncology*, vol. 17, no. 12, pp. 1533–1544, 2021.
- [24] L. Lu, L. P. Liu, Q. Q. Zhao, R. Gui, and Q. Y. Zhao, "Identification of a ferroptosis-related lncRNA signature as a novel prognosis model for lung adenocarcinoma," *Frontiers in Oncology*, vol. 11, article 675545, 2021.
- [25] F. Huang, Y. Zheng, X. Li, H. Luo, and L. Luo, "Ferroptosis-related gene AKR1C1 predicts the prognosis of non-small cell lung cancer," *Cancer Cell International*, vol. 21, no. 1, p. 567, 2021.
- [26] S. Joshi, S. Campbell, J. Y. Lim et al., "Subcellular localization of FANCD2 is associated with survival in ovarian carcinoma," *Oncotarget*, vol. 11, no. 8, pp. 775–783, 2020.
- [27] J. Moes-Sosnowska, I. K. Rzepecka, J. Chodzyska et al., "Clinical importance of FANCD2, BRIP1, BRCA1, BRCA2 and FANCF expression in ovarian carcinomas," *Cancer Biology & Therapy*, vol. 20, no. 6, pp. 843–854, 2019.
- [28] H. Liu, L. Gao, T. Xie, J. Li, T. S. Zhai, and Y. Xu, "Identification and validation of a prognostic signature for prostate cancer based on ferroptosis-related genes," *Frontiers in Oncology*, vol. 11, article 623313, 2021.
- [29] J. Nie, D. Shan, S. Li et al., "A novel ferroptosis related gene signature for prognosis prediction in patients with colon cancer," *Frontiers in Oncology*, vol. 11, article 654076, 2021.
- [30] Y. Weijiao, L. Fuchun, C. Mengjie et al., "Immune infiltration and a ferroptosis-associated gene signature for predicting the prognosis of patients with endometrial cancer," *Aging (Albany NY)*, vol. 13, no. 12, pp. 16713–16732, 2021.
- [31] D. H. Kim, W. D. Kim, S. K. Kim, D. H. Moon, and S. J. Lee, "TGF- β 1-mediated repression of SLC7A11 drives vulnerability to GPX4 inhibition in hepatocellular carcinoma cells," *Cell Death & Disease*, vol. 11, no. 5, p. 406, 2020.
- [32] K. Chansky, F. C. Detterbeck, A. G. Nicholson et al., "The IASLC lung cancer staging project: external validation of the revision of the TNM stage groupings in the eighth edition of the TNM classification of lung cancer," *Journal of Thoracic Oncology*, vol. 12, no. 7, pp. 1109–1121, 2017.
- [33] S. H. Feng and S. T. Yang, "The new 8th TNM staging system of lung cancer and its potential imaging interpretation pitfalls and limitations with CT image demonstrations," *Diagnostic and Interventional Radiology*, vol. 25, no. 4, pp. 270–279, 2019.
- [34] B. Pulendran and M. M. Davis, "The science and medicine of human immunology," *Science*, vol. 369, no. 6511, 2020.
- [35] L. Galluzzi, J. Humeau, A. Buqué, L. Zitvogel, and G. Kroemer, "Immunostimulation with chemotherapy in the era of immune checkpoint inhibitors," *Nature Reviews. Clinical Oncology*, vol. 17, no. 12, pp. 725–741, 2020.
- [36] W. S. Wold and K. Toth, "Adenovirus vectors for gene therapy, vaccination and cancer gene therapy," *Current Gene Therapy*, vol. 13, no. 6, pp. 421–433, 2014.
- [37] Z. Wang, Z. Wu, Y. Liu, and W. Han, "New development in CAR-T cell therapy," *Journal of Hematology & Oncology*, vol. 10, no. 1, p. 53, 2017.
- [38] G. P. Dunn, A. T. Bruce, H. Ikeda, L. J. Old, and R. D. Schreiber, "Cancer immunoediting: from immunosurveillance to tumor escape," *Nature Immunology*, vol. 3, no. 11, pp. 991–998, 2002.
- [39] L. Liu, Y. Wang, R. Yan et al., "BMP-7 inhibits renal fibrosis in diabetic nephropathy via miR-21 downregulation," *Life Sciences*, vol. 238, article 116957, 2019.
- [40] P. Darvin, S. M. Toor, V. Sasidharan Nair, and E. Elkord, "Immune checkpoint inhibitors: recent progress and potential biomarkers," *Experimental & Molecular Medicine*, vol. 50, no. 12, pp. 1–11, 2018.