The cytokines interleukin-8 (IL-8) and granulocyte-macrophage colony-stimulating factor (GM-CSF) enhanced the extracellular release of arachidonate metabolites from ionophore-stimulated neutrophils by 145 ± 10% (mean ± S.E.M., n = 13) and 182 ± 11% (n = 16), respectively. To determine whether enhanced leukotriene production mediates the effects of these cytokines on neutrophil activity, two different specific arachidonate 5-lipoxygenase (5-LO) inhibitors, piriprost and MK-886, were used to inhibit leukotriene synthesis. Neither inhibitor affected the upregulation of CD11b β2-integrin expression or priming of superoxide generation stimulated by IL-8 and GM-CSF. It is concluded that leukotrienes do not mediate either the direct or priming effects of these cytokines and that these classes of anti-inflammatory drugs are therefore unlikely to inhibit the effects of IL-8 and GM-CSF on neutrophil activation.

Key words: Granulocyte-macrophage colony-stimulating factor, β2-Integrin, Interleukin-8, Leukotrienes, 5-Lipoxygenase inhibitors, Neutrophil, Superoxide

The effect of inhibition of leukotriene synthesis on the activity of interleukin-8 and granulocyte-macrophage colony-stimulating factor

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Materials and Methods

Materials: Recombinant human (rh) IL-8 from monocytes was a kind of gift of Sandoz Forschungsinstitut, Vienna, Austria. A stock solution at 100 µg/ml was prepared in sterile PBS with 2% foetal calf serum and stored at −80°C. A
P. J. Roberts, A. R. Pizzey and D. C. Linch

1 μg/ml stock solution of rhGM-CSF (expressed in E. coli and provided by Hoechst UK, Beringwerke, Marburg, Germany) was prepared in RPMI medium with 2% (v/v) foetal calf serum and stored at -20°C. Lymphoprep (density = 1.077 g/ml) was obtained from Nycomed Pharma AS, Oslo, Norway. Dulbecco’s phosphate buffered saline (PBS) was from Gibco BRL, Paisley, Scotland. The Mo1 CD11b and control IgM antibodies (Coulter clone) were from Coulter Electronics Ltd., Luton, Bedfordshire, UK. [5,6,8,9,11,12,14,15-3H]arachidonic acid was from Amersham International, Marburg, Germany) was prepared in RPMI 114 medium with 2% (v/v) foetal calf serum and stored at -20°C. Lymphoprep (density = 1.077 g/ml) was obtained from Nycomed Pharma AS, Oslo, Norway. Dulbecco’s phosphate buffered saline (PBS) was from Gibco BRL, Paisley, Scotland. The Mo1 CD11b and control IgM antibodies (Coulter clone) were from Coulter Electronics Ltd., Luton, Bedfordshire, UK. Piriprost (6,9-deepoxy-6,9-(phenylimino)-delta-6,8-prostaglandin E1) was a gift from Dr Michael Bach, The Upjohn Laboratories, Kalamazoo, MI, USA. The FLAP antagonist, MK-886 (L-663,536) 3-[1-(p-chloro-benzyl)-5-isopropyl-3- r-butylthioindol-2-yl]-2,2'-dimethylpropanoic acid, was a gift from Dr Gillard, Merck-Frosst Canada Inc., Pointe Claire-Dorval, Quebec, Canada. C18 (300 mg) Maxi-Clean columns were from Alltech, Carnforth, Lancashire, UK. Leukotriene standards were from Cascade Biochem Ltd, Reading, Berkshire, UK.

Neutrophil purification: Neutrophils were purified from venous blood anticoagulated with 2 mM EDTA, pH 7.4 by dextran sedimentation, Lymphoprep density gradient separation and erythrocyte lysis, as described previously. The purified neutrophils were suspended in PBS supplemented with 0.9 mM CaCl2, 0.5 mM MgCl2 and 5 mM D-glucose (PBSG).

CD11b expression: 50 μl aliquots of purified neutrophils (1 × 10^6/ml) were incubated with either diluent, IL-8 (100 ng/ml), or GM-CSF (10 ng/ml) for 30 min at 37°C. The samples were placed on ice and CD11b expression was measured by flow cytometry using direct immunofluorescence with a mouse IgM FITC-conjugated CD11b monoclonal antibody (Mo1) as described previously.

Superoxide generation: Cells (1 × 10^6/ml) were equilibrated in 1 ml plastic spectrophotometer cuvettes for 5 min before addition of cytokines or diluent. Cells were stimulated with either 1 μM formyl-methionyl-leucyl-phenylalanine (fMLP) or 1.67 μM 12- O-tetradecanoylphorbol-13-acetate (TPA). Superoxide production by purified neutrophils at 37°C was measured by the continuous assay of superoxide dismutase-inhibitable reduction of ferricytochrome c, as described previously.

Extracellular release of [3H]arachidonate-labelled metabolites from intracellular arachidonate: Purified neutrophils (5 × 10^6/ml PBSG, 0.1% FCS) were labelled with [3H]arachidonate (202 Ci/mmol) at a final concentration of 0.5 μCi/ml for 2 h at 37°C with occasional mixing. Under these conditions approximately 80% of the isotope was taken into the cells. The radiolabelled cells were centrifuged (180 × g, 5 min) and the supernatant containing unincorporated isotope was removed. The cell pellet was washed four times in PBSG and finally resuspended to 2 × 10^6 cells/ml PBSG. Control experiments measuring superoxide production showed that this labelling procedure did not itself cause priming of neutrophils. 0.5 ml aliquots were equilibrated to 37°C, mixed with either diluent, IL-8 (100 ng/ml), or GM-CSF (10 ng/ml) and incubated for 30 min at 37°C. Arachidonate metabolism was stimulated with Ca^{2+} ionophore A23187 (1 μM) for 7 min and the reaction was terminated by placing the samples on ice. The samples were centrifuged (12,000 × g, 2 min) and 0.4 ml aliquots of the supernatants were assayed for radioactivity by liquid scintillation spectroscopy. The cell pellets were lysed with 0.1% Triton-X-100, made up to 0.5 ml volume, and 0.4 ml aliquots were also assayed for radioactivity.

Inhibitors: An 8.7 mM stock solution of the 5-LO inhibitor, piriprost, in distilled water and a 100 μM stock solution of the FLAP antagonist, MK886, in DMSO, were prepared immediately prior to use. In all experiments described neutrophils were incubated for 5 min with inhibitors before priming with cytokines or stimulation with 1 μM A23187. Dose–response experiments measuring the release of metabolites labelled with [3H]arachidonate were performed to determine the optimal final concentration of inhibitors. Control experiments showed that the final concentration of DMSO used (0.1% v/v) had no effect on neutrophil function.

Measurement of leukotriene synthesis: Neutrophils (1 × 10^7/ml, 4 × 10^7/sample) were warmed to 37°C and incubated with MK-886 or diluent as described above and then incubated with or without GM-CSF (10 ng/ml) for 30 min before stimulation with 1 μM A23187. The reaction was stopped after 10 min by addition of 2 vol of ice cold ethanol, followed by 200 ng prostaglandin B2 as internal standard. The samples were cooled on ice. The samples were diluted with water to give a final ethanol concentration of 15% (v/v) and centrifuged at 400 × g for 10 min at 4°C. The supernatants were acidified with formic acid to pH 3.0. Solid phase extraction of leukotrienes was performed using C18 minicolumns following the method of Powell. Eicosanoids were eluted with 5 ml ethyl acetate. The samples were dried by centrifugal evaporation, resuspended in 0.5 ml ethyl acetate and transferred to a 1.5 ml polypropylene container. The samples were again dried and the pellet was dissolved in 50 μl methanol–water (70:30). Eicosanoids were separated by reverse-phase HPLC (RP-HPLC) using a Spherisorb C18 column (ODS2, 250 ×
Primed of neutrophil 5-LO activity by IL-8 and GM-CSF: The major metabolites of endogenous arachidonate in neutrophils stimulated with the Ca\(^{2+}\) ionophore, A23187, are the 5-LO products, 5-HETE and LTB\(_4\). The synthesis of metabolites from endogenous [\(^3\)H]arachidonate pools can be estimated by counting the extracellular radioactivity released from stimulated cells. Pilot experiments using thin layer chromatography showed that under the labelling conditions used 63% of incorporated [\(^3\)H]arachidonate was in glycerophospholipid pools with no detectable free intracellular [\(^3\)H]arachidonate. Table 1 shows the effects of the cytokines, IL-8 and GM-CSF on the release of radioactivity from both resting and ionophore-stimulated neutrophils. The release of radioactivity stimulated by IL-8 was not different from control, whereas the release stimulated by GM-CSF was greater than control by 200 cpm/10\(^6\) cells (representing 0.5% of total incorporated radioactivity), a very minor but statistically significant difference (\(p = 0.003\), \(n = 13\)). In samples preincubated for 30 min with cytokine diluent, 1 \(\mu\)M A23187 stimulated the release of about 4000 cpm/10\(^6\) cells greater than background, which represented on average 9.5% of total incorporated radioactivity. When neutrophils were incubated with IL-8 and GM-CSF before stimulation, the release of radioactivity in response to A23187 was enhanced by 145 \(\pm\) 10% \((n = 13)\) and 182 \(\pm\) 11% \((n = 16)\), respectively, a statistically significant increase.

### Results

#### Priming of neutrophil 5-LO activity by IL-8 and GM-CSF

The major metabolites of endogenous arachidonate in neutrophils stimulated with the \(\text{Ca}^{2+}\) ionophore, A23187, are the 5-LO products, 5-HETE and LTB\(_4\). The synthesis of metabolites from endogenous [\(^3\)H]arachidonate pools can be estimated by counting the extracellular radioactivity released from stimulated cells. Pilot experiments using thin layer chromatography showed that under the labelling conditions used 63% of incorporated [\(^3\)H]arachidonate was in glycerophospholipid pools with no detectable free intracellular [\(^3\)H]arachidonate. Table 1 shows the effects of the cytokines, IL-8 and GM-CSF on the release of radioactivity from both resting and ionophore-stimulated neutrophils. The release of radioactivity stimulated by IL-8 was not different from control, whereas the release stimulated by GM-CSF was greater than control by 200 cpm/10\(^6\) cells (representing 0.5% of total incorporated radioactivity), a very minor but statistically significant difference (\(p = 0.003\), \(n = 13\)). In samples preincubated for 30 min with cytokine diluent, 1 \(\mu\)M A23187 stimulated the release of about 4000 cpm/10\(^6\) cells greater than background, which represented on average 9.5% of total incorporated radioactivity. When neutrophils were incubated with IL-8 and GM-CSF before stimulation, the release of radioactivity in response to A23187 was enhanced by 145 \(\pm\) 10% \((n = 13)\) and 182 \(\pm\) 11% \((n = 16)\), respectively, a statistically significant increase.

**Table 1. Effect of IL-8 and GM-CSF on the extracellular release of radioactivity from neutrophil phospholipids with [\(^3\)H]arachidonate**

<table>
<thead>
<tr>
<th>Stimulus</th>
<th>Release of [(^3)H]arachidonate metabolites (cpm/10(^6) cells)</th>
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<tbody>
<tr>
<td></td>
<td>IL-8 (100 ng/ml)</td>
</tr>
<tr>
<td>Dilluent + 0.01% DMSO</td>
<td>990 (\pm) 116 (p = 0.18)</td>
</tr>
<tr>
<td>Cytokine + 0.01% DMSO</td>
<td>1 062 (\pm) 142 (p = 0.001)</td>
</tr>
<tr>
<td>Dilluent + 1 (\mu)M A23187</td>
<td>4 514 (\pm) 692</td>
</tr>
<tr>
<td>Cytokine + 1 (\mu)M A23187</td>
<td>5 977 (\pm) 815 (p = 0.001)</td>
</tr>
</tbody>
</table>

*Radiolabelled neutrophils were incubated at 37°C for 30 min with either IL-8, GM-CSF or 2% FCS diluent, and then stimulated with either A23187 or DMSO diluent.

The authors have previously shown using RP-HPLC that total cellular leukotriene synthesis and extracellular release by ionophore-stimulated neutrophils is completely inhibited by 87 \(\mu\)M piriprost. In the present study total eicosanoid production was similarly measured in ionophore-stimulated cells by extracting both the cells and supernatants together as described in the methods and then separating the leukotrienes by RP-HPLC. Neutrophils were incubated with either 100 nM MK-886 or 0.1% DMSO for 5 min before exposure to GM-CSF (10 ng/ml) or diluent for 30 min. All samples were stimulated with 1 \(\mu\)M A23187 for 10 min. In samples not exposed to MK-886, peaks were identified that had the spectral characteristics of LTB\(_4\) and its oxidation products, however the
Peptidoleukotrienes, LTC₄, LTD₄ and LTE₄ were not detected. Figure 2 shows the data for GM-CSF primed neutrophils and similar data (not shown) were obtained with unprimed cells. Figure 2 also shows the complete inhibition of leukotriene production in GM-CSF primed neutrophils pre-incubated with 100 nM MK-886. Similar inhibition of leukotriene synthesis by MK-886 was seen in unprimed neutrophils (data not shown).

The effect of 5-LO inhibitors on the activity of IL-8 and GM-CSF:

a) Priming of fMLP-stimulated superoxide generation: Neutrophil superoxide production was measured by the superoxide dismutase-inhibitable reduction of ferricytochrome c. Initial experiments showed that the 5-LO inhibitors, piriprost (87 μM) and MK-886 (100 nM), did not inhibit respiratory burst activity stimulated by 1.67 μM TPA, demonstrating that the doses used were not toxic to the cells and that these inhibitors did not inhibit protein kinase c (data not shown). Neither did piriprost nor MK-886 inhibit the generation of superoxide stimulated by 1 μM fMLP (Table 2). Table 2 also shows that in experiments where GM-CSF and IL-8 increased fMLP-stimulated superoxide production by between three to five-fold, there was no reduction in this priming activity when replicate samples were preincubated with either piriprost or MK-886 before exposure to the cytokines, conditions which were demonstrated in parallel experiments to give greater than 90% inhibition of 5-LO activity.

b) Upregulation of CD11b expression: Neutrophil CD11b expression was measured by direct immunofluorescence, using flow cytometry to determine the mean cell fluorescence (MCF). Figure 3 shows that IL-8 rapidly stimulated the increased expression of CD11b antigen on purified neutrophils. CD11b expression was upregulated more slowly by GM-CSF, reaching a maximum by 20 min. When cells were pre-incubated with either 100 nM MK-886 or 87 μM piriprost before exposure to cytokines, there was no change in either the kinetics or magnitude of the response (Fig. 3).
Table 2. Effect of 5-lipoxygenase inhibitors on the priming of the fMLP-stimulated neutrophil respiratory burst by IL-8 and GM-CSF

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Superoxide generation (nmol/10^6 cells)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Cytokine + Cytokine</td>
</tr>
<tr>
<td>0.2% DMSO ± GM-CSF</td>
<td>5.4 ± 1.1^b 27.3 ± 2.7 (506%)^c</td>
</tr>
<tr>
<td>MK-886 ± GM-CSF</td>
<td>4.8 ± 0.7 26.1 ± 1.9 (544%)</td>
</tr>
<tr>
<td>H2O ± GM-CSF</td>
<td>5.2 ± 1.2 14.3 ± 1.8 (275%)</td>
</tr>
<tr>
<td>Piriprost ± GM-CSF</td>
<td>5.0 ± 1.0 18.3 ± 3.2 (366%)</td>
</tr>
<tr>
<td>0.2% DMSO ± IL-8</td>
<td>5.6 ± 2.0 20.2 ± 7.4 (381%)</td>
</tr>
<tr>
<td>MK-886 ± IL-8</td>
<td>6.4 ± 0.8 19.8 ± 7.1 (309%)</td>
</tr>
</tbody>
</table>

^aCells were incubated with either 87 μM piriprost, 100 nM MK-886 or diluent before the addition of GM-CSF (10 ng/ml) or IL-8 (100 ng/ml), followed by stimulation with 1 μM fMLP. ^bThe absolute data shown are the mean ± S.E. of seven experiments with GM-CSF and three experiments with IL-8. ^cSuperoxide production by samples treated with cytokine was expressed as a percentage of control (−cytokine) and is given in parentheses.

Discussion

The authors have shown that both IL-8 and GM-CSF enhance or ‘prime’ the Ca^{2+}-ionophore-stimulated extracellular release of [H]-arachidonate metabolites from endogenous phospholipid pools. This confirms previous studies using GM-CSF. Daniels et al. have recently shown that IL-8 can prime the release of [H]arachidonate from cytochalasin B-treated neutrophils, however this fungal metabolite grossly alters neutrophil phospholipid metabolism and is itself a priming agent.

Using specific inhibitors of arachidonate 5-lipoxygenase, piriprost, and the indole derivative, MK-886, which specifically binds to the membrane protein FLAP, inhibiting translocation of 5-LO and leukotriene production in intact cells, it has been demonstrated that the release of [H]arachidonate metabolites from unprimed or IL-8 primed neutrophils could be blocked by greater than 90%. That these compounds inhibited both the intracellular synthesis and extracellular release of leukotrienes in unprimed and GM-CSF primed cells was confirmed by RP-HPLC. In this study the Ca^{2+} ionophore A23187 was used to demonstrate the efficacy of these compounds because it is the most potent stimulus for neutrophil leukotriene production. The inhibition observed with these compounds was not due to any toxic effect because at the same concentrations neither inhibited the generation of superoxide stimulated by phorbol ester or fMLP.

The authors sought to determine whether leukotrienes mediate the signal transduction or priming activities of GM-CSF and IL-8. Previous studies have shown little evidence for the involvement of 5-LO in IL-8 signalling, but studies of Rapoport et al. have suggested a role for this enzyme in GM-CSF signal transduction. Our data exclude the possibility that leukotrienes function in this respect, as the upregulation of cell adhesion molecules directly stimulated by IL-8 and GM-CSF was not altered by complete inhibition of 5-LO. Priming of the fMLP stimulated respiratory burst by IL-8 and GM-CSF was similarly unaffected by inhibition of 5-LO.

It is concluded that leukotriene synthesis does not mediate either the direct activity or the priming effects of IL-8 and GM-CSF on mature neutrophils. The study shows that while therapeutic treatment of inflammation with 5-lipoxygenase inhibitors will inhibit leukotriene production by neutrophils exposed to cytokines, it is unlikely to inhibit other activities of these cytokines such as those associated with upregulation of cell adhesion proteins, degranulation, adhesion and migration, or increased oxygen free-radical production.
References


ACKNOWLEDGEMENTS. This work was financially supported by the Kay Kendall Leukaemia Fund.

Received 9 February 1993; accepted in revised form 11 March 1993