The serum concentration of two pro-angiogenic cytokines: basic fibroblast growth factor (bFGF) and transforming growth factor beta1 (TGF-β1), and anti-angiogenic factor endostatin in the serum of 80 never treated B-cell chronic lymphocytic leukemia (CLL) patients and 27 healthy volunteers was measured using an enzyme linked immunosorbent assay. The serum levels of both bFGF and TGF-β1 were found to be significantly higher in the CLL group (median 40.5 pg/ml and 38.6 ng/ml respectively) when compared to the control group (median 9.4 pg/ml and 18.9 ng/ml, respectively) (p < 0.001). The levels of endostatin were not significantly different in CLL and control groups (median 12.3 ng/ml and 8.4 ng/ml, respectively) (p > 0.05). In the group of CLL patients the level of bFGF was significantly higher in patients with progressive disease as compared with patients with stable disease (median 90.5 pg/ml and 40.5 pg/ml respectively) (p < 0.001). Patients in Rai stage III and IV also had significantly higher levels of bFGF than patients in Rai stage 0–II (median 100.1 pg/ml and 29.3 pg/ml respectively) (p < 0.001). The levels of both TGF-β1 and endostatin were lower in patients in Rai stage III and IV (median 28.9 ng/ml and 9.1 ng/ml respectively) than in patients in Rai stage 0–II (42.8 ng/ml and 13.1 ng/ml respectively) (p < 0.001 and p = 0.002 respectively). The level of endostatin was also lower in the group of CLL patients with progressive disease (median 10.0 ng/ml) as compared to patients with stable disease (median 20.5 ng/ml) (p = 0.008). In conclusion, the disturbance in the balance between pro- and anti-angiogenic factors may have an important influence on the course of CLL.

Key words: Angiogenesis, bFGF, TGF-β1, Endostatin, Chronic lymphocytic leukemia

Introduction

Angiogenesis is a multi-step process involving endothelial cell proliferation, differentiation, migration and organisation to form tubules. It has been well established that angiogenesis is involved in the expansion and dissemination of primary solid tumors. Many studies have demonstrated an association between microvessel density and prognosis in different cancers.1 Angiogenesis is regulated by many substances with pro-angiogenic activity and a number of inhibitors. Basic fibroblast growth factor (bFGF) and transforming growth factor beta (TGFβ1) are potent inducers of angiogenesis, enhancing the proliferation of epithelial cells and fibroblasts and are involved in the regulation of cellular replication and synthesis of many components of the extracellular matrix.2 Endostatin is a recent discovered, a 20 kDa fragment of the C-terminal, noncollagenous domain of collagen XVIII with antiangiogenic activity. It is able to block mitogen-activated protein kinase activation in endothelial cells and specifically inhibits endothelial proliferation.3

Recent studies indicate that angiogenesis may also play an important role in the pathogenesis of hematological malignancies, including lymphoma, myeloma and leukemia.4–6 Evidence of the role of this process in chronic lymphocytic leukemia (CLL) originates from the finding that increased vessel density was observed in the bone marrow of CLL patients.7 There are also studies that have revealed that CLL B cells spontaneously secrete a variety of pro and anti-angiogenic factors including vascular endothelial growth factor (VEGF), TGFβ1, bFGF and endostatin.8–11 The shift in balance between production of anti-angiogenic factors and pro-angiogenic factors is a process that may have significant influence on disease progression.

The aim of our study was to compare the serum levels of the pro-angiogenic factors: bFGF and TGFβ1 and the anti-angiogenic factor endostatin, between healthy blood donors and a group of never treated
CLL patients. We have also investigated if the levels of pro- and anti-angiogenic factors correlate with the stage of the disease.

Patients and methods

Patients

The study involved 80 B-CLL patients and 27 healthy volunteers. All CLL patients fulfilled the National Cancer Institute-Sponsored Working Group diagnostic criteria for CLL.12 There were 48 male patients and 32 female patients with a mean age of 64 years (range 40–78). Forty-four patients had stable disease and 36 had progressive disease (including lymphocytosis doubling time < 1 year, massive splenomegaly, bulky adenopathy, recurrent disease-related infections, weight loss > 10% in a 6-month period, temperature of 38°C related to disease, extreme fatigue, III and IV Rai stage disease).13 According to Rai classification14 18 patients were in 0 stage, 15 in I, 19 in II, 6 in III and 22 in IV stage of the disease. Fifty-two patients (Rai 0–II) were included in good prognosis group and 28 (Rai II–IV) in bad prognosis group.15 All patients included in this study were previously untreated. Characteristics of the patients are shown in Table 1.

An assessment of history and a physical examination was performed as initial diagnostic procedure. The laboratory tests included: complete blood count, immunoglobulins level, liver and renal function tests, bone marrow aspiration for morphology and immunophenotyping. Surface marker analysis was performed to confirm B-cell origin and monoclonal proliferation, including immunoglobulins heavy- and light-chains, CD5, CD10, CD19, CD20 and CD23.

The controls included 18 men and 9 women, with a mean age of 52 years (range 40–75).

This project was performed in accordance with the Helsinki Declaration. Informed consent was obtained from all patients participating in the study. The local Ethics Committee approved the project.

Serum sampling and cytokine determination

The serum obtained was stored at −80°C until assayed for cytokines. The cytokines serum concentrations were assayed by specific, commercially available, enzyme linked assay kits. The kits for bFGF and TGFβ1 were the products of R&D Systems Inc. (Minneapolis, MN, USA) and the endostatin was a product of Oncogene Research Products (Boston, MA, USA). Standards and samples were assayed as duplicates, and interassay variations were within the range given by manufacture. Assay sensitivity was 1.0 pg/ml for bFGF, 5.0 ng/ml for TGFβ1 and 1.953 ng/ml for endostatin.

Statistical analysis

Comparisons of values were made with Mann–Whitney tests. Comparisons were considered significant when \( p < 0.05 \). The linear correlations between serum cytokine levels as compared with the lymphocyte number were evaluated using the Sperman rank-sum correlation method.

Results

The serum level of TGFβ1 and endostatin were measurable in all normal individuals and all CLL patients and bFGF was measurable in 24 out of 27 healthy volunteers and in all CLL patients.

The result of the measurements of TGFβ1, bFGF and endostatin are shown in Table 2.

The serum levels of both bFGF and TGFβ1 were found to be significantly higher in the CLL group (median 40.5 pg/ml and 38.6 ng/ml, respectively) when compared to the control group (median 9.4 pg/ml and 18.9 ng/ml, respectively; \( p < 0.001 \)). The levels of endostatin were not significantly different between the control and the CLL group (median 8.4 ng/ml and 12.3 ng/ml, respectively; \( p = 0.09 \)). In the group of CLL patients the level of bFGF was significantly higher in patients with progressive disease as compared to patients with stable disease (median 90.5 pg/ml and 40.5 pg/ml, respectively; \( p = 0.001 \)). Patients in Rai stage III and IV also had significantly higher levels of bFGF than patients in Rai stage 0–II (median 100.1 pg/ml and 29.3 pg/ml, respectively; \( p < 0.001 \)). The levels of both TGF-β1 and endostatin were lower in patients in Rai stage III and IV (median 28.9 ng/ml and 9.1 ng/ml, respectively) than in patients in Rai stage 0–II (42.8 ng/ml
Table 2. Comparison of serum levels of TGFβ1, bFGF and endostatin in the healthy control group and in 80 untreated B-cell CLL patients

<table>
<thead>
<tr>
<th>Cytosine</th>
<th>Control (n = 27) (A)</th>
<th>All CLL patients (n = 80) (B)</th>
<th>Stable CLL (n = 43) (C)</th>
<th>Progressive CLL (n = 37) (D)</th>
<th>Rai 0–II (n = 52) (E)</th>
<th>Rai III–IV (n = 28) (F)</th>
<th>Statistically significant comparison</th>
</tr>
</thead>
<tbody>
<tr>
<td>TGF-beta (ng/ml)</td>
<td>18.9 (7.0–38.5)</td>
<td>36.6 (13.9–89.0)</td>
<td>13.0 (15.8–89.0)</td>
<td>32.3 (13.9–63.2)</td>
<td>42.8 (13.9–89.0)</td>
<td>28.9 (13.9–42.0)</td>
<td>A vs. B: p = 0.001</td>
</tr>
<tr>
<td>bFGF (pg/ml)</td>
<td>9.4 (0–20.1)</td>
<td>40.5 (5.1–230.5)</td>
<td>41.9 (5.1–150.0)</td>
<td>90.5 (10.1–230.5)</td>
<td>29.3 (5.1–150.0)</td>
<td>100.1 (10.1–230.5)</td>
<td>A vs. B: p = 0.001</td>
</tr>
<tr>
<td>Endostatin (ng/ml)</td>
<td>8.4 (2.1–21.5)</td>
<td>12.3 (2.9–58.0)</td>
<td>20.5 (6.4–34.0)</td>
<td>10.0 (2.9–58.0)</td>
<td>13.1 (4.4–34.0)</td>
<td>9.1 (2.9–58.0)</td>
<td>A vs. B: p = 0.09</td>
</tr>
</tbody>
</table>

and 13.1 ng/ml, respectively; p < 0.001 and p = 0.002, respectively). The level of endostatin was also lower in the group of CLL patients with progressive disease (median 10.0 ng/ml) as compared to patients with stable disease (median 20.5 ng/ml; p = 0.008).

In the entire group of CLL patients we have found a negative correlation between the serum level of TGFβ1 and bFGF (p = 0.019, R = −0.26) and the serum levels of endostatin and bFGF (p = 0.0004, R = 0.39). We have also found a positive correlation between the number of lymphocytes in the peripheral blood of CLL patients and the level of bFGF (p < 0.0001, R = 0.48) and endostatin (p = 0.0003, R = 0.39).

Discussion

The angiogenic switch in tumor cells is characterised at a molecular level by an imbalance in the production of pro- and anti-angiogenic factors. There is more and more evidence that B-CLL cells themselves may be an important source of these factors, but their exact role in the biology of the disease remains unclear.10,16 In order to determine if the levels of pro- and anti-angiogenic factors are associated with the course of the disease we compared the levels of bFGF, TGFβ1 and endostatin to the Rai risk stage and whether a patient has stable or progressive disease.

CLL cells-derived bFGF acts as a paracrine endothelial cells mitogen in angiogenesis. It enhances the proliferation of a variety of ectoderm- and mesoderm-derived cells, such as epithelial cells and fibroblasts.1 It also acts in an autocrine fashion on CLL cells behavior such as prolong survival due to inhibition of apoptosis.17 Thus, in progressive phase of CLL, when the lymphocytosis doubling time is quick and the expansion of lymphoid tissue and neovascularisation take place, bFGF could play an important role. Our investigations revealed significantly higher serum bFGF levels in CLL patients as compared to the healthy control group (p < 0.001). Furthermore we revealed that bFGF concentration is significantly higher in the serum of patients with more advanced Rai stage and in those with progressive disease (p < 0.001). We also observed the positive correlation between the serum level of this cytokine and the number of lymphocytes in the peripheral blood. These observations are in agreement with other studies.17,19 Most of the studies revealed a positive correlation between the increased bFGF concentrations and tumor progression or patient survival. High pre-treatment level of bFGF has been found an independent predictor of poor prognosis in non-Hodgkin lymphomas.19 Our previous study revealed that serum levels of bFGF and TGFβ1 were significantly higher in progressive CLL patients than in patients in complete remission.20 Menzel et al. revealed a high level of bFGF in lymphocytes of CLL patients, especially those with high-risk disease.17 Molica et al. analysed 81 Binet stage A CLL patients and found that serum levels of bFGF positively correlated with Rai sub-stages but failed to demonstrate any correlation between the bFGF level and disease progression. In contrast to our results they did not find correlation between bFGF concentration and peripheral blood lymphocytosis.18

CLL-B cells were shown to express TGFβ1 RNA and to release bioactive TGFβ1 into culture supernatants.21 Autocrine secretion of TGFβ1 exhibits an antiproliferative effect on CLL-B cells. This effect is most relevant in B cells cultured in direct contact with activated CD4+ T cells suggesting an indirect mode of action.22 On the other hand TGFβ1 is a well known pro-angiogenic factor.23 It also has been shown to potentate the expression of VEGF, the most potent pro-angiogenic factor in CLL.24 In our studies we observed a significantly higher concentration of TGFβ1 in the serum of untreated CLL patients when compared to the control group (p < 0.001). An increased TGFβ1 level in comparison with healthy controls has been demonstrated in patients with renal, colorectal and nasopharyngeal carcinomas.25–27 There was also a correlation between high TGFβ1 serum level and advanced tumor stage. In contrast
we observed significantly lower serum levels of this cytokine in patients with advanced CLL stages ($p < 0.001$). It is not unexpected because although TGFβ1 is a molecule with pro-angiogenic activity it is also known as an autocrine growth inhibitor of lymphocyte proliferation. 28 TGFβ1 appears to function in CLL as a negative regulator of B lymphocytes but loss of responsiveness to this cytokine accompanied by a decrease of TGFβ1 receptor expression, might provide a selective advantage to B CLL lymphocytes. 29–31 Moreover this increased TGFβ1 production could be responsible for the inhibiting activity on hematopoietic precursors and bone marrow failure seen in advances stages of CLL. 32

In addition to pro-angiogenic growth factors we evaluated plasma levels of endostatin an endogenous inhibitor of angiogenesis. 3 However we did not find a difference between serum levels of this cytokine in CLL patients and the healthy control group ($p = 0.09$). Other authors have reported endostatin levels similar to healthy control in patients with squamous cell vulvar carcinoma and hepatocellular carcinoma. 33,34

The same observation was made in patients with rheumatoid arthritis and systemic lupus erythematosus (SLE). 35,36 In contrast Feldman et al. found significantly elevated serum levels of endostatin in patients with advanced Rai stages or progressive disease as compared to lower Rai stages and stable disease ($p < 0.001$). We also found a positive correlation between the serum concentration of endostatin and peripheral blood lymphocytosis and a negative correlation between this cytokine level and bFGF serum concentration. Similar to our results, a negative correlation between endostatin and bFGF was also revealed in patients with SLE. 36 In contrast Bertolini et al., in series of 36 non-Hodgkin lymphoma patients, did not find a difference in endostatin level between patients in complete remission and patients with progressive disease nor a correlation between endostatin levels and event-free survival rate. 39 Feldman et al. revealed that higher endostatin levels were associated with an increased risk of tumour recurrence after resection of soft tissue sarcoma. 37 They also found the positive correlation of endostatin and pro-angiogenic VEGF.

In conclusion, the results of the current study showed that the levels of anti-angiogenic endostatin and pro-apoptotic TGFβ1 are lower in the serum of CLL patients in advanced stages or with progressive disease while the level of pro-angiogenic bFGF is significantly higher in these groups of patients. The imbalance between these cytokines may have an influence on the course of the disease.

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