Research Article

Antibacterial Activity of Hydrophobic Composite Materials Containing a Visible-Light-Sensitive Photocatalyst

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The conventional superhydrophobic surface offered by polytetrafluoroethylene (PTFE) provides no sterilization performance and is not sufficiently repellent against organic matters. Thus, there is potential risk for bacteria to adhere to its surface more readily in ambient air, as well as organic matters is considered to reduce the durability of the superhydrophobic performance. To remedy this shortcoming, anatase titanium dioxide (TiO2), a UV light-sensitive photocatalyst added water repellent composite material was developed to demonstrate the inactivation of various bacteria, such as Escherichia coli (E. coli), methicillin-resistant Staphylococcus aureus (MRSA), Pseudomonas aeruginosa (P. aeruginosa), Legionella pneumophila (L. pneumophila) [6–8], and Clostridium difficile spores [8]. The inclusion of TiO2 to the PTFE coating was expected to generate antimicrobial and self-cleaning properties, which would expand its scope of application. Anatase TiO2-added PTFE composite material is not only water-repellent but also exhibits self-cleaning properties. However, because anatase

1. Introduction

It is well known that the conventional superhydrophobic surface offered by polytetrafluoroethylene (PTFE) provides no sterilization performance and is not sufficiently repellent against organic matters. Thus, there is potential risk for bacteria to adhere to its surface more readily in ambient air, as well as organic matters is considered to reduce the durability of the superhydrophobic performance. To remedy this shortcoming, anatase titanium dioxide (TiO2), a UV light-sensitive photocatalyst added water repellent composite material has been developed in our previous study [1–5]. TiO2 was used to demonstrate the inactivation of various bacteria, such as Escherichia coli (E. coli), methicillin-resistant Staphylococcus aureus (MRSA), Pseudomonas aeruginosa (P. aeruginosa), Legionella pneumophila (L. pneumophila) [6–8], and Clostridium difficile spores [8]. The inclusion of TiO2 to the PTFE coating was expected to generate antimicrobial and self-cleaning properties, which would expand its scope of application. Anatase TiO2-added PTFE composite material is not only water-repellent but also exhibits self-cleaning properties. However, because anatase
TiO$_2$ is catalytically active only under ultraviolet irradiation, its application is limited to outdoor usage [9–12].

In recent years, nosocomial infections have become a very important issue, with the appearance of resistant bacteria and have thus attracted much great attention [13]. Improving the hospital environment is recognized as a highly effective countermeasure to combat this problem. In response to the problem, we have developed a hydrophobic, self-cleaning composite material by adding N-doped TiO$_2$, a visible-light-sensitive photocatalyst [14], to a hydrophobic composite material containing dispersed PTFE particles in order to obtain antibacterial properties. This novel coating is expected to be useful in both as exterior or interior wall coating. The surface characteristics of the newly developed composite are examined in this paper. Furthermore, the antibacterial activities of the developed composite against E. coli and MRSA were evaluated under visible-light irradiation with intensities ranging from 100 to 2,000 lx, which is equivalent to indoor and outdoor conditions.

2. Materials and Methods

2.1. Development of Hydrophobic Composite Material. The samples were composed of commercial PTFE particles, a fluorinated binder, and fluoro oil with varying amounts of N-doped TiO$_2$ (0, 3, 8, and 12 wt%). N-doped TiO$_2$ was provided by courtesy of Sumitomo Chemical Co., Ltd. (Japan). The PTFE particles, fluorinated binder, fluoro oil and N-doped TiO$_2$ mixture was sprayed over a substrate, together with butyl acetate. A dispersed particulate composite material about 5 $\mu$m in thickness was formed with a single spraying. Particulate composite materials about 15 $\mu$m in thickness were formed by spraying the entire substrate three times. The composite material was ready for the various measurements 24 hours after the butyl acetate dried.

2.2. Characteristics

2.2.1. Surface Characteristics. Contact angles were measured with an automatic contact angle meter (CA-Z; Kyowa Interface Science Ltd., Saitama Japan). A drop of water was deposited on the sample using a syringe. The contact angle was measured after the tip of the needle was separated from the drop. The measured contact angles, therefore, were equilibrium contact angles and not advancing or receding contact angles. The contact angles were determined at five different positions for each sample.

The surface free energy of the sample was obtained by the following procedure. The contact angles $\theta_{SL}$ of $\alpha$-bromonaphthalene (90°), methylene iodide (87°), and water (152°) were substituted into the Young-Dupre equation:

$$W_{SL} = y_{LV}(1 + \cos \theta_{SL}),$$

(1)

where $W_{SL}$ is the work of adhesion and $y_{LV}$ is the surface free energy of the liquid.

The dispersion, polar, and hydrogen-bonding components of $W_{SL}$ provided the dispersion $y_{SV}^d$, polar $y_{SV}^p$, and hydrogen-bonding $y_{SV}^h$ components of the surface free energy using the Kitazaki-Hata method [15]. The surface free energy $y_{SV}$ of the sample was obtained via the following formula:

$$y_{SV} = y_{SV}^d + y_{SV}^p + y_{SV}^h.$$  

(2)

The samples were observed using a JSM-6300F scanning electron microscope. The transmittance IR spectrum of the PTFE powder was obtained using an MFT-2000 FT-IR spectrometer. Crushed potassium bromide (KBr) crystal and PTFE powders were mixed and pressed into a disk, which was then placed in the FT-IR spectrometer for measurement.

2.2.2. Antibacterial Testing. The bacterial activity of the PTFE composite against E. coli (NBRC 3972; Naitinol Institute of Technology and Evaluation, Biological Resource Center, Japan) and MRSA (clinically isolated by the Graduate School of Medicine, Yokohama City University) was evaluated, after some adjustments, in accordance with Japanese Industrial Standard methods (JIS R 1702) [16]. The bacterial strains were cultured twice on nutrient agar plates (E-MC35, Eikenkizai, Japan) at 37°C for 16 ~ 24 hours. The strains were then suspended in a 1/500 nutrient broth (NB) solution (E-MC35, Eikenkizai, Japan) with a pH of 7.0, and diluted to approximately $10^6$ colony-forming units (CFUs) per milliliter to be utilized for bactericidal testing.

Before testing, all the water-repellent composite samples ($50 \times 50 \text{mm}$) were presterilized by UV light with an irradiance of 1 mW $\cdot$ cm$^{-2}$ overnight. A sterilized moisture-control paper filter was placed in a sterilized Petri dish, 5 mL of sterilized distilled water was added, and then a glass tube was placed on the paper filter to avoid contact between the test material and the paper filter. The sterilized test material was then placed on the glass tube, and 100 $\mu$L of the bacterial suspension (approximately $10^5$ CFUs) was deposited on the surface of the test material. Because the test surface is hydrophobic, a covering quartz glass plate ($40 \times 40 \text{mm}$) was applied to spread out the suspension drip. Then, another quartz glass plate ($100 \times 100 \text{mm}$) was placed on top of the Petri dish. The paper filter, distilled water, and quartz glasses plates were all employed to prevent the evaporation the test bacterial suspension (Figure 1). The Petri dishes containing the specimens were illuminated by 20 W tubular-type white fluorescent lamps (Hitachi FL20SSW/18-B; Tokyo, Japan) for 24 hours. The influence of the light intensity on the bactericidal activity was examined for 100, 300, and 2000 lx. Only the intensity of 2000 lx included light in the UV region with wavelengths less than 400 nm at an intensity of 0.01 mW $\cdot$ cm$^{-2}$. The light intensity incident on the center of the specimen surface was adjusted by a UD-40 radiometer head (Topcon Corporation; Tokyo, Japan).

After illumination, the specimens were washed with 10 mL of soybean-casein digest broth including lecithin and polysorbate 80 (SCDLP) (Nihon Pharmaceutical co., LTD, Japan). The washing solution was diluted with phosphate-buffered saline (PBS) in a 10-fold dilution series. At each dilution stage, aliquots (1 mL) of the SCDLP solution were mixed with the nutrient agar medium (14 mL) at 45°C in a 10 cm Petri dish
and allowed to cool to room temperature. After the medium solidified, the dishes were incubated at 35°C for 24 to 48 hours prior to determining the number of CFUs.

Bacterial tests were conducted by three parallel analyses for each sample, and the viable cell count was obtained from the average value of the product of the CFUs multiplied by the dilution rate and by ten as shown as follows:

\[
\text{Viable bacterial cells} = N \times \text{dilution rate} \times 10,
\]

where \( N \) is the number of CFUs on the N-doped TiO\(_2\)-PTFE composite films after visible-light illumination for a period of time. The value 10 indicates the sample washing solution (SCDLP liquid) volume.

When no CFUs were observed in undiluted SCDLP liquid, this indicated that the detection limit was no more than 10 CFUs. In such a case, the viable cell count was expressed as “≤10” to indicate the detection limit. The determination of the disinfectant property is given in terms of log decrease. N-doped TiO\(_2\)-free PTFE composite films were used as contrast samples.

### 2.2.3. Assessment of Self-Cleaning Performance

To assess the oxidative self-cleaning performance of the N-doped TiO\(_2\)-PTFE composite coatings, measurement of the contact angles of the composite coatings contaminated with oleic acid (C\(_{18}\)H\(_{34}\)O\(_2\)) during exposure to UV irradiation was performed according to the JIS R 1703-1 standard method [17]. The light source selected here is based on assumption that N-doped TiO\(_2\)-PTFE composite coating used outdoor is more susceptible to contamination by organic pollutants and sunlight including strong ultraviolet light. Prior to measuring the contact angles, the samples were cleaned for 24 hours by 2 mW \( \cdot \) cm\(^{-1} \) of UV illumination incident on the testing surface to remove organic pollutants. Then, the samples were contaminated by dip coating (60 cm \( \cdot \) min\(^{-1} \)) with 0.5 vol% oleic acid diluted in \( n \)-heptane. After contamination, the samples were dried at 70°C for 15 minutes and were then irradiated for 230 hours using an unfiltered UV light source (Toshiba FL10BLB; Tokyo, Japan). The N-doped TiO\(_2\)-free PTFE composite was again used for contrast testing.

### 3. Results and Discussion

#### 3.1. Surface Characteristics

**3.1.1. Contact Angles and Surface Free Energy**. The results of water contact angles and surface free energy of the N-doped TiO\(_2\)-PTFE composite material are presented in Table 1.

All composite samples exhibited contact angles exceeding 150°, which is consistent with their hydrophobicity despite the presence of the hydrophilic N-doped TiO\(_2\) photocatalyst. The contact angles of similar materials are usually expected to lie between those of PTFE and those of the binder with additives, including hydrophilic N-doped TiO\(_2\). Instead, these angles (150°) were significantly greater than those of pure PTFE (110°). This is consistent with results previously reported by Yamauchi et al. [18]. In that study, a particulate composite model was analysed by the Wenzel equation, which takes into consideration surface roughness [19], and the Cassie equation, which considers heterogeneous surfaces [20]. They concluded that the binder wets more easily with water than the PTFE in PTFE particulate composite materials. In such a system, the contact angle increases with increasing PTFE concentration, however, and beyond the contact angle of the PTFE. When the concentration of PTFE particles dispersed in an appropriate binder reached 80 wt%, more than 78% of the surface area between the water droplets and composite surface was covered by air due to the surface roughness of the PTFE. Thus, we consider that the same tendency may be observed in the N-doped TiO\(_2\)-PTFE composite materials if a part of the binder is replaced with N-doped TiO\(_2\). Nakajima et al. [1] also reported a similar result; TiO\(_2\) nanoparticles added to superhydrophobic thin films, at a concentration below 20 wt%, did not significantly increase the hydrophobicity of the films significantly even after UV illumination for 800 hours at an intensity of 1.7 mW \( \cdot \) cm\(^{-2} \).

The surface free energy of the 8 wt% N-doped TiO\(_2\) composite material was 5.8 mN \( \cdot \) m\(^{-1} \), similar to that of CF\(_3\) (6 mN \( \cdot \) m\(^{-1} \)) [21]. This hydrophobicity reflects the low surface energy of the N-doped TiO\(_2\)-PTFE composite coating, which is expected to inhibit the adhesion of contaminants.

**Table 1**: Contact angles of water and surface free energy on the PTFE hydrophobic composite films with the addition of N-doped TiO\(_2\).

<table>
<thead>
<tr>
<th>Without N-doped TiO(_2)</th>
<th>152</th>
<th>5.8</th>
</tr>
</thead>
<tbody>
<tr>
<td>With N-doped TiO(_2)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3 wt%</td>
<td>152</td>
<td>—</td>
</tr>
<tr>
<td>8 wt%</td>
<td>151</td>
<td>5.0</td>
</tr>
<tr>
<td>12 wt%</td>
<td>152</td>
<td>—</td>
</tr>
</tbody>
</table>

### 3.2. Bacterial Inactivation Performance

**3.2.1. Determination of the Disinfectant Property**. The results of bacterial tests were conducted by three parallel analyses for each sample, and the viable cell count was obtained from the average value of the product of the CFUs multiplied by the dilution rate and by ten as shown as follows:

\[
\text{Viable bacterial cells} = N \times \text{dilution rate} \times 10,
\]

where \( N \) is the number of CFUs on the N-doped TiO\(_2\)-PTFE composite films after visible-light illumination for a period of time. The value 10 indicates the sample washing solution (SCDLP liquid) volume.

When no CFUs were observed in undiluted SCDLP liquid, this indicated that the detection limit was no more than 10 CFUs. In such a case, the viable cell count was expressed as “≤10” to indicate the detection limit. The determination of the disinfectant property is given in terms of log decrease. N-doped TiO\(_2\)-free PTFE composite films were used as contrast samples.

**Table 1**: Contact angles of water and surface free energy on the PTFE hydrophobic composite films with the addition of N-doped TiO\(_2\).
Figure 2: Scanning electron micrograph of the surface of a sample composed of 80 wt% PTFE, 8 wt% N-doped TiO₂, fluoro-oil and the remainder of fluorine binder. The water repellent property of the sample is attributed to the surface roughness created by the PTFE particles.

3.1.2. Scanning Electron Microscope Observation. A scanning electron micrograph of the sample containing 80 wt% PTFE and 8 wt% N-doped TiO₂ is shown in Figure 2. The micrograph reveals many small spherical aggregates of PTFE particles, indicating the heterogeneous morphology of the composite coating. Therefore, any analysis of surface wetting properties must consider the surface heterogeneity resulting from the PTFE particles and the binder. Such an analysis has previously been conducted by Yamauchi et al. [18]; they attributed the extraordinary hydrophobicity to the presence of air between the water droplets and the substrate surface.

3.1.3. Fourier Transform Infrared Spectrometer Observation. The Fourier Transform Infrared Spectrometer Observation (FT-IR) spectrum of the PTFE powder used for the samples is shown in Figure 3. The FT-IR spectra show clear peaks for the CF₃ (1213 cm⁻¹) and CF₂ (1155 and 639 cm⁻¹) groups, which are responsible for the hydrophobicity of the composite material. No peak was observed for C=O (1800 cm⁻¹) which causes the hydrophilic behaviour reported by Yamauchi et al. [4]. This finding accounts for the preventing of deterioration in the hydrophobicity of the composite material.

3.2. Bactericidal Properties. It is well known that superhydrophobicity is not sufficient to repel organic matter. Thus, there is potential for bacteria to adhere to the composite more readily in ambient air. The addition of N-doped TiO₂ to the PTFE coating was expected to generate antimicrobial properties, which would expand its scope of application. We analyzed the bactericidal properties of the N-doped TiO₂-PTFE composite by the test method described in Section 2.2.2 and shown in Figure 1.

Although the E. coli and MRSA cells had to be artificially attached to the surface of N-doped TiO₂-PTFE composite, the results revealed a very high level of bactericidal activity. Figure 4 shows the decrease in viable E. coli cells on the PTFE hydrophobic composite films with N-doped TiO₂. It can be seen in Figure 4 that under 1000 and 2000 lx visible-light illumination for 24 h, the number of viable bacterial cells on the composite coating containing 12 wt% of N-doped TiO₂ decreased from ~10⁵ CFUs to a value below the detection limits of the colony-formation method set at 10 CFUs. Also, even under very low-intensity illumination (100 lx), the number of viable E. coli cells decreased two logs (Figure 4). As shown in Figure 5, by using 2000 lx visible-light irradiation, the number of viable MRSA cells decreased from ~10⁵ cells to less than 10 CFUs, for all samples containing differing amounts of N-doped TiO₂. It can be seen from Figures 4 and 5 that the bactericidal
activity on \( E. \text{coli} \) is lower than that on MRSA, at the surface of 3 wt\% N-TiO\(_2\)/PTFE under 2000 lx of visible light for 24 h. This is consistent with a normal understanding: that is, the photocatalytic killing effect on Gram-positive strain is faster than that on Gram-negative strain [6, 8].

The data shows a possible approach to lower the incidence of hospital-related bacterial infection, since MRSA is the major pathogenic organism and is frequently resistant to many other agents [22, 23]. As has been previously reported, the bactericidal activity of the N-doped TiO\(_2\)-PTFE composite was dependent on the intensity of the visible light and the proportion of N-doped TiO\(_2\) (Figure 4). No bactericidal effect was observed in the control sample (PTFE only). These revealed that N-doped TiO\(_2\) nanoparticles are unique materials that contribute to the bactericidal action of the composite material. Furthermore, compared with the N-doped TiO\(_2\)-only coating, the bactericidal activity of N-doped TiO\(_2\)-PTFE composite was enhanced (Figure 6).

Numerous studies have explained the mechanism of photocatalytic antimicrobial activity as the loss of cell membrane integrity caused by electrons/holes or by reactive oxygen species (ROS) [24–26]. In the present investigation, we deem that the greater surface roughness (Figure 3) afforded by PTFE results in more air pockets that provide additional oxygen for the photocatalytic reaction, which in turn leads to further ROS generation.

We also examined the rapid adhesion of bacterial cells to the 8 wt\% N-doped TiO\(_2\)-PTFE composite coating by rinsing the samples with nutrient broth solution containing \( 10^8 \) CFU/mL of \( E. \text{coli} \) for 10 sec. \( 10^3 \) \( E. \text{coli} \) cells were adsorbed on the surface of PTFE hydrophobic composite films. The viable \( E. \text{coli} \) were all inactivated with visible-light illumination at 2000 lx for 24 hours. Only 100 CFUs of \( E. \text{coli} \) were detected on the rinsed sample. These results reveal that bacterial cells are more readily adsorbed onto superhydrophobicity surface contaminated with organic material. However, although a small amount of \( E. \text{coli} \) was found to be adsorbed onto the N-doped TiO\(_2\)-PTFE composite surface, the cells were inactivated by using 1000 lx of visible-light illumination, as expected (Table 2).

### Table 2: Adhesion of \( E. \text{coli} \) onto the PTFE hydrophobic composite films with and without the addition of N-doped TiO\(_2\) (NT).

<table>
<thead>
<tr>
<th>Visible-light illuminating time (h)</th>
<th>Viable ( E. \text{coli} ) count/cells</th>
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<tbody>
<tr>
<td></td>
<td>Control</td>
</tr>
<tr>
<td>0</td>
<td>( 1 \times 10^3 )</td>
</tr>
<tr>
<td>8</td>
<td>( 1 \times 10^3 )</td>
</tr>
<tr>
<td>24</td>
<td>( 1 \times 10^3 )</td>
</tr>
</tbody>
</table>

By rinsing the samples with nutrient broth solution containing \( 10^8 \) CFU/mL of \( E. \text{coli} \) for 10 sec, \( 10^3 \) \( E. \text{coli} \) cells were adsorbed on the surface of PTFE hydrophobic composite films. The viable \( E. \text{coli} \) were all inactivated with visible-light illumination at 2000 lx for 24 hours.

#### 3.3. Self-Cleaning Properties.

The results of oxidative degradation of the PTFE composite coating on oleic acid under visible light, with and without N-doped TiO\(_2\), are shown in Figure 7. From the data, large decreases in the contact angles could be observed in the composites coating both with and without N-doped TiO\(_2\) when they were contaminated with oleic acid. It is noteworthy that the oleic acid was only
removed successfully on the N-doped TiO$_2$-PTFE composite coating surface by 1 mW $\cdot$ cm$^{-2}$ of UV irradiation for 230 hours, after which the contact angle recovered to its initial value. The mechanism of the photocatalytic degradation of oleic acid on TiO$_2$ films has been analyzed by Rathousky et al. [27]. In their study, nonanal and 9-oxononanoic acid and azelaic and nonanoic acid were detected as primary and secondary intermediates, respectively. They proposed that oxidative degradation on the TiO$_2$ film was induced by an attack of hydroxyl radicals on the double bond. Eliminated hydroxyl radicals could act catalytically and thus accelerate the degradation of unsaturated compounds including fatty acids, which can be expected to be proportional to the degree of degradation (i.e., its transformation into more hydrophilic compounds, as well as into gaseous products such as CO$_2$ and H$_2$O). These published propositions and our results clarify that, although the superhydrophobicity of such films is not sufficient to repel organic matter, especially oils, photocatalytic oxidation could eliminate these organic stains and preserve superhydrophobic surface after they have been exposed to outdoor conditions for a long period of time.

4. Conclusion

In this investigation, we developed a novel water-repellent and antimicrobial composite coating by combining an N-doped TiO$_2$ photocatalyst (a visible-light-sensitive photocatalyst) with polytetrafluoroethylene (PTFE) particles, a binder, and fluoro oil. This composite coating exhibited a water contact angle greater than 150 degrees, and a surface energy lower than 6 mN/m, indicating superhydrophobicity. The composite coating also exhibited a high level of bactericidal activity against *E. coli* and MRSA by visible-light illumination at both high and low intensity. In particular, greater than four logs of inactivation was observed on MRSA, which indicates that this newly developed superhydrophobic composite material may be resistant to the bacteria that cause nosocomial infections. Additionally, bacterial inactivation was enhanced, and a comparison with the N-doped TiO$_2$-only coating revealed that this enhancement was attributed to the surface roughness offered by the PTFE; more air pockets provided additional oxygen for the photocatalytic reaction leading to greater reactive oxygen species generation.
We also confirmed that organic stains adsorbed on the composite surface could be successfully removed utilizing photocatalytic oxidation, which resulted in sustainable high hydrophilic states. These results demonstrate potential applications of the new N-doped TiO₂ photocatalyst hydrophobic composite material not only for antibacterial action indoor, but also to prevent contamination outdoors.

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References


