Review Article

Marine Diterpenoids as Potential Anti-Inflammatory Agents

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The inflammatory response is a highly regulated process, and its dysregulation can lead to the establishment of chronic inflammation and, in some cases, to death. Inflammation is the cause of several diseases, including rheumatoid arthritis, inflammatory bowel diseases, multiple sclerosis, and asthma. The search for agents inhibiting inflammation is a great challenge as the inflammatory response plays an important role in the defense of the host to infections. Marine invertebrates are exceptional sources of new natural products, and among those diterpenoids secondary metabolites exhibit notable anti-inflammatory properties. Novel anti-inflammatory diterpenoids, exclusively produced by marine organisms, have been identified and synthetic molecules based on those structures have been obtained. The anti-inflammatory activity of marine diterpenoids has been attributed to the inhibition of Nuclear Factor- κ B activation and to the modulation of arachidonic acid metabolism. However, more research is necessary to describe the mechanisms of action of these secondary metabolites. This review is a compilation of marine diterpenoids, mainly isolated from corals, which have been described as potential anti-inflammatory molecules.

1. Introduction

Inflammation is a complex biological response against pathogens or tissue damage characterized by vasodilation, increased blood flow, vascular permeability, and cellular extravasation [1]. Macrophages, mast cells, and dendritic cells, resident in the tissues, are the first cells of innate immunity that detect and recognize the pathogen and initiate the inflammatory response [1]. Acute inflammation is an early response in which innate immune cells such as polymorphonuclear cells and monocytes are recruited to the site of irritation and secrete inflammatory mediators (e.g., cytokines, chemokines, and free radicals), which amplify the response [2]. Chronic inflammation, in turn, is the long-term inflammatory process that occurs as a dysregulation of acute inflammation often due to extended exposure to the initial irritant, persistent injury, or autoimmune disease. Chronic inflammation is associated with many pathological diseases including cancer, autoimmune diseases, atherosclerosis, rheumatoid arthritis, asthma, and cardiovascular diseases [3–5].

The search for new anti-inflammatory agents is challenging due to the complexity of the inflammatory process and its role in host defense. However, the progress attained in understanding the mechanisms involved in inflammation has made the identification of new targets possible, opening the range of search for new compounds with potential therapeutic effects on acute or chronic inflammatory diseases. Several drug discovery and development programs are focused on the search for bioactive compounds obtained from natural sources. Many drugs used today for the treatment of several diseases have been developed from natural products. The studies in terrestrial organisms have been extended to the marine environment, a resource with an enormous potential for drug discovery [6–8].

In the world of natural products, terpenoids are one of the largest and most studied groups of molecules. Terpenoids are secondary metabolites containing a C₅ isoprene unit derived from a biosynthetic pathway based on mevalonate, which is essential for diverse cellular functions [9]. Terpenoids can be classified into hemi, mono, sesqui, di, sester, or tri based

on the number of isoprene C5 units. These compounds are found largely in higher plants, but also in lower invertebrates including marine organisms.

Diterpenoids, in particular, are a promising class of molecules of secondary metabolites with a range of activity including antiviral, antibacterial, antiparasite, anticancer, and anti-inflammatory [10]. Diterpenes are comprised of four isoprene units with the chemical structure $C_{20}H_{32}$. Several studies have demonstrated a variety of diterpenoid structures presenting anti-inflammatory capacity. This review discusses the potential anti-inflammatory role of several diterpenoids derived from marine organisms.

2. Current Anti-Inflammatory Drugs

There are several classes of anti-inflammatory drugs available today, including nonsteroidal anti-inflammatory drugs (NSAIDs), glucocorticoids, and immunomodulatory drugs. NSAIDs, including aspirin, ibuprofen, and naproxen, are a widely administered class of drugs used for anti-inflammatory and analgesic purposes. Drugs in the NSAID category differ significantly in structure, but all share common mechanisms of action. These drugs prevent the release of prostaglandins through the inhibition of cyclooxygenase (COX) by covalently modifying the enzyme or by competing with the substrate for the active site [11, 12]. Side effects of these engineered drugs, however, are often severe and range from gastric ulcers to kidney damage and death.

Two COX isozymes are encoded in the human genome, COX-1 and COX-2. COX-1 is expressed in nearly all organs and cells but is most prominent in the stomach and in platelets, whereas COX-2 is an inducible, inflammation-specific isoform and regulates the synthesis of prostaglandins during inflammation [13–15]. Prostaglandins modulate different immune cell types including macrophages, dendritic cells, and T and B lymphocytes, leading to pro- and anti-inflammatory effects. Prostaglandins have several functions, including augmenting the blood flow and vascular permeability, regulating the expression of cytokines by innate immune cells, and inducing the expression of costimulatory molecules [16].

When it was discovered that the gross reduction or elimination of prostaglandins achieved by nonselective inhibition of COX enzymes often resulted in gastric ulcers [17, 18], researchers sought drugs that selectively inhibited COX-2. The resulting set of COX-2 selective drugs are collectively called coxibs and exhibit much lower rates of gastric ulcers than nonselective COX inhibitors [19, 20]. Unfortunately, it was then discovered that many coxibs also carry a higher risk of cardiovascular events such as coronary heart disease, heart attack, and stroke [21].

Glucocorticoids (GCs) are steroidal hormones that are naturally produced by vertebrates and function to control inflammation [22]. Because of their native role, synthetic GCs have been produced and used to treat a variety of inflammatory-related diseases including asthma [23], inflammatory bowel disease [24], rheumatoid arthritis [25], and systemic lupus erythematosus [26]. Both synthetic and natural GCs act by binding to and activating the glucocorticoid

receptor (GR), a transcription factor that acts as an activator or repressor of several genes by direct binding with specific DNA sequences or by interfering with the transcriptional activity of other transcription factors [27]. Several mechanisms have been proposed to explain the inhibitory effect of GCs on the transcription of inflammatory genes. Many of them are related to the inhibition of Nuclear Factor- κ B (NF κ B) activation at different levels, including a direct physical association of GR with NF κ B and the induction of the expression of the regulatory protein I κ B α [28].

NF κ B is a constitutively expressed protein present in nearly all cell types. It has been implicated in the regulation of apoptosis genes, cell adhesion molecules, stress responses, cancer, immune system, and inflammatory responses [29, 30]. In inflammation, NF κ B regulates the transcription of inflammatory genes induced by a variety of intra- and extracellular stimuli. The activation of NF κ B and its translocation to the nucleus depends on the phosphorylation and degradation of the I κ B proteins [31, 32]. It has been shown that several NSAIDs also inhibit NF κ B activation independently of their effect on COX inhibition [33–36]. These agents include aspirin, salicylates, sulindac, and sulphasalazine. However, until now inhibitors of NF κ B with a comparable anti-inflammatory capacity as glucocorticoids have not been identified. Although glucocorticoids, like dexamethasone, prednisone, and hydrocortisone, are successful at treating many inflammatory based diseases, continued use may lead to adverse events such as bruising, cataracts, muscle weakness, skin changes, sleep disturbances, weight gain, or more severe side effects such as type II diabetes mellitus, osteoporosis, and psychiatric symptoms [37–39].

Immunomodulatory drugs, such as thalidomide and its analogs, are also inhibitors of NF κ B activation [40]. These drugs have anticancer, anti-inflammatory, and antiangiogenic actions by modulating the secretion of cytokines such as Tumor Necrosis Factor Alpha (TNF- α) and interleukins IL-6 and IL-12 [41–43]. It has been proposed that the anti-inflammatory effect of these drugs occurs by inhibition of I κ B degradation and downregulation of NF κ B DNA-binding activity. Inhibitors of TNF- α are also being used currently for the treatment of inflammatory diseases such as rheumatoid arthritis, Crohn's disease, and asthma [44]. These molecules act by inhibiting the binding of TNF- α to its receptor or by neutralizing the soluble and the membrane-bound forms of TNF- α [44, 45]. However, several adverse effects for these drugs have been described, such as heart failure, increased predisposition to infection, and exacerbation of latent tuberculosis [44, 46].

There are many natural remedies for inflammation and pain, such as curcumin and green tea, which act via similar mechanisms but exhibit limited, if any, unwanted side effects [47]. Curcumin, a compound found in turmeric, has also been described to confer anti-inflammatory effects through a combination of mechanisms including inhibition of COX-2, lipoxygenase, and the NF κ B pathway [48–51]. In addition to anti-inflammatory effects, curcumin has also been attributed with antitumor [52, 53], antiviral [54], and antibacterial [55] effects. Curcumin is being tested for efficacy in patients with ulcerative colitis [56–58].

TABLE 1: Anti-inflammatory effect of eunicellane diterpenoids.

	Families name	Biological source	COX-2	iNOS	Superoxide anion generation	Elastase release	Other
Krempfelins	<i>Cladiella kremphi</i>		B-C, D* [73] E*, G, I [77]		K, M [82] N, P [84]	
	Hirsutalins	<i>C. hirsuta</i>	B [71]	B-D, H [71] K [78]		N [86] S [87]	
	Cladieunicellins	<i>Cladiella</i> sp.			C-E [80] 6- <i>epi</i> F [81]	A, C-D [80] 6- <i>epi</i> F [81]	
	Klymollins	<i>Klyxum molle</i>	F-G [74]	C-H [74]	M [83]	M [83]	X [79]
	Klysimplexins	<i>K. simplex</i>	R-S [72]	J-N, R-S [75]			
	Klysimplexin sulfoxides	<i>K. simplex</i>	C [72]	A-C [72]			
	Simplexin	<i>K. simplex</i>	E [70]	A, D-E [70]			

Data refer to compounds with percentage of inhibition > 50% for COX-2 and iNOS and >25% for superoxide anion generation and elastase release.

*Percentage of inhibition 40–50%.

Epigallocatechin-3-gallate (EGCG) is the main component in green tea that is responsible for conferring not only anti-inflammatory effects but also antiviral [59, 60], antibacterial [61], and anticancer effects [62, 63]. The anti-inflammatory effects are achieved most notably through COX-2 inhibition at the RNA and protein level [64]. Interestingly, EGCG has not been found to have an effect on COX-1 expression.

3. Marine-Derived Diterpenoids as Anti-Inflammatory Compounds

De las Heras and Hortelano in 2009 compiled a comprehensive list of the most promising anti-inflammatory diterpenoids, almost all of which were extracted from plants [10]. In their compilation, they describe the mechanisms of action associated with inhibition of the NF κ B signaling pathway of most families of diterpenoids. Bioactive diterpenoids act in the NF κ B pathway by blocking a range of activities including DNA-binding, IKK complex activation, and I κ B phosphorylation. Clinical studies have shown that commercial extracts from medicinal plants that contain large concentrations of diterpenoids that inhibit the NF κ B pathway are effective in reducing symptoms of rheumatoid arthritis [65, 66]. These extracts have also been tested in the treatment of other autoimmune and inflammatory diseases showing efficacy with variable mild side effects [67–69]. In this regard we discuss different families of diterpenoids isolated from marine organisms with anti-inflammatory capacity. Several of those molecules are promising candidates for further anti-inflammatory drug development.

3.1. Eunicellane Diterpenoids. Anti-inflammatory activity for eunicellin-based diterpenoids has been reported in the last few years. This class of compounds is secondary metabolites that present the cladiellane skeleton with a C2-C9 or C2-C6 oxygen bridge. Eunicellin-based diterpenes include krempfelins, hirsutalins, klymollins, klysimplexin, klysimplexin

sulfoxide, simplexin, and cladieunicellin and have been isolated and identified from soft corals belonging to the genera *Cladiella* or *Klyxum*. Some of these compounds have shown the capacity to inhibit the upregulation of inducible nitric oxide synthases (iNOS), COX-2, or IL-6 proteins in RAW 246.7 macrophages stimulated with lipopolysaccharide (LPS) [70–79]. Other compounds of these types were identified as inhibitors of superoxide generation and elastase release by N-formyl-methionyl-leucyl-phenylalanine/cytochalasin B (FMLP/CB) induced human neutrophils [80–87]. The release of superoxide and elastase by immune cells, mainly neutrophils, is important for the killing of host invading microorganisms but also contributes to host tissue damage during a chronic inflammatory disease. The mechanisms of action by which these compounds exert their anti-inflammatory effect have not been elucidated yet. Interestingly, most of these compounds selectively influenced certain inflammatory responses without affecting others (Table 1 and Supplementary Material available online at <http://dx.doi.org/10.1155/2015/263543>).

Reports have proposed that an epoxy group on C-11/C-17 present in some members of the klymollins is important for the inhibitory activity on iNOS expression [74]. However, some compounds of this family, the klymollins F and G (Figure 1, e.g., 1), were significant inhibitors of both iNOS and COX-2, suggesting that the modulation of these enzymes might be due to the inhibition of a common molecule upstream in the signaling pathway that governs their expression. These two compounds, in addition to the epoxy, present a fatty acid residue at C-6 position attributing a micelle-like feature to the structure that might be important for membrane diffusion. Other authors have attributed the capacity of inhibiting elastase release and superoxide generation of klymollin M (2) to the presence of a phenylacetate group at C-6 (Figure 1) [83]. Comparing the structure of klymollin M with other eunicellin-based diterpenoid inhibitors of elastase release, it appears that the presence of a butyric acid at C-3, a common feature of these molecules, might be also important for this activity (Figure 1

FIGURE 1: Structures of klymollin G [1] and klymollin M [2]. The epoxy group at C-17 for klymollin G and phenylacetate group at C-6 are labeled.

FIGURE 2: Structures of briarellin S [3] and *seco*-briarellinone [4]. The opening of the 10-member ring and the presence of carbonyl groups in the *seco*-briarellinone that have been suggested as being responsible for the higher anti-inflammatory effect than briarellin S are labeled.

and Supplementary Material). Further studies are necessary to identify the structural components that play a role in the anti-inflammatory effect of these molecules and to describe their mechanisms of action.

Briarellins are another class of eunicellane diterpenoid. Most of the briarellins have been isolated from corals of the genera *Briareum* and *Pachyclavularia*. The anti-inflammatory activity of this family has been little explored. Our group recently showed that briarellin S (3) inhibits the production of nitric oxide (NO) by primary murine macrophages stimulated with LPS. This effect was smaller than the one observed when cells were exposed to LPS in the presence of *seco*-briarellinone (4). Differences in the IC_{50} of briarellin S (20.3 μ M) and *seco*-briarellinone (4.7 μ M) might be due to the opening of the 10-member ring and the presence of carbonyl groups in the *seco*-briarellinone, which is the main structural difference with the briarellin S [88] (Figure 2). The ester moiety present in the molecule of briarellin S could also be interfering with the activity of this compound. Structural modifications of these molecules would give a clue about the groups responsible for the anti-inflammatory effect.

3.2. Briarane Diterpenoids. Briarane diterpenoids form a family of compounds that present a basic chemical structure of a [8.4.0] bicycle carbon skeleton with most members containing a γ -lactone moiety. These compounds have been exclusively isolated from soft corals belonging to the order Gorgonacea (reviewed by [89]) and genera including *Briareum*, *Dichotella*, *Junceella*, and *Verrucella*. Around 600 briarane diterpenoids have been identified with a variety of

bioactivities including antimicrobial, cytotoxic, and in some cases anti-inflammatory effects. Briaranes, such as frajunolides, juncenolides, and the briarenolides, are inhibitors of the superoxide generation and elastase released by human neutrophils stimulated with FMLP/CB [90–98].

Compounds isolated from *Junceella juncea*, the juncenolides, have shown moderate inhibition in the release of elastase [99] and juncenol has presented weak inhibitory effects on neutrophil superoxide generation [100, 101]. However, neither the mechanisms of action nor the structural components involved in these differences in anti-inflammatory activity have been described. The inhibitory effect of briarane compounds on COX-2 and iNOS expression induced by LPS in macrophages has been also reported [102, 103]. Table 2 shows a compilation of briarane diterpenoids with anti-inflammatory properties.

Excavatolide B (BrD1) (5), a briarane diterpenoid isolated from the coral *Briareum excavatum*, demonstrates *in vitro* and *in vivo* anti-inflammatory activity [104]. This compound inhibited vascular permeability and edema and decreased the expression of iNOS, COX-2, and matrix metalloproteinase (MMP-9) when topically applied in the skin of mice with 12-O-tetradecanoylphorbol-13-acetate- (TPA-) induced dermatitis. This effect might occur by a mechanism involving the inhibition of NF κ B and Akt activation observed in the skin of the animals. Comparing the effect on IL-6 secretion induced by LPS in bone marrow derived dendritic cells (BMDC) of different briarane diterpenoids isolated from the same coral and semisynthetic analogs of BrD1, the authors concluded that 8,17-epoxide and 12-hydroxyl groups are essential for the inhibition of IL-6 secretion by BrD1 [104] (Figure 3).

TABLE 2: Anti-inflammatory effect of briarane diterpenoids.

	Families name	Biological source	iNOS	Superoxide anion generation	Elastase release
Frajunolide	<i>Junceella fragilis</i>		P-Q [97]	P-Q [97]
	Juncenolide	<i>J. juncea</i>		H [92] O [99]	N-O [99]
	Junceol	<i>J. juncea</i>		A-C [101] E [100]	
	Briarenolides	<i>Briareum</i> sp.	K-L [103]	F [95] I [96] J [98]	E [94] F [95] J [98]

Data refer to compounds with percentage of inhibition > 50% for iNOS and >25% superoxide anion generation and elastase release.

FIGURE 3: Structure of excavatolide B [5]. The 8,17-epoxide and 12-hydroxyl groups that have been suggested as being responsible for the anti-inflammatory effect are marked.

3.3. Cembrane Diterpenoids. Cembranes are a large family of diterpenoids isolated from terrestrial and marine organisms that exhibit a range of biological activities including antibacterial, antitumor, anti-inflammatory, and antiviral effects [105]. The basic structure of cembrane diterpenoids is constituted by a common 14-membered carbocyclic skeleton and usually presents cyclic ether, lactone, or furan moieties around this nucleus (reviewed by [106]). Unconventional cembranoids with 12-, 13-, or 14-membered variants have also been described [107, 108]. Cembranoids from marine organisms are mainly isolated from corals of the genera *Sinularia*, *Lobophytum*, *Eunicea*, and *Sarcophyton*.

Anti-inflammatory activity for different groups of cembrane diterpenoids has been reported. Cembranoids such as gibberosenes, grandilobatin, querciformolides, sarcocrascolides, crassumolides, crassarines, sinularolides, durumolides, and columnariols have shown a capacity to inhibit the expression of iNOS and/or COX-2 by LPS-stimulated RAW 264.7 cells [109–121] (Table 3 and Supplementary Material). The presence of a α -methylene- γ -lactone in cembranoids has been suggested to be essential for the inhibition of iNOS expression [119] (Figure 4, e.g., 6).

Some cembranoids have been identified as modulators of NF κ B signaling pathway [122–126]. Compounds from the crassumolide and laevigatol groups have shown dose-dependent inhibitory effects on the mRNA expression of

iNOS and COX-2 induced by TNF- α in HepG2 cells by a mechanism that involved the inhibition of NF κ B transcriptional activation [123, 124]. The cembrane lobohedleolide (6) isolated from *Sarcophyton* sp. showed inhibitory activity on the production of TNF- α in LPS-stimulated RAW 264.7 cells [127]. This effect was later attributed to the ability of this compound to inhibit the degradation of I κ B α and the binding of NF κ B to the DNA [122]. However, lobohedleolide also induced an increase in the production of IL-8 in LPS-stimulated THP-1 cells through the activation of the IL-8 promoter region [122]. High levels of IL-8 have been found in some human cancers and have been associated with tumor progression and metastasis [128–130]. Thus, the identification of new anti-inflammatory molecules must be accompanied by a rigorous description of the mechanisms involved in the effect. Considering the pharmacological properties of lobohedleolide in the inhibition of NF κ B pathway, synthetic analogs could be produced with structural modifications that might favor the anti-inflammatory properties.

Members of the cembrane diterpenoids, lobocrasols isolated from *Lobophytum crassum*, have also shown inhibitory activity on NF κ B activation in TNF- α stimulated HepG2 with consequent decreases in COX-2 and iNOS gene expression [125]. The presence of an epoxy group at C-1/C-15 in the active compounds appears to be essential for the anti-inflammatory effect (Figure 4, e.g., 7). Cembrane sinumaximols B and C isolated from *Sinularia maxima* were identified as potent inhibitors of IL-12 secretion by dendritic cells stimulated with LPS [131]. This activity could be attributed to the lactone moiety present in these molecules. Later, it was demonstrated that the sinumaximols A, B, and G inhibited the transcriptional activity of NF κ B induced by TNF- α in HepG2 cells and the expression of the intracellular adhesion molecule (ICAM-1) and iNOS [126]. Authors suggested that hydroxyl groups at C-7 and/or C-8 are responsible for the anti-inflammatory activity of these compounds. One of those compounds, sinumaximol B (8), exhibited inhibitory activity in both dendritic and HepG2 cells (Figure 4). It is important to note that only sinumaximol B contains the lactone and the hydroxyl at C-7 and C-8.

3.4. Diterpene Glycosides. Marine diterpene glycosides are derivatives exclusively produced by Gorgonian corals [132].

TABLE 3: Anti-inflammatory effect of cembrane diterpenoids.

	Families name	Biological source	COX-2	iNOS	NFκB
Cembrane diterpenoids	Crassarines	<i>Sinularia crassa</i>		H [118]	
	Grandilobatins	<i>S. grandilobata</i>		D [110]	
	Querciformolides	<i>S. querciformis</i>	C [111]	C [111] E [115]	
	Sinumaximols	<i>S. maxima</i>			A-C, G, I [126]
	Sarcocrassocolides	<i>Sarcophyton crassocaule</i>	I [117] Q [120]	A-D [116] F-L [117] M-O [119] P-R [120]	
	Crassocolides	<i>S. crassocaule</i>	A, E [120]	A-B, D-E [120]	
	Crassumolides	<i>Lobophytum crassum</i>	A, C [112]	A-C, F [112]	
	Crassumols	<i>L. crassum</i>			E [124]
	Lobocrasols	<i>L. crassum</i>			A-C [125]
	Durumolides	<i>L. durum</i>	C [113] F [114]	A-E [113] F-L [114]	
	Laevigatols	<i>L. laevigatum</i>			A-B [123]
	Columnariols	<i>Nephthea columnaris</i> (cultured coral)	A-B [121]	A-B [121]	

Data refer to compounds with percentage of inhibition > 50% for COX-2 and iNOS and IC₅₀ values < 50 μM in NFκB.

FIGURE 4: Cembrane diterpenoids: lobohedleolide [6], lobocrasol A [7], and sinumaximol B [8]. The presence of a α -methylene- γ -lactone in lobohedleolide (and cembranolides); an epoxy group at C-1/C-15 in lobocrasol A and a hydroxyl group at C-7 and/or C-8 in sinumaximol B are labeled.

A diterpene aglycone core and a carbohydrate moiety characterize this class of compounds. Among the marine diterpenes glycosides, eleutherobins, fuscoidins, and pseudopterogens are the most studied compounds [132]. The pseudopterogens (Ps) have been described as molecules with important anti-inflammatory and analgesic properties and were the first to be isolated from *Pseudopterogorgia elisabethae* [133, 134]. Pseudopterogen A (9) was identified as a potent anti-inflammatory agent, with a greater effect than the NSAID indomethacin, in the phorbol myristate acetate- (PMA-) induced topical inflammation animal model [133]. Pseudopterogen A also inhibited prostaglandin E₂ and leukotriene C₄ secretion in zymosan-stimulated murine peritoneal macrophages [135]. This molecule inhibited phagosome formation and triggered intracellular calcium release by a mechanism that involved its binding to a G protein coupled receptor [136]. Other pseudopterogens with exceptional anti-inflammatory activity also have been identified [137, 138] and are suggested to

inhibit the synthesis of leukotrienes and the degranulation of human neutrophils [135, 137] (Table 4 and Supplementary Material). Several analogs of Ps such as *seco*-pseudopterogens and amphilectosins reduced the mouse ear edema induced by different inflammatory stimuli [138, 139] and the levels of myeloperoxidase at the inflammation site [139].

Due to the relevant anti-inflammatory properties of Ps, they have attracted great attention from the organic chemistry community and new synthetic pseudopterogens have been obtained. Discussions of Ps syntheses are out of the scope of this review but they can be found elsewhere [reviewed by [132]]. It appears that the location and identity of carbohydrate moiety are not relevant for the anti-inflammatory activity; instead, the intact diterpene glycoside is needed for the Ps biological effect [140]. However, nonglycosylated compounds structurally related to the aglycone component of Ps, such as elisabethadione (10) and elisabethatrienol (11), have shown anti-inflammatory activity

TABLE 4: Glycosides diterpenes.

Families name	Biological source	Bioactive compounds
Pseudopterosins	<i>Pseudopterogorgia elisabethae</i>	Pseudopterosins A [133], E [137], Q [134], P, T, U [139]
Fuscosides	<i>Eunicea fusca</i>	Fuscosides A-B [145]
Eleutherobins	<i>Eunicea</i> sp.	Calyculaglycoside B [149]

Data refer to glycosides diterpenoids with anti-inflammatory activity.

FIGURE 5: Pseudopterosin A [9], elisabethadione [10], and elisabethatrienol [11]. Glycoside diterpene, pseudopterosin A, nonglycoside diterpenes, elisabethadione, and elisabethatrienol.

[138, 139] (Figure 5). Simplified structural analogs of the Ps and *seco*-Ps have been synthesized, which conserve the anti-inflammatory effect, suggesting that a more accessible aglycone would be sufficient for the activity [141, 142]. A semisynthetic derivative of pseudopterosin A maintaining the anti-inflammatory capacity has been obtained [143]. Due to their anti-inflammatory properties natural extracts from *P. elisabethae* rich in pseudopterosins are used in commercial skin care products [144].

Fuscosides have been isolated from the coral *Eunicea fusca*. Fuscosides A and B exhibit anti-inflammatory activity [145, 146]. Both compounds, when topically applied, reduce PMA-induced edema in mouse ears by inhibiting neutrophil infiltration. Fuscoside B inhibits the synthesis of leukotriene C_4 in calcium ionophore-activated murine macrophages [145, 146]. It was demonstrated using cultures of human leukocytes that fuscoidin B is a selective inhibitor of 5-lipoxygenase [147] (Table 4 and Supplementary Material). The aglycone precursor of fuscoidin B, the fuscoidol, and other compounds as eunicol and the analogous eunicidiol, isolated from *E. fusca*, have also shown anti-inflammatory activity by reducing the edema induced by PMA in mouse ear [148]. Different approaches for the synthesis of naturally occurring fuscosides, conserving the anti-inflammatory capacity, have been attempted unsuccessfully.

Other members of the diterpene glycosides compounds have also shown anti-inflammatory activity. A calyculaglycoside isolated from *Eunicea* sp. exhibited topical anti-inflammatory activity in two *in vivo* assays, and it was suggested as a nonselective inhibitor of the 5-lipoxygenase and COX pathways [149]. It is relevant to note that compounds belonging to this family have the same aglycone (dilophol) and only differ in the identity of the carbohydrate moieties. Anti-inflammatory activity has not been reported for the eleutherobin compounds; however, two nonglycoside compounds, the valdivones A and B, which are related

to the eleutherobin aglycone, inhibited chemically induced inflammation in mouse ear [150]. These findings question the relevance of carbohydrate moiety for the biological activity of glycoside compounds.

3.5. Other Diterpenoids. Pseudopteranes are only found in corals of the genera *Pseudopterogorgia*. Their ring system could be originated from a ring contraction reaction of a cembrane precursor [151]. Pseudopterolide 1 was the first compound identified and isolated from *Pseudopterogorgia acerosa* [152]. Other pseudopterane compounds include kallolides and isogorgiacerodiol isolated from *P. kallos* and *P. acerosa*, respectively [153, 154]. Pseudopterolide 1 and some kallolides have shown anti-inflammatory capacity in topical skin inflammation induced by PMA [152, 153]. Our group has recently demonstrated that pseudopterolide 1 derivative (12) also exhibits anti-inflammatory capacity. This compound inhibited the secretion and/or mRNA expression of a variety of inflammatory mediators (TNF- α , IL-6, NO, IP-10, iNOS, COX-2, and MCP-1) induced by TNF- α and ligands of TLRs in mouse peritoneal macrophages [155]. This effect was due to the capacity of this compound to inhibit I κ B α phosphorylation and the subsequent activation of NF κ B. The compound also inhibited the expression of macrophages activation markers such as CD80 and CD86 suggesting a role in the modulation of a variety of processes occurring during macrophage activation. The methoxyl group at C-9 appears to be important in the anti-inflammatory effect as it was more potent than the isogorgiacerodiol pseudopterane (13), which has a hydroxyl group at the same position [155] (Figure 6). It is notable how subtle structural differences in small molecules are essential for modifying their immune modulation activities.

Smaller groups of diterpenoids called verticillane-based and norditerpenoids, isolated from coral of the genera

FIGURE 6: Pseudopterolide derivative [12] and isogorgiacerodiol [13]. The methoxyl group at C-9 in pseudopterolide derivative that has been suggested as being responsible for the higher anti-inflammatory effect than isogorgiacerodiol is labeled.

Cespitularia and *Sinularia*, respectively, have been recently identified to have anti-inflammatory capacity [156, 157]. It has been reported that members of these families, for example, cespitularin (verticillane-based diterpenoid), isolated from *C. hypotentaculata* and a series of norcembranolid, gyrosanolides, and other norditerpenoids isolated from *S. gyrosa*, inhibit the expression of iNOS in LPS-stimulated RAW 264.7 cells [156, 157].

The neorogioltriol, a tricyclic brominated diterpenoid isolated from the red algae *Laurencia glandulifera*, showed anti-inflammatory effects *in vitro* and *in vivo* [158]. This compound inhibited the activation of NF κ B and the production of TNF- α , COX-2, and NO in RAW 264.7 macrophages stimulated with LPS. The systemic administration of neorogioltriol reduced the edema formation in an animal model of carrageenan-induced local inflammation.

Dolabellane diterpenoids have been isolated mainly from plants but are also present in marine organisms. These compounds have a 5,11-bicyclic skeleton and exhibit antiviral, antiprotozoa, and antibacterial properties [159]. Recently, it has been suggested that the dolabelladienetriol, isolated from the brown marine alga *Dictyota paffii*, downregulates the production of TNF- α and NO through the inhibition of NF κ B activation in *Leishmania amazonensis* infected and uninfected macrophages, conferring an anti-inflammatory activity to this compound [160]. To our knowledge, this is the first report of anti-inflammatory capacity described for a marine-derived dolabellane diterpenoid.

4. Conclusions

Many efforts have been made to identify new anti-inflammatory molecules from natural sources. Terrestrial organisms are commonly used in traditional medicine to treat inflammatory diseases and have often been ascribed diterpenoid compounds to the anti-inflammatory effects. Marine invertebrates are exceptional sources of new molecules with therapeutic potential including diterpenoids secondary metabolites, which exhibit notable anti-inflammatory properties.

The anti-inflammatory capacity of some diterpenoids isolated from marine organisms is due to the inhibition of the

NF κ B signaling pathway at different levels [122, 155]. NF κ B plays a crucial role in regulating the inflammatory responses and in the development of various human pathological conditions. Hence, this transcription factor constitutes a suitable target for the development of new anti-inflammatory drugs. Moreover, some marine diterpenoids have been shown to be inhibitors of prostaglandins and leukotrienes secretion and in some cases found to be selective inhibitors of 5-lipoxygenase and COX enzymes [147, 149]. Together, this evidence demonstrates that marine diterpenoids show a capacity of inhibiting different pathways involved in inflammation, supporting their potential for anti-inflammatory drugs development. However, little is known about the molecular mechanisms involved in the anti-inflammatory characteristics of marine diterpenoids. Thus, further studies are necessary to better understand their mechanisms of action.

The largest limitation for the study of natural products is the small amount of compounds that are obtained and the variations on their production that are influenced by the environmental changes to which marine organisms are exposed. Due to the potential applications of coral-derived compounds, coral aquaculture has been proposed as a way to establish a stable supply of bioactive materials for the extraction of natural products [161]. Some laboratories use this approach for the production of marine invertebrates with bioprospecting purposes. Importantly, natural growth rate of these organisms is not enough to sustain pharmaceutical exploitation. Many researchers have developed new strategies for the synthesis of compounds that conserve the biological activities of their natural analogs; nonetheless, it remains a challenging area.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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References

- [1] C. N. Serhan and J. Savill, "Resolution of inflammation: the beginning programs the end," *Nature Immunology*, vol. 6, no. 12, pp. 1191–1197, 2005.
- [2] C. N. Serhan, S. D. Brain, C. D. Buckley et al., "Resolution of inflammation: state of the art, definitions and terms," *The FASEB Journal*, vol. 21, no. 2, pp. 325–332, 2007.
- [3] L. M. Coussens and Z. Werb, "Inflammation and cancer," *Nature*, vol. 420, no. 6917, pp. 860–867, 2002.
- [4] P. Libby, P. M. Ridker, and A. Maseri, "Inflammation and atherosclerosis," *Circulation*, vol. 105, no. 9, pp. 1135–1143, 2002.
- [5] P. Libby, "Inflammation and cardiovascular disease mechanisms," *The American Journal of Clinical Nutrition*, vol. 83, no. 2, pp. 456S–460S, 2006.
- [6] R. A. Medina, D. E. Goeger, P. Hills et al., "Coibamide A, a potent antiproliferative cyclic depsipeptide from the panamanian marine cyanobacterium *Leptolyngbya* sp.," *Journal of the American Chemical Society*, vol. 130, no. 20, pp. 6324–6325, 2008.
- [7] C. C. Hughes, J. B. MacMillan, S. P. Gaudêncio, P. R. Jensen, and W. Fenical, "The ammosamides: structures of cell cycle modulators from a marine-derived *Streptomyces* species," *Angewandte Chemie International Edition*, vol. 48, no. 4, pp. 725–727, 2009.
- [8] T. P. Kondratyuk, E.-J. Park, R. Yu et al., "Novel marine phenazines as potential cancer chemopreventive and anti-inflammatory agents," *Marine Drugs*, vol. 10, no. 2, pp. 451–464, 2012.
- [9] J. L. Goldstein and M. S. Brown, "Regulation of the mevalonate pathway," *Nature*, vol. 343, no. 6257, pp. 425–430, 1990.
- [10] B. de las Heras and S. Hortelano, "Molecular basis of the anti-inflammatory effects of terpenoids," *Inflammation and Allergy—Drug Targets*, vol. 8, no. 1, pp. 28–39, 2009.
- [11] J. R. Vane and R. M. Botting, "Anti-inflammatory drugs and their mechanism of action," *Inflammation Research*, vol. 47, supplement 2, pp. S78–S87, 1998.
- [12] O. Llorens, J. J. Perez, A. Palomer, and D. Mauleon, "Differential binding mode of diverse cyclooxygenase inhibitors," *Journal of Molecular Graphics & Modelling*, vol. 20, no. 5, pp. 359–371, 2002.
- [13] W. L. Xie, J. G. Chipman, D. L. Robertson, R. L. Erikson, and D. L. Simmons, "Expression of a mitogen-responsive gene encoding prostaglandin synthase is regulated by mRNA splicing," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 88, no. 7, pp. 2692–2696, 1991.
- [14] S. H. Lee, E. Soyoola, P. Chanmugam et al., "Selective expression of mitogen-inducible cyclooxygenase in macrophages stimulated with lipopolysaccharide," *The Journal of Biological Chemistry*, vol. 267, no. 36, pp. 25934–25938, 1992.
- [15] C. S. Williams, M. Mann, and R. N. DuBois, "The role of cyclooxygenases in inflammation, cancer, and development," *Oncogene*, vol. 18, no. 55, pp. 7908–7916, 1999.
- [16] E. Ricciotti and G. A. FitzGerald, "Prostaglandins and inflammation," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 31, no. 5, pp. 986–1000, 2011.
- [17] T. D. Warner, F. Giuliano, I. Vojnovic, A. Bukasa, J. A. Mitchell, and J. R. Vane, "Nonsteroid drug selectivities for cyclo-oxygenase-1 rather than cyclo-oxygenase-2 are associated with human gastrointestinal toxicity: a full *in vitro* analysis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 96, no. 13, pp. 7563–7568, 1999.
- [18] J. L. Wallace, W. McKnight, B. K. Reuter, and N. Vergnolle, "NSAID-induced gastric damage in rats: requirement for inhibition of both cyclooxygenase 1 and 2," *Gastroenterology*, vol. 119, no. 3, pp. 706–714, 2000.
- [19] C.-C. Chan, S. Boyce, C. Brideau et al., "Rofecoxib [vioxx, MK-0966; 4-(4'-methylsulfonylphenyl)-3-phenyl-2-(5H)-furanone]: a potent and orally active cyclooxygenase-2 inhibitor. Pharmacological and biochemical profiles," *The Journal of Pharmacology and Experimental Therapeutics*, vol. 290, no. 2, pp. 551–560, 1999.
- [20] J. L. Goldstein, F. E. Silverstein, N. M. Agrawal et al., "Reduced risk of upper gastrointestinal ulcer complications with celecoxib, a novel COX-2 inhibitor," *The American Journal of Gastroenterology*, vol. 95, no. 7, pp. 1681–1690, 2000.
- [21] W. A. Ray, C. M. Stein, J. R. Daugherty, K. Hall, P. G. Arbogast, and M. R. Griffin, "COX-2 selective non-steroidal anti-inflammatory drugs and risk of serious coronary heart disease," *The Lancet*, vol. 360, no. 9339, pp. 1071–1073, 2002.
- [22] J. P. Tuckermann, A. Kleiman, K. G. McPherson, and H. M. Reichardt, "Molecular mechanisms of glucocorticoids in the control of inflammation and lymphocyte apoptosis," *Critical Reviews in Clinical Laboratory Sciences*, vol. 42, no. 1, pp. 71–104, 2005.
- [23] V. M. Keatings, A. Jatakanon, Y. M. Worsdell, and P. J. Barnes, "Effects of inhaled and oral glucocorticoids on inflammatory indices in asthma and COPD," *The American Journal of Respiratory and Critical Care Medicine*, vol. 155, no. 2, pp. 542–548, 1997.
- [24] P. Zakrojsky, W. Thai, R. C. Deaño et al., "Steroid exposure, acute coronary syndrome, and inflammatory bowel disease: insights into the inflammatory milieu," *The American Journal of Medicine*, vol. 128, no. 3, pp. 303–311, 2015.
- [25] M. Ibañez, A. M. Ortiz, I. Castrejón et al., "A rational use of glucocorticoids in patients with early arthritis has a minimal impact on bone mass," *Arthritis Research & Therapy*, vol. 12, no. 2, article R50, 2010.
- [26] M. Mosca, C. Tani, L. Carli, and S. Bombardieri, "Glucocorticoids in systemic lupus erythematosus," *Clinical and Experimental Rheumatology*, vol. 29, supplement 68, no. 5, pp. S126–S129, 2011.
- [27] L. I. McKay and J. A. Cidlowski, "Molecular control of immune/inflammatory responses: interactions between nuclear factor- κ B and steroid receptor-signaling pathways," *Endocrine Reviews*, vol. 20, no. 4, pp. 435–459, 1999.
- [28] K. De Bosscher, W. Vanden Berghe, and G. Haegeman, "The interplay between the glucocorticoid receptor and nuclear factor- κ B or activator protein-1: molecular mechanisms for gene repression," *Endocrine Reviews*, vol. 24, no. 4, pp. 488–522, 2003.
- [29] M. S. Hayden and S. Ghosh, "Signaling to NF- κ B," *Genes & Development*, vol. 18, no. 18, pp. 2195–2224, 2004.
- [30] M. Karin, "Nuclear factor- κ B in cancer development and progression," *Nature*, vol. 441, no. 7092, pp. 431–436, 2006.
- [31] H. Häcker and M. Karin, "Regulation and function of IKK and IKK-related kinases," *Science's STKE*, vol. 2006, no. 357, p. rel3, 2006.

- [32] A. Oeckinghaus, M. S. Hayden, and S. Ghosh, "Crosstalk in NF- κ B signaling pathways," *Nature Immunology*, vol. 12, no. 8, pp. 695–708, 2011.
- [33] J. W. Pierce, M. A. Read, H. Ding, F. W. Luscinskas, and T. Collins, "Salicylates inhibit I κ B- α phosphorylation, endothelial-leukocyte adhesion molecule expression, and neutrophil transmigration," *The Journal of Immunology*, vol. 156, no. 10, pp. 3961–3969, 1996.
- [34] M.-J. Yin, Y. Yamamoto, and R. B. Gaynor, "The anti-inflammatory agents aspirin and salicylate inhibit the activity of I(kappa)B kinase-beta," *Nature*, vol. 396, no. 6706, pp. 77–80, 1998.
- [35] Y. Yamamoto, M.-J. Yin, K.-M. Lin, and R. B. Gaynor, "Sulindac inhibits activation of the NF- κ B pathway," *The Journal of Biological Chemistry*, vol. 274, no. 38, pp. 27307–27314, 1999.
- [36] K. S. Berman, U. N. Verma, G. Harburg, J. D. Minna, M. H. Cobb, and R. B. Gaynor, "Sulindac enhances tumor necrosis factor- α -mediated apoptosis of lung cancer cell lines by inhibition of nuclear factor- κ B," *Clinical Cancer Research*, vol. 8, no. 2, pp. 354–360, 2002.
- [37] A. D. Adinoff and J. R. Hollister, "Steroid-induced fractures and bone loss in patients with asthma," *The New England Journal of Medicine*, vol. 309, no. 5, pp. 265–268, 1983.
- [38] J. R. Curtis, A. O. Westfall, J. Allison et al., "Population-based assessment of adverse events associated with long-term glucocorticoid use," *Arthritis Care & Research*, vol. 55, no. 3, pp. 420–426, 2006.
- [39] W. Ericson-Neilsen and A. D. Kaye, "Steroids: pharmacology, complications, and practice Delivery Issues," *The Ochsner Journal*, vol. 14, no. 2, pp. 203–207, 2014.
- [40] M. Karin, Y. Yamamoto, and Q. M. Wang, "The IKK NF- κ B system: a treasure trove for drug development," *Nature Reviews Drug Discovery*, vol. 3, no. 1, pp. 17–26, 2004.
- [41] J. A. Keifer, D. C. Guttridge, B. P. Ashburner, and A. S. Baldwin Jr., "Inhibition of NF- κ B activity by thalidomide through suppression of I κ B kinase activity," *The Journal of Biological Chemistry*, vol. 276, no. 25, pp. 22382–22387, 2001.
- [42] S. Majumdar, B. Lamothe, and B. B. Aggarwal, "Thalidomide suppresses NF-kappa B activation induced by TNF and H₂O₂, but not that activated by ceramide, lipopolysaccharides, or phorbol ester," *The Journal of Immunology*, vol. 168, no. 6, pp. 2644–2651, 2002.
- [43] N. Mitsiades, C. S. Mitsiades, V. Poulaki et al., "Apoptotic signaling induced by immunomodulatory thalidomide analogs in human multiple myeloma cells: therapeutic implications," *Blood*, vol. 99, no. 12, pp. 4525–4530, 2002.
- [44] M. A. Palladino, F. R. Bahjat, E. A. Theodorakis, and L. L. Moldawer, "Anti-TNF-alpha therapies: the next generation," *Nature Reviews Drug Discovery*, vol. 2, no. 9, pp. 736–746, 2003.
- [45] A. T. Paul, V. M. Gohil, and K. K. Bhutani, "Modulating TNF- α signaling with natural products," *Drug Discovery Today*, vol. 11, no. 15-16, pp. 725–732, 2006.
- [46] S. J. Bickston, G. R. Lichtenstein, K. O. Arseneau, R. B. Cohen, and F. Cominelli, "The relationship between infliximab treatment and lymphoma in Crohn's disease," *Gastroenterology*, vol. 117, no. 6, pp. 1433–1437, 1999.
- [47] C. D. Lao, M. T. Ruffin, D. Normolle et al., "Dose escalation of a curcuminoid formulation," *BMC Complementary and Alternative Medicine*, vol. 6, article 10, 2006.
- [48] C. V. Rao, "Regulation of COX and LOX by curcumin," *Advances in Experimental Medicine and Biology*, vol. 595, pp. 213–226, 2007.
- [49] M. Shakibaei, T. John, G. Schulze-Tanzil, I. Lehmann, and A. Mobasheri, "Suppression of NF- κ B activation by curcumin leads to inhibition of expression of cyclo-oxygenase-2 and matrix metalloproteinase-9 in human articular chondrocytes: implications for the treatment of osteoarthritis," *Biochemical Pharmacology*, vol. 73, no. 9, pp. 1434–1445, 2007.
- [50] C. Csaki, A. Mobasheri, and M. Shakibaei, "Synergistic chondroprotective effects of curcumin and resveratrol in human articular chondrocytes: inhibition of IL-1beta-induced NF-kappaB-mediated inflammation and apoptosis," *Arthritis Research and Therapy*, vol. 11, no. 6, article R165, 2009.
- [51] S. C. Gupta, J. H. Kim, S. Prasad, and B. B. Aggarwal, "Regulation of survival, proliferation, invasion, angiogenesis, and metastasis of tumor cells through modulation of inflammatory pathways by nutraceuticals," *Cancer and Metastasis Reviews*, vol. 29, no. 3, pp. 405–434, 2010.
- [52] T. Kawamori, R. Lubet, V. E. Steele et al., "Chemopreventive effect of curcumin, a naturally occurring anti-inflammatory agent, during the promotion/progression stages of colon cancer," *Cancer Research*, vol. 59, no. 3, pp. 597–601, 1999.
- [53] A.-L. Chen, C.-H. Hsu, J.-K. Lin et al., "Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions," *Anticancer Research*, vol. 21, no. 4, pp. 2895–2900, 2001.
- [54] Y. Lv, Z. An, H. Chen, Z. Wang, and L. Liu, "Mechanism of curcumin resistance to human cytomegalovirus in HELF cells," *BMC Complementary and Alternative Medicine*, vol. 14, article 284, 2014.
- [55] J. A. Cho and E. Park, "Curcumin utilizes the anti-inflammatory response pathway to protect the intestine against bacterial invasion," *Nutrition Research and Practice*, vol. 9, no. 2, pp. 117–122, 2015.
- [56] P. R. Holt, S. Katz, and R. Kirshoff, "Curcumin therapy in inflammatory bowel disease: a pilot study," *Digestive Diseases and Sciences*, vol. 50, no. 11, pp. 2191–2193, 2005.
- [57] H. Hanai, T. Iida, K. Takeuchi et al., "Curcumin maintenance therapy for ulcerative colitis: randomized, multicenter, double-blind, placebo-controlled trial," *Clinical Gastroenterology and Hepatology*, vol. 4, no. 12, pp. 1502–1506, 2006.
- [58] S. Kumar, V. Ahuja, M. J. Sankar, A. Kumar, and A. C. Moss, "Curcumin for maintenance of remission in ulcerative colitis," *Cochrane Database of Systematic Reviews*, vol. 10, Article ID CD008424, 2012.
- [59] J. M. Weber, A. Ruzindana-Umunyana, L. Imbeault, and S. Sircar, "Inhibition of adenovirus infection and adenain by green tea catechins," *Antiviral Research*, vol. 58, no. 2, pp. 167–173, 2003.
- [60] J.-M. Song, K.-H. Lee, and B.-L. Seong, "Antiviral effect of catechins in green tea on influenza virus," *Antiviral Research*, vol. 68, no. 2, pp. 66–74, 2005.
- [61] K. Kono, I. Tatara, S. Takeda, K. Arakawa, and Y. Hara, "Antibacterial activity of epigallocatechin gallate against methicillin-resistant *Staphylococcus aureus*," *Kansenshogaku Zasshi. The Journal of the Japanese Association for Infectious Diseases*, vol. 68, no. 12, pp. 1518–1522, 1994.
- [62] J. Jankun, S. H. Selman, R. Swiercz, and E. Skrzypczak-Jankun, "Why drinking green tea could prevent cancer," *Nature*, vol. 387, no. 6633, p. 561, 1997.
- [63] Y.-C. Wang and U. Bachrach, "The specific anti-cancer activity of green tea (–)-epigallocatechin-3-gallate (EGCG)," *Amino Acids*, vol. 22, no. 2, pp. 131–143, 2002.

- [64] T. Hussain, S. Gupta, V. M. Adhami, and H. Mukhtar, "Green tea constituent epigallocatechin-3-gallate selectively inhibits COX-2 without affecting COX-1 expression in human prostate carcinoma cells," *International Journal of Cancer*, vol. 113, no. 4, pp. 660–669, 2005.
- [65] R. A. Burgos, J. L. Hancke, J. C. Bertoglio et al., "Efficacy of an *Andrographis paniculata* composition for the relief of rheumatoid arthritis symptoms: a prospective randomized placebo-controlled trial," *Clinical Rheumatology*, vol. 28, no. 8, pp. 931–946, 2009.
- [66] Q.-W. Lv, W. Zhang, Q. Shi et al., "Comparison of *Tripterygium wilfordii* Hook F with methotrexate in the treatment of active rheumatoid arthritis (TRIFRA): a randomised, controlled clinical trial," *Annals of the Rheumatic Diseases*, vol. 74, no. 6, pp. 1078–1086, 2015.
- [67] J. T. Coon and E. Ernst, "*Andrographis paniculata* in the treatment of upper respiratory tract infections: a systematic review of safety and efficacy," *Planta Medica*, vol. 70, no. 4, pp. 293–298, 2004.
- [68] J. Ren, Q. Tao, X. Wang, Z. Wang, and J. Li, "Efficacy of T2 in active Crohn's disease: a prospective study report," *Digestive Diseases and Sciences*, vol. 52, no. 8, pp. 1790–1797, 2007.
- [69] A. M. Brinker, J. Ma, P. E. Lipsky, and I. Raskin, "Medicinal chemistry and pharmacology of genus *Tripterygium* (Celastraceae)," *Phytochemistry*, vol. 68, no. 6, pp. 732–766, 2007.
- [70] S.-L. Wu, J.-H. Su, Z.-H. Wen et al., "Simplexins A-I, eunicellin-based diterpenoids from the soft coral *Klyxum simplex*," *Journal of Natural Products*, vol. 72, no. 6, pp. 994–1000, 2009.
- [71] B.-W. Chen, S.-M. Chang, C.-Y. Huang et al., "Hirsutalins A-H, eunicellin-based diterpenoids from the soft coral *Cladiella hirsuta*," *Journal of Natural Products*, vol. 73, no. 11, pp. 1785–1791, 2010.
- [72] B.-W. Chen, C.-H. Chao, J.-H. Su, Z.-H. Wen, P.-J. Sung, and J.-H. Sheu, "Anti-inflammatory eunicellin-based diterpenoids from the cultured soft coral *Klyxum simplex*," *Organic & Biomolecular Chemistry*, vol. 8, no. 10, pp. 2363–2366, 2010.
- [73] C.-J. Tai, J.-H. Su, M.-S. Huang, Z.-H. Wen, C.-F. Dai, and J.-H. Sheu, "Bioactive eunicellin-based diterpenoids from the soft coral *Cladiella krempfi*," *Marine Drugs*, vol. 9, no. 10, pp. 2036–2045, 2011.
- [74] F.-J. Hsu, B.-W. Chen, Z.-H. Wen et al., "Klymollins A-H, bioactive eunicellin-based diterpenoids from the formosan soft coral *Klyxum molle*," *Journal of Natural Products*, vol. 74, no. 11, pp. 2467–2471, 2011.
- [75] B.-W. Chen, C.-H. Chao, J.-H. Su et al., "Klysimplexins I-T, eunicellin-based diterpenoids from the cultured soft coral *Klyxum simplex*," *Organic and Biomolecular Chemistry*, vol. 9, no. 3, pp. 834–844, 2011.
- [76] S.-L. Wu, J.-H. Su, C.-Y. Huang et al., "Simplexins P-S, eunicellin-based diterpenes from the soft coral *Klyxum simplex*," *Marine Drugs*, vol. 10, no. 6, pp. 1203–1211, 2012.
- [77] C.-J. Tai, J.-H. Su, C.-Y. Huang et al., "Cytotoxic and anti-inflammatory eunicellin-based diterpenoids from the soft coral *Cladiella krempfi*," *Marine Drugs*, vol. 11, no. 3, pp. 788–799, 2013.
- [78] B.-W. Chen, S.-Y. Wang, C.-Y. Huang, S.-L. Chen, Y.-C. Wu, and J.-H. Sheu, "Hirsutalins I-M, eunicellin-based diterpenoids from the soft coral *Cladiella hirsuta*," *Tetrahedron*, vol. 69, no. 10, pp. 2296–2301, 2013.
- [79] F.-Y. Chang, F.-J. Hsu, C.-J. Tai, W.-C. Wei, N.-S. Yang, and J.-H. Sheu, "Klymollins T-X, bioactive eunicellin-based diterpenoids from the soft coral *Klyxum molle*," *Marine Drugs*, vol. 12, no. 5, pp. 3060–3071, 2014.
- [80] Y.-H. Chen, C.-Y. Tai, Y.-H. Kuo et al., "Cladieunicellins A-E, new eunicellins from an Indonesian soft coral *Cladiella* sp.," *Chemical & Pharmaceutical Bulletin*, vol. 59, no. 3, pp. 353–358, 2011.
- [81] Y.-H. Chen, T.-L. Hwang, Y.-D. Su et al., "New 6-hydroxyeunicellins from a soft coral *Cladiella* sp.," *Chemical & Pharmaceutical Bulletin*, vol. 60, no. 1, pp. 160–163, 2012.
- [82] Y.-N. Lee, C.-J. Tai, T.-L. Hwang, and J.-H. Sheu, "Krempfielins J-M, new eunicellin-based diterpenoids from the soft coral *Cladiella krempfi*," *Marine Drugs*, vol. 11, no. 8, pp. 2741–2750, 2013.
- [83] M.-C. Lin, B.-W. Chen, C.-Y. Huang, C.-F. Dai, T.-L. Hwang, and J.-H. Sheu, "Eunicellin-based diterpenoids from the formosan soft coral *Klyxum molle* with inhibitory activity on superoxide generation and elastase release by neutrophils," *Journal of Natural Products*, vol. 76, no. 9, pp. 1661–1667, 2013.
- [84] Y.-N. Lee, C.-J. Tai, T.-L. Hwang, and J.-H. Sheu, "Krempfielins N-P, new anti-inflammatory eunicellins from a Taiwanese soft coral *Cladiella krempfi*," *Marine Drugs*, vol. 12, no. 2, pp. 1148–1156, 2014.
- [85] C.-J. Tai, U. Chokkalingam, Y. Cheng et al., "Krempfielins Q and R, two new eunicellin-based diterpenoids from the soft coral *Cladiella krempfi*," *International Journal of Molecular Sciences*, vol. 15, no. 12, pp. 21865–21874, 2014.
- [86] T.-Z. Huang, B.-W. Chen, C.-Y. Huang, T.-L. Hwang, C.-F. Dai, and J.-H. Sheu, "Eunicellin-based diterpenoids, hirsutalins N-R, from the formosan soft coral *Cladiella hirsuta*," *Marine Drugs*, vol. 12, no. 5, pp. 2446–2457, 2014.
- [87] T.-Z. Huang, B.-W. Chen, C.-Y. Huang et al., "Eunicellin-based diterpenoids, hirsutalins S-V, from the formosan soft coral *Cladiella hirsuta*," *Marine Drugs*, vol. 13, no. 5, pp. 2757–2769, 2015.
- [88] J. F. Gómez-Reyes, A. Salazar, H. M. Guzmán et al., "secobriarellinone and briarellin S, two new eunicellin-based diterpenoids from the panamanian octocoral *Briareum asbestinum*," *Marine Drugs*, vol. 10, no. 11, pp. 2608–2617, 2012.
- [89] J.-H. Sheu, Y.-H. Chen, Y.-H. Chen et al., "Briarane diterpenoids isolated from gorgonian corals between 2011 and 2013," *Marine Drugs*, vol. 12, no. 4, pp. 2164–2181, 2014.
- [90] Y.-C. Shen, Y.-H. Chen, T.-L. Hwang, J.-H. Guh, and A. T. Khalil, "Four new briarane diterpenoids from the gorgonian coral *Junceella fragilis*," *Helvetica Chimica Acta*, vol. 90, no. 7, pp. 1391–1398, 2007.
- [91] C.-C. Liaw, Y.-C. Shen, Y.-S. Lin, T.-L. Hwang, Y.-H. Kuo, and A. T. Khalil, "Frajunolides E-K, briarane diterpenes from *Junceella fragilis*," *Journal of Natural Products*, vol. 71, no. 9, pp. 1551–1556, 2008.
- [92] S.-S. Wang, Y.-H. Chen, J.-Y. Chang et al., "Juncenolides H-K, new briarane diterpenoids from *Junceella juncea*," *Helvetica Chimica Acta*, vol. 92, no. 10, pp. 2092–2100, 2009.
- [93] C.-C. Liaw, Y.-H. Kuo, Y.-S. Lin, T.-L. Hwang, and Y.-C. Shen, "Frajunolides L-O, four new 8-Hydroxybriarane diterpenoids from the Gorgonian *Junceella fragilis*," *Marine Drugs*, vol. 9, no. 9, pp. 1477–1486, 2011.
- [94] P.-H. Hong, Y.-D. Su, N.-C. Lin et al., "Briarenolide E: the first 2-ketobriarane diterpenoid from an octocoral *Briareum* sp. (Briareidae)," *Tetrahedron Letters*, vol. 53, no. 14, pp. 1710–1712, 2012.
- [95] P.-H. Hong, Y.-D. Su, J.-H. Su et al., "Briarenolides F and G, new briarane diterpenoids from a *Briareum* sp. octocoral," *Marine Drugs*, vol. 10, no. 5, pp. 1156–1168, 2012.

- [96] Y.-D. Su, T.-L. Hwang, N.-C. Lin et al., "Briarenolides H and I: new 8-hydroxybriarane diterpenoids from a formosan octocoral *Briareum* sp. (Briareidae)," *Bulletin of the Chemical Society of Japan*, vol. 85, no. 9, pp. 1031–1036, 2012.
- [97] C.-C. Liaw, Y.-C. Lin, Y.-S. Lin, C.-H. Chen, T.-L. Hwang, and Y.-C. Shen, "Four new briarane diterpenoids from Taiwanese Gorgonian *Junceella fragilis*," *Marine Drugs*, vol. 11, no. 6, pp. 2042–2053, 2013.
- [98] Y.-D. Su, C.-H. Cheng, W.-F. Chen et al., "Briarenolide J, the first 12-chlorobriarane diterpenoid from an octocoral *Briareum* sp. (Briareidae)," *Tetrahedron Letters*, vol. 55, no. 44, pp. 6065–6067, 2014.
- [99] J.-Y. Chang, C.-C. Liaw, A. E. Fazary, T.-L. Hwang, and Y.-C. Shen, "New briarane diterpenoids from the gorgonian coral *Junceella juncea*," *Marine Drugs*, vol. 10, no. 6, pp. 1321–1330, 2012.
- [100] P.-J. Sung, C.-H. Pai, T.-L. Hwang et al., "Junceols D-H, new polyoxygenated briaranes from sea whip gorgonian coral *Junceella juncea* (Ellisellidae)," *Chemical & Pharmaceutical Bulletin*, vol. 56, no. 9, pp. 1276–1281, 2008.
- [101] P.-J. Sung, C.-H. Pai, Y.-D. Su et al., "New 8-hydroxybriarane diterpenoids from the gorgonians *Junceella juncea* and *Junceella fragilis* (Ellisellidae)," *Tetrahedron*, vol. 64, no. 19, pp. 4224–4232, 2008.
- [102] A. Bahl, S. M. Jachak, K. Palaniveloo, T. Ramachandram, C. S. Vairappan, and H. K. Chopra, "2-acetoxyverecynarmin C, a new briarane COX inhibitory diterpenoid from *Pennatula aculeata*," *Natural Product Communications*, vol. 9, no. 8, pp. 1139–1141, 2014.
- [103] Y.-D. Su, T.-R. Su, Z.-H. Wen et al., "Briarenolides K and L, new anti-inflammatory briarane diterpenoids from an octocoral *Briareum* sp. (briareidae)," *Marine Drugs*, vol. 13, no. 2, pp. 1037–1050, 2015.
- [104] W.-C. Wei, S.-Y. Lin, Y.-J. Chen et al., "Topical application of marine briarane-type diterpenes effectively inhibits 12-O-tetradecanoylphorbol-13-acetate-induced inflammation and dermatitis in murine skin," *Journal of Biomedical Science*, vol. 18, no. 1, article 94, 2011.
- [105] B. Yang, X.-F. Zhou, X.-P. Lin et al., "Cembrane diterpenes chemistry and biological properties," *Current Organic Chemistry*, vol. 16, no. 12, pp. 1512–1539, 2012.
- [106] W.-C. Wei, P.-J. Sung, C.-Y. Duh, B.-W. Chen, J.-H. Sheu, and N.-S. Yang, "Anti-inflammatory activities of natural products isolated from soft corals of Taiwan between 2008 and 2012," *Marine Drugs*, vol. 11, no. 10, pp. 4083–4126, 2013.
- [107] Z. Xi, W. Bie, W. Chen et al., "Sarcophyolides B–E, new cembranoids from the soft coral *Sarcophyton elegans*," *Marine Drugs*, vol. 11, no. 9, pp. 3186–3196, 2013.
- [108] S.-K. Wang, M.-K. Hsieh, and C.-Y. Duh, "New diterpenoids from soft coral *Sarcophyton ehrenbergi*," *Marine Drugs*, vol. 11, no. 11, pp. 4318–4327, 2013.
- [109] A. F. Ahmed, Z.-H. Wen, J.-H. Su et al., "Oxygenated cembranoids from a Formosan soft coral *Sinularia gibberosa*," *Journal of Natural Products*, vol. 71, no. 2, pp. 179–185, 2008.
- [110] A. F. Ahmed, S.-H. Tai, Z.-H. Wen et al., "A C-3 methylated isocembranoid and 10-oxocembranoids from a formosan soft coral, *Sinularia grandilobata*," *Journal of Natural Products*, vol. 71, no. 6, pp. 946–951, 2008.
- [111] Y. Lu, C.-Y. Huang, Y.-F. Lin et al., "Anti-inflammatory cembranoids from the soft corals *Sinularia querciformis* and *Sinularia granosa*," *Journal of Natural Products*, vol. 71, no. 10, pp. 1754–1759, 2008.
- [112] C.-H. Chao, Z.-H. Wen, Y.-C. Wu, H.-C. Yeh, and J.-H. Sheu, "Cytotoxic and anti-inflammatory cembranoids from the soft coral *Lobophytum crassum*," *Journal of Natural Products*, vol. 71, no. 11, pp. 1819–1824, 2008.
- [113] S.-Y. Cheng, Z.-H. Wen, S.-F. Chiou et al., "Durumolides A–E, anti-inflammatory and antibacterial cembranoids from the soft coral *Lobophytum durum*," *Tetrahedron*, vol. 64, no. 41, pp. 9698–9704, 2008.
- [114] S.-Y. Cheng, Z.-H. Wen, S.-K. Wang et al., "Anti-inflammatory cembranoids from the soft coral *Lobophytum durum*," *Bioorganic & Medicinal Chemistry*, vol. 17, no. 11, pp. 3763–3769, 2009.
- [115] Y. Lu, J.-H. Su, C.-Y. Huang et al., "Cembranoids from the soft corals *Sinularia granosa* and *Sinularia querciformis*," *Chemical & Pharmaceutical Bulletin*, vol. 58, no. 4, pp. 464–466, 2010.
- [116] W.-Y. Lin, J.-H. Su, Y. Lu et al., "Cytotoxic and anti-inflammatory cembranoids from the Dongsha Atoll soft coral *Sarcophyton crassocaule*," *Bioorganic & Medicinal Chemistry*, vol. 18, no. 5, pp. 1936–1941, 2010.
- [117] W.-Y. Lin, Y. Lu, J.-H. Su et al., "Bioactive cembranoids from the dongsha atoll soft coral *Sarcophyton crassocaule*," *Marine Drugs*, vol. 9, no. 6, pp. 994–1006, 2011.
- [118] C.-H. Chao, K.-J. Chou, C.-Y. Huang et al., "Bioactive cembranoids from the soft coral *Sinularia crassa*," *Marine Drugs*, vol. 9, no. 10, pp. 1955–1968, 2011.
- [119] W.-Y. Lin, Y. Lu, B.-W. Chen et al., "Sarcocrassocolides M–O, bioactive cembranoids from the Dongsha Atoll soft coral *Sarcophyton crassocaule*," *Marine Drugs*, vol. 10, no. 3, pp. 617–626, 2012.
- [120] W.-Y. Lin, B.-W. Chen, C.-Y. Huang et al., "Bioactive cembranoids, sarcocrassocolides P–R, from the Dongsha Atoll soft coral *Sarcophyton crassocaule*," *Marine Drugs*, vol. 12, no. 2, pp. 840–850, 2014.
- [121] T.-H. Hsiao, C.-S. Sung, Y.-H. Lan et al., "Anti-inflammatory cembranes from the cultured soft coral *Nephthea columnaris*," *Marine Drugs*, vol. 13, no. 6, pp. 3443–3453, 2015.
- [122] T. Oda, W. Wewengkang, M. M. Kapojos, R. P. Mangindaan, J.-S. Lee, and M. Namikoshi, "Lobohedleolide induces interleukin-8 production in LPS-stimulated human monocytic cell line THP-1," *International Journal of Applied Research in Natural Products*, vol. 4, no. 3, pp. 16–21, 2011.
- [123] T. H. Quang, T. T. Ha, C. V. Minh et al., "Cytotoxic and anti-inflammatory cembranoids from the Vietnamese soft coral *Lobophytum laevigatum*," *Bioorganic and Medicinal Chemistry*, vol. 19, no. 8, pp. 2625–2632, 2011.
- [124] N. X. Cuong, N. P. Thao, B. T. T. Luyen et al., "Cembranoid diterpenes from the soft coral *lobophytum crassum* and their anti-inflammatory activities," *Chemical & Pharmaceutical Bulletin*, vol. 62, no. 2, pp. 203–208, 2014.
- [125] N. P. Thao, B. T. T. Luyen, N. T. T. Ngan et al., "New anti-inflammatory cembranoid diterpenoids from the Vietnamese soft coral *Lobophytum crassum*," *Bioorganic & Medicinal Chemistry Letters*, vol. 24, no. 1, pp. 228–232, 2014.
- [126] N. P. Thao, N. H. Nam, N. X. Cuong et al., "Inhibition of NF- κ B transcriptional activation in HepG2 cells by diterpenoids from the soft coral *Sinularia maxima*," *Archives of Pharmacological Research*, vol. 37, no. 6, pp. 706–712, 2014.
- [127] M. M. Kapojos, J.-S. Lee, T. Oda et al., "Two unprecedented cembrene-type terpenes from an Indonesian soft coral *sarcophyton* sp.," *Tetrahedron*, vol. 66, no. 3, pp. 641–645, 2010.

- [128] B. König, F. Steinbach, B. Janocha et al., "The differential expression of proinflammatory cytokines IL-6, IL-8 and TNF- α in renal cell carcinoma," *Anticancer Research*, vol. 19, no. 2 C, pp. 1519–1524, 1999.
- [129] G. Galfy, K. A. Mohammed, P. A. Dowling, N. Nasreen, M. J. Ward, and V. B. Antony, "Interleukin 8: an autocrine growth factor for malignant mesothelioma," *Cancer Research*, vol. 59, no. 2, pp. 367–371, 1999.
- [130] A. R. Green, V. L. Green, M. C. White, and V. Speirs, "Expression of cytokine messenger RNA in normal and neoplastic human breast tissue: identification of interleukin-8 as a potential regulatory factor in breast tumours," *International Journal of Cancer*, vol. 72, no. 6, pp. 937–941, 1997.
- [131] N. P. Thao, N. H. Nam, N. X. Cuong et al., "Diterpenoids from the soft coral *Sinularia maxima* and their inhibitory effects on lipopolysaccharide-stimulated production of pro-inflammatory cytokines in bone marrow-derived dendritic cells," *Chemical & Pharmaceutical Bulletin*, vol. 60, no. 12, pp. 1581–1589, 2012.
- [132] F. Berru, M. W. B. McCulloch, and R. G. Kerr, "Marine diterpene glycosides," *Bioorganic & Medicinal Chemistry*, vol. 19, no. 22, pp. 6702–6719, 2011.
- [133] S. A. Look, W. Fenical, G. K. Matsumoto, and J. Clardy, "The pseudopterogens: a new class of antiinflammatory and analgesic diterpene pentosides from the marine sea whip *Pseudopterogorgia elisabethae* (Octocorallia)," *The Journal of Organic Chemistry*, vol. 51, no. 26, pp. 5140–5145, 1986.
- [134] I. I. Rodrguez, Y.-P. Shi, O. J. Garca et al., "New pseudopterogen and seco-pseudopterogen diterpene glycosides from two Colombian isolates of *Pseudopterogorgia elisabethae* and their diverse biological activities," *Journal of Natural Products*, vol. 67, no. 10, pp. 1672–1680, 2004.
- [135] A. M. S. Mayer, P. B. Jacobson, W. Fenical, R. S. Jacobs, and K. B. Glaser, "Pharmacological characterization of the pseudopterogens: novel anti-inflammatory natural products isolated from the caribbean soft coral, *Pseudopterogorgia elisabethae*," *Life Sciences*, vol. 62, no. 26, pp. 401–407, 1998.
- [136] C. E. Moya and R. S. Jacobs, "Pseudopterogen A inhibits phagocytosis and alters intracellular calcium turnover in a pertussis toxin sensitive site in *Tetrahymena thermophila*," *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, vol. 143, no. 4, pp. 436–443, 2006.
- [137] V. Roussis, Z. Wu, W. Fenical, S. A. Strobel, G. D. Van Duyne, and J. Clardy, "New antiinflammatory pseudopterogens from the marine octocoral *Pseudopterogorgia elisabethae*," *The Journal of Organic Chemistry*, vol. 55, no. 16, pp. 4916–4922, 1990.
- [138] A. Ata, R. G. Kerr, C. E. Moya, and R. S. Jacobs, "Identification of anti-inflammatory diterpenes from the marine gorgonian *Pseudopterogorgia elisabethae*," *Tetrahedron*, vol. 59, no. 23, pp. 4215–4222, 2003.
- [139] H. Correa, A. L. Valenzuela, L. F. Ospina, and C. Duque, "Anti-inflammatory effects of the gorgonian *Pseudopterogorgia elisabethae* collected at the Islands of Providencia and San Andrs (SW Caribbean)," *Journal of Inflammation*, vol. 6, article 5, 2009.
- [140] W. Zhong, C. Moya, R. S. Jacobs, and R. D. Little, "Synthesis and an evaluation of the bioactivity of the C-glycoside of pseudopterogen A methyl ether," *The Journal of Organic Chemistry*, vol. 73, no. 18, pp. 7011–7016, 2008.
- [141] V. M. Tanis, C. Moya, R. S. Jacobs, and R. D. Little, "Synthesis and evaluation of the bioactivity of simplified analogs of the seco-pseudopterogens; progress toward determining a pharmacophore," *Tetrahedron*, vol. 64, no. 47, pp. 10649–10663, 2008.
- [142] F. Flachsmann, K. Schellhaas, C. E. Moya, R. S. Jacobs, and W. Fenical, "Synthetic pseudopterogen analogues: a novel class of antiinflammatory drug candidates," *Bioorganic & Medicinal Chemistry*, vol. 18, no. 23, pp. 8324–8333, 2010.
- [143] D. S. Scherl, J. Afflitto, and A. Gaffar, "Influence of OAS-1000 on mediators of inflammation," *Journal of Clinical Periodontology*, vol. 26, no. 4, pp. 246–251, 1999.
- [144] A. Kijjoa and P. Sawangwong, "Drugs and cosmetics from the sea," *Marine Drugs*, vol. 2, no. 2, pp. 73–82, 2004.
- [145] J. Shin and W. Fenical, "Fuscoides A-D: antiinflammatory diterpenoid glycosides of new structural classes from the Caribbean gorgonian *Eunicea fusca*," *The Journal of Organic Chemistry*, vol. 56, no. 9, pp. 3153–3158, 1991.
- [146] P. B. Jacobson and R. S. Jacobs, "Fuscoid: an anti-inflammatory marine natural product which selectively inhibits 5-lipoxygenase. Part I: physiological and biochemical studies in murine inflammatory models," *Journal of Pharmacology and Experimental Therapeutics*, vol. 262, no. 2, pp. 866–873, 1992.
- [147] P. B. Jacobson and R. S. Jacobs, "Fuscoid: an anti-inflammatory marine natural product which selectively inhibits 5-lipoxygenase. Part II: biochemical studies in the human neutrophil," *Journal of Pharmacology and Experimental Therapeutics*, vol. 262, no. 2, pp. 874–882, 1992.
- [148] D. H. Marchbank, F. Berru, and R. G. Kerr, "Eunicidiol, an anti-inflammatory dilophol diterpene from *Eunicea fusca*," *Journal of Natural Products*, vol. 75, no. 7, pp. 1289–1293, 2012.
- [149] O. M. Cbar, A. D. Rodrguez, O. L. Padilla, and J. A. Snchez, "The calyculaglycosides: dilophol-type diterpene glycosides exhibiting antiinflammatory activity from the Caribbean gorgonian *Eunicea* sp.," *The Journal of Organic Chemistry*, vol. 62, no. 21, pp. 7183–7188, 1997.
- [150] Y. Lin, C. A. Bewley, and D. J. Faulkner, "The valdivones, anti-inflammatory diterpene esters from the South African soft coral alcyonium valdivae," *Tetrahedron*, vol. 49, no. 36, pp. 7977–7984, 1993.
- [151] W. Fenical, "Marine soft corals of the genus pseudopterogorgia: a resource for novel anti-inflammatory diterpenoids," *Journal of Natural Products*, vol. 50, no. 6, pp. 1001–1008, 1987.
- [152] M. M. Bandurraga, W. Fenical, S. F. Donovan, and J. Clardy, "Pseudopterolide, an irregular diterpenoid with unusual cytotoxic properties from the Caribbean sea whip *Pseudopterogorgia acerosa* (Pallas) (Gorgonacea)," *Journal of the American Chemical Society*, vol. 104, no. 23, pp. 6463–6465, 1982.
- [153] S. A. Look, M. T. Burch, W. Fenical, Z. Qi-tai, and J. Clardy, "Kallolide A, a new antiinflammatory diterpenoid, and related lactones from the Caribbean octocoral *Pseudopterogorgia kallos* (Bielschowsky)," *The Journal of Organic Chemistry*, vol. 50, no. 26, pp. 5741–5746, 1985.
- [154] W. F. Tinto, L. John, W. F. Reynolds, and S. McLean, "Novel pseudopteroids of *Pseudopterogorgia acerosa*," *Tetrahedron*, vol. 47, no. 41, pp. 8679–8686, 1991.
- [155] Y. Gonzlez, D. Doens, R. Santamara et al., "A pseudopterane diterpene isolated from the octocoral *Pseudopterogorgia acerosa* inhibits the inflammatory response mediated by TLR-ligands and TNF- α in macrophages," *PLoS ONE*, vol. 8, no. 12, Article ID e84107, 2013.
- [156] S.-Y. Cheng, E.-H. Lin, Z.-H. Wen, M. Y.-N. Chiang, and C.-Y. Duh, "Two new verticillane-type diterpenoids from the formosan soft coral *Cespitularia hypotentaculata*," *Chemical & Pharmaceutical Bulletin*, vol. 58, no. 6, pp. 848–851, 2010.

- [157] S.-Y. Cheng, C.-T. Chuang, Z.-H. Wen et al., "Bioactive norditerpenoids from the soft coral *Sinularia gyrosa*," *Bioorganic & Medicinal Chemistry*, vol. 18, no. 10, pp. 3379–3386, 2010.
- [158] R. Chatter, R. B. Othman, S. Rabhi et al., "In vivo and in vitro anti-inflammatory activity of neorogioltriol, a new diterpene extracted from the red algae *Laurencia glandulifera*," *Marine Drugs*, vol. 9, no. 7, pp. 1293–1306, 2011.
- [159] J. P. Barbosa, R. C. Pereira, J. L. Abrantes et al., "In vitro antiviral diterpenes from the Brazilian brown alga *Dictyota pfaffii*," *Planta Medica*, vol. 70, no. 9, pp. 856–860, 2004.
- [160] D. C. Soares, T. C. Calegari-Silva, U. G. Lopes et al., "Dolabella-dienetriol, a compound from *Dictyota pfaffii* algae, inhibits the infection by *Leishmania amazonensis*," *PLoS Neglected Tropical Diseases*, vol. 6, no. 9, Article ID e1787, 2012.
- [161] M. C. Leal, R. Calado, C. Sheridan, A. Alimonti, and R. Osinga, "Coral aquaculture to support drug discovery," *Trends in Biotechnology*, vol. 31, no. 10, pp. 555–561, 2013.