**Met-Enkephalin Involvement in Morphine-Modulated Peritonitis in Swiss Mice**

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Morphine coinjection with zymosan inhibits pain and leukocyte accumulation during peritonitis in several strains of mice, and affects systems of endogenous opioids. Present investigations focus on Met-enkephalin (Met-ENK) in the inflamed peritoneal cavity and brain centers of Swiss mice. Males of Swiss mice were IP injected with zymosan or zymosan supplemented with morphine. At the selected time points the peritoneal leukocytes were counted and the Met-ENK level was measured in exudatory fluid and leukocytes, striatum, hypothalamus, and pituitary gland. The Met-ENK content in peritoneal fluid rised sharply after zymosan injection, which corresponded with its decline in exudatory leukocytes, hypothalamus, and striatum. Morphine coinjection enhanced intraperitoneal accumulation of Met-ENK and its release from exudatory leukocytes, but inhibited its early fluctuations in hypothalamus and striatum. Effects of morphine-modulated inflammation on the Met-ENK system lasted longer than 7 days.

**INTRODUCTION**

The immune system plays an important role in the pain control, especially under inflammatory conditions. As evidenced in series of experiments on adjuvant-induced paw inflammation in the rat, opioid receptors on peripheral sensory nerve terminals in the inflamed tissue are upregulated and activated by opioid peptides derived from immunocytes accumulated in the focus of inflammation resulting in potent analgesia [1, 2].

We wish to draw attention to another model for investigations of the opioid system participation in the inflammatory processes, namely to experimental peritonitis induced by intraperitoneal (IP) injection of a sterile stimulant, for example, zymosan. The convenience of this model consists in the possibility of a precise quantification of inflammation-related cells and soluble factors in samples of exudatory fluid quantitatively retrieved from the control or inflamed peritoneal cavity [3, 4, 5]. We have shown that the supplementation of a stimulant with exogenous opioid, morphine, morphine, affects inflammation in a naltrexone-reversed manner, that is, through the binding of opioid receptors. Morphine co-administration significantly reduces the levels of chemotactic factors and the number of inflammatory leukocytes in several (but not all) strains of mice [3, 5, 6, 7], in the two investigated fish species, goldfish and Atlantic salmon [3, 8, 9], but in none of the three investigated species of anuran amphibians [9, 10] with experimental peritonitis. The morphine-induced inhibition of early stages of peritonitis corresponds with an inhibited in vitro migration of mice and fish (but not frogs) morphine-treated leukocytes to zymosan-activated serum [11].

In mice, early stages of zymosan-induced peritonitis are accompanied by characteristic body writhes considered to be visceral pain symptoms [12]. Morphine supplementation of the irritant completely eliminates body writhes even at the low doses (5 or 10 mg/kg BW) while, at the high dose (20 mg/kg BW), additionally inhibits influx of leukocytes in four out of the five investigated strains of mice, including the Swiss [5, 6, 7]. Therefore, local morphine administration might offer double profits during planned surgeries, being both antinociceptive and anti-inflammatory [5]. Such a morphine-dependent modification of the inflammatory process would seem to be advantageous for the host, since some inflammation-related cells and molecules at the high concentration and/or during prolonged action might be detrimental [5].

We may speculate that morphine added to an inflammatory stimulant can interplay with endogenous opioids so as to support their antinociceptive function in the focus of inflammation and, as a consequence, lead to the inhibition of influx and accumulation of new leukocytes. In a series of the ongoing experiments we investigate the effects of exogenous morphine on systems of endogenous opioids...
opioids during experimental peritonitis in Swiss mice. It turned out that morphine coinjection affects the proopio-
 melanocortin (POMC) and prodynorphin (PDYN) sys-
tems on the levels of mRNAs for respective peptides and
their receptors on inflammatory leukocytes as well as the
accumulation of peptides in the focus of inflammation
[13]. In the case of the proenkephalin (PENK) system
we detected the upregulated level of PENK mRNA in ex-
udatory leukocytes [14], but despite the strong efforts we
were unable to detect mRNA for delta opioid receptors
on Swiss mice peritoneal leukocytes (PTLs) [13]. We have
also shown a local increase in the Met-enkephalin (Met-
ENK) level in the focus of inflammation with its paral-
lel decrease in the lymph nodes and distant neurohor-
monal centers 4 hours after zymosan-induced peritonitis
in Swiss mice. This implies the dual origin of this peptide
from both locally accumulated leukocytes and distal neu-
rohormonal centers [15]. The goal of the present study
was to extend those preliminary observations by record-
ing the time course of Met-ENK changes in the focus of
inflammation and some brain areas. The second goal was
to check if the coinjection of the anti-inflammatory dose
of morphine would affect zymosan-induced fluctuations
in the local and distant Met-ENK levels.

MATERIALS AND METHODS

Animals

Adult male Swiss mice, purchased from the commer-
cial supplier (Breed ing of Laboratory Animals, Collegium
Medicum, Krakow, Poland), were kept in 20×13×18 cm³
cages (four mice per cage) in a room with controlled
temperature (22°C) and lighting (lights on 8:00–20:00).
Food (standard mouse laboratory chow) and water were
available ad libitum. Animals were 6–8 weeks of age (25–
30 g body weight) at the beginning of the investigations.
The experiments were conducted according to License no.
16/OP/2001 from the Local Ethical Committee.

Schedule of drug administration
and experimental protocol

Animals were either left untreated (0 time) or in-
jected IP with freshly prepared zymosan (Z groups)
(2 mg/mL, 0.5 mL/25 g BW) (zymosan A, Sigma, St Louis,
Mo) in sterile PBS, or with zymosan supplemented with
morphine hydrochloride (ZM groups) (20 mg/kg BW;
Polfa, Kutno, Poland). Animals were killed at selected
time points and their peritoneal cavities were lavaged
with 1 mL of PBS. Peritoneal cells were Turk-stained
and counted with a hemocytometer. Peritoneal fluid
(PF) and peripheral blood were centrifuged (15 minutes,
1500 rpm), then supernatants and blood plasma were
stored at −20°C for future analysis. Selected brain ar-
areas (hypothalamus, pituitary gland, striatum) were dis-
sected and kept frozen (−70°C) until used. All experi-
ments were repeated at least twice with at least 3 animals
per group.

Nociceptive activity

Nociceptive activity was tested using the writhing
model [12]. Characteristic body writhes (consisting of
a contraction of the abdominal muscles together with
a stretching of hind limbs) were counted during five-
minute intervals for each mouse.

Met-ENK detection

Met-ENK levels were measured in homogenates of
the brain structures (striatum, hypothalamus, pituitary
glands), pellets of peritoneal cells, and PF. Brain structures
were quickly removed and frozen on dry ice. Weighed tis-
ues were homogenized in chilled phosphate buffer (pH
= 6.5, 1:10 w/v) using Ultra-Turrax T8, then centrifuged
for 30 minutes, 4000 rpm, at 4°C, samples were stored at
−70°C until performing the assay.

Native enkephalin was purified on Porapak columns
comprised of Porapak Q 100–120 mesh (Waters, Mil-
ford, Mass) in 3 mL of absolute ethanol. Porapak slurry
was prepared by degassing overnight 25 g of the mate-
rial in 350 mL of absolute ethanol. Shortly before applying
the samples, columns were washed with 6 mL of absolute
ethanol and equilibrated with 9 mL of doubly distilled wa-
ter. Columns loaded with the samples were washed with
6 mL of doubly distilled water and native enkephalin was
eluted with 3 mL of absolute ethanol, then lyophilized and
held at 4°C until assay within 2 days.

Met-ENK immunoreactivity was quantitated using
commercial antiserum developed in rabbit (Bachem,
Baden-Wuerttemberg, Germany), 125I-Met-enkephalin
(New England Nuclear, Boston, Mass) and Met-ENK
standard (Peninsula, San Carlos, Calif). The antiserum
was used in a final dilution of 1:12,000; it showed
30% cross-reactivity of 100% with Met-enkephalin sulfoxide,
2% with Leu-enkephalin and less than 1% with Met-
enkephalin-Arg-Phe, Met-enkephalin–Arg-Gly-Leu, or β-
endorphin. Intra-assay and interassay coefficients of vari-
ation for the assay are 7 and 11 percent, respectively.
Recovery of standard Met-ENK added to the samples
and carried through the entire extraction and radioim-
unoassay procedures was 79% [16].

Statistical analysis

The results were statistically analyzed by 2-way
ANOVA (to check if the time course of ZM groups dif-
sers significantly from that of Z groups) with post hoc
Tukey’s test (indicating the time points with means sig-
nificantly different). The differences were considered sta-
ristically significant at P < .05.

RESULTS

Time course of peritonitis induced by
injection with Z or ZM in Swiss mice

Values at particular time points were compared with
the level at time 0 as well as between the Z and ZM groups
of animals. See Figure 1.
**Temporal Activity**

Body writhes (pain symptoms) were compared between the animals from ZM and Z groups. The number of body writhes in the Z groups was high during the first 30 minutes after zymosan injection, while thereafter these symptoms were sporadic only. Body writhes were absent in the ZM groups of Swiss mice (Figure 1a).

**Time course of leukocyte accumulation**

The number of resident PTLs was low in the intact animals (time 0) while the total number of PTLs increased sharply between 2 and 6 hours after the zymosan injection and remained at a similarly high level for at least 7 days after injection (Figure 1b). Among them, PMNs dominated during the first 24 hours while on the 7th day intraperitoneal exudate consisted of mainly the mononuclear cells (Figure 1c).

The PTLs influx was significantly delayed in animals injected with zymosan supplemented with morphine (Figure 1b). Such a delay in PTLs accumulation in the focus of inflammation concerned mainly PMNs, which, nevertheless, eventually reached the level characteristic for the Z group 24 hours after injection (Figure 1c). The effects of morphine supplementation on the accumulation of mononuclear leukocytes were less drastic (not shown).

**Met-ENK fluctuations in the focus of inflammation**

After the irritant injection, the Met-ENK content was significantly increased in PF, significantly more in the ZM than in the Z groups of animals (Figure 2a). The content of Met-ENK in the total pellet of exudatory cells in the Z group was relatively stable with a peak only at the 4th hour after the Z injection. This was apparently due to a sustained IP influx of new leukocytes that balanced the peptide release into PF. In contrast, the Met-ENK level was significantly decreased in PTLs of ZM animals that corresponded with the delayed influx of cells. Despite the similar PTLs numbers in the Z and ZM groups of animals on the 7th day of peritonitis, the Met-ENK content of PTLs was higher in the Z than ZM groups, which suggests its enhanced release to PF of ZM mice (Figure 2b). The Met-ENK levels in the same numbers (10^6) of PTLs decreased significantly, more in the ZM than Z groups of animals (Figure 2c).

**Met-ENK fluctuations in brain centers**

A sudden drop of Met-ENK in the brain centers (striatum and hypothalamus) of zymosan-injected animals was reversed by the morphine coinjection (Figures 2d and 2e). On the 7th day after injection, the level of Met-ENK was still different from that in the control in the striatum (lower in the Z and higher in ZM groups) and in hypothalamus (decreased in both Z and ZM animals) (Figures 2d and 2e). Drastic fluctuations of Met-ENK in the pituitary gland of the Z group of animals were smoothed in the ZM groups of mice (Figure 2f). Met-ENK was at the control level on the 7th day in the pituitary gland (Figure 2f).
FIGURE 2. Time course of Met-enkephalin levels in males of Swiss mice either intact (time 0) or IP injected with zymosan (Z, solid lines and circles) or zymosan with morphine (ZM, broken lines and triangles) in (a) peritoneal fluid (PF); (b) total pool of PTLs; (c) 10⁶ PTLs; (d) striatum; (e) hypothalamus; (f) pituitary gland. The results are expressed as X + SE, n = 4–8. Solid symbols indicate means significantly different from time 0 according to ANOVA at P < .05; asterisks between significantly different means of Z and ZM groups at the given time point according to post hoc Tukey’s test denote that P < .05 (**); P < .01 (***); P < .001 (****).
DISCUSSION

The zymosan-induced peritonitis described here follows a typical pattern of inflammatory reaction with a rapid accumulation of PMNs during the initial 24 hours followed by mononuclear leukocytes dominating in peritoneal cavity on the 7th day (Figure 1). The early influx of leukocytes is accompanied with a pronounced IP accumulation of cytokines, both classical proinflammatory factors such as IL-1β, TNF, IL-6, KC, MCP-1 [17], and also endogenous opioids, namely, β-endorphin, dynorphin [13], and Met-ENK [14, 15], which make up the best characterized endogenous analgesic system [18]. Therefore it is not surprising that behavioral episodes reflecting visceral pain, namely, the characteristic body writhes of mice with peritonitis, are relatively short lasting, as they are frequent mostly during the first 30 minutes after zymosan injection (Figure 1a, [7, 12]). The endogenous peptides can act antinociceptively on the opioid receptors on the local nerve endings, as evidenced in a case of adjuvant-induced paw inflammation in rats [18]. The opioid receptors on the nerve endings are gradually upregulated in a focus of inflammation. The initial low availability of opioid receptors may be partly responsible for the delay in the analgesic effects of rapidly accumulating opioid peptides (cf Figures 1a and 2a), in addition to the early abundance of a variety of proanalgesic factors at the initiation of inflammation [1].

The results of the present and previous studies of experimental peritonitis indicate that opioids that accumulate in the inflamed peritoneal cavity may have dual origin, being released from both exudatory leukocytes and distant neurohormonal centers (Figure 2). Figure 2 illustrates that an IP zymosan injection triggers a rapid opioid release from the PTLs, striatum, hypothalamus, and pituitary gland. Apart from the opioids from the neurohormonal centers it seems that the wave of neutrophil-derived peptides (dominating during the first 24 hours of peritonitis, Figure 1c) is followed by the peptides derived from the mononuclear leukocytes (dominating on the 7th day), similar to the sequence detected by immunohistochemistry during the rat paw inflammation [2].

Exogenous morphine shares receptors with opioid peptides [18] thus it is not surprising that the coinjection of morphine with zymosan affects the system of endogenous opioids in a different way than that induced by the injection of zymosan only (Figure 2, [13]). In general, an IP coinjection of a high dose of morphine inhibits the Met-ENK release from the brain centers or even induces their temporal accumulation. On the other hand, the morphine coinjection enhances the zymosan-induced accumulation of Met-ENK in PF on both the 1st and 7th days of peritonitis (Figure 2), which implies the leukocyte origin of the IP Met-ENK. The latter phenomenon is surprising as the IP influx of leukocytes is delayed in the ZM versus Z groups of animals, while on the 7th day the numbers of PTLs (both PMNs and mononuclear cells) are similar in the Z and ZM groups of mice (Figure 1). The accumulation of free Met-ENK in exudatory fluid may be connected with the occupation of opioid receptors by exogenous morphine. This phenomenon may explain the enhanced accumulation of free endogenous opioids soon after the morphine injection but not 7 days later. Therefore at present we may only conclude that morphine interplay with endogenous opioids during inflammation has long-lasting effects.

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REFERENCES


