Th1 and Th2 cytokine profile in patients with early onset periodontitis and their healthy siblings

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Introduction

Early onset periodontitis (EOP) is a chronic inflammatory periodontal disease affecting individuals aged 17 to 25.¹ Pathological changes of the periodontal tissue are characterised by a mild resorption of alveolar bone and the presence of Actinobacillus actinomycetemcomitans (A. a.) in the gingival crevice area. Functional studies have demonstrated polymorphonuclear leukocytes (PMN) chemotactic defects in some 70% of subjects with EOP.

Periodontal disease is caused by bacteria in dental plaque, and evidence is accumulating that specific periodontal pathogens are associated with the progressive form of the disease.¹² However, some individuals harbour these specific bacteria but do not appear to show evidence of disease progression. Patient susceptibility is most likely genetically determined, although a number of local and environmental factors are thought to influence disease expression.³

Immunological mechanisms have been implicated in the pathogenesis of periodontal disease for over 30 years. In the 1970s, attention focused on the participation of peripheral blood T lymphocytes in response to the action of dental plaque bacteria and on their role in the development of inflammation.⁴ Since then many papers have been published that concern the relationship between peripheral blood mononuclear cells (PBMC) blastic transformation and periodontitis.⁵⁻⁷ The significance of the dental plaque bacteria on polyclonal B lymphocyte activation was really only studied later.⁷⁻⁸ Results showed that both specific and non-specific mechanisms of B lymphocyte activation can play a role in the pathogenesis of periodontal disease and that they can be highly influenced by the treatment.⁸⁻¹³ At the same time, immunological studies clearly established the B cell/plasma cell dominance of the periodontitis lesion.¹¹,¹²

Both specific and non-specific activation are regulated by T lymphocytes. Based on the concept of functional Th1/Th2 and Tc1/Tc2 subsets,¹⁴,¹⁵ a model for the pathogenesis of periodontal disease has been constructed.¹⁶,¹⁷ According to the hypothesis of Seymour et al.,¹⁶ patients with progressive disease activate mainly Th2 lymphocytes in response to an antigenic stimulus whereas patients with the slowly progressive disease activate Th1 lymphocytes. In the latter response it is proposed that a local expansion of the B cell population would result, on the one hand, in the production of IL-1 and hence subsequent

Early onset periodontitis (EOP) is a chronic inflammatory periodontal disease with a strong genetic link affecting individuals aged 17 to 25. In the familial studies we tested the hypothesis about the role of Th1 and Th2 cytokines in the pathogenesis of EOP disease. The study involved 6 individuals with EOP disease and their 6 siblings with healthy periodontium. Actinobacillus actinomycetemcomitans (A. a.), a bacterium typical for EOP, was detected in all people studied. Th1 and Th2 cytokine production was measured after in vitro stimulation. Peripheral blood mononuclear cells (PBMC) were isolated and cultivated for 24 h and 7 days with PWM, A. a. or Escherichia coli. The levels of IL–4, IFN-gamma, IgA, IgG and IgM were measured by ELISA methods. After in vitro stimulation of PBMC, a significantly higher production of IL–4 and significantly lower production of IFN-gamma were found in the group of patients compared with their healthy siblings. The increased level of IL–4 in patients was in good agreement with an increased level of IgM after stimulation of lymphocytes with E. coli. These results support Seymour's hypothesis according to which patients with progressive disease primarily activate Th2 lymphocytes while non-susceptible individuals activate Th1 lymphocytes.

Key words: Early onset periodontitis, Th1, Th2, Cytokine profile, Immunoglobulin, Peripheral blood lymphocytes, Familial study, Actinobacillus actinomycetemcomitans, Escherichia coli
tissue destruction and, on the other hand, in the production of specific antibodies. It is further proposed that if protective antibody is produced it could result in the elimination of the organism and disease progression would cease. Production of antibody that is non-protective would, however, result in continuous connective tissue breakdown.

Our present study concerns the effect of short-term cultivation of peripheral blood lymphocytes with pokeweed mitogen (PWM), \textit{A. a.} and \textit{Escherichia coli} on the formation of IL–4 and IFN-gamma in patients with early onset periodontitis and their healthy siblings. \textit{A. a.} has been suggested to play a pathogenic role in the development of this disease.\textsuperscript{1} In our previous studies we have shown that polyclonal activation of B cells with a suspension of \textit{E. coli} leads to high production of immunoglobulins by PBMC of patients with periodontitis.\textsuperscript{18} The results should contribute to finding the origins of the susceptibility to early onset periodontitis.

Materials and methods

Two years before beginning this study the group of 47 patients with EOP was collected from a set of 9225 young (16–19 years) individuals at the Institute of Dental Research. These patients met the basic clinical diagnosis criterion of periodontitis: they had at least one periodontal pocket deeper than 3 mm, situated in the region of the molars or incisors, inflammation of gingiva and the presence of the suspect pathogen—the bacterium \textit{A. a.} in the sulcular region. There were no individuals with systemic disease or pregnant women in the group of patients.

During the familial study\textsuperscript{19} we found that among the total of 47 patients and their 54 siblings only 6 patients had siblings with healthy periodontium and presence of \textit{A. a.} in sulcular region. These families were included in this study.

In agreement with Helsinki conventions (1964), informed consent for blood collection for immunological examination was obtained from all persons examined.

Clinical examination

The examination of patients at the beginning of therapy included the determination of gingival inflammation using the gingival index (GI),\textsuperscript{20} ascertaining the depth of the pockets with the calibrated probe and X-ray. This was followed immediately by conservative treatment. The dental calculus was removed, unsatisfactory fillings or crowns were restored and correct tooth brushing technique was demonstrated. All patients were thoroughly instructed in oral hygiene including teeth cleaning and gingival massage. They were also warned that oral hygiene is the decisive factor in periodontitis therapy. The conservative part of the therapy lasted 4 weeks and was followed by a 10-day tetracycline treatment. Two weeks after the antibiotic therapy the patients were invited for a control clinical examination, with further check-ups at 3 and then 6-month intervals.

This study reports on the results of an immunological examination 2 years after the start of the therapy.

Bacterial suspension and mitogens

\textit{A. a.} was isolated from a subgingival locality from the patients of the experimental group. Bacterial culture was incubated for 48 h in Brain Heart Infusion (Oxoid) medium and then inactivated for 20 min at 120°C. The bacterial suspension was washed 4 times with physiological saline, centrifuged at 600\textsuperscript{g} and adjusted to the required concentration (10\textsuperscript{9} cells/ml) with RPMI 1640 tissue culture medium.

The suspension of heat-killed bacteria of the \textit{E. coli} 086 strain was obtained from the Department of Immunology, Institute of Microbiology, Academy of Sciences of the Czech Republic, and diluted to the experimentally optimal concentration (10\textsuperscript{9} cells/ml) with RPMI medium.

PWM (\textit{Phytolacca americana} mitogen, Sigma) and ConA (Concanavalin A, Sigma) were diluted to the experimentally optimal concentration (PWM 2 \textmu g/ml, PWM 2 \textmu g/ml + ConA 10 \textmu g/ml) with RPMI 1640 tissue culture medium.

Stimulation of lymphocytes to cytokine and immunoglobulin production

Mononuclear cells isolated from the peripheral blood of the patients and their siblings with healthy periodontium were washed, diluted with supplemented tissue culture medium RPMI 1640 to a concentration of 10\textsuperscript{6} cells/ml and cultivated with mitogens and heat-killed bacteria (\textit{A. a.} 10\textsuperscript{8} /ml and \textit{E. coli} 10\textsuperscript{8} /ml), as described earlier.\textsuperscript{21} Supernatants were taken after 24 h and after 7 days, which were then stored at –20°C until further assay.

Assay of cytokines and immunoglobulins in culture media

Cytokine assay.

Cytokines IL–4 and IFN-gamma were assayed in the tissue culture supernatants using commercial Immunotech France kits. The technique is based on the sandwich method in which monoclonal antibody is used as a binding antibody, and another biotinylated monoclonal antibody is used for detection. This second antibody reacts with the streptavidin-peroxidase conjugate. The binding enzymatic activity is determined by adding a chromogen (without TMB), and the colour intensity is measured on an SLT Spectra II microreader.
Enzyme linked immunosorbent assay of immunoglobulins ELISA assay of immunoglobulins.

Polyclonal antibodies Q-SwaHu-IgM, Q-SwaHu-IgG or Q-SwaHu-IgA (Sevac, Prague) were used as the binding antibodies. After incubation and washings, control sera with defined immunoglobulin contents and the supernatants were applied. Following incubation and washing, peroxidase-labelled antibodies against human immunoglobulin (Q-SwaHu-IgM, Q-SwaHu-IgG or Q-SwaHu-IgA) were added. A 1-h incubation at room temperature was followed by washing the plates and the redox reaction with OPD in phosphate ELISA buffer (pH = 6.2) was developed using hydrogen peroxide. The reaction was stopped with 1 M sulphuric acid and read on an SLT Spectra II ELISA reader. Individual samples were evaluated in ng/ml using the KIME E program.

Statistical evaluation

Student’s pair t-test at a probability level of \(P=0.05\) was used to evaluate the significance of differences between patients and their siblings. The results are presented as mean ± SE.

Results

Cytokine production by PBMC after a 24-h stimulation by mitogen and bacteria suspensions is shown in Table 1. The IFN-gamma formation was found to be significantly higher in healthy siblings than in the patients after stimulation by \(A. a\). and \(E. coli\). On the other hand, a significantly higher formation of IL–4 was detected in patients after stimulation by \(A. a\). and \(E. coli\) as compared with their healthy siblings.

The release of immunoglobulins after a 7-day stimulation of PBMC by mitogen and bacteria suspensions is shown in Table 2. Compared to their siblings, patients exhibited a significantly higher IgM level after polyclonal stimulation with \(E. coli\). The formation of IgA was significantly higher in the patients than in their siblings in cultures stimulated by both PWM and ConA. No significant differences were found after stimulation of PBMC by other mitogens and in unstimulated cultures. No significant differences were found in the level of IgG.

Discussion

Evidence based on microbiological, immunological and animal studies has shown that some forms of periodontal disease in adults can remain stable over many years and not endanger the life of the dentition, whereas other forms, despite extensive treatment, continue to break down, leading ultimately to tooth loss.22

The regulatory activity of T lymphocytes and the identification of which cytokines are present have been studied intensively in order to shed some light on the pathogenesis of periodontal disease. It has been shown that individuals susceptible to periodontal disease predominantly activate Th2 lymphocytes, while non-susceptible individuals activate Th1 lymphocytes.16

The verification of this hypothesis required a correct selection of patients. Patients with early onset periodontitis (susceptible individuals) and their siblings with healthy periodontium (non-susceptible individuals), both exhibiting presence of \(A. a\)., seem to be ideal for the study of the role of Th1 and Th2 lymphocytes in the pathogenesis of the disease. Despite the similar genetic backgrounds, similar age and similar hygiene habits in families, we found different production of cytokines after \textit{in vitro} stimulation.

The majority of immune responses occur locally rather than systemically within a small area of tissue: therefore most authors have concentrated on the \(T\) cells isolated from inflamed tissues of patients.16,23–26 This tissue can be obtained during surgical treatment of seriously affected patients. Lymphocytes isolated from peripheral blood of patients are rarely used.27–29

The use of PBMC is favourable for analysing the transformation of the immune response during the disease’s remission, when it is not possible to use cells from the tissue.

PBMC were capable of expressing mRNA for IL–2, IL–4, IL–5, IL–6, IL–10, IL–13 and IFN-gamma

<table>
<thead>
<tr>
<th>Mitogens and activators</th>
<th>IFN-gamma (U/ml × 1000)</th>
<th>(\text{IL–4 (ng/ml)})</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Patients</td>
<td>Siblings</td>
</tr>
<tr>
<td>PWM</td>
<td>30 ± 12</td>
<td>28 ± 12</td>
</tr>
<tr>
<td>PWM+ConA</td>
<td>34 ± 11</td>
<td>861 ± 824</td>
</tr>
<tr>
<td>(A. a).</td>
<td>6 ± 2</td>
<td>178 ± 14</td>
</tr>
<tr>
<td>(E. coli)</td>
<td>4 ± 1</td>
<td>125 ± 10</td>
</tr>
<tr>
<td>Unstimulated</td>
<td>740 ± 734</td>
<td>54 ± 4</td>
</tr>
</tbody>
</table>

*Student’s pair \(t\)-test at a probability level of \(P=0.05\) was used to evaluate the significance of differences between patients and their siblings. The results are presented as mean ± SE.
Table 2. Release of immunoglobulins after the stimulation (7 days) of PBMC by mitogens and antigens (results in ng/ml)\(^a\)

<table>
<thead>
<tr>
<th>Mitogens and activators</th>
<th>IgM Patients</th>
<th>IgM Siblings</th>
<th>IgM Significance</th>
<th>IgA Patients</th>
<th>IgA Siblings</th>
<th>IgA Significance</th>
<th>IgG Patients</th>
<th>IgG Siblings</th>
<th>IgG Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>PWM</td>
<td>122 ± 85</td>
<td>32 ± 17</td>
<td></td>
<td>7626 ± 2195</td>
<td>5520 ± 1992</td>
<td>P&lt;0.05</td>
<td>3510 ± 1307</td>
<td>1567 ± 310</td>
<td></td>
</tr>
<tr>
<td>PWM+ConA</td>
<td>11 ± 5</td>
<td>6 ± 3</td>
<td></td>
<td>1168 ± 367</td>
<td>378 ± 203</td>
<td></td>
<td>1120 ± 485</td>
<td>3372 ± 1279</td>
<td></td>
</tr>
<tr>
<td>A. a.</td>
<td>4 ± 2</td>
<td>7 ± 3</td>
<td></td>
<td>2948 ± 2674</td>
<td>298 ± 256</td>
<td></td>
<td>2307 ± 1295</td>
<td>2220 ± 1305</td>
<td></td>
</tr>
<tr>
<td>E. coli</td>
<td>7 ± 2</td>
<td>2 ± 1</td>
<td>P&lt;0.05</td>
<td>1494 ± 1040</td>
<td>1884 ± 1627</td>
<td></td>
<td>1490 ± 603</td>
<td>2325 ± 1320</td>
<td></td>
</tr>
<tr>
<td>Unstimulated</td>
<td>4 ± 1</td>
<td>4 ± 3</td>
<td></td>
<td>1194 ± 1039</td>
<td>971 ± 723</td>
<td></td>
<td>4576 ± 1434</td>
<td>3108 ± 1379</td>
<td></td>
</tr>
</tbody>
</table>

\(^a\) Student’s pair t-test at a probability level of P<0.05 was used to evaluate the significance of differences between patients and their siblings. The results are presented as mean ± SE.
following stimulation by ConA for 2 h. Cells isolated from affected tissue only expressed IL–10, IL–6, IL–13 and IFN-gamma. It seems that PBMC were capable of producing a wide spectrum of cytokines, but after homing of T cells into inflamed tissue, they were only capable of expressing either Th1 or Th2 cytokines.

By immunohistochemical analysis a higher percentage of IL–4 and IL–6 producing cells was found in the tissues affected by inflammation. Following in vitro stimulation by Porphyromonas gingivalis and Fusobacterium nucleatum, most lymphocytes obtained from biopsies of periodontitis affected tissues were of the Th2 phenotype, i.e. they formed high amounts of IL–4 and only small amounts of IL–2 and IFN-gamma. Manhart et al. proved that altered IL–2/IL–4 activities were founded in lesion of patients with early onset periodontitis.

After activation by PHA, CD8+CD28+ cells isolated from inflamed lesions of patients with adult periodontitis produced high amounts of IFN-gamma and were shown to have cytotoxic activity, whereas CD8+CD28–cells produced predominantly IL–4 and IL–5 and they showed a suppressive effect. We found a significantly higher formation of IL–4 and significantly lower formation of IFN-gamma after stimulation by ConA for 2 h. Following stimulation by ConA for 2 h. et al.

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