

THE present study was performed to examine whether residues 36–62 of TNF α contain the chemotactic domain of TNF α , and whether the p55 and p75 TNF receptors are involved in TNF α induced chemotaxis. The chemotactic effect of TNF α on PMN was inhibited by the mAbs Hrt-7b and Utr-1, against the p55 and p75 TNF receptors, respectively. Both receptors may therefore be required for mediating the chemotactic effect of TNF α . The synthetic TNF α 36–62, similar to TNF α , had chemotactic effects on both PMN and monocytes. The chemotactic activity of the TNF α 36–62 peptide on PMN, was inhibited by Htr-7b, Utr-1 and soluble p55 receptor, which shows that the peptide possessed the ability to induce chemotaxis through the TNF receptors. In contrast to TNF α , the peptide did not show a cytotoxic activity against WEHI 164 fibrosarcoma cells. It is suggested that different domains of the TNF α molecule induce distinct biological effects.

Key words: Chemotaxis, Molecular modelling, Synthetic peptide, TNF α , TNF receptors

The TNF receptors p55 and p75 mediate chemotaxis of PMN induced by TNF α and a TNF α 36–62 peptide

Ø. Rekdal,^{1,CA} Z. Konopski,¹ J. S. Svendsen,² J.-O. Winberg,³ T. Espevik⁴ and B. Østerud¹

¹Institute of Medical Biology, ²Institute of Mathematical and Physical Sciences and ³Polar Institute of Medical Genetics, University of Tromsø, 9037 Tromsø, and ⁴Institute of Cancer Research, University of Trondheim, N-7006 Trondheim, Norway

^{CA} Corresponding Author

Introduction

Circulating polymorphonuclear cells (PMN) and monocytes are activated by chemotactic factors for recruitment to sites of inflammation.¹ The pleiotropic cytokine, tumour necrosis factor- α (TNF α), has been reported to be chemotactic, as it induces directional locomotion of PMN and monocytes *in vitro*.^{2–5} Furthermore, *in vivo* studies show that TNF α plays a crucial role in the recruitment of neutrophils at an early stage, and monocytes at a later stage of immune complex-induced inflammatory reactions.^{6,7} Many biological effects induced by TNF α ^{8–11} have been shown to involve binding to the 55 kDa (p55) and the 75 kDa (p75) TNF receptors, which are expressed on almost all cell types.¹⁰ However, the involvement of these receptors in the TNF α induced chemotaxis has not been studied.

An interesting approach for the study of distinct TNF α activities is the use of TNF α peptides. Different TNF α peptides have recently been reported to induce distinct TNF α effects,^{12–14} and inhibit binding of TNF α to the TNF receptors.¹³ The authors have performed molecular dynamic calculations¹⁵ combined with studies on the three-dimensional structure of TNF α ¹⁶ in order to design TNF α peptides which could interact with TNF receptors, and induce TNF α effects. It was found that a peptide including residues 36–62 had conformational properties which could be related to the corresponding parent molecule. This

sequence is also one of the most homologous domains between TNF α and TNF β , which both bind to the TNF receptors.¹⁶ In the present study, this peptide was investigated for two crucial TNF α effects, chemotaxis and cytotoxicity. The involvement of p55 and p75 TNF receptors in the chemotactic response of TNF α and TNF α 36–62, was also studied.

Materials and Methods

Molecular modelling: The molecular modelling studies of TNF α peptides were performed using the Molecular Simulation Inc. Quanta 3.2/CHARMm 21.2 program package, on a Silicon Graphics Personal IRIS 4D/30 EG (USA). The peptide atom coordinates were obtained from the TNF α X-ray structure (pdb1 tnf), and peptide candidates were minimized by molecular mechanics using 2000 steps of adopted basis set Newton–Raphson minimization, before the minimized structures were subjected to molecular dynamic calculations with a total simulation of 250 ps at 300 K.

Synthesis of TNF α peptides: TNF α peptides were synthesized as described previously,¹⁷ using Fmoc chemistry on a semi-automatic peptide synthesizer (Milligen, Model 9020). The peptides were purified and analysed using reverse-phase HPLC, and FIB–MS on a VG Tribid MS instrument (VG Analytical, Manchester, UK).

Materials: Human recombinant TNF α (Hr TNF α), with a specific activity of 1.0×10^8 U/mg, was purchased from Boeringer (Mannheim, Germany). The generation of the mAbs Utr-1 and Htr-7b, specific for p75 and p55 respectively, is described elsewhere,¹⁸ and soluble p55 was kindly provided by Dr Hansruedi Loetscher, Hoffman–La Roche (Basel, Switzerland). Anti-IL-8 and anti-MCP-1 were purchased from British Biotechnology (UK). Formylmethionyl-leucyl phenylalanine (FMLP) was purchased from Sigma Chemical Co. (St Louis, MO). Endospey from Seikagaku Co. (Tokyo, Japan) was used to check endotoxin contamination.

PMN isolation: Polymorphonuclear cells (PMN) were isolated as follows: 2 ml of freshly drawn heparinized blood (10 U/ml) from healthy adults was applied on top of a bilayer consisting of 3 ml polymorphoprep and 3 ml lymphoprep in polycarbonate tubes (Nycomed Pharma AS, Norway). After centrifugation at $530 \times g$ for 20 min, the PMN band in the polymorphoprep layer was isolated. The cells were washed once with ice cold sterile 0.15 M NaCl, and centrifuged at $185 \times g$ for 10 min. Contaminating erythrocytes in the PMN band were lysed with ice cold 0.2% NaCl for 90 s. The cells were resuspended at 10^6 /ml in ice cold RPMI-1640 and used immediately. The PMN preparation contained at least 95% neutrophils.

Monocyte isolation: The monocyte band was isolated using the method previously described by Bøyum.¹⁹ In brief, mononuclear cells (PMBC) from healthy adults, from either freshly drawn heparinized blood or buffy coats (10 U/ml), were centrifuged on lymphoprep, isolated and washed with 0.15 M NaCl. The PMBC, resuspended at 10^6 /ml in RPMI-1640 were used directly for chemotaxis studies.

Assay for chemotaxis: TNF α and TNF α peptides were tested for chemotactic activity on PMN and PBMC. Chemotactic activity was assayed in a 48-well microchemotaxis chamber (Neuro Probe Inc. Cabin John, MD, USA), as described previously.²⁰ In brief, the upper wells were filled with 50 μ l of cells, and 25 μ l of the compounds tested for chemotaxis were filled in the bottom wells. For checkerboard analysis, the stimulants were also placed in the upper wells. In the pretreatment studies, the anti-TNF receptor antibodies, or other antibodies were added to the cells for 10 min at 4°C, before they were placed in the upper wells. The soluble p55 was mixed with 0.5 μ M TNF α 36–62 in a 1:1 molar ratio at 20°C for 10 min, before addition to the lower wells. A polycarbonate–polyvinyl pyrrolidone (PVP) filter with 5 μ m pore size was used in the PBMC chemotaxis assay, while a PVP-free polycarbonate filter, with the same pore size, was used for the PMN chemotaxis assay. Chemotaxis chamber assemblies

were incubated at 37°C in humidified 95% air and 5% CO₂ for 3 h in assays with PBMC and 40 min in assays with PMN. Then the filters were removed, fixed in 2.5% glutaraldehyde (Merck, Damstadt, Germany), and stained with Giemsa (Sigma, Cleveland, USA) for 30 min. Cells that had migrated through to the bottom of the filter were counted in 6–10 high-power fields (HPF) ($\times 60$ or 100 objective). Chemotactic bioactivity was expressed as the mean number of cells per HPF. Variations in response to the tested agents, were dependent on the blood donor.

Assay for cytotoxicity: Cytotoxicity of TNF α 36–62 was tested using the fibrosarcoma cell line WEHI 164 clone 13, as described by Espevik *et al.*²¹ Cell viability in the assay was measured colorimetrically, by using tetrazolium salt (MTT), as described by Mosmann *et al.*²²

Results

Molecular modelling of TNF α 36–62 peptide: Starting from a minimized X-ray structure, the conformational properties of TNF α 36–62 in a vacuum environment were calculated using molecular dynamics. The results from the calculations suggested that the peptide would possess a partially conserved tertiary structure similar to the conformation in the minimized crystal structure of TNF α (Fig. 1). Parts of β -strands from each of the two β -sheets in the TNF α monomer are included in TNF α 36–62 (Figs. 1A and 1B). The β -strands in TNF α 36–62 were stabilized by hydrogen bonds, but some of these were different from the corresponding hydrogen bonds observed in the TNF α structure. A β -strand interruption in TNF α could also be recognized in the peptide. The two loops (38–41 and 50–54) located at the base of TNF α , and important residues surrounding a shallow depression which are suggested to be involved in receptor binding,¹⁶ were all exposed in a similar manner in the peptide as in TNF α . The distance between the carbon in the C-terminal carboxyl and the nitrogen in the N-terminal amino group in TNF α 36–62 was only 2.89 Å compared to 18.19 Å in TNF α , indicating an attraction between the oppositely charged C- and N-terminals. The effect of this electrostatic attraction is, however, expected to be of much less importance in aqueous environment.

Chemotactic effects of TNF α and TNF α 36–62 peptide on PMN: TNF α and the TNF α 36–62 peptide have been tested for the ability to attract PMN. The experiments were repeated at least three times, and similar results were obtained despite donor variations. Both TNF α and TNF α 36–62 showed a dose dependent chemotactic effect on PMN. Migration of approximately 120 cells was achieved with either 2 nM TNF α or 10 μ M TNF α 36–62 (Fig. 2). That a higher concen-

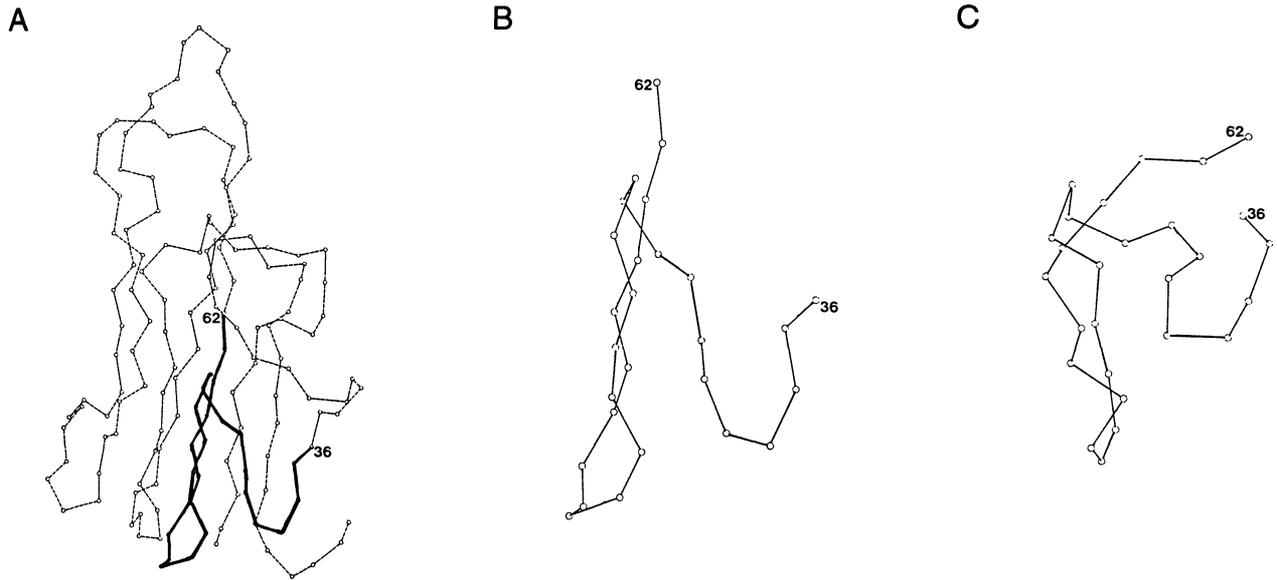


FIG. 1. Drawing of the TNF α monomer and the TNF α 36–62 peptide. (A) Drawing of the α -carbon backbone of the minimized crystal structure of TNF α . The sequence consisting of residues 36–62 are blackened. (B) Drawing of the α -carbon backbone of the TNF α sequence 36–62. (C) Drawing of the minimized structure of the TNF α 36–62 α -carbon backbone based on molecular modelling, using Computer Graphics.

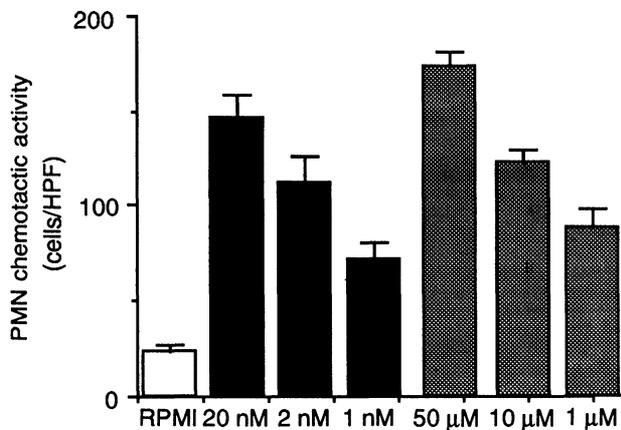


FIG. 2. Dose dependent effect of TNF α and TNF α 36–62 on PMN migration. Indicated concentrations of TNF α (■), and TNF α 36–62 (▨) were tested for chemotactic effect on PMN with RPMI (□) as a control. Migrated cells were counted in high-power fields (HPF) ($\times 60$ objective). Results are presented as means \pm S.E.M. ($n = 9$).

Table 1. Effect of varying concentrations of TNF α 36–62 peptide on PMN migration.

TNF α 36–62 concentration in lower compartment	TNF α 36–62 concentration in upper compartment			
	0	1 μ M	10 μ M	50 μ M
1 μ M	91 \pm 11	63 \pm 13*	54 \pm 13	61 \pm 11
10 μ M	129 \pm 10	89 \pm 7	64 \pm 8*	57 \pm 14
50 μ M	197 \pm 9	161 \pm 16	111 \pm 8	51 \pm 9*

The indicated concentrations of TNF α 36–62 peptide or medium alone were added to the upper compartments of the chemotaxis chamber, to neutralize the chemotactic effect of TNF α 36–62 used in the lower compartments. Migrated PMN were counted in high-power fields (HPF) ($\times 60$ objective). The data represent the mean \pm S.E.M. ($n = 9$). *The number of migrating PMN with the peptide present at the same concentration in both compartments.

tration of TNF α 36–62 was needed to induce migration of the same number of cells as induced by TNF α , is probably due to lack of domains necessary for optimal binding of TNF α 36–62.¹² FMLP was used as a positive control at 10^{-7} M, and gave an 11-fold migration of PMN compared to the negative control RPMI. As for TNF α , a Zigmond–Hirsch checkerboard analysis confirmed that TNF α 36–62 displayed chemotactic, and not chemokinetic effects on PMN (Table 1). A different TNF α peptide including residues 78–96, used at the same concentrations as TNF α 36–62, did not exhibit any chemotactic effect on PMN. We also tested whether TNF α and TNF α 36–62 had chemotactic effects on PBMC, and observed that both stimulants induced chemotactic activities on PBMC in a dose dependent manner and at similar concentrations as for PMN (data not shown). Both stimulants with their buffers were tested and confirmed free of LPS using the Endospey assay.

Inhibition of TNF α and TNF α 36–62-induced chemotaxis on PMN: Both mAb, Utr-1 and Htr-7b, specific for p75 and p55 respectively, significantly inhibited the chemotactic response of TNF α on PMN (Fig. 3A), when used separately or in combination. Likewise, the chemotactic effect of TNF α 36–62 peptide was also significantly inhibited by Utr-1 or Htr-7b (Fig. 3B) or a combination of both mAb. These mAb alone, or in combination, did not induce chemotaxis on PMN. FMLP induced migration was not inhibited by these mAb alone or in combination. The experiments were repeated four times, and similar results were obtained despite donor variations. Antibodies against IL-8 or MCP-1 did not show any inhibitory effect on either the TNF α , TNF α 36–62 or

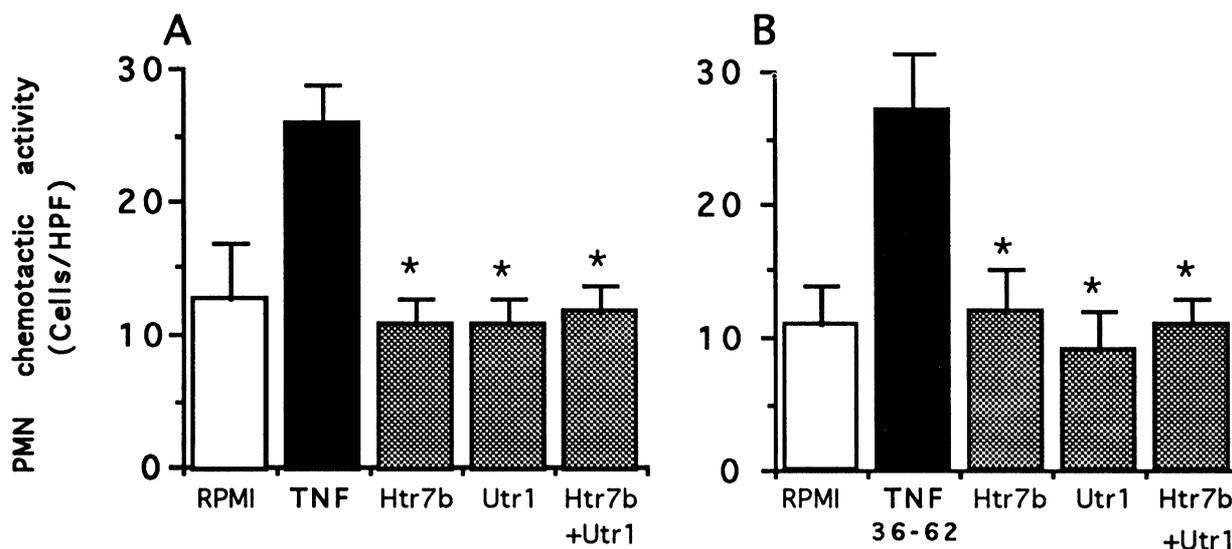


FIG. 3. Inhibitory effect of anti-p55 and anti-p75 antibodies on the chemotactic effect on PMN, induced by TNF α and TNF α 36-62. Utr-1 (10 mg/ml) and Htr-7b (10 mg/ml) (anti-p75 and anti-p55, respectively) were preincubated with cells for 10 min at 4°C. The PMN were then tested for chemotaxis toward (A) 0.6 nM TNF α and (B) 10 μ M TNF α 36-62 with (■) or without (□) antibodies (□:control). Migrated cells were counted in high-power fields (HPF) (\times 100 objective). The data represent the mean \pm S.E.M. (n = 8). * p < 0.001 compared to TNF α or TNF α 36-62 induced chemotactic effect without antibodies.

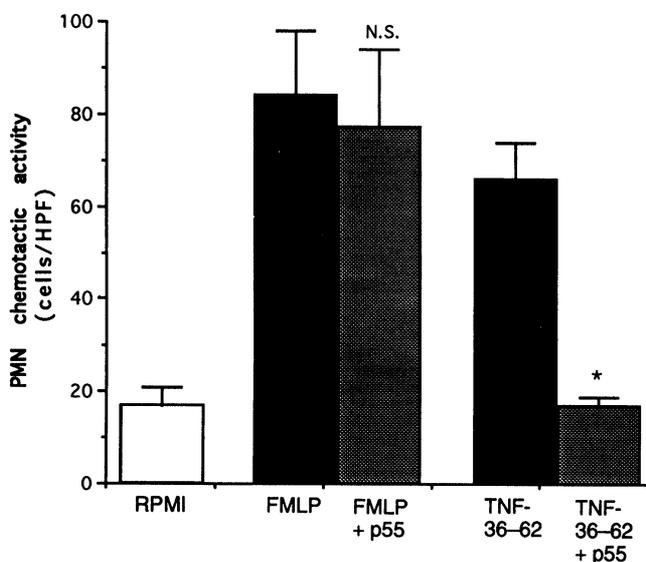


FIG. 4. Inhibitory effect of the soluble p55 receptor on PMN chemotaxis induced by TNF α 36-62. TNF α 36-62 or FMLP were tested for their ability to induce migration of PMN with (■) or without (□) soluble p55 (□: control). Soluble p55 was preincubated with TNF α 36-62 or FMLP for 10 min at 20°C. Migrated cells were counted in high-power fields (HPF) (\times 100 objective). The results are expressed as mean \pm S.E.M. (n = 6). * p < 0.001 compared with TNF α 36-62 alone. N.S. = not significant.

FMLP induced chemotaxis on PMN when used in the same concentrations as Utr-1 and Htr-7b.

TNF α 36-62 interacted with soluble p55: The soluble p55 was able to significantly inhibit the chemotactic response of the TNF α 36-62 peptide on PMN (Fig. 4). Soluble p55 did not inhibit the chemotactic effect of FMLP (Fig. 4), indicating a specific binding of TNF α 36-62.

TNF α 36-62 had no cytotoxic effect: TNF α 36-62 was tested for cytotoxic activity. In contrast to TNF α ,

TNF α 36-62 did not show any cytotoxic effect on the WEHI 164 clone 13 fibrosarcoma cell line, when tested up to a 10⁴-fold higher concentration than TNF α .

Discussion

In the present study it is shown that both TNF α 36-62 and TNF α is a chemoattractant to PMN *in vitro*. It is also shown that the chemotaxis induced by TNF α and TNF α 36-62 is mediated through both p55 and p75 TNF receptors. The finding regarding TNF α as a chemoattractant is in line with previous reports showing that antibodies against TNF α inhibit TNF α induced chemotaxis *in vitro* on PMN³ and monocytes,⁵ respectively. It is noticeable that there have been conflicting reports about the chemotactic property of TNF α .^{2-5,23,24} In the authors' opinion the discrepancy might be explained by variations in experimental conditions, donor variations and sensitivities of the test systems. TNF α has also been shown to stimulate production of the chemoattractant IL-8 by granulocytes and monocytes.²⁵ Since interleukin-8 (IL-8) shows chemotactic activity for PMN,^{20,26} TNF α might induce release of IL-8 by PMN, which in turn could be responsible for the chemotactic effect observed in our experiment. However, the present work shows that anti-IL-8 and anti-MCP-1 antibodies did not inhibit the chemotactic response of TNF α . This excludes IL-8 and MCP-1 as responsible for the chemotactic activity observed.

It is shown that the chemotactic activity TNF α on PMN involves both p55 and p75 receptors. Even non-redundancy in the function of the two receptors for

some effects has been observed,²⁷ a simultaneous involvement of both p55 and p75 receptors has been reported for a series of TNF α activities, such as induction of differentiation of ML-1 cells, NF- κ B activation, cytotoxicity on U937 cells and IL-6 production by endothelial cells.^{27–29} Tartaglia *et al.*³⁰ have suggested that the high affinity p75 receptor may regulate the rate of TNF α association with the p55 receptor, by increasing the local concentration of TNF α through rapid ligand association and dissociation. In contrast to this, Brouckaert *et al.*³¹ have proposed an alternative cooperation between the two receptors where p75 interferes with the p55 signalling pathway. This hypothesis is based on the fact that p75 triggering is not sufficient to initiate the redundant signals and that p75 triggering can diminish p55 mediated c-fos induction. However, it remains to be explored how the two receptors cooperate in TNF α induced chemotaxis.

Similar to TNF α , the TNF α 36–62 peptide also induced chemotaxis on PMN and PBMC. The finding that soluble p55 inhibited this effect (Fig. 3), indicates that the peptide possesses the conformation needed for interaction with the receptor. This supports the results of the molecular dynamic calculations (Fig. 1). That TNF α 36–62 is able to interact with soluble p55, is also in line with the recent work by Ratjen *et al.*¹³ who found that several bioactive TNF α peptides were able to inhibit binding of TNF α to the TNF receptors. Antibodies against p55 and p75 inhibited TNF α 36–62 induced chemotaxis on PMN, which indicates that this peptide, like TNF α , induces the chemotactic activity through both TNF receptors. It is noticeable that Postlewaite *et al.*¹² have observed chemotaxis on fibroblasts, by using another TNF α peptide, which also included the sequence 36–62. Although these authors did not show that the chemotaxis induced by the peptide was TNF receptor mediated, desensitisation studies suggested involvement of TNF receptors. The present work supports and extends this hypothesis.

TNF α 36–62 was not cytotoxic, suggesting the existence of distinct TNF α regions for the cytotoxic and chemotactic effects. It is noteworthy that two other TNF α peptides, which overlap with only four residues (54–58), were shown to be cytotoxic,¹³ suggesting a critical domain for TNF α cytotoxicity. This sequence is included in our TNF α 36–62 peptide and the molecular calculations of the peptide suggested that this specific domain encompassing residues 54–58 did not possess the conformation needed for optimal interaction with the TNF receptors. This was due to the attraction between the oppositely charged C- and N-terminals (Fig. 1).

Crystallographic studies on the TNF β /TNF receptor complex have shown that three TNF β monomers bind three TNF receptors in a symmetrical fashion.³² It has been suggested that a crosslinking of TNF

receptor is also necessary for signal transduction leading to TNF α effects,^{33–35} and is based on studies with antibodies and TNF α mutants against TNF receptors.^{33–36} Our and other investigations with synthetic TNF α peptide fragments,^{12–14} however, suggest that at least some of the TNF α effects are not dependent on crossreaction of TNF receptors with three TNF α monomers.

In conclusion, different TNF α peptides may induce distinct activities, indicating that TNF α possess distinct domains critical for different TNF α activities. This property opens the possibility of designing TNF α fragments with specific TNF α effects. We are currently investigating this hypothesis by studying TNF α 36–62 and several other TNF peptides for their bioactivities and specificities to target cells.

References

- Yoshimura T, Leonard EJ. Human monocyte chemoattractant protein-1 (MCP-1). In: Westwick J, Lindley IJD, Kunkel SL, eds. *Advances in Experimental Medicine and Biology. Chemotactic cytokines*. New York: Plenum Press. 1991; **305**: 47–53.
- Ming WJ, Bersani L, Mantovani A. Tumor necrosis factor is chemotactic for monocytes and polymorphonuclear leukocytes. *J Immunol* 1987; **138**: 1469–1474.
- Newman I, Wilkinson PC. Chemotactic activity of lymphotoxin and tumour necrosis factor alpha for human neutrophils. *Immunology* 1989; **66**: 318–320.
- Figari I, Mori NA, Palladino MA. Regulation of neutrophil migration and superoxide production by recombinant tumor necrosis factor- α and - β : comparison to recombinant interferon- γ and interleukin-1 α . *Blood* 1987; **70**: 979–984.
- Wang MJ, Walter S, Mantovani A. Reevaluation of the chemotactic activity of tumor necrosis factor for monocytes. *Immunology* 1990; **71**: 364–367.
- Zhang Y, Ramos BF, Jakchik BA. Neutrophil recruitment by tumor necrosis factor from mast cells in immune complex peritonitis. *Science* 1992; **258**: 1957–1959.
- Issekutz AC, Issekutz TB. Quantitation and kinetics of blood monocyte migration to acute inflammatory reactions, and IL-1 α , tumor necrosis factor- α and IFN- γ . *J Immunol* 1993; **151**: 2105–2115.
- Goeddel DV, Aggarwall BB, Gray PW, *et al.* Tumor necrosis factors: gene structure and biological activities. *Cold Spring Harbor Symp Quant Biol* 1986; **51**: 597–609.
- Beutler B, Cerami A. Tumor necrosis, cachexia, shock, and inflammation: A common mediator. *Ann Rev Biochem* 1988; **57**: 505–518.
- Vilcek J, Lee TH. Tumor necrosis factor. New insights into the molecular mechanisms of its multiple actions. *J Biol Chem* 1991; **266**: 7313–7316.
- Halvorsen H, Olsen JO, Østerud B. Granulocytes enhance LPS-induced tissue factor activity in monocytes via an interaction with platelets. *J Leukoc Biol* 1993; **54**: 275–282.
- Postlewaite AE, Seyer JM. Stimulation of fibroblast chemotaxis by human recombinant tumor necrosis factor α (TNF- α) an a synthetic TNF- α peptide. *J Exp Med* 1990; **172**: 1749–1756.
- Rathjen DA, Ferrante A, Aston R. Differential effects of small tumour necrosis factor peptides on tumour cell cytotoxicity, neutrophil activation and endothelial cell procoagulant activity. *Immunology* 1993; **80**: 293–299.
- Kapas L, Hong L, Cady AB, *et al.* Somnogenic, pyrogenic, and anorectic activities of tumor necrosis factor- α and TNF- α fragments. *Am J Physiol* 1992; **263**: R708–R715.
- Glen RC. Recent progress in computational chemistry and molecular graphics applied to drug design. *Drug News and Perspectives* 1990; **3**: 332–336.
- Eck JE, Sprang RS. The structure of tumour necrosis factor- α at 2.6 Å resolution. *J Biol Chem* 1989; **264**: 17595–17605.
- Bartnes K, Rekdal Ø, Briand JP, Hannestad K. Th1 clones that suppress IgG2a^b specifically recognize an allopeptide determinant comprising residues 435–451 of γ 2a^b. *Eur J Immunol* 1993; **23**: 2655–2660.
- Brockhaus M, Schoenfeld HJ, Schlaeger EJ, Hunziker W, Lesslauer W, Loetscher H. Identification of two types of tumor necrosis factor receptors on human cell lines by monoclonal antibodies. *Proc Natl Acad Sci USA* 1990; **87**: 3127–3131.
- Bøyum A. Isolation of mononuclear cells and granulocytes from human blood. *Scand J Clin Lab Invest* 1968; **21** (suppl): 77–89.
- Helset E, Sildnes T, Konopski ZS. Endothelin stimulates monocytes *in vitro* to release chemotactic activity identified as interleukin-8 and monocyte chemoattractant protein-1. *Mediators of Inflammation* 1994; **3**: 155–160.
- Espvik T, Nissen-Meyer J. A highly sensitive cell line, WEHI-164 clone 13, for measuring cytotoxic factor/tumor necrosis factor from human monocytes. *J Immunol Methods* 1986; **95**: 99–105.
- Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL. Two types of murine helper T cell clone I. Definition according to profiles of lymphokine activation and secreted proteins. *J Immunol* 1986; **137**: 2348–2357.
- Kownatzki E, Kapp A, Uhrich S. Modulation of human neutrophilic granulocyte functions by recombinant human tumor necrosis factor and recombinant human

- lymphotoxin. Promotion of adherence, inhibition of chemotactic migration and superoxide anion release from adherent cells. *Clin Exp Immunol* 1988; **74**: 143–148.
24. Mrowietz U, Schröder JM, Christophers E. Recombinant human tumor necrosis factor- α lacks chemotactic activity for human peripheral blood neutrophils and monocytes. *Biochem Biophys Res Commun* 1988; **153**: 1223–1228.
25. Van Damme J. Granulocyte and monocyte chemotactic factors: stimuli and producer cells. In: Westwick J, Lindley IJD, Kunkel SL, eds. *Advances in Experimental Medicine and Biology. Chemotactic cytokines*. New York: Plenum Press. 1991; **305**: 1–9.
26. Baglioni M, Walz AO, Kunkel SL. Neutrophil-activating peptide/interleukin 8, a novel cytokine that activates neutrophils. *J Clin Invest* 1989; **84**: 1045–1049.
27. Tartaglia LA, Goeddel DW. Two TNF receptors. *Immunology Today* 1992; **13**: 151–153.
28. Takeda K, Iwamoto S, Takeda M. Roles of two tumor necrosis factor receptors in induction of differentiation of ML-1 cells. *Anticancer Research* 1993; **13**: 883–886.
29. Shalaby MR, Sundan A, Loetscher H, Brockhaus M, Lesslauer W, Espevik T. Binding and regulation of cellular functions by monoclonal antibodies against human tumor necrosis factor receptors. *J Exp Med* 1990; **172**: 1517–1520.
30. Tartaglia LA, Pennica D, Goeddel DV. Ligand passing: The 75-kDa tumor necrosis factor (TNF) receptor recruits TNF for signalling by the 55-kDa TNF receptor. *J Biol Chem* 1993; **268**: 18542–18532.
31. Brouckaert P, Libert C, Everaerd B, Takahashi N, Cauwles A, Fiers W. Tumor necrosis factor, its receptors and the connection with interleukin 1 and interleukin 6. *Immunobiol* 1993; **187**: 317–329.
32. Banner DW, D'Arcy A, Janes W, et al. Crystal structure of the soluble human 55 kDa TNF receptor–human TNF- β complex: Implication for TNF receptor activation. *Cell* 1993; **73**: 431–445.
33. Zhang XM, Weber I, Chen MJ. Site-directed mutational analysis of human tumor necrosis factor- α receptor binding site and structure–functional relationship. *J Biol Immunol* 1992; **267**: 24069–24075.
34. Engelmann H, Holtmann H, Brakebusch C, et al. Antibodies to a soluble form of a tumor necrosis factor (TNF) receptor have TNF-like activity. *J Biol Chem* 1990; **265**: 14497–14504.
35. Ostade XV, Tavernier J, Prange T, Fiers W. Localization of the active site of human tumor necrosis factor by mutational analysis. *The EMBO Journal* 1991; **10**: 827–836.
36. Espevik T, Brockhaus M, Loetscher H, Nonstad U, Shalaby R. Characterization of binding and biological effects of monoclonal antibodies against a human tumor necrosis factor receptor. *J Exp Med* 1990; **171**: 415–426.

ACKNOWLEDGEMENTS. We thank Eli Berg and Liv Tone Eliassen for excellent technical assistance. The contribution of Arnfinn Kvarnsnes in performing the FAB-MS analysis of the synthetic peptides is gratefully acknowledged. The cost of the MS instrument is partially paid by the Norwegian Council for Science and the Humanities (NAVF), and the Norwegian Council for Agriculture Research (NLVF).

Received 29 March 1994;
accepted in revised form 25 May 1994



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

