Effect of Nano-ZnO Particle Suspension on Growth of Mung (Vigna radiata) and Gram (Cicer arietinum) Seedlings Using Plant Agar Method

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1. Introduction

Nanotechnology is a versatile field and has found application in almost all existing fields of science. The use of nanoparticles in the growth of plants and for the control of plant diseases is a recent practice [1–5]. However, whether beneficial or harmful to plant growth is an unresolved issue [6, 7]. Various studies had been carried out to understand the effect of nanoparticles on the growth of plants. For example, Lu et al. [8] studied the effect of mixtures of nano-SiO2 and nano-TiO2 on soybean seed. They found that the mixture of nanoparticles increases nitrate reductase in soybean increasing its germination and growth. Hong et al. [9, 10] and Yang et al. [11] reported that a proper concentration of nano-TiO2 was found to improve the growth of spinach by promoting photosynthesis and nitrogen metabolism. Canas et al. [12] found that nanofunctionalized carbon nanotubes enhanced root elongation in onion and cucumber.

Nano-ZnO has been widely used in industry for several decades. However, no study had been made on its potential use in agricultural. Zinc is one of the micronutrient of plants, and limited studies have been done on its beneficial effect on plant growth. However, its phytotoxicity has been reported [13]. The Present study deals with the effect of nano-ZnO particle suspension, as micro-nutrient on the growth of mung and gram seedlings at various concentrations by plant agar method [7]. The possible toxic effect at higher concentration was also studied. The concentration of nano-ZnO particles in root was determined by inductively coupled plasma/atomic emission spectroscopy. The distribution of nano-ZnO particle and the architecture of root were studied using light microscopy and scanning electron microscopy.

2. Materials and Methods

2.1. Preparation of Nano-ZnO Particle Suspension. Nano-ZnO particle (size 20 nm), used for this study was synthesized in our laboratory. Particle size was characterized using transmission electron microscopy (TEM, Philips CM200 electron microscope). Nanoparticles were directly suspended in deionized water and were dispersed using mechanical stirrer and ultrasonicator (100 W, 40 khz) for 30 min. Nano-particle suspensions of different concentration were prepared separately by weighing particles and dispersing them in deionized water.
2.2. Seeds. Seeds of plant species mung (Vigna radiata) and gram (Cicer arietinum) were purchased locally. Seeds were kept in dry place in the dark under room temperature prior to use.

2.3. Germination of Seeds. Mung and gram seeds were sterilized in 5% sodium hypochlorite solution for 10 min to ensure surface sterility and were rinsed thoroughly with deionised water several times. Mung and Gram seeds were allowed to germinate in wet cotton, at a controlled temperature of 25 ± 1°C in the dark for 24 hours, respectively. The seeds were checked for germination and sprouted seeds were used for further study.

2.4. Plant Agar Method. The effect of nano-ZnO particle suspension on the growth of seedlings was conducted in Petri dish test unit (87 mm × 18 mm). Each Petri dish containing 30 mL of dual agar culture media (20 mL of 2.5% agar covered with 10 mL of 1% agar) were used in the test. To these Petri dish test units, specific concentrations of nano-ZnO particles suspension were added (0, 10, 20, 50, 100, 500, 1000, and 2000 ppm for mung seeds and 0, 1, 2, 5, 10, 20, 50, 100, 500, 1000, and 2000 ppm for gram seeds). For preparing the test media, 20 mL of 2.5% agar solution was poured into a Petri dish test unit and immediately solidified in freezer to avoid the possible precipitation of nanoparticles. Again, 10 mL of 1% agar solution was poured evenly over 2.5% agar media. To each Petri dish test unit, 10 plant seedlings were gently placed above the surface of agar media. The test units were placed in incubator at a controlled temperature of 25 ± 1°C in dark. For each concentration, three replicates were prepared and the test units were exposed for 60 hr. After exposure period, the seedlings were separated from agar media and their growth response was measured with ruler. Agar media without nano-ZnO particles were used as control. Based on our preliminary observations, the test units showing highest and lowest seedling growth for corresponding nano-ZnO concentrations with respect to control were selected for further studies.

2.5. Sample Preparation for Microscopy. For light microscopy, the root sample sections were taken manually and were observed under light microscope (Leica, DM500 B, Germany). For scanning electron microscopy, the control and nano-ZnO particle treated seedlings were cut to separate root and shoot. The roots were frozen in liquid nitrogen and then cut into smaller pieces with blunt knife. The pieces were freeze-dried overnight at −30°C under vacuum (15–25 torr) using freeze dry system (ALPHA1-2 LDplus, Germany). The morphology of roots was examined by scanning electron microscope, while quantitative elemental analysis for presence of nanoparticles was carried out with electron diffraction spectrum (SEM/EDS, Model no. S3400, Hitachi).

2.6. Biomass Assay and Zinc Concentration Determination. At the end of the experiment, seedlings were washed with deionised water. Roots and shoots were separated from seedlings for biomass determination. Initially, the fresh weight of roots and shoots was measured, dried in oven at 70°C for 24 hr and dry weights were recorded. Zn content in the root was also determined by inductively coupled plasma/atomic emission spectroscopy (ICP-AES, Model Ultima, Jobin-Yvon Company, France) after HNO3 digestion of dry roots.

2.7. Statistical Analysis. The experiment was conducted in three replicates and the results were presented as the mean ± SD (standard deviation). The statistical analysis of experimental data utilized Student’s t-test. Each of the experimental value was compared to its corresponding reference. Statistical significance was accepted when the probability of result assuming the null hypothesis (P) was less than 0.05.

3. Result and Discussion

3.1. TEM Study of Nano-ZnO Particles. The transmission electron microscopy (TEM) image and selected area electron diffraction (SAED) pattern of ZnO particles are shown in Figures 1(a) and 1(b), respectively. The TEM micrographs indicated that the zinc oxide particles were monodispersed with a narrow size distribution and near spherical morphology. Analysis of particles in TEM monograph indicates hexagonal particles with the average size of 20 nm. The SAED pattern shows that the rings are composed of dots suggesting the crystalline nature of ZnO particles.

3.2. Effect of Nano-ZnO Particles Suspension on Root and Shoot Growth. The dose response curves of nano-ZnO on
root and shoot of mung and gram seedlings are shown in Figures 2(a) and 2(b), respectively. It was observed that with increase in nano-ZnO concentration, the root and shoot growth also increases. However, after certain concentration the growth of root and shoot was found to decline. For mung seedlings, the best growth response for root (42.03%, \( P \) value 0.0498) and shoot (97.87%, \( P \) value 0.0444) was observed at a concentration of 20 ppm over control. At highest concentration, 2000 ppm the retardation in root (93.28%, \( P \) value 1.736 \( \times \) 10\(^{-14} \)) and shoot (14.85%, \( P \) value 2.46 \( \times \) 10\(^{-11} \)) growth of mung seedlings were observed. Similarly, for gram seedlings, the dose of 1 ppm promotes significant increase in root (53.13%, \( P \) value 1.125 \( \times \) 10\(^{-7} \)) and shoot (6.38%, \( P \) value 0.026) growth as compared to control. However, beyond this concentration retardation in growth of root and shoot was observed. At a dose of 2000 ppm, significant reduction in the root growth (74.18%, \( P \) value 1.32 \( \times \) 10\(^{-10} \)) and shoot growth (22.52%, \( P \) value 0.020) was observed. The reduction in root and shoot growth at higher doses may be attributed to toxic level of nanoparticles. This is good evidence for demonstrating that mung and gram seedlings respond to added nanoparticles in a limited range, above which toxic levels are reached causing subsequent declines in growth.

3.3. Effect of Nano-ZnO Particles Suspension on Biomass. The graph of biomass estimated with respect to concentration of nanoparticles for mung and gram seedlings is shown in Figures 3(a) and 3(b), respectively. Root and shoot biomass production were found to be in accordance with the root and shoot length for corresponding nano-ZnO treatment. For nano-ZnO at 20 ppm treatment, mung seedlings showed 40.89% increase the root biomass and 76.04% increase in shoot biomass over control. In case of gram seedlings at 1 ppm treatment, 37.15% increase in root biomass and 26.61% increase in shoot biomass was observed. At the highest concentration of 2000 ppm, 44.75% decrease of root biomass and 31.95% in shoot biomass for mung seedling was observed. While for gram seedlings, 32.25% decrease in root biomass and 52.84% decrease in shoot biomass were observed. The increase in biomass at certain concentration suggests the optimum dose limit for the growth of mung and gram seedlings. However, the decrease in biomass beyond this concentration suggests the toxic effect of nano-ZnO particles.

3.4. Elemental Determination. Table 1 shows the Zn content in the roots of mung and gram seedlings at optimum and highest concentration. It was observed that the Zn content of the roots in mung and gram increased significantly with the
Figure 4: Transverse section of mung-seedling roots (a) control, (b) at 20 ppm, and (c) at 2000 ppm of nano-ZnO treatment.

Figure 5: Transverse section of gram-seedling roots (a) control, (b) at 1 ppm, and (c) at 2000 ppm of nano-ZnO treatment.

Table 1: Weight of zinc element in (a) mung and (b) gram seedling roots determined by ICP-AES.

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The effect of ZnO nanoparticles to the mung and gram roots were further examined by light microscope. Figures 4 and 5 shows the transverse section of mung and gram roots, respectively. At a treatment of 20 ppm (Figure 4(b)) and 1 ppm (Figure 5(b)) concentration dose of nano-ZnO for mung and gram, respectively, the root developed very well with the usual three tissue system (epidermis, cortex, and vascular cylinder) as observed in control (Figures 4(a) and 5(a)). However, the disruption of tissue system in both mung and gram roots (Figures 4(c) and 5(c)) was observed at higher concentration. The cortical cells were highly vacuolated and collapsed, while vascular cylinder was shrunk. The damage to the basic architecture of roots at higher concentration may be a direct reason for the growth inhibition in mung and gram seedlings [14].

Figures 6 and 7 shows the surface micrographs of roots of mung and gram seedlings, respectively. At a treatment of 20 ppm (Figure 6(b)) and 1 ppm (Figure 7(b)) concentration of nano-ZnO particles for mung and gram, respectively, no structural alteration on the root surface was observed when compared with respective control samples (Figures 6(a) and 7(a)). However, structural alteration in root surface was observed for seedlings treated at higher concentrations. Damage to the epidermal layer of roots at higher concentration was evident from Figures 6(e) and 7(e). The adsorption and aggregation of nano-ZnO particles were observed on root surface of treated seedling, which was located and identified on the root surface by SEM-EDEX analysis (Figures 6(c), 6(d), 6(f), 6(g), 7(c), 7(d), 7(f), and 7(g)).

3.5. Microscopic Analysis. The increase of nano-ZnO application. It indicates that the ZnO nanoparticles are absorbed and translocated in seedlings resulting in increase in Zn content at corresponding nano-ZnO treatment.

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Figure 6: Scanning electron micrographs of roots of mung (Vigna radiata) (a) control, (b) at 20 ppm with EDAX (c) and (d) and (e) at 2000 ppm with EDAX (f) and (g) of nano-ZnO treatment. The energy dispersive spectroscopy shows the Zn peaks obtained from points indicated by the arrow.
Figure 7: Scanning electron micrographs of roots of gram (Cicer arietinum) (a) control, (b) at 1 ppm with EDAX (c) and (d) and (e) at 2000 ppm with EDAX (f) and (g) of nano-ZnO treatment. The energy dispersive spectroscopy shows the Zn peaks obtained from points indicated by the arrow.
4. Conclusion

The presence of ZnO nanoparticles affects the growth of mung and gram seedlings at different concentrations. The maximum effect was found at 20 ppm for mung (Vigna radiata) and 1 ppm for gram (Cicer arietinum) seedlings. Beyond this concentration, the growth was inhibited. The statistically determined $P$ value for root and shoot growth at optimum concentration and at highest concentration was found to be less than 5%. The effective growth at certain optimum concentration and inhibited growth beyond this concentration may be attributed to the accumulation and uptake of nano-ZnO particle by the roots. It was found that the accumulation and uptake of nanoparticles was dependent on the exposure concentrations.

Acknowledgment

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References

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