Reduction of ferric hemoglobin from *Trematomus bernacchii* in a partial bis-histidyl state produces a deoxy coordination even when encapsulated into the crystal phase

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Abstract. Crystallographic and spectroscopic evidences on Antarctic fish hemoglobins (AFHbs) have revealed that their ferric tetramers at physiological pH are in a mixed α(aquo-met)/β(bis-histidyl) coordination state and show a quaternary structure intermediate between the classical R and T states (H state). Ferric bis-histidyl adducts (hemichromes) have been also observed in some mammalian Hbs. In order to clarify whether hemichrome in AFHbs can be converted into a ferrous bis-histidyl adduct (hemochrome), at least in the crystal phase, chemical reduction of ferric hemoglobins from *Trematomus bernacchii* (HbTb) single crystals has been followed via Raman microscopy. The results of this analysis reveal that in HbTb, upon reduction, the bis-histidyl coordination is disrupted in favor of a penta-coordinated ferrous deoxy state, with no evidence of hemochrome. These data are in agreement with UV/Vis absorption spectra in solution. Furthermore, our data are also indirectly supported by the observation that upon reduction with dithionite, the ferric HbTb crystals crack and lose their diffraction power: in the crystalline state, the quaternary structure transition from the H to the T state is not compatible with the crystal packing. Altogether these data indicate that if bis-histidyl adducts have a functional significance in AFHbs, this function refers to a stable ferric state, or to a transient, though never detected, ferrous species.

Keywords: Roman crystallography, hemicrome, Antarctic fish, hemoglobins

Abbreviations

AFHb, Antarctic fish hemoglobin;
Hb, hemoglobin;
HbA, human adult hemoglobin;

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HbTb, major hemoglobin of *Trematomus bernacchii*; ox-HbTb, ferric hemoglobin of *Trematomus bernacchii* at pH 7.6; RR, resonance Raman; 6cHS, hexa-coordinate high spin; 6cLS, hexa-coordinate low spin; 5cHS, penta-coordinate high spin.

1. Introduction

Tetrameric hemoglobins (Hbs) deserve a special position in the field of protein allostery [1–5]. The pioneering achievements of Perutz led to the identification of two distinct Hb structures: T (tense) and R (relaxed) [6]. More recent crystallographic analyses have clearly shown that tetrameric Hbs possess a larger repertoire of structural states [7–14]. Many of these observations are based on studies of Antarctic fish Hb (AFHb).

Cold-adapted proteins generally differ from their mesophilic or thermophilic homologues only for limited structural characteristics and/or relatively few amino-acid substitutions in functional regions, which can lead to enhanced structural flexibility. Thus, a systematic study of these molecules may facilitate the identification and characterization of accessible conformational substates that result in a better understanding of their functional properties. Tetrameric Hbs from Antarctic fish are particularly well suited to this purpose, as they form a highly correlated system with peculiar structural and functional similarities and differences. These Hbs have T and R states very similar to those characterized in mammalian Hb [15–17]. However, they follow a peculiar oxidation pathway, that is characterized by a distinct behaviour of the α and β chains [18,19]. Intermediate species with the α chain in the oxy, carbomonoxy or met state and the β chain in the hexa-coordinate low-spin, 6cLS, bis-histidyl form (hemichromes) [18–20] or in the penta-coordinate high-spin, 5cHS, ferric form have been detected [20,21]. Interestingly, these mixed forms also present a quaternary organization somewhat intermediate between the classical R and T states (H state) [18–20]. Recently, these ferric states have been identified also in polar and polar-related fish Hbs by using a combination of Raman spectroscopy [22–24], EPR [20,21] and X-ray crystallography [18–20,24].

The ferric forms of Hb are physiologically inert to further oxygenation, but several subsequent side reactions in the Hb auto-oxidation may interfere with or merge into other biochemical pathways, including the formation of a hemichrome. The relevance of this species spans from biomedical to physiological aspects [23]. For example, auto-oxidation is a serious problem that limits the storage time of acellular Hb-based blood substitutes [25]; also, hemichrome detection has been suggested as a valuable tool for tumour diagnosis [26]. The reaction of acetylphenyldrazine (APH) with erythrocytes leads to hemichrome formation in healthy people and not in breast-cancer patients [26]. Some of the hypotheses on the functional role of 6cLS hemichrome were reviewed in [27]. The bis-histidyl complex can be involved in ligand binding [28,29], in the *in vivo* reduction of met-Hb, in Heinz body formation [27] and in nitric oxide scavenging.

Reduction of the ferric form of AFHb in solution produces a deoxy state [19]. Interestingly, in human Hb (HbA), thermally-generated hemichrome in trehalose glass [25], when thermally-reduced by glucose, converts to a ferrous deoxy [25] or to a hemochrome state [30], depending on the heating time.

In order to establish whether AFHb hemichromes encapsulated in the crystal phase can be converted into hemochromes, we chemically reduced the ferric HbTb crystals and followed the process by Raman microscopy. Similar to results obtained by short thermal reduction of trehalose-glass encapsulated ferric
HbA and myoglobin [25], but unlike the long thermal reduction in trehalose glass of HbA and myoglobin [30], the crystal environment does not favour the formation of a ferrous 6cLS bis-histidyl adduct in HbTb [19]. As observed in solution, in the crystal state HbTb hemichrome converts into a 5cHS deoxy state. The transition from the H to the T quaternary structure produces severe surface cracks on protein crystals that cannot allow the X-ray diffraction data collection, hampering structural characterization of the process at the atomic level.

2. Materials and methods

2.1. Purification and crystallization

Purification and storage of Hb from T. bernacchii (HbTb) was performed as previously reported [31]. HbTb was oxidized with K$_3$Fe(CN)$_6$ and the excess was removed by gel filtration on a Sephadex G-25 column. Crystallization of oxidized HbTb (ox-HbTb) was carried out at pH 7.6 and room temperature by liquid-diffusion, using a capillary, according to a previously described procedure [19]. These crystals correspond to the previously characterized $\alpha$(aquo-met)/$\beta$(hemichrome) structure [19].

2.2. Reduction of ox-HbTb crystals

In order to obtain the structure of reduced ox-HbTb, the crystals have been soaked for a few hours in a stabilizing solution containing 500 mM dithionite. Unfortunately, the soaking procedure produced cracks on the crystals surface that prevented data collection. To reveal at least the first events in the reducing pathway of ox-HbTb, diffraction data on two crystals, soaked in a solution of dithionite for just few min (10 and 30 min), have been collected at high resolution (1.60 and 1.76 Å, respectively) at the XRD1 beam-line of Elettra synchrotron. Data were collected at 100 K using glycerol as cryo-protectant and processed with the program suite HKL2000. The coordinates of the structure of the $\alpha$(aquo-met)/$\beta$(hemichrome) form of HbTb (Protein Data Bank (PDB) code 2PEG), solved from crystals isomorphous to those here reported, were used as a starting model to refine the new structures. The refinement runs, performed using the program CNS [32], were followed by manual intervention using the molecular graphic program O [33] to correct minor errors in the position of few side chains. At convergence, the R-factor (R-free) values for the two structures were 0.211 (0.234) and 0.195 (0.222), respectively. Further diffraction and refinement statistics are listed in Table 1.

2.3. Raman microscopy

The reduction of ferric HbTb was also followed by resonance Raman (RR) microscopy. In the RR experiments carried out in solution the protein was kept in 50 mM Tris-HCl (pH 7.5). The heme concentration was 0.5 mM. A confocal Raman microscope (Jasco, NRS-3100) was used to record Raman spectra. One of the three available lines (488 and 514 nm of an air-cooled Ar$^+$ laser, and 532 nm of a NdYAG laser) was injected into an integrated Olympus microscope and focused to a spot size of approximately 2 µm by a 20 × objective. A holographic notch filter was used to reject the excitation laser line. Raman scattering was dispersed through a monochromator (2400 grooves/mm grating) and collected by a peltier-cooled 1024 × 128 pixel CCD photon detector (Andor DU401BVI). Typically, several 15 min solution spectra were recorded and averaged (3–4 cm$^{-1}$ resolution) by a standard software routine. Frequency shifts were calibrated by using indene. Microscopy experiments were conducted on ferric HbTb
Table 1

<table>
<thead>
<tr>
<th>Data collection</th>
<th>ox-HbTb 10 min in dithionite</th>
<th>ox-HbTb 30 min in dithionite</th>
</tr>
</thead>
<tbody>
<tr>
<td>Space group</td>
<td>C2</td>
<td>C2</td>
</tr>
<tr>
<td>Cell parameters</td>
<td></td>
<td></td>
</tr>
<tr>
<td>a (Å)</td>
<td>87.158</td>
<td>87.405</td>
</tr>
<tr>
<td>b (Å)</td>
<td>87.762</td>
<td>87.687</td>
</tr>
<tr>
<td>c (Å)</td>
<td>55.385</td>
<td>55.413</td>
</tr>
<tr>
<td>$\beta$ (°)</td>
<td>97.76</td>
<td>97.73</td>
</tr>
<tr>
<td>Resolution limits (Å)</td>
<td>30.00–1.60</td>
<td>30.0–1.76</td>
</tr>
<tr>
<td>Highest resolution shell (Å)</td>
<td>1.65–1.60</td>
<td>1.81–1.76</td>
</tr>
<tr>
<td>No. of observations</td>
<td>387,063</td>
<td>267,769</td>
</tr>
<tr>
<td>No. of unique reflections</td>
<td>57,841</td>
<td>43,414</td>
</tr>
<tr>
<td>Completeness (%)</td>
<td>98.8 (98.5)</td>
<td>98.9 (98.5)</td>
</tr>
<tr>
<td>$I/\sigma(I)$</td>
<td>14.7 (3.0)</td>
<td>23.7 (3.2)</td>
</tr>
<tr>
<td>R-merge (%)</td>
<td>7.4 (24.1)</td>
<td>5.5 (39.0)</td>
</tr>
<tr>
<td>Mosaicity</td>
<td>0.53</td>
<td>0.53</td>
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<tr>
<td>Refinement results</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Resolution limits (Å)</td>
<td>20.00–1.60</td>
<td>20.0–1.76</td>
</tr>
<tr>
<td>Number of reflections used in the refinement</td>
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<td>38,218</td>
</tr>
<tr>
<td>No. of reflections in working set</td>
<td>46,038</td>
<td>34,368</td>
</tr>
<tr>
<td>No. of reflections in test set</td>
<td>5205</td>
<td>3850</td>
</tr>
<tr>
<td>R-working/R-free (%)</td>
<td>21.1/23.4</td>
<td>19.5/22.2</td>
</tr>
<tr>
<td>No. of protein atoms</td>
<td>2199</td>
<td>2199</td>
</tr>
<tr>
<td>No. of water molecules</td>
<td>319</td>
<td>324</td>
</tr>
<tr>
<td>RMSD from ideal values</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bond lengths (Å)</td>
<td>0.009</td>
<td>0.010</td>
</tr>
<tr>
<td>Bond angles (°)</td>
<td>1.21</td>
<td>1.21</td>
</tr>
</tbody>
</table>

Note: Values in parentheses correspond to the highest resolution shells.

(ox-HbTb) crystals by previously reported procedures [34]. A complete data set on HbTb crystals was registered in 60 s. Raman crystallography measurements on crystals mounted onto a goniometer head were performed by means of a modified Ventracon accessory.

3. Results

3.1. X-ray diffraction data

Crystals of ferric HbTb (ox-HbTb) were grown at pH 7.6 according to a previous procedure [19, 20]. Single crystals of HbTb grew as large as 0.2 × 0.2 × 0.2 mm³. The crystal structure revealed an $\alpha$(aquo-met)/$\beta$(hemichrome) state [19,20]. The quaternary structure of this form at pH 7.6 is intermediate between R and T state. This new quaternary structure will be hereafter denoted as H state. The quaternary structure of the deoxy HbTb crystals from pH 6.0 up to 8.4 showed a classical T state [16].

To characterize the reduction of the ox-HbTb structure at atomic level, crystals of ox-HbTb were soaked in a stabilizing solution containing 100 mM dithionite, from a few min to a few hours. Diffrac-
tion data on two HbTb crystals soaked in dithionite for 10 and 30 min were collected at 1.60 and 1.76 Å resolution, respectively (Table 1). The structures obtained from these crystals reveal no significant differences from ox-HbTb (the root mean square deviation (RMSD) calculated superimposing the C\(^\alpha\) atoms is only 0.13 Å in both cases). The two structures are also very similar to each other (RMSD < 0.1 Å). The overall structure adopts the H-state quaternary shape. The \(\alpha\) heme is in 6cHS aquo-met/6cLS hydroxy-met state, with the centre of the electron-density peak about 2.0 Å away from the iron and about 4 Å from the \(N\varepsilon_2\) atom of distal His-59\(\alpha\). In the \(\beta\) chains, a 6cLS bis-histidyl coordination of the irons is observed. These data indicate that a few-minute dithionite soaking is not sufficient to significantly reduce the \(\alpha\)-heme iron of ox-HbTb crystals. Furthermore, no indication on the \(\beta\)-heme oxidation state is provided, since the 6cLS state for iron Fe\(^{2+}\) and Fe\(^{3+}\) are not expected to show relevant stereochemical differences. On the other hand, dithionite-soaking times of a few hours, caused severe surface cracks on the HbTb crystals that hamper data collection. This evidence indirectly suggests that the reduction of ox-HbTb involves a quaternary structure transition that is not compatible with the crystal packing of HbTb.

3.2. Raman microscopy

Since the reduction of ox-HbTb cannot be properly investigated by X-ray crystallography studies, we followed this process using Raman microscopy. Ox-HbTb crystals, before and after reduction by dithionite, were characterized. The high-frequency region (1300–1700 cm\(^{-1}\)) of the RR spectrum includes the porphyrin in-plane vibrational modes (that are sensitive to the electron density of the macrocycle), oxidation, coordination and spin state of the iron atom [35]. During most of the analysis, crystals were kept into a 1-µl drop of their mother liquor. High-frequency resonance Raman spectra of a solution and single crystal of ox-HbTb are reported in Fig. 1. The two spectra, except for an intensity factor, are indistinguishable. Therefore, the starting point of the reduction followed by Raman microscopy is the same as in solution. A tentative assignment of the Raman bands, based on previous studies [35], is reported in Table 2. The spectra of ox-HbTb crystals are characterized by a mixture of two ferric states, a 6cHS aquo-met state (\(\nu_1\) and \(\nu_2\) bands at 1476 and 1561 cm\(^{-1}\)) and a 6cLS hemichrome state (\(\nu_3\) and \(\nu_2\) bands at 1505 and 1578 cm\(^{-1}\)). The X-ray crystal structure of ox-HbTb has assigned equal amounts to aquo-met and hemichrome, due to selective binding to the \(\alpha\) and \(\beta\) heme, respectively [19,20]. High-frequency Raman spectra of ferric HbTb single crystals have been collected using three excitation wavelengths (Fig. 2). The enhancement of Raman bands corresponding to the two ferric states is different when using the three wavelengths around the zone of the Q-bands, that should particularly enhance B-terms [36]. The 532-nm line particularly enhances the 6cLS hemichrome bands (\(\nu_{10} = 1506, \nu_{19} = 1587\) and \(\nu_{10} = 1640\) cm\(^{-1}\)), and it is expected to also enhance the bands of the 6cLS hemochrome state, namely the main target of this investigation. Therefore, we chose the 532 nm line to follow ox-HbTb reduction encapsulated into the crystal state.

Raman spectra of single crystals can be affected by the crystal orientation [37]. Therefore, prior to reduction of ferric HbTb crystals, we transferred a single crystal into a capillary (0.5 mm-diameter) for X-ray diffraction, which was mounted onto a goniometer head for Raman crystallography analysis. The high-frequency spectra were recorded at different crystal orientations, showing that the crystal orientation does not affect the Raman spectrum of ox-HbTb (data not shown).

Reduction of the ferric HbTb crystals was initiated by adding a small volume of stabilizing solution containing dithionite to the drop of the crystal. Raman spectra at different dithionite additions are reported in Fig. 3. The quality of the spectra deteriorate with dithionite addition, but this is not surprising,
Fig. 1. RR spectra of ferric HbTb in crystal form (1-min acquisition) and solution (15-min acquisition) with 488-nm excitation line (2.7 mW at the sample).

### Table 2

<table>
<thead>
<tr>
<th>Mode</th>
<th>Ferric 6cLS</th>
<th>Ferric 6cHS</th>
<th>Ferrous 5cHS</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\nu_2$</td>
<td>1578</td>
<td>1561</td>
<td>1565</td>
</tr>
<tr>
<td>$\nu_3$</td>
<td>1505</td>
<td>1476</td>
<td>1471</td>
</tr>
<tr>
<td>$\nu_4$</td>
<td>1373</td>
<td>1371</td>
<td>1355</td>
</tr>
<tr>
<td>$\nu_{10}$</td>
<td>1640</td>
<td>1610</td>
<td>1604</td>
</tr>
<tr>
<td>$\nu_{19}$</td>
<td>1588</td>
<td>1561</td>
<td>1555</td>
</tr>
<tr>
<td>$\nu(C=\text{C})$</td>
<td>1621</td>
<td>1621</td>
<td>1619</td>
</tr>
<tr>
<td>$d(=\text{CH}_2)$</td>
<td>1435</td>
<td>1429</td>
<td>1426</td>
</tr>
<tr>
<td>$\nu_{38}$</td>
<td>1543</td>
<td>1518</td>
<td>1524</td>
</tr>
</tbody>
</table>

*Bands overlapping.*

since visual inspection in an optical microscope reveals wrinkles on the crystal surfaces. Upon dithionite addition, the oxidation marker $\nu_4$ and the core size band $\nu_3$ and $\nu_{10}$ show the appearance of the ferrous 5cHS deoxy state ($\nu_4 = 1355, \nu_3 = 1471, \nu_{11} = 1548, \nu_{19} = 1555$ and $\nu_{10} = 1604$ cm$^{-1}$). Concomitantly, the ferric species ($\nu_4 = 1373$ cm$^{-1}$), and more specifically the 6cLS ferric form ($\nu_3 = 1505$ and $\nu_{10} = 1640$ cm$^{-1}$), tends to disappear. The putative 6cLS hemochrome state should appear at $\nu_4 = 1361$.
Fig. 2. RR spectra of ferric HbTb in crystal form (1-min acquisition) with 488-nm (2.7 mW at the sample), 514-nm (3 mW at the sample) and 532-nm (0.7 mW at the sample) excitation lines.

and $\nu_3 = 1493$ cm$^{-1}$ [38,39]. On the contrary, the final spectrum in Fig. 4 contains bands associated with non-totally symmetric skeletal modes ($\nu_{10}$, $\nu_{19}$ and $\nu_{11}$), and it matches well the spectrum obtained at 532 nm for deoxy HbA [40]. Indeed, this reference spectrum contains a broad overlapped band ($\nu_{19}$ plus $\nu_{11}$) with a maximum at 1550 cm$^{-1}$, and another band $\nu_{10}$ at 1604 cm$^{-1}$ [40]. Though the final spectrum is deteriorated by partial crystal dissolution, it mostly corresponds to that of a deoxy state; if present, only a minor amount of a ferrous 6cLS species is formed. The Raman microscopy data on crystals are in perfect agreement with the spectroscopic data in solution [19].

4. Discussion

In principle, when reduced by dithionite, the $\alpha$(aquo-met)/$\beta$(hemichrome) crystals may be converted to a mixture of high-spin penta-coordinate (5cHS) deoxy state and low-spin hexa-coordinate (6cLS) hemochrome state. In fact, the crystal phase could hamper the scissor-like motion that helices EF undergo to disrupt the endogenous bis-histidyl coordination.

We have characterized the reduction of HbTb hemichrome in a crystal phase. Unfortunately, X-ray diffraction experiments cannot be performed on reduced ox-HbTb crystals, given that the addition of dithionite produces severe crystal damage that hamper data collection. However, experiments on these
crystals suggest that the reduction process of ox-HbTb is a cooperative phenomenon, involving a conformational variation that cannot be accommodated into the protein crystal. These indirect indications are in full agreement with the Raman microscopy data, showing that the 6cLS bis-histidyl coordination is loosened in favour of a 5cHS ferrous deoxy state, with no evidence of ferrous 6sLS bis-histidyl state. Upon reduction, the hemichrome converts to a deoxy ferrous state; the quaternary structure of ox-HbTb must change from the H [20] to the T state [16]. This large conformational variation produces increased disorder in the protein crystal that decreases its diffraction power. The conversion of hemichrome in HbTb to the deoxy state is also in agreement with the data on reduction of ferric HbTb in solution [19]. Altogether these analyses suggest that in Antarctic fish Hbs (AFHb), the bis-histidyl adduct is typical only of the ferric state (hemichrome). In several Hbs, both hemochrome and hemichrome occur, and in other systems only hemochrome has been observed [23]. Despite the structural similarities in the local stereochemistry of the two compounds, and the restrictions caused by the crystal packing, the hemochrome does not replace efficiently the hemichrome species. The interpretation of such a result requires a quantum-mechanical approach that is under way in our laboratory.

The results of this work indicate that, even though bis-histidyl adducts have a functional significance in AFHbs, this function refers to a stable ferric state, or to a transient, though never detected, ferrous species.
Acknowledgements

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