

MACROPHAGES harvested from the peritoneal cavities of rats release a neutrophil chemotactic factor (MNCF) in response to stimulation with Gram-negative bacterial lipopolysaccharide (LPS). MNCF has been shown to be active in rats treated with dexamethasone, a glucocorticoid that usually inhibits the neutrophil migration induced in this species by interleukin (IL)-1, tumour necrosis factor alpha (TNF α), IL-8, C5a and leukotriene B $_4$ (LTB $_4$). Here we report that macrophages harvested from peritoneal cavities of mice, and stimulated *in vitro* with LPS, also release a factor that induces neutrophil migration in dexamethasone-treated animals. This chemotactic activity was neutralized by the incubation of the LPS-stimulated macrophage supernatants with a purified polyclonal IgG anti-mouse TNF α . In addition, significant amounts of TNF were detected in the supernatants. The neutrophil migration induced by intraperitoneal administration of recombinant murine TNF α was also unaffected by pretreatment of the mice with dexamethasone. Moreover, neutrophil migration induced by intraperitoneal injection of LPS was completely blocked by pretreatment of the mice with a monoclonal antibody against murine TNF α . In conclusion, our results support the hypothesis that, in contrast to the role of TNF in rats (where it indirectly induces neutrophil migration), in mice, it may be an important mediator in the recruitment of neutrophils to inflammatory sites.

Key words: Glucocorticoids, Neutrophil migration, Tumour necrosis factor alpha

The neutrophil migration induced by tumour necrosis factor alpha in mice is unaffected by glucocorticoids

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Introduction

Neutrophil migration is a key event in acute inflammatory reactions. This is a complex and multimediaged event and remains not completely understood. Chemotactic mediators are generally derived either from plasma components or from multiple cellular sources,^{1,2} and one of the most important cellular sources of neutrophil chemotactic factors is resident macrophages.^{1,3} In fact, it has been shown that neutrophil migration into the peritoneal cavity, induced by carrageenin, endotoxin, latex beads or zymozan, is increased when the number of resident macrophages is augmented by pretreatment with thioglycollate, and reduced after depletion of the peritoneal macrophage population through previous washing of the cavity.⁴

The chemotactic mediators thought to be involved in neutrophil migration into inflamed

tissues include the fifth complement component fragment, C5a,⁵ leukotriene B $_4$ (LTB $_4$),⁶ and the pleiotropic cytokines interleukin (IL)-1,⁷ tumour necrosis factor alpha (TNF α),⁷ and IL-8.⁸ Some of these substances, such as C5a, may have a direct chemotactic effect on neutrophils;⁵ others may act indirectly through mechanisms which depend upon the resident cells of the tissues, and the endothelium.^{7,8} IL-1, TNF and LTB $_4$, for example, not only increase the expression of adhesion molecules but also stimulate the release of chemotactic factors by rat resident macrophages.⁷ Furthermore, neutrophil migration into the peritoneal cavities of rats induced by these substances is also dependent on the number of resident macrophages.⁷ Another potent leukocyte chemoattractant cytokine is IL-8, which has been shown to induce neutrophil migration by means of a mast cell dependent mechanism, since its activity cannot

be observed in rat peritoneal cavities depleted of mast cells or in mice lacking this cellular population.⁸ Furthermore, rat mast cells stimulated with IL-8 release a neutrophil chemotactic activity into the supernatant.⁸

It has been demonstrated that macrophages incubated *in vitro* with either LPS³ or IL-1 and TNF α ⁷ release a neutrophil chemotactic factor named MNCF (macrophage-derived neutrophil chemotactic factor) into the supernatant. This factor has a mol.wt of 54 kDa, a PI < 4.0 and lectin-like activity.⁹ The neutrophil migration induced by MNCF in rats is unaffected by the variation of the number of resident peritoneal cells, as well as by pretreatment of animals with glucocorticoids. Based upon these results, it was suggested that MNCF may be a mediator involved in the recruitment of neutrophil induced by various inflammatory stimuli, such as the cytokines IL-1 and TNF α in rats.

The fact that the glucocorticoids did not affect MNCF-induced neutrophil migration conflicts with the hypothesis that glucocorticoids block the expression of adhesion molecules in the neutrophil and endothelium.¹⁰ It thus seems reasonable to speculate that certain stimuli, such as MNCF, can bypass one or more critical steps which are sensitive to glucocorticoids, inducing leukocyte migration, whereas other stimuli, even though capable of bypassing some of these steps, cannot cause *in vivo* neutrophil migration in glucocorticoid-treated animals. Thus, it has been shown that in the hamster chick-pouch model of inflammation, dexamethasone failed to block the LTB₄-induced neutrophil adherence to and migration through endothelial cells.¹¹ In this model the neutrophils were halted in the lamina basal, between the endothelial cells and the pericyte, never reaching the interstitial space.

The present study shows that LPS-stimulated mouse macrophage monolayers release an MNCF-like mediator into culture supernatants since these supernatants induced neutrophil migration into the peritoneal cavities of animals pretreated with dexamethasone. Not only were significant amounts of TNF α detected in the supernatants, but their *in vivo* neutrophil chemotactic activity was also neutralized by preincubation with a purified polyclonal IgG anti-mouse TNF α . Furthermore, the neutrophil recruitment induced by the intraperitoneal administration of recombinant murine TNF α was not affected by the treatment of the mice with dexamethasone. A monoclonal antibody against TNF was also shown to be able to block the LPS-induced neutrophil migration to the peritoneal cavities of mice completely.

Materials and Methods

Reagents

LPS isolated from *Escherichia coli* O111B4 was purchased from Sigma (St Louis, MO). Recombinant murine TNF α (rmTNF α) was donated by Dr M. A. Palladino (Genentech Inc., South San Francisco, CA). The rat monoclonal antibody against rmTNF α (XT22.11) was purified from mice ascitic peritoneal fluids by affinity chromatography on Sepharose-protein G (Sigma).

Animals

Male 8- to 10-week-old C57BL/6 mice (Centro Multinstitucional de Bioterismo-CEMIB/UNICAMP, Campinas, SP, Brazil) were housed in a temperature-controlled room and received water and food *ad libitum* throughout the period of study.

Preparation of anti-TNF sera

Polyclonal rabbit antisera against recombinant murine TNF α were produced in New Zealand rabbits, as described elsewhere.¹² Briefly, on day 0, 100 μ g of rmTNF α were diluted in 500 μ l of a phosphate buffered saline solution (PBS) and mixed with an equal volume of Freund's complete adjuvant, and the emulsion was injected intradermically into the rabbits. Boosting with intradermal injections of emulsions containing 50, 10, and 10 μ g of rmTNF α in 50% Freund's incomplete adjuvant, occurred on days 15, 30 and 45, respectively. The rabbits were bled 10 days after the final injection. Specific antibodies against TNF present in the sera were detected by immunoblotting and capacity to block the effect of TNF on WEHI cells. In some experiments, antibodies obtained by chromatography of the sera in DEAE-Sepharose, according to the manufacturer's recommendations, were used as an IgG fraction.

LPS stimulated murine macrophage supernatants

Mouse peritoneal cavities were stimulated by the injection of a 3% thioglycollate solution (DIFCO Laboratories; Detroit, MI) (3 ml/animal). Four days later, the peritoneal cells were harvested with RPMI 1640 (Flow Laboratories, McLean, VA) and seeded on plastic tissue culture dishes at a density of 2×10^6 cells/5 ml/plate. The cells were allowed to adhere for 1 h at 37°C, in an atmosphere of air with 5% CO₂. Adherent cells (95% macrophages) were washed

three times with PBS (pH 7.2), and incubated for 30 min with 5 mg/ml of LPS in RPMI or with RPMI alone. The supernatants were discarded and the cells were extensively washed with PBS, followed by a final 2 h incubation at 37°C with 5 ml of RPMI medium per dish (without LPS or fetal calf serum). The cell-free incubation fluids were subsequently ultradialyzed through a YM 10 membrane with a cut-off 10 Kd (Amicon Corp., Lexington, MA, USA). The final volumes (corresponding to 5% of the original volume) were filtered on 0.22 µm membranes (Millipore; Bedford, MA) and the samples were kept at -70°C for TNF detection and neutrophil migration assays.

Detection of TNF in the supernatants

The TNF content in macrophage supernatants was measured using a highly TNF-sensitive cell line, WEHI 164 clone 13, as described elsewhere.¹³ Briefly, WEHI cells were seeded on microplates (Costar 3596, Cambridge, MA) at a density of 4×10^4 cells/well in 50 µl of growth medium (RPMI 1640 medium containing 10% fetal calf serum (Gibco)). Fifty µl of diluted supernatants were added to the cells, and the experiments were conducted in triplicate. The plates were incubated for 20 h at 37°C in a 5% CO₂ incubator; 10 µl of a 3,4,5-dimethylthiazol-2,4,5-diphenylformazan solution (5 mg/ml PBS) (MIT, Sigma) were added to each well and the plates incubated for an additional 4 h. One hundred µl of isopropanol containing 0.04 N HCl were added to each well. Fifteen min later, the degree of cell lysis was determined spectrophotometrically (570 nm) using an enzyme-linked immunoassay analyser (Multiskan MCC/340 MKII, Flow Laboratories). TNF content in the assayed materials was calculated by comparison with standard curves which were determined with rmTNF (Genentech Inc.). The results correspond to the averages obtained in at least three independent experiments.

The presence of the TNF in the supernatants was confirmed by ELISA assays using the monoclonal antibody against TNF (XT 22.11) as the coating antibody (10 µg/ml) and the rabbit polyclonal antibodies (serum dilution 1:200) as the detecting antibody. The bound rabbit antibodies were revealed using an immunoperoxidase conjugate anti-rabbit IgG, and the chromogenic substrat OPD (Sigma). In our system, it was possible to detect amounts of TNF varying between 50 to 0.25 ng/ml, according to standard curves performed with rmTNFα.

In vivo neutrophil migration

The method for testing *in vivo* neutrophil migration has been described elsewhere.³ Briefly, LPS solutions (8–1000 ng/0.2 ml of saline solution/animal), rmTNFα (10–100 µg/0.2 ml/animal) and either LPS-stimulated or non-stimulated macrophage supernatants (equivalent to the products released by 2×10^6 adherent cells/0.2 ml/animal) were injected into the peritoneal cavities of mice pretreated subcutaneously (s.c.) with dexamethasone (0.5 mg/kg) 1 h previously. Controls received intraperitoneal (i.p.) injections of sterile saline solution. Six hours after these i.p. injections, the mice were sacrificed in a CO₂ chamber, and their peritoneal cavities washed with 3 ml of PBS containing 5 IU/ml of heparin; total and differential cell counts in the washing fluid were conducted as described elsewhere.¹⁴ The results are expressed as means \pm standard error of the mean (SEM) for the number of neutrophils per ml of peritoneal wash.

In some experiments, rmTNFα was incubated for 30 min at 37°C with polyclonal IgG anti-TNF (100 µg IgG/100 ng TNF) prior to the i.p. injections. The control groups for these experiments received i.p. injections of the same quantity of the antibody diluted in saline solution.

In other experiments, monoclonal antibody against TNF (XT22.11) was administrated intra-peritoneally to mice (100 µg/cavity) 1 h before the i.p. injections of saline solution (0.2 ml/cavity) or LPS (200 ng/cavity), with control group receiving i.p. injections of saline solution prior to the inflammatory stimulus.

Protein measurement

The protein contents of purified materials was determined according to the method of Hartree.¹⁵

Statistical analysis

The data are presented as the means \pm SEM, with statistical comparisons between the groups performed using the Student *t*-test for unpaired samples.

Results

Time course and dose dependence of LPS-induced neutrophil migration in mice

The intraperitoneal injection of increasing doses of LPS in naive mice induced a dose-dependent neutrophil migration with a bell-shaped curve

(Fig. 1A). The maximal migration was observed with a dose of 200 ng LPS/cavity, peaking between 8 and 12 h after the LPS administration, and returning to control levels after 48 h (Fig. 1B); the neutrophil migration induced by the dose of 1000 ng/cavity was less than that observed with the other doses tested.

Effects of dexamethasone

pretreatment of mice and pre-incubation of the LPS-stimulated macrophage supernatants with TNF specific antibodies on capacity to induce neutrophil migration

The supernatants from LPS-stimulated macrophages induced a significant neutrophil migration into the peritoneal cavities of naive mice, which was not significantly inhibited by the pretreatment of the animals with dexamethasone (Fig. 2A). The dexamethasone treatment, however, reduced the neutrophil migration induced by saline solution. On the other hand, the pre-incubation of these supernatants (from 2×10^6 adherent cells) with the IgG fraction of anti-rmTNF α serum (100 μ g) eliminated their capacity to induce neutrophil migration into peritoneal cavities of dexamethasone-pretreated mice (Fig. 2B). The same quantity of IgG did not interfere with neutrophil migration in the control group.

TNF concentration in the LPS-stimulated macrophage supernatants

The TNF concentration for five different samples of LPS-stimulated macrophage super-

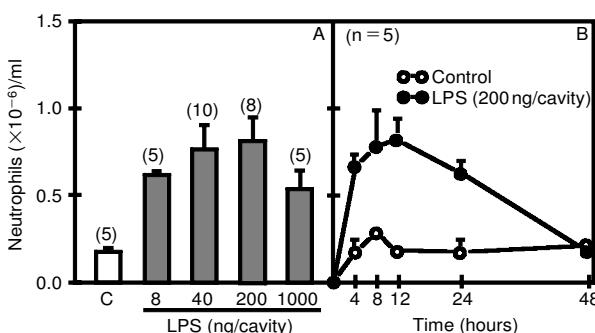


FIG. 1. Dose response and time course of neutrophil migration induced by LPS. (A) Bars represent neutrophil migration induced by the indicated doses of LPS into peritoneal cavities of mice. The control (C) represents neutrophil migration induced by saline solution. The neutrophil migration was analysed 6 h after the injection of the stimuli. The number of animals per experimental group is indicated at the top of the bars. (B) Time course of neutrophil migration induced by either LPS (200 ng/cavity) or saline solution (control group). The number of animals per group was five. In both panels, the results are reported as mean \pm SEM.

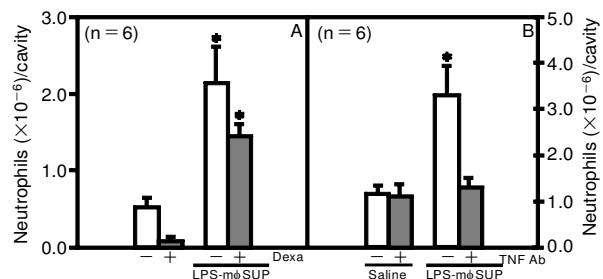


FIG. 2. Effects of dexamethasone pretreatment and preincubation of LPS-stimulated mouse macrophage supernatants with specific antibody to rmTNF α on capacity to induce neutrophil migration. (A) Bars represent neutrophil migration induced by saline solution or LPS-stimulated mouse macrophage supernatants (LPS-M ϕ SUP) into peritoneal cavities of mice, pretreated (+, hatched bars) or not (–, open bars) with dexamethasone (Dexa). (B) Bars represent neutrophil migration induced by LPS-M ϕ SUP, incubated (+) or not (–) with 100 μ g of IgG anti-rmTNF α , into peritoneal cavities of dexamethasone-pretreated mice. Control groups received i.p. injections of saline solution or saline solution containing IgG anti-TNF α (100 μ g/0.2 ml/animal). In both panels, the results are reported as mean \pm SEM for six animals in the group. *P < 0.05 (Student's t-test).

natants, derived from the culture of 2×10^5 adherent cells, varied from 1 to 14 ng as detected by activity and/or ELISA assays.

Effect of dexamethasone pretreatment on neutrophil migration induced by recombinant murine TNF α in mice

The intraperitoneal administration of rmTNF α in naive mice induced a dose-dependent neutrophil migration which peaked 5–10 h after the injection. In dexamethasone-pretreated mice, rmTNF α also induced a dose-dependent neutrophil migration, which was determined 6 h later; the migration induced by a dose of 0.1 μ g TNF/cavity in naive mice was similar to that observed for dexamethasone-pretreated animals (Fig. 3A). Fig. 3B shows that the neutrophil chemotactic activity of the rmTNF α was completely eliminated by pre-incubation with an anti-TNF α IgG.

Effect of treatment with dexamethasone or TNF specific antibody on LPS-induced neutrophil migration in mice

The intraperitoneal injection of LPS in naive mice induced significant neutrophil migration which was suppressed by pretreatment with dexamethasone (Fig. 4A). This LPS-induced neutrophil migration was completely eliminated by pretreatment of the animals with a monoclonal antibody against TNF α (Fig. 4B).

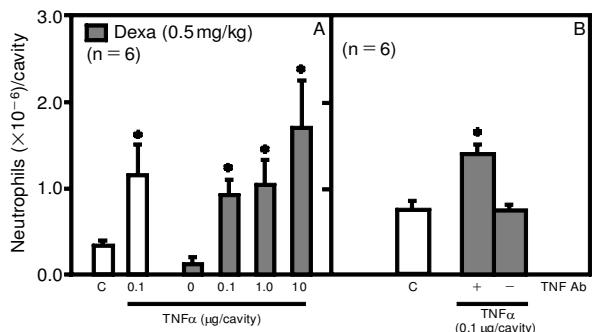


FIG. 3. Effect of dexamethasone pretreatment on the neutrophil migration induced by rmtTNF α in mice. (A) Bars represent neutrophil migration induced i.p. injection of saline solution (SAL) or of the indicated amounts (in $\mu\text{g}/\text{animal}$) of rmtTNF α into peritoneal cavities of dexamethasone-pretreated or naive mice (hatched and open bars, respectively). (B) Bars represent neutrophil migration induced by 0.1 μg rmtTNF α , preincubated (+) or not (-) with 100 μg of IgG anti-TNF α , into peritoneal cavities of dexamethasone-pretreated mice. The control animals (SAL) received i.p. injections of saline solution containing IgG anti-TNF α (100 $\mu\text{g}/0.2 \text{ ml}/\text{animal}$). In both panels, the results are reported as means \pm SEM for six animals in the group. *P < 0.05 (Student's t-test).

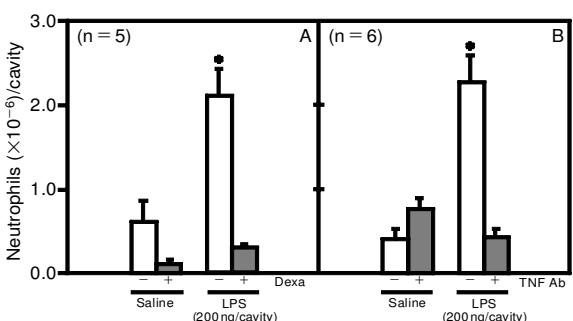


FIG. 4. Effects of pretreatment of mice with either dexamethasone or monoclonal antibody against murine TNF α on neutrophil migration induced by LPS. (A) Bars represent neutrophil migration induced by i.p. injection of saline solution (0.2 ml/cavity) or of LPS (200 ng/cavity) in dexamethasone-pretreated or non-treated mice (+, hatched and -, open bars, respectively). (B) Bars represent neutrophil migration induced by saline solution (0.2 ml/animal) or by LPS (200 ng/animal) into peritoneal cavities of mice either pretreated (+, hatched bars) or not (-, open bars) with monoclonal antibody against TNF α (100 $\mu\text{g}/\text{animal}$). In both panels, the results are reported as means \pm SEM for five animals in the group. *P < 0.05 (Student's t-test).

Discussion

The intraperitoneal injection of LPS in mice caused a significant, dose and time-dependent, neutrophil migration with a bell-shaped dose-response curve, as was found in other species.^{16,17} This may be due to the liposolubility of the LPS and the fact that when it is injected in high concentrations into extravascular tissue, the amount that enters the circulatory system is sufficient to block neutrophil migration. It has, for example, been demonstrated

that intravenous administration of LPS inhibits the neutrophil migration induced by various stimuli, including LPS itself.¹⁶ It has recently been demonstrated that macrophages harvested from the peritoneal cavities of rats and stimulated *in vitro* with LPS release MNCF, a neutrophil chemotactic factor.¹⁸ Rat macrophages stimulated *in vitro* with IL-1 and TNF also release MNCF.⁷ The neutrophil migration induced by such MNCF is unaffected by the number of resident cells or by the treatment of the rats with dexamethasone.¹⁹ The fact that this glucocorticoid did not affect the neutrophil migration induced by MNCF made the factor distinguishable from the other known neutrophil chemotactic mediators, since it has been demonstrated that the neutrophil migration induced by IL-1, TNF, IL-8, FMLP, C5a and LTB₄ in rats is inhibited by treatment with dexamethasone.^{7,8} It is thus suggested that in rats MNCF may be a late mediator in the recruitment of neutrophil induced by various stimuli, including the cytokines IL-1 and TNF.

In the present study we demonstrated that macrophages harvested from peritoneal cavities of mice and stimulated with LPS for a brief period of time also released a neutrophil chemotactic factor into the supernatants, and the activity of this factor was not significantly inhibited by pre-treatment of animals with dexamethasone, at least at doses that block the neutrophil migration induced by LPS.

The incubation of the supernatant with an antibody against murine TNF blocked the neutrophil chemotactic activity, thus suggesting that the presence of the TNF was responsible for this activity. This fact also suggested that the neutrophil chemotactic activity of the supernatant was due to the presence of the TNF. The presence of TNF in the supernatant was then confirmed, and dose-dependent neutrophil migration in dexamethasone-treated mice was induced by the intraperitoneal administration of recombinant murine TNF. This neutrophil migration was not statistically different from that observed in untreated mice. The antibody that blocked the neutrophil chemotactic activity of the LPS-stimulated macrophage supernatant also eliminated it for rmTNF. The fact that the quantity of mrTNF necessary to induce neutrophil migration was greater than that found in the supernatant might be explained by the lack of glycosylation in the recombinant form, since it is well established that the level of glycosylation affects the specific activity of this cytokine. These results are in contrast to those obtained in rats, where the neutrophil migration induced by TNF is blocked by pre-treatment with

dexamethasone; moreover, in this case TNF induces neutrophil migration via MNCF.⁷

In mice, the TNF generation upon LPS injection was suppressed by *in vivo* administration of a monoclonal antibody against TNF (XT22.11), which blocked the neutrophil migration induced by the endotoxin. It has recently been demonstrated that TNF is also involved in neutrophil migration into the lung induced by intratracheal challenge of presensitized mice with *Schistosoma mansoni* egg antigen.²⁰

It is well documented that glucocorticoids inhibit the release of several cytokines, including TNF.²¹⁻³⁰ This fact could explain the inhibitory effect of dexamethasone on the neutrophil migration induced by LPS. The blockage of neutrophil-endothelium adhesion has been suggested as a mechanism by which the glucocorticoids inhibit the migration, although *in vitro* experimental evidences does not support this interpretation.³¹ The *in vivo* monitoring of different stages of neutrophil migration showed that dexamethasone did not inhibit rolling or adhesion to the venular endothelium induced by LTB₄ and fMLP in hamsters.³² Moreover, glucocorticoid treatment did not inhibit neutrophil penetration through the endothelial cell junctions. It was thus concluded that, in addition to the effect of corticoids on the release of chemotactic mediators, they blocked the final step of migration because the neutrophils remained between the endothelium and the basal membrane without reaching the perivascular tissue.³² Haptotaxis seems to be an important mechanism in this final step, and the binding of the chemoattractant to a sub-endothelial or tissue matrix seems to be an important requirement for the process. In support of this hypothesis, *in vitro* evidence is provided to show that MNCF and IL-8 are able to induce migration through a haptotatic gradient, although they recognize different ligands.³³ Since dexamethasone blocks the neutrophil migration induced by IL-8 but does not affect the MNCF activity, it is postulated that glucocorticoids may be able to block the expression of the IL-8 ligand while not affecting that of MNCF.³³ Thus, TNF probably induces neutrophil migration in mice through a mechanism similar to that described for MNCF in rats, although the expression of the extravascular ligand to TNF in mice is not affected by glucocorticoid treatment. It is important to point out that, despite the fact that MNCF is not released in significant quantities by mouse macrophages, as demonstrated in the present study, when MNCF from rat macrophages was injected in dexamethasone pre-treated mice, the neutrophil migration observed

was similar to that observed in normal mice (data not shown).

In conclusion, the results reported here support the hypothesis that, in contrast to rats, in which TNF induces neutrophil migration through an indirect mechanism, in mice TNF may be a late mediator involved in the recruitment of neutrophil into an inflammatory site.

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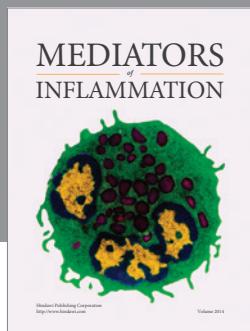
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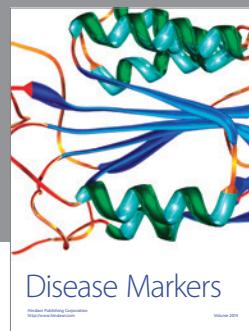
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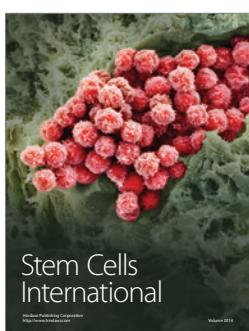
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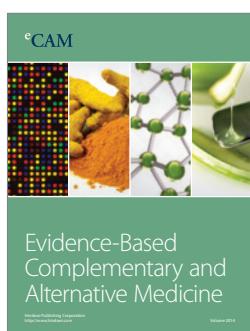
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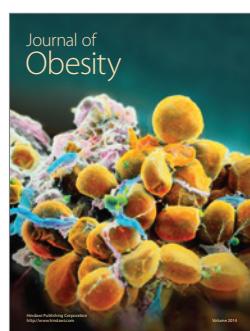
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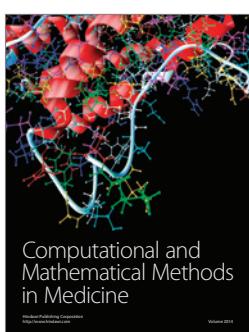
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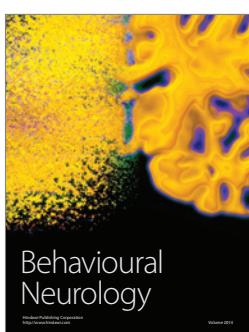
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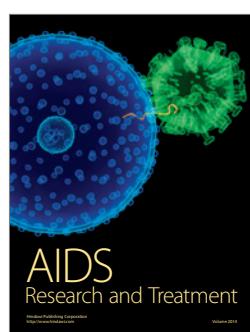
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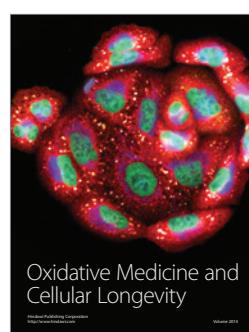
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