Multidose Streptozotocin Induction of Diabetes in BALB/c Mice Induces a Dominant Oxidative Macrophage and a Conversion of $T_{H1}$ to $T_{H2}$ Phenotypes During Disease Progression

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Macrophages (Mp) are implicated in both early and late phases in type 1 diabetes development. Recent study has suggested that a balance between reductive Mp (RMp) and oxidative Mp (OMp) is possible to regulate $T_{H1}/T_{H2}$ balance. The aim of this study is to investigate the redox status of peritoneal Mp and its cytokine profile during the development of autoimmune diabetes induced by multiple low-dose streptozotocin in BALB/c mice. Meanwhile, the polarization of $T_{H1}/T_{H2}$ of splenocytes or thymocytes was also examined. We found that peritoneal Mp appeared as an “incomplete” OMp phenotype with decreased icGSH along with disease progression. The OMp showed reduced TNF-$\alpha$, IL-12, and NO production as well as defective phagocytosis activity compared to nondiabetic controls; however, there was no significant difference with IL-6 production. On the other hand, the levels of IFN-$\gamma$ or IL-4 of splenocytes in diabetic mice were significantly higher compared to the control mice. The ratio of IFN-$\gamma$ to IL-4 was also higher at the early stage of diabetes and then declined several weeks later after the occurrence of diabetes, suggesting a pathogenetic $T_{H1}$ phenotype from the beginning gradually to a tendency of $T_{H2}$ during the development of diabetes. Our results implied that likely OMp may be relevant in the development of type 1 diabetes; however, it is not likely the only factor regulating the $T_{H1}/T_{H2}$ balance in MLD-STZ-induced diabetic mice.

INTRODUCTION

Macrophages (Mp) play a pivotal role in specific and nonspecific immunity, and the physiological status of Mp may contribute to the overall regulation of the host defense system. A number of studies have showed the functional heterogeneity of Mp with different cytokine propensity or metabolic activities, therefore inducing distinct immune response such as $T_{H1}$-type versus $T_{H2}$-type ($T_{H1}$, T helper). Very recently, Murata et al proposed the functional discrimination of two classes of Mp, namely the reductive Mp (RMp) with a high intracellular content of glutathione (icGSH) and oxidative Mp (OMp) with a reduced content [1]. It was found that $T_{H1}/T_{H2}$ balance might be regulated by the altered balance between RMp and OMp through the distinctive production of TNF-$\alpha$, IL-12, and NO (nitric oxide) versus IL-6 and IL-10. This classification cast light on the potential role of the predominant RMp or OMp in inflammatory and autoimmune diseases, such as IDDM (insulin-dependent diabetes mellitus) in NOD (nonobese diabetic) mice [2], spontaneous inflammatory bowed disease [3], cancer immunotherapy [4], corneal allograft [5], and aging-related autoimmune disease [6]. To date, the predominant presence of RMp or OMp in the development of multiple low-dose streptozotocin (MLD-STZ) induced diabetes has been little studied.

For our experiments, we used the MLD-STZ animal model. Recently this model has been attractive for its little or no influence of genetic background, as most new cases of type 1 diabetes are sporadic and occur in families with no previous history of diabetes. Furthermore, as the term “latent autoimmune diabetes mellitus in adults” (LADA) has been introduced to describe an important minority of adult-onset patients with diabetes [7], STZ-induced diabetic mice may serve to be a better model of it since diabetes can be initiated at a relatively older age, when animals had all reached maturity, with a negligible weight loss.

Using this murine model, we found that during the development of diabetes, peritoneal Mp consistently skewed to OMp with gradually decreased icGSH.
The OMP showed reduced TNF-α, IL-12, and NO production, and also defective phagocytosis activity compared to nondiabetic controls, although there was no significant difference with IL-6 production. On the other hand, the levels of IFN-γ or IL-4 of splenocytes in diabetic mice were significantly higher than the controls. The ratio of IFN-γ to IL-4 level was higher at the early stage of diabetes, then declined several weeks after the occurrence of diabetes, concomitant with the sequential change of Th1/Th2 skewing in the thymus. Thus, the aberrant OMP might play a relevant role in the pathological progression of diabetes, however, it is not likely the only factor regulating the Th1/Th2 balance in MLD-STZ-induced diabetes mellitus.

MATERIALS AND METHODS

Experimental animals

Male BALB/c mice were initially purchased from Shandong Laboratory Animal Center and bred in our laboratory animal facility. The mice were housed with free access to water and a standard laboratory diet. Diabetes was induced in mice with STZ (Sigma, St Louis, Mo) as described previously [8]. Briefly, STZ was reconstituted in 25 mM sodium citrate (pH 4.5). Injections were made intraperitoneally (40 mg/kg, IP) within 15 minutes of preparation. Mice were treated with five consecutive daily injections. Nonfasting blood glucose levels were measured once a week, following the administration of STZ by a blood glucose monitor drawing blood from the tail vein (Lifescan, Inc, Milpitas, Calif). Mice with a blood glucose of at least 11.1 mmol (300 mg/DL) were induced gradually after 3 weeks following MLD-STZ of BALB/c mice; those that became diabetic within 1 week were used as diabetic group in the experiment (n = 6; blood glucose: 15±2.1 mmol; body weight: 28±1.9 g), while those that became diabetic after 4 weeks were used as advanced diabetic group (n = 6; blood glucose: 20±2.5 mmol; body weight: 25±2.9 g). Mice given the same amount of 25 mM citrate buffer were used as the control group (n = 6; blood glucose: 5+/−0.4 mmol; body weight: 28+/−1.5 g). Even to advanced diabetes mice, there was no significant loss in body weight compared with the controls.

Proliferation assay

The thymus cells or spleen cells proliferation assay on stimulation of ConA (Concanavalin A) was measured using MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium-bromide) reduction assay as previously described [9]. Briefly, Single cell suspensions of either thymocytes or splenocytes were prepared and viability was assessed by trypan blue exclusion. Thymocytes (8×10⁵ cells/well) or splenocytes (4×10⁵ cells/well) were plated in 96-well plates in RPMI-1640 (medium named after Roswell Park Memorial Institute) complete medium and were stimulated with ConA (30 μg/ml, Sigma) for 20 hours or 44 hours. Sterile MTT solution (5 mg/ml MTT in RPMI-1640) was then added into the wells and incubated for another 4 hours until purple precipitate was visible. After moving the medium by centrifugation, the converted dye was solubilized with 200 UL dimethyl sulfoxide, and the absorbance of the converted dye was measured at a wavelength of 490 nm with background subtraction as 630 nm. The stimulation index (SI) is determined by the absorbance with ConA/the absorbance without ConA.

Peritoneal macrophages

Peritoneal cells (PC) were harvested by injecting total 10 mL of an ice-cooled Hanks-10% FBS (fetal bovine serum)-heparin (10 U/mL) solution into the peritoneal cavity of mice. The collected PCs were added to a microplate at 1–3 × 10⁵ cells/200 μL RPMI-1640 medium. The adherent cells after a 2-hour incubation were used as resident peritoneal Mp for production of cytokines and NO by culturing for 48 hours.

Determination of intracellular GSH

Peritoneal cells adherent to dishes were collected by D-Hanks (2.5 mmol/L EDTA) and washed 3 times with cold D-hanks buffer. The cell pellet was immediately lysed with ultrasonic, and after centrifugation, some of the supernatants was assayed for the total protein content using the Coomassie protein assay kit (Jiancheng Co, Nangjing, China). Thereafter, 10% sulfosalicylic acid was added to the remained supernatants to precipitate protein. After centrifugation, supernatants were collected for GSH assay. The cellular GSH concentration was assayed using the GSH kit (Jiancheng Co), and the icGSH was determined as mg GSH/g protein.

Nitrite assay

The accumulation of NO₂⁻ was taken as a parameter for nitrite (NO) production. NO production by Mp was measured in supernatants collected after 48 hours of culture. Briefly, cell-free supernatants were incubated with the Griess reagent for 10 minutes at room temperature and absorbance at 550 nm was measured. The concentration of NO₂⁻ was determined by the square linear regression analysis of a sodium nitrite standard that was measured in each experiment.

Measurement of cytokine levels by ELISA

The IL-4, IL-6, TNF-α, and IFN-γ concentration was determined using an ELISA (enzyme-linked immunosorbent assay) kit (Alpha Diagnostic Intl, Inc, San Antonio, Tex) according to the indication of the manufacturer.

Phagocytosis assays

The phagocytosis activity of Mp was measured by the neutral fuchsin uptake method. 0.075% neutral fuchsain solution was added to adherent Mp (5×10⁶ cells/mL), and the samples were incubated for 1 hour at 37°C to study phagocytosis. Cells were then subjected to lysis by adding...
Figure 1. The periodic change of peritonealMp. (a) Mp count. There was significant difference between the diabetic and control groups \((P < .05)\), and the count increased significantly after the onset of diabetes \((P < .01)\). (b) The amount of intracellular GSH in adherent Mp of the mice. Mp collected from the diabetic mice showed significantly decreased content compared to that of control Mp. Results shown are representative of three experiments, and the Mp were all resting Mp.

**Statistical analysis**

The results of icGSH levels, NO content, macrophage phagocytosis, ConA-stimulated index, and cytokine levels were expressed as mean \(+/-\) SE. This data was analyzed by Student’s \(t\) test, and the difference was judged at \(P < .05\).

**RESULTS**

The Mp count was significantly increased in the peritoneal cavity of diabetic mice \((4.5+/−2.21 × 10^6 cells/mouse)\) as compared to nondiabetic controls \((1.1+/−0.59 × 10^6 cells/mouse)\) \((P < .05)\), and the Mp continued to increase 4 weeks after the incidence of diabetes \((5.4+/−1.46 × 10^6 cells/mouse)\) (Figure 1a). In addition, the redox status of Mp, indexed by icGSH, was significantly decreased in either early \((28+/−6.6 mg/g protein)\) or advanced diabetic mice \((12+/−3.3 mg/g protein)\) as compared to the controls \((62+/−3.4 mg/g protein)\) (Figure 1b). Significant difference was also found between the (early) diabetic mice and the advanced diabetic ones. Therefore, along with the diabetic progression, the peritoneal Mp showed OMP phenotypes with gradually decreased icGSH.

To characterize the cytokine expression profile by the OMP in diabetic mice, we assessed cytokines that have been found to be related to OMP or RMP, namely, TNF-\(\alpha\), IL-12, IL-6, and NO. As expected, levels of TNF-\(\alpha\), IL-12, and NO exhibited a significant reduction in diabetic mice compared with the controls, however, IL-6 remained unchanged instead of being upregulated (Figure 2). The observed reduced cytokine expression was consistent with the decreased amount of intracellular GSH in diabetic Mp except IL-6, which would show an elevated production in OMP according to its definition [1].

There is clear evidence that Mp play an essential role in the provocation of autoimmune diabetes. In order to determine whether the OMP in diabetic mice are associated with aberrant Mp function, we examined the phagocytic ability of the peritoneal Mp using the neutral fuchsin uptake method. It was found that STZ treatment suppressed the phagocytosis activity of the peritoneal Mp markedly (Figure 3), though there was no marked difference between the early and the advanced phases of diabetes.

Besides the alteration in immune mediator release, we also observed evident morphological changes in cultured Mps. During culture (within 7 days), normal Mp increased in size and differentiated from a homogeneous round shape into a morphologically heterogeneous population of fusiform- and epithelioid-shaped Mp. But in diabetic mice, the majority of Mp remained undifferentiated (data not shown).
Figure 3. Spleen cell and thymus cell proliferation and cytokine secretion. (a) Spleen cell and thymus cell proliferation (assessed at 24 hours) in response to mitogen stimulation was affected by STZ treatment, and the stimulation indexes in the diabetic groups are all higher than the control, though not always significant. (b) Thymocytes or (c) splenocytes were stimulated with ConA (30 µg/mL) for 48 hours before collection of conditioned medium. IL-4 and IFN-γ concentrations following stimulation were determined by ELISA. (d) The periodic change of TH1/TH2 balance during disease progression.

The balance of TH1/TH2 responses has been recognized as a critical factor in the development of type 1 diabetes, and it is well established that diabetes is associated with the development of a pathogenic TH1 response [10]. However, it remains to be determined whether the TH1 response varies during disease progression, and whether the variation is concomitant with the change of Mp phenotype in (multiple low-dose streptozotocin) MLD-STZ-induced diabetes model. In the succeeding experiments, the TH1/TH2 balance of T cells in the spleen and thymus was analyzed. The unfractionated splenocytes or thymocytes were in vitro stimulated with ConA, and the proliferative response of cells and the cytokine levels were measured. After stimulation, the proliferative response of splenocytes or thymocytes was found to be higher in some sort than that of control mice (Figure 4a), accompanied by

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secretion of IL-4 and IFN-γ (Figure 4b, 3c). Though it appeared rather complicated in the cytokines change, the ratio of IFN-γ to IL-4 showed a similar tendency in the thymus in diabetic mice injected 72 hours earlier with liquid para.

**DISCUSSION**

In line with the increasing evidence claiming the central role of innate over adaptive immune responses, a paradigm has been proposed on the presence of functional heterogeneity of Mps. Murata et al have divided Mps at least into two activated states based on their icGSH [1], and the role of intracellular redox status of Mps in regulating T111/T112 balance has been discussed widely [2, 3, 4, 5, 6]. However, to the author’s knowledge, there is little information available in literature about the redox status of Mps in diabetic mice induced by MLD-STZ. In the present study, we conducted this study to determine whether the redox status of Mps and their cytokine production change during the development of diabetes in BALB/c mice and, additionally, if the change of the redox status of Mps is concomitant with the change of T111/T112 skewing in the spleen or the thymus. The results indicated that the peritoneal Mps were consistently skewed to OMP dominance during the disease progression. T111/T112 skewing in the spleen or the thymus. The results indicated that the peritoneal Mps were consistently skewed to OMP dominance during the disease progression, tending to produce declined cytokine expression of TNF-α, IL-12, and NO compared to those from nondiabetes controls. However, the diabetic OMP phenotype showed its abnormalities somehow characterized by its unchanged IL-6 levels and significantly reduced phagocytic activity compared with the defined OMP. On the other hand, the thymus cells and spleen cells exhibited a different T111/T112 balance shift, with a T111 type on the onset of diabetes, converting to T112 phenotypes several weeks later after diabetes occurred. The data reported here might imply that multifactor dysfunction, rather than aberrant OMP alone, may cooperate to precipitate T111/T112 imbalance of diabetic mice induced by MLD-STZ.

Streptozotocin (STZ), a potential source of oxidative stress, can penetrate into the organism generating NO and thus inducing genotoxicity [11]. It has appeared that STZ treatment generally induces an oxidative predominance in murine tissues, including liver, kidney, heart, lungs, spleen, brain, muscles, and pancreas, with decreased GSH values and increased lipid peroxidation [12]. We examined the redox status of the peritoneal Mps in MLD-STZ diabetes model and gained the similar results that OMP phenotype was dominant during the development of diabetes, consistent with the progressive decrease in GSH concentration of the serum (data not shown). This phenomenon is compatible with the findings of Carmen that a significant decline is observed in blood GSH content at the recent onset of type 1 diabetes patients, and progressive GSH depletion during diabetes evolution [13]. It is noteworthy that OMP induced by STZ treatment is not completely in accordance with the defined OMP in consideration of the rather unaltered IL-6 production. We presumed that Mps from diabetic mice might be damaged resulting from the toxic STZ, thus leading to an aberrant or “incomplete” OMP accompanied by the lower GSH content. Though the pathogenic mechanism of diabetic OMP remains elusive, one possibility is the forced infiltration of Mps into pancreas or other tissues by modulating the chemotaxis of Mps. The redox status of Mps is suggested to be a crucial determinant in the regulation of the chemokine system. Saccani et al have described that H2O2 and the GSH-depleting drug, buthionine sulfoximine, can increase CCR2, CCR5, and CXCR4 mRNA expression to different extents and the cell migration (3-fold) in response to macrophage inflammatory protein-Iβ and human monocytes [14]. Similar to it, a certain subset of Mps selectively produces C-C chemokines [15] and the redox status affects the expression of adhesion molecules critical for trans-endothelial migration of the inflammatory cell [16].

There is a growing body of evidence that aberrant cytokine production in Mps is part of a complex pathway mediating autoimmune diabetes. However, previous studies exploring inflammatory cytokine production by
Mp have not always reported consistent results [10, 17, 18]. Our data showed the generally reduced cytokine production of TNF-α, IL-12, and NO from diabetic OMP upon in vitro LPS (lipopolysaccharide) stimulation. However, in consideration of the increasing peritoneal Mp count in the diabetic mice, the total amount of each inflammatory mediator above is significantly higher than that of the control mice (data not shown). As larger amount of immune cells, including Mp, is also found in the islets of Langerhans in autoimmune diabetes, it is necessary to be determined whether the deficient cytokine expression itself or its relatively excessive production is relevant to the pathogenesis in diabetic Mp.

Deficiencies in phagocytosis have been associated with and may participate in the pathogenesis of both systemic and organ-specific autoimmune diseases [19]. In our animal model, we also found a significant decline in the phagocytosis activity of Mp, a possible outcome resulting from the oxidative status in diabetic Mp [20]. Yamada et al assessed the relationship between the proportion of oxidative peritoneal exudate cells (PEC) and the β-cell destruction, and suggested the proportion of the oxidative PEC as a novel marker of disease activity [21]. As clearance of dying cells is critical to the control of inflammation, engulfment of cells dying by apoptosis must occur prior to cell lysis to prevent the release of intracellular contents and possibly the generation of new antigens. So a defect in the phagocytosis may be contributory to the initiation of autoimmunity.

A number of studies have correlated diabetes with TH1 phenotype development in the MLD-STZ diabetic animal model [8, 22, 23, 24, 25]. However, similar to other models of type 1 diabetes, some reports argued against this oversimplification. Muller et al demonstrated that MLD-STZ stimulated the production of IL-4 and IL-10, but significantly reduced IFN-γ and TNF-α levels in islets of BALB/c mice of both genders [26]. Similarly, Sitasawad indicated that both IFN-γ and IL-4 mRNA expression increased in the MLD-STZ diabetic pancreas [27]. Ins-IFN-γ transgenic mice showed apparent resistance to the induction of severe diabetes after STZ treatment compared to the control BALB/c mice [28]. In our study, the Th polarization in the spleen or the thymus varied during disease progression, with TH1 phenotype dominant on the onset of disease converting to TH2 phenotype several weeks after diabetes occurred. The factors dictating polarization to TH1 phenotypes at the early stage of diabetes remain elusive. Also, it is uncertain whether the OMP participate in the conversion of TH1 to TH2 along with disease progression. Previous study in NOD mice showed a similar TH1/TH2 conversion while with a consistent sequential change of RMP/OMP skewing [2]. In contrast to the change style, another report suggested a converse TH1/TH2 skewing in NOD or BB model, as TH2-mediated attack is responsible for the early phase of IDDM, while TH1-driven responses are responsible for the persistent and sustained attacks [29]. These findings together may suggest that the TH1/TH2 shift contributes to a pathological process, which may be dictated by multiple factors, which would include the factors precipitating the disease, the local APC (antigen-presenting cell) function, and the genetic background of a given strain.

Besides the controversial TH1/TH2 polarization, there have been also conflicting reports regarding lymphocyte proliferative responses to mitogen either in NOD mice or in MLD-STZ-induced diabetic mice [9, 30, 31]. Here we showed a higher proliferative response of T cells (unfractionated splenocytes or thymocytes) in the diabetic mice, though not always significant, compared to the controls, in accordance with the exuberant cytokine production. One possibility is the toxic effect of STZ treatment on the subset of lymphocytes. Further experiments should be conducted to examine the expression of cell surface markers, especially the regulatory T-cell (Treg) marker of CD4+CD25+, for a loss in the number of Treg may contribute to the highly activated proliferative response. It is also under discussion whether the defective function in Mp or other APC might lead to the defective stimulation of Treg cells, thus affecting the proliferative response of lymphocytes [32]. Interestingly, a recent report suggested that the all of the thiol pool in T cells may play a role as an amplifying mechanism for TCR/CD3 signals in immune response [33]. It may thus provide a new insight into the abnormal lymphocyte functions under the oxidative stress in MLD-STZ-induced diabetes. To clarify this point, experiments exploring the alteration of lymphocyte function with altered iGSH in lymphocytes of the STZ-treated mice should be carried out in the future.

In conclusion, our results presented the progressive OMP phenotype in the STZ-induced diabetic mice, and the change of TH1/TH2 skewing during the disease progression in the thymus and spleen. The data here implied that OMP may be relevant in the development of type 1 diabetes, however, it is not likely the only factor regulating the TH1/TH2 balance in MLD-STZ-induced diabetic mice. Previous studies regarding the pathogenesis of oxidative stress mainly focus on the β-cell destruction by reactive oxygen species (ROS). However, our study showed that the oxidative stress induced by STZ treatment could convert the peritoneal Mp to OMP phenotype with a generalized decrease in cytokine production and defective function. In addition, significant difference was also observed in either the blastogenesis of the splenocytes or the cytokine expression profile in the thymus and the spleen during the development of diabetes. This implies that the STZ-treatment damage in the immune cells besides Mp mediated by oxidative stress might also involve in the pathogenesis of type 1 diabetes. In future studies it is needed to be elucidated that to which extent the abnormal immune cells contribute to the pathogenesis and whether the redox regulation of the immune cells could prevent the incidence of type 1 diabetes.
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