

Research Article

Involvement of the IL23/Th17 Pathway in the Pathogenesis of Tunisian Pemphigus Foliaceus

M. Ben Jmaa ¹, O. Abida,¹ R. Fakhfakh,¹ E. Bahloul,² Kh. Sellami,² L. Gaddour,³ N. Elloumi,¹ M. Ben Ayed,¹ A. Masmoudi,² M. Dhouib,⁴ M. Abdelmoula,⁴ N. Mahfoudh,³ H. Makni,³ H. Turki,² and H. Masmoudi¹

¹Immunology Department, Habib Bourguiba Hospital, University of Sfax, Sfax, Tunisia

²Dermatology Department, Hedi Chaker Hospital, University of Sfax, Sfax, Tunisia

³Immunology Department, Hedi Chaker Hospital, University of Sfax, Sfax, Tunisia

⁴Maxillofacial Surgery Department, Habib Bourguiba Hospital, Sfax, Tunisia

Correspondence should be addressed to M. Ben Jmaa; mariem.benjmaa@yahoo.fr

Received 15 March 2018; Revised 9 May 2018; Accepted 5 June 2018; Published 12 July 2018

Academic Editor: Carlo Cervellati

Copyright © 2018 M. Ben Jmaa et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Pemphigus foliaceus (PF) is a rare autoimmune skin disease caused by anti-Dsg1 pathogenic autoantibodies. It is considered as a Th2-mediated disease. Likewise, Th17 cells were recently described in the pathogenesis of the disease but their role is still unclear. We aimed to unravel the eventual implication of the IL23/Th17 pathway in the development of PF. A case-control study was conducted on 115 PF patients and 201 healthy controls using PCR-RFLP and AS-PCR methods. SNPs in *IL23R*, *RORγt*, *IL17A*, *IL17F*, *IL17AR*, *TNFA*, and *STAT3* genes were genotyped. mRNA expression of *IL23R* and *RORγt* was evaluated using Q-PCR. The frequency of circulating Th17 cells was analyzed by flow cytometry. Genetic associations between *IL23R*>rs11209026, *IL17A*>rs3748067, *IL17F*>rs763780, and *TNFA*>rs1800629 and the susceptibility to PF were reported. Moreover, we revealed a significant increased frequency of circulating CD4⁺IL17⁺ cells as well as higher mRNA levels of *RORγt* and *IL23R* in PBMCs of patients. However, no significant increase of *RORγt* and *IL23R* mRNA expression was observed in lesional skin biopsies. In spite of the little size of specimens, our results provide converging arguments for the contribution of the IL23/Th17 pathway in the pathogenesis of PF.

1. Introduction

Pemphigus foliaceus (PF) is an autoimmune skin disease with a complex etiology involving genetic and environmental factors [1]. It is mediated by pathogenic auto-antibodies (–Abs) that bind to desmoglein 1 (Dsg1) and lead to acantholysis and consequent blister formation in the subcorneal layer of the epidermis [2–4]. The progression from pre-clinical phase to clinical active disease is associated with the subclass switching of these auto-Abs from IgG1/IgG2 to IgG4 [5–7]. Therefore, and since Th2 cytokines induce B cells to secrete IgG4, PF was considered as a Th2 disease [8, 9]. However, this hypothesis was rapidly contradicted by other researchers who detected Dsg1-responsive Th1 and Th2 cells simultaneously in PF patients' blood [10]. More recently, a

significant high frequency of CD4⁺IL17⁺ cells in pemphigus patients' PBMCs was reported, particularly in acute onset stage and active chronic stage [11]. Moreover, in PF patients' skin biopsies, while the ratio of IL17⁺ to CD4⁺ cell count was 1.8%, IL17-positive cells were undetectable in healthy controls' skin [12]. These data that are in favor of a possible implication of Th17 cells in the pathogenesis of PF were based only on the Th17 common marker to characterize this lymphocyte subset whose differentiation and function involve several other specific markers such as the lineage-specific transcription factor *RORγt* and the IL23 receptor (*IL23R*).

Indeed, the differentiation of Th17 cells from CD4⁺CD161⁺ naïve Th cells requires specific cytokines such as TGF-β, IL6, IL1β, IL21, and IL23 [13, 14]. In fact, the synergic action of the different cytokines induces the expression of

IL23R, which in turn activates the STAT signaling pathway leading to the expression of the transcription factor ROR γ t and then the production of Th17-specific cytokines, mainly IL17A and IL17F [15, 16]. Th17 cells play an important role in the clearance of fungal and extracellular bacterial infections by the recruitment of several cell types such as neutrophils, macrophages, and dendritic cells and by the induction of the production of cellular inflammatory mediators, such as TNF α [17–19].

The field of action of Th17 cells is not limited to these infections, and there is increasing evidence for the contribution of these cells in the pathogenesis of various autoimmune diseases (AIDs). In fact, IL23 which is considered as the stabilizing factor for Th17 cell commitment seems to play a key role in the acquisition of the pathogenic character of these cells [20]. IL23 signaling through IL23R can amplify Th17 cell response by inducing proinflammatory cytokines, such as IL1 β , TNF α , and IL6, in innate immune cells and by enhancing the activation of STAT3, which is coordinated with ROR γ t to stabilize them and maintain their function [15]. Moreover, it was reported that the pathogenicity of Th17 cells was associated with the over activation of the IL23/IL23R signaling pathway [16, 21]. Thus, the IL23/Th17 axis is becoming a leading interesting pathway in the field of AIDs such as systemic lupus erythematosus [22] and pemphigus vulgaris [23] and is one of the most promising targets for AID therapy.

In light of these data and to unfold the involvement of the IL23/Th17 pathway in the pathogenesis of Tunisian PF, we aimed to unravel the eventual genetic contribution of IL23/Th17 genes' polymorphisms and functional association of Th17 cells through the expression of their specific markers.

2. Material and Methods

2.1. Subjects. One hundred and fifteen PF patients have been recruited since 2002 at the department of Dermatology in Hedi Chaker University Hospital of Sfax, Tunisia. The diagnosis of PF was confirmed by clinical presentation, histopathology (acantholysis in the upper epidermis either in the granular layer or immediately below with subcorneal bullous formation), direct immunofluorescence (IgG and C3 deposits most often located on the whole epidermis and less frequently predominant in the upper layers of the epidermis), indirect immunofluorescence (IgG Abs directed against the epithelial cell surface), and ELISA test for circulatory anti-Dsg1 Abs that was positive for all patients [24]. Two hundred and one healthy controls (HC), who did not suffer from any autoimmune or inflammatory disease, were also recruited (see Table 1).

The genetic study is a case-control retrospective study enrolled in the whole population described above. The patient group included 8 men and 107 women, with a mean age of 35 years ranging from 18 years to 84 years. All patients originate from the center and south of Tunisia. The control group included 21 men and 180 women, with a mean age of 39 years ranging from 14 years to 73 years.

Related patients and/or HC were excluded from the genetic study.

TABLE 1: Characteristics of study populations.

Features	PF patients	Controls
Number	115	201
Sex ratio	1/14	1/9
Mean age	35 (18–84)	39 (14–73)
Origin	Center-southern regions of Tunisia	
Anti-Dsg1	Positive	Negative

The functional study is a perspective one, conducted on subjects from the population described above that were recruited since 2014. Pemphigus disease area index (PDAI) was measured for all retained patients. We enrolled 13 PF patients and 6 HC for mRNA expression analysis on biopsies, 5 PF women patients and 4 women HC for mRNA expression analysis on PBMCs, and 9 women patients in active stage and 5 women HC for flow cytometry analysis.

The patient group was divided into two groups depending on the stage of the disease: active stage and chronic stage. The active stage of PF is defined as de novo development of new blisters on the skin; none of these patients had yet received immunosuppressive therapy. The chronic stage of PF is defined as the expansion and/or the persistence of existing blisters on the skin. Patients in this stage had already received immunosuppressive treatment.

PF patients suffering from other AIDs were excluded. Healthy subjects were selected based on additional criteria; they have not taken any medicines for at least three weeks before the sampling. All HC were negative for anti-Dsg1Abs.

2.1.1. Sample Collection. After obtaining a written informed consent, samples were collected. Our project was approved by the ethical committee of the Habib Bourguiba University Hospital of Sfax (protocol number of the ethical committee, 4/12).

(1) Skin Biopsy. 13 biopsies were taken from 10 patients in the active stage of the disease and from 3 patients in the chronic stage. In addition, 6 control biopsies were taken from healthy subjects undergoing surgeries for a trichilemmal cyst or plastic surgery.

(2) Blood. A total of 10 ml of blood was taken from patients and HC in sterile endotoxin-free vacutainers with EDTA as anticoagulant for DNA extraction or in heparinized tubes for mRNA expression analysis and cytometry analysis.

2.2. SNP Selection. Tag single-nucleotide polymorphisms (tag SNPs) in the IL23/Th17 axis genes were selected according to their susceptibility to other AIDs and to the genotyping data from the CEU available from the International HapMap project. Selection was undertaken using minor allele frequency (greater than 5%) in Caucasian and sub-Saharan populations.

Four candidate SNPs in the *IL23R* gene in chromosome 1: rs1884444, rs7517847, rs11209026 and rs10889677, two SNPs in the *IL17A* gene in chromosome 6: rs3748067 and rs2275913, rs763780 in *IL17F* in chromosome 6, rs4819554

TABLE 2: Primary information of genotyped SNPs in Th17/IL23 pathway's genes.

Gene SNPs	Base change	MAF	Chromosome regions	Localization	Primers	Enzyme
IL23R rs1884444 ^a	G/T nonsens	0.47		Exon2	F:Fam [*] TGCTCTGTTTCCTTCCTTCC R1:CATCCCATTGAATAGTGACC R2:T ₄ CATCCCATTGAATAGTGACA	—
IL23R rs7517847	G/T	0.36	Chr1 1p31.3	Intron6	F:ATTTCTGGATGCCCTTTCCT R:ACATGAATTTGAGGGGCCTA	BbvI
IL23R rs11209026	A/G Arg-gln	0.022		Exon8	F:TTAGACAACAGAGGAGACATTGGA R:CATGTAGTCTAAATCAGAAAACAGAAA	Hpy188I
IL23R rs10889677	A/C	0.39		3'UTR	F:TGCTGGGCCATATGATAAGC R:TCCACCTTCGGGACCTTAAT	MnII
ROR γ t rs9645406 ^a	T/C		Chr1 1q21.3	Intron	F1:T4CCTCACAGCAAATCTTTTCTC F2:CCTCACAGCAAATCTTTTCTT R:NED [*] AAAAACCGCTGTAGGGATCA	—
IL17A rs2275913	A/G	0.29	Chr6 6p12.2	5'UTR	F:ATTTCTGCCCTTCCCATTTT R:TTGTGCCTGCTATGAGATGG	EarI
IL17A rs3748067	T/C	0.077		3'UTR	R:GGGGCGAAAATGGTTACGAT F:AAGGCCCTCAGAGATCAAC	ApoI
IL17F rs763780	C/T	0.093	Chr6 6p12.2	Exon3	F:TGGGTAAGGAGTGGCATTTC R:CTGCATCAATGCCTCAAGGAA	NlaIII
IL17RA rs4819554	G/A	0.233	Chr22 22q11.1	5'UTR	F:TGAAATGTGTAATTCGCTGGC R:TGCTTTCCTTGGCTTTGCTT	PvuII
TNF α rs1800629	A/G	0.090	Chr6 6p21.33	5'UTR	F:AGGCAATAGTTTTGAGGGGCAT R:TCTCCCTGCTCCGATTCC	NcoI
STAT3 rs744166	C/T	0.482	Chr17 17q21.2	Intron	F:GCTGGAGTACAAACCCTGAA R:TGGTATTACAGATGGCGGTCA	AluI

^aGenotyped using the AS-PCR method.

in the *IL17RA* gene in chromosome 22, rs9645406 in *ROR γ t* within chromosome 1, rs1800629 in the *TNF α* gene within chromosome 6, and rs744166 in the *STAT3* gene within chromosome 17, were selected. All primers and enzymes used in this study are presented in Table 2.

2.3. SNP Genotyping. Genomic DNA was extracted from whole blood samples using a standard proteinase K digestion and phenol/chloroform extraction procedure. Genotyping was performed using the PCR-RLFP method for all SNPs except for rs1884444 in the *IL23R* gene and rs9645406 in *ROR γ t* which were genotyped using the AS-PCR method (see Table 2).

2.3.1. PCR-RFLP. The PCR amplification was carried out in a volume of 25 μ l including 1x buffer, 2 mM MgCl₂, 0.2–0.4 μ mol of each primer (Invitrogen[®], CA, USA), 0.12 mM dNTP (Invitrogen, CA, USA), 1 U Taq polymerase (Invitrogen, CA, USA), and 50 ng of DNA template. Enzymatic digestion was performed in a total of 10 μ l mixture reaction containing 1x buffer, 0.1x BSA, and 2 U restriction enzyme (Thermo Fisher[®], MA, USA). Primers were designed using primer3 software (<http://primer3.ut.ee/>). Restriction enzymes were selected using the NEBcutter software (<http://nc2.neb.com/NEBcutter2/>).

2.3.2. AS-PCR. The primers were designed using WASP software (<http://bioinfo.biotech.or.th/WASP>). Common primer

was labeled with 6-FAM fluorescent labels for the rs1884444 and 6-NED fluorescent labels for rs9645406. The mutant primer containing the mutant base was labeled with a poly(A) tail. The PCR amplification was carried out in a volume of 10 μ l including 1x buffer, 2.5 pmol of each primer (Invitrogen, CA, USA), 10 mM dNTP (Invitrogen, CA, USA), 1 U AmpliTaq Gold[™] DNA Polymerase (Applied Biosystems[™], CA, USA), and 1 ng of DNA template. Amplified products were run on ABI prism DNA sequencer (PerkinElmer[®], CT, USA) and the output file was analyzed using GeneScan software analysis.

2.4. Quantitative mRNA Expression of *IL23R* and *ROR γ t* in the Skin and PBMCs by Q-PCR. Peripheral blood mononuclear cells (PBMCs) were isolated by Histopaque-1077 density-gradient method. Total RNA from PBMCs was extracted using One-step RNA reagent (Bio Basic[®], Canada) following the manufacturer's instructions and then reversely transcribed using the High-Capacity cDNA Reverse Transcription kit (Applied Biosystems[®], CA, USA). The real-time quantitative PCR was performed via the StepOnePlus Real-Time PCR systems (Applied Biosystems, CA, USA). TaqMan analysis was conducted using pre-designed and optimized assays from Applied Biosystems: *IL23R* (ID: Hs 00332759_m1) and *ROR γ t* (ID: Hs 01076122_m1). PCR reaction parameters were as follows: 95°C for 10 min, followed by 40 cycles of 95°C for 15 s, 60°C for 1 min. All measurements were performed in triplicate. For the relative

quantification, data were analyzed by the comparative $2^{-\Delta\Delta Ct}$ method and normalized to the average of housekeeping gene GAPDH.

2.5. Flow Cytometry Analysis. PBMCs were seeded into 24-well plates at a final concentration of 10^6 cells/ml, in RPMI1640 complete medium (containing 10% of the fetal bovine serum, 200 g/ml penicillin, and 200 U/ml streptomycin), and were stimulated with 12-myristate 13-acetate (50 ng/ml) and calcium ionomycin ($1 \mu\text{g/ml}$), in the presence of GolgiStop protein transport inhibitor (BD Biosciences®, CA, USA) at 37°C for four hours in a 5% of CO_2 incubator. Cells were subsequently stained for surface markers using PerCP-Cy5.5-labeled anti-human CD4 and PE-labeled anti-human IL17 for intracellular cytokine (BD Bioscience, CA, USA). $\text{CD4}^+\text{T}$ cells were gated, and the percentages of these cells producing IL17 were calculated. The BD FACS Canto II System (BD Biosciences, USA) and Kaluza Analysis 1.5a software (Beckman Coulter, USA) were used for analysis.

2.6. Statistical Analysis. A case-control analysis was performed using SHESIS software (<http://analysis.bio-x.cn>) for each SNP and haplotype. Hardy-Weinberg equilibrium (HWE) was assessed in controls using a χ^2 test with one degree of freedom. A threshold $P < 0.05$ was regarded to indicate deviation from HWE. Odds ratios (OR) and 95% confidence intervals (CI) were calculated for each allele using 2×2 contingency tables to estimate the magnitude of association. Binary logistic regression was used to make age and gender adjustment. The linkage disequilibrium (LD) coefficients $D' = D/D_{\max}$ and r^2 values for the pair of the most common alleles at each site were also estimated, and high values of LD were defined as $r^2 > 0.33$ and $D' > 0.7$. The significance level of $P < 0.05$ and odds ratios (OR) with 95% confidence intervals (95% CI) was chosen for all sets.

For functional experiments, statistical analyses were carried out using SPSS20.0 software (IBM SPSS® Inc., IL, USA). As the number of samples was less than twenty, the nonparametric Mann-Whitney U test was used to analyze flow cytometry results and mRNA expression of IL23R and ROR γ t. Spearman correlation rho “ r ” was used to determine the possible correlation between different results.

3. Results

3.1. Genetic Association of Investigated SNPs with Susceptibility to PF. Genotype frequencies of all SNPs tested in control subjects were consistent with those expected from the Hardy-Weinberg equilibrium (HWE) except for rs1884444 polymorphism in the *IL23R* gene and rs763780 in *IL17F* gene ($P < 0.001$). Minor allele frequency of all the polymorphisms was consistent with that reported in the HapMap database. The genotypic and allelic distributions of the *IL23/Th17* gene polymorphisms as well as their associations with the risk to PF are shown in Table 3.

Concerning the *IL23R* gene, the rs11209026 A>G polymorphism seems to be closely associated with the PF's risk. In fact, the homozygous GG genotype was significantly overexpressed in patients (89.7%) compared to HC

(76.2%) ($P = 0.004$, OR = 2.37, 95% CI 1.35–5.53), contrary to the AG genotype which was more expressed in controls ($P = 0.01$, OR = 0.42, 95% CI 0.21–0.87). Likewise, the decreased expression of the A allele observed in the patient group compared to HC suggests its protective role against PF ($P = 0.001$, OR = 0.35, 95% CI 0.18–0.69). No association was found either with rs1884444, with rs7517847, or with rs10889677.

For the *IL17A* gene, a strong association of rs3748067 T>C with the occurrence of PF was revealed. The CC homozygous genotype and the C allele seem to increase the risk of the development of the disease ($P = 1.17e - 04$, OR = 5.03, 95% CI 2.34–10.78; $P = 8.29e - 06$, OR = 4.57, 95% CI 2.23–9.36). In addition, the heterozygous genotype CT appears as a PF protection genotype ($P = 9.47e - 04$, OR = 0.23, 95% CI 0.11–0.50). No significant association was observed for rs2275913.

For *IL17RA* gene polymorphism, no statistical significant difference in genotype and allele frequencies was observed between patients and controls.

As for the *IL17F* gene, a significant increase of the rs763780>C allele was observed in the patient group (10.8%) compared to HC (4.9%) ($P = 0.007$, OR = 2.35, 95% CI 1.24–4.42). On the other hand, the TT genotype was significantly less expressed in patients (89.2%) than in HC (95.1%) ($P = 0.02$, OR = 0.44, 95% CI 0.22–0.91), suggesting its protective role.

Regarding the *TNFA* gene, the allelic distribution of rs1800629 A>G in patients' cohort revealed a great significant increase of the A allele ($P = 4.06e - 5$, OR = 2.33, 95% CI 1.55–3.52). In addition, while the homozygous AA and heterozygous AG genotypes are associated with the PF's risk ($P = 0.02$, OR = 2.76, 95% CI 1.12–6.82; $P = 0.008$, OR = 2.05, 95% CI 1.20–3.51), the GG genotype appeared to be a protector genotype ($P = 0.0002$, OR = 0.38, 95% CI 0.22–0.63).

No significant association was found with rs9645406 in the ROR γ t gene or with rs744166 in the STAT3 gene.

All significant associations were conserved after age and gender adjustment using the binary logistic regression.

3.2. Linkage Disequilibrium (LD), Haplotype, and Combination Analysis. The LD analysis among the patient group was conducted by pairwise comparison of the 4 polymorphisms studied in the *IL23R* gene within chromosome 1 (see Figure 1).

An evidence for LD was revealed between rs11209026 and rs10889677 ($D' = 0.74$) and with rs1884444 ($D' = 0.71$) suggesting that the presumed haplotype of susceptibility to Tunisian PF is rs1884444>T, rs7517847>T, rs11209026>G, rs10889677>A, which contains the rs11209026>G susceptibility allele. This haplotype was more expressed in patients (23.4%) than in controls (16.3%) and, therefore, appears as a risk haplotype to the disease ($P = 1.71e - 04$, OR = 2.41, 95% CI 1.51–3.85) (see Table 4).

Since *IL17A* (rs3748067 and rs2275913), *IL17F* (rs763780), and *TNFA* (rs1800629) genes are located on nearby regions 6p12.2 and 6p21.33, respectively, LD values may explain the possible effect and interaction between their different alleles (see Figure 2).

TABLE 3: Genotype and allele frequencies of IL23/Th17 pathway's genes polymorphisms in Pemphigus foliaceus patients and matched healthy controls.

Gene/SNP	Genotype/allele	Case N (%)	Control N (%)	P	OR	95% CI
IL23R rs11209026	AA	0	5 (2.6)	NS	—	—
	AG	11 (10.3)	41 (21.2)	0.016	0.42	0.21–0.87
	GG	96 (89.7)	147 (76.2)	0.004	2.37	1.35–5.53
	A	11 (5.1)	51 (13.2)	0.001	0.35	0.18–0.69
	G	203 (94.9)	335 (86.8)			
IL23R rs1884444	GG	3 (3)	11 (8.3)	NS	—	—
	GT	80 (79.2)	106 (79.7)	NS	—	—
	TT	18 (17.8)	16 (12)	NS	—	—
	G	86 (42.6)	128 (48.1)	NS	—	—
	T	116 (57.4)	138 (51.9)			
IL23R rs7517847	GG	10 (9.8)	14 (7.4)	NS	—	—
	GT	43 (42.2)	75 (39.7)	NS	—	—
	TT	49 (48)	100 (52.9)	NS	—	—
	G	63 (30.9)	103 (27.2)	NS	—	—
	T	141 (69.1)	276 (72.8)			
IL23R rs10889677	AA	20 (19.8)	40 (22.1)	NS	—	—
	AC	41 (40.6)	71 (39.2)	NS	—	—
	CC	40 (39.6)	70 (38.7)	NS	—	—
	A	81 (40.1)	151 (41.7)	NS	—	—
	C	121 (59.9)	211 (58.3)			
IL17A rs2275913	AA	1 (1)	1 (0.6)	NS	—	—
	AG	9 (9.2)	10 (6.2)	NS	—	—
	GG	88 (89.8)	150 (93.2)	NS	—	—
	A	11 (5.6)	12 (3.7)	NS	—	—
	G	185 (94.4)	310 (96.3)			
IL17A rs3748067	TT	0	4 (3.6)	NS	—	—
	CT	10 (11.1)	38 (35)	9.47e – 4	0.23	0.11–0.50
	CC	79 (87.7)	66 (60.8)	1.17e – 4	5.03	2.34–10.78
	T	10 (5.5)	46 (21.1)	8.29e – 6	0.218	0.10–0.44
	C	170 (94.4)	171 (78.8)			
IL17AR rs4819554	AA	78 (74.3)	137 (71.8)	NS	—	—
	AG	24 (22.9)	48 (25.1)	NS	—	—
	GG	3 (2.9)	6 (3.1)	NS	—	—
	A	180 (85.7)	322 (84.3)	NS	—	—
	G	30 (14.3)	60 (15.7)			
IL17F rs763780	CC	5 (4.7)	3 (1.6)	NS	—	—
	CT	13 (12.3)	13 (6.7)	NS	—	—
	TT	88 (83)	177 (91.7)	0.02	0.44	0.22–0.91
	C	23 (10.8)	19 (4.9)	0.007	2.35	1.24–4.42
	T	189 (89.2)	367 (95.1)			
TNFa rs1800629	AA	12 (12.8)	9 (5)	0.022	2.76	1.12–6.82
	AG	37 (39.4)	43 (24)	0.008	2.05	1.20–3.51
	GG	45 (47.9)	127 (70.9)	0.0002	0.38	0.22–0.63
	A	61 (32.4)	61 (17)	4.06e – 5	2.33	1.55–3.52
	G	127 (67.6)	297 (83)			

TABLE 3: Continued.

Gene/SNP	Genotype/allele	Case N (%)	Control N (%)	P	OR	95% CI
RORyt rs9645406	TT	0	0	NS	—	—
	CT	9 (7.9)	13 (6.5)	NS	—	—
	CC	106 (92.1)	188 (93.5)	NS	—	—
	T	11 (4.7)	13 (3.3)	NS	—	—
	C	219 (95.3)	389 (96.7)	NS	—	—
STAT3 rs744166	TT	22 (23.1)	29 (18.1)	NS	—	—
	CT	47 (49.5)	77 (48.1)	NS	—	—
	CC	26 (27.4)	54 (33.8)	NS	—	—
	T	91 (47.9)	135 (42.2)	NS	—	—
	C	99 (52.1)	185 (57.8)	NS	—	—

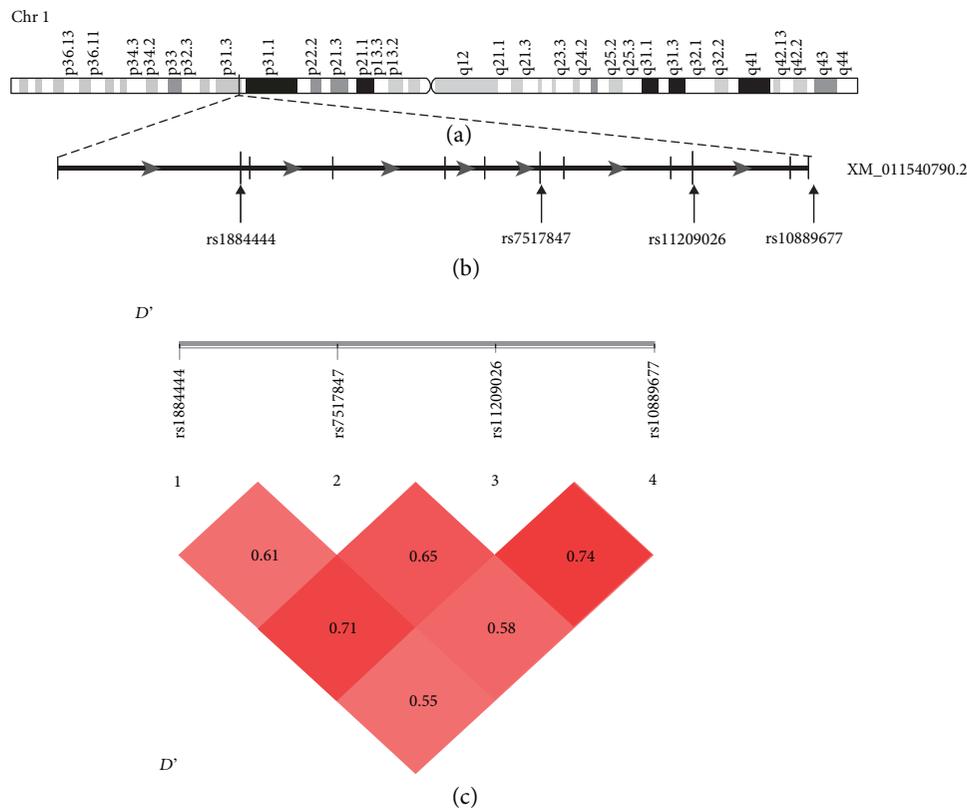


FIGURE 1: Overview and linkage disequilibrium (LD) on chromosome 1q31.1. (a) Overview of chromosome 1. (b) Schematic structure of the *IL23R* gene. Exons are depicted by black vertical lines. The positions of the four SNPs are shown by the arrows. (c) LD prime charts generated using SHEsis software summarize LD (D') patterns between the 4 SNP. $D' > 0.7$ is considered as a high value of LD.

TABLE 4: Haplotypes of genotyped SNPs on chromosomes 1 and 6.

Chromosome	Haplotype	Case N (%)	Control N (%)	P	OR (95% CI)
1	TTGA	44 (23.4)	39 (16.3)	$1.71e - 04$	2.409 (1.508~3.846)
	TGGC	33 (17.5)	24 (10)	$1.16e - 04$	2.871 (1.648~5.001)
6	AGGT	42 (0.291)	30(0.168)	$7.67e - 06$	3.027 (1.832~5.003)
	GGAT	6 (0.041)	38 (0.213)	0.002	0.276 (0.114~0.664)

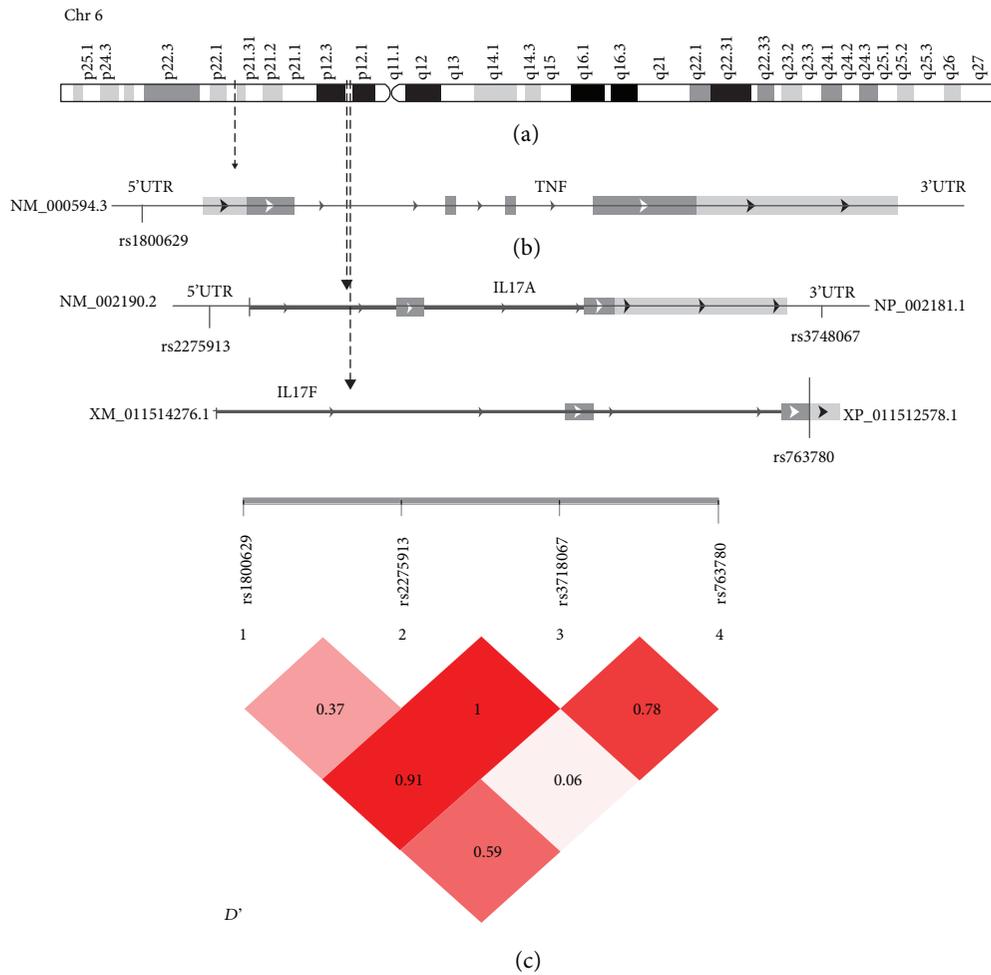


FIGURE 2: Overview and linkage disequilibrium (LD) on chromosome 6. (a) Overview of chromosome 6. (b) Schematic structure of the *IL17A*, *IL17F*, and *TNFA* genes. The positions of the four SNPs are shown by the arrows (c). LD prime charts generated using SHEsis software summarize LD (D') patterns between the 4 SNPs. $D' > 0.7$ is considered as a high value of LD.

Thus, an evidence for LD between the SNPs investigated in *IL17A*, *F*, and *TNFA* was observed. Indeed, strong LD between rs3748067 and rs2275913 ($D' = 1$) and rs3748067 and rs1800629 ($D' = 0.91$) were found, indicating that the association observed with these SNPs must be interdependent. Additionally, the estimated frequencies of haplotypes were statistically different between PF patients and controls (global $\chi^2 = 23.17$, global Pearson's $P = 1.17e - 04$). In fact, haplotypes AGCT and GGCT which contain the associated risk allele rs3748067>C ($P < 0.0001$) were associated with an increased risk to PF. In contrast, the GGTT haplotype seems to confer protection against PF ($P = 0.001$) (see Table 4).

On the other hand, possible combinations between the different SNPs were analyzed and a strong evidence for gene-gene interactions between them was revealed (see Table 5). Thus, three associated combinations that differ statically between patients and controls were observed; TTGAAG CTAC ($P = 0.018$, OR = 4.49, 95% CI 1.15–17.57), TGGCGG CTAC ($P = 0.003$, OR = 4.87, 95% CI 1.51–15.74) and TTGAGGCTAT ($P = 0.017$, OR = 0.017, 95% CI 1.16–8.00).

3.3. Relative mRNA Expression of *IL23R* and *RORγt* in the Skin and Blood. In patients' blood, the relative mRNA expression levels of *RORγt* and *IL23R* were significantly higher ($P < 0.05$) compared to the HC (9.64 ± 12.96 versus 0.778 ± 0.382 and 5.01 ± 1.00 versus 0.75 ± 0.22 , resp.) as shown in Figure 3. Moreover, a strong positive correlation was revealed between the expression of these two markers ($r = 0.943$; $P < 0.005$).

In spite of the insignificant difference in *RORγt* and *IL23R* expression levels in skin biopsies between the patient and control groups (1.09 ± 1.74 versus 0.95 ± 0.96 and 2.99 ± 3.89 versus 1.18 ± 0.90 , resp.), the *IL23R* mRNA was slightly more expressed in specimens from patients in the chronic stage than those de novo (7.427 ± 6.25 versus 1.661 ± 1.734) (see Figure 3).

3.4. Increased Frequency of *CD4⁺IL17⁺* Circulatory Cells in Patients' Blood. After stimulation, the frequency of *CD4⁺IL17⁺* cells was significantly higher in patients compared to controls as shown in Figure 4 ($P = 0.001$). In patients, the percentage of Th17 cells varied from 1.36 to 3.57% of

TABLE 5: Combinations of genotyped SNPs in *IL23R*, *IL17A*, *IL17F*, *TNFA*, *ROR γ t*, and *STAT3* showing significant differences between PF patients and controls.

Combinations	Case N (%)	Control N (%)	P	OR (95% CI)
TTGAAGCTAC	7 (5.5)	3 (2.3)	0.018	4.497 (1.151~17.574)
TGGCGGCTAC	10 (7.9)	4 (3.1)	0.003	4.877 (1.511~15.742)
TTGAGGCTAT	11 (8.7)	7 (5.4)	0.017	3.057 (1.167~8.006)

CD4⁺ cells; however, in the control group, the percentage does not exceed 1% (see Figure 4).

4. Discussion

The IL23/Th17 pathway has taken a lot of interest recently, as it is pivotal both in tissue immunosurveillance and autoimmunity mechanisms. Based on genetic and functional studies, strong evidence for its implication in the pathogenesis of several autoimmune diseases, such as systemic lupus erythematosus [22] and rheumatoid arthritis [23], has emerged. Bearing in mind that substitutions of one base pair (SNPs) may have functional importance, we investigated, in the present data, an eventual functional contribution of the most important IL23/Th17 axis markers in the pathogenesis of PF.

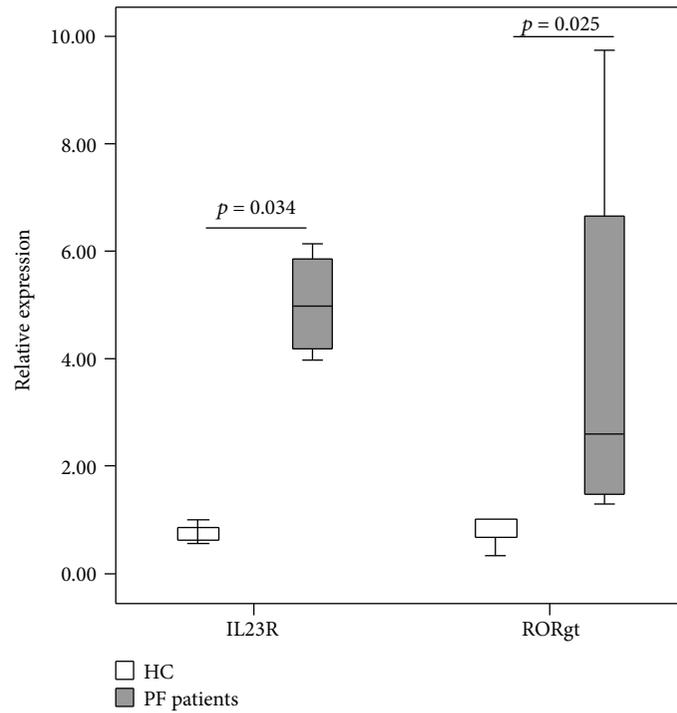
Concerning the four Tag SNPs genotyped in the *IL23R* gene, only rs11209026>G was found to be strongly associated with PF susceptibility ($P = 0.001$, OR = 2.81). Moreover, this SNP in strong LD with the others generates the susceptibility haplotype rs1884444>T-rs7517847>T-rs11209026>G-rs10889677>A. Lesional skin biopsies from PF patients showed a slight increase of IL23R mRNA expression as compared to those from HC. Interestingly, specimens of patients in the chronic stage showed a higher mRNA level than those of de novo patients. Furthermore, this relative expression was more significantly pronounced in PF's PBMCs compared to HC ones ($P = 0.03$). The critical role of the IL23/IL23R signaling pathway was proved in favoring terminal differentiation, maintenance, and pathogenicity of Th17 cells [16]. In addition, strong data demonstrate that the lack of the proinflammatory cytokine IL23 in mice makes them resistant to experimental models of arthritis and multiple sclerosis (MS) [25, 26]. Thus, overexpression of this cytokine was reported in several AIDs such as psoriasis [27] and PV [28] but not in PF. The genetic association of rs11209026 was widely described in the context of AID. This variant is located between the transmembrane domain and putative JAK2 binding site in the cytoplasmic portion of IL23R protein and is extremely conserved across different species [29]. Consequently, a change in the highly conserved Arg to Gln might have functional consequences on the IL23R transduction pathway. Several functional data have proved that the protective effect of A allele against autoimmune disease is exerted by the impairment of IL23-induced Th17 cell functions without interfering with Th17 differentiation nor

affecting the IL17 production [21, 30]. Unfortunately, as all our patients express the GG genotype, the analysis of the expression in patients with GG genotypes versus others with AG genotypes was not possible.

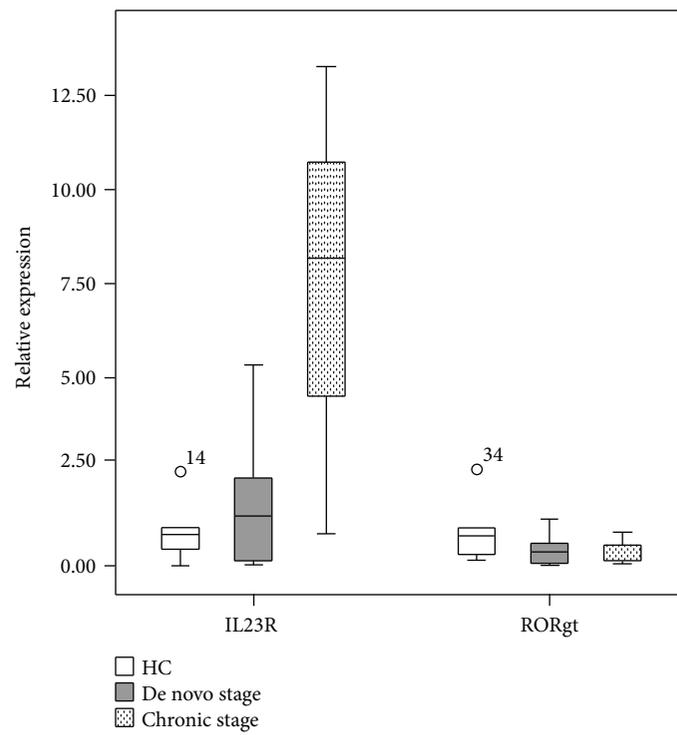
On the other hand, we found significant associations with genotyped SNPs in the 6p12.2 region in chromosome 6: rs3748067>C in *IL17A* and rs763780>C in *IL17F*. While few studies have explored the association of rs3748067 in *IL17A* with the cancer risk but not with AIDs [31, 32], rs763780 in *IL17F* was associated with many AIDs such rheumatoid arthritis in Tunisian population [33].

Little amount of ROR γ t, the specific transcription factor of Th17 cells, was observed in PF patients' skin biopsies, but the difference when compared to HC was not statistically significant. PBMCs from PF patients showed a significantly higher level of ROR γ t mRNA, when compared to specimens from HC. A possible explanation for the lack of correlation may reside in the sample in which mRNA was measured and/or in the little size of the skin sample group. Thus, Th17 cells have low-frequency lesional tissue. So, it is possible that ROR γ t mRNA is expressed at an undetectable low level in skin biopsies as compared to PBMC. Besides, cellular infiltration in PF includes essentially eosinophils and a little amount of CD4⁺ cells as compared to other autoimmune intraepidermal bullous diseases, such as that in PV [24]. In fact, Arakawa et al. revealed the presence of IL17⁺ cells in PF biopsies with a ratio count to CD4⁺ cell of 1.8%, but the difference compared to HC was not statistically significant. On the contrary, a higher frequency of IL17⁺ cells reaching a level of 5.2% was reported in skin biopsies of PV patients, in which cellular infiltration is more important [12]. As was expected, correlation analysis showed a positive correlation between the expression of ROR γ t and IL23R, as the expression of this receptor is necessary for the induction of ROR γ t expression. Several data showed that cytokines, including TGF- β , IL6, IL1, and IL21, are important in the differentiation of Th17 cells and have proved the necessity of IL23 for the maintenance and the proliferation of established Th17 cells.

For the *TNFA* gene, we found a strong association with rs1800629>A to the PF risk, similar to the result found by Torzecka et al. on Polish population [34]. However, other studies in Indian and Egyptian population estimated no significant genotype and/or allelic frequency difference between PF patients and HC [35, 36]. The investigation of the relation between this polymorphism and secreted levels of TNF α showed inconsistent results. Thus, some studies revealed that A allele is associated with a higher level of TNF α compared to the common G allele [37, 38], whereas others failed to demonstrate any functional change related to allele change [39, 40]. Previously, we reported the association of an STR within the *TNFA* gene with the PF risk [41]. As the *TNFA* locus is located in the HLA area near the DR and DQ regions, several data suggest that the *TNFA* association with PF could be explained by its linkage disequilibrium with DRB1 locus. However, after multivariate analysis since multiple regression analysis, we showed that, in our population, TNF and HLA DR susceptibility alleles were independently associated with PF [41].



(a)



(b)

FIGURE 3: mRNA expression of IL23R and RORyt in peripheral blood (a) and skin biopsies (b). PBMCs were extracted from 5 de novo patients and 4 HC. Biopsies from de novo patients ($N = 10$), patients in the chronic stage ($N = 3$), and from HC ($N = 5$) were analyzed using Q-PCR. The relative expression was estimated using $2^{-\Delta\Delta Ct}$ method and normalized to the average of the GAPDH housekeeping gene. P value was calculated using Mann–Whitney U test.

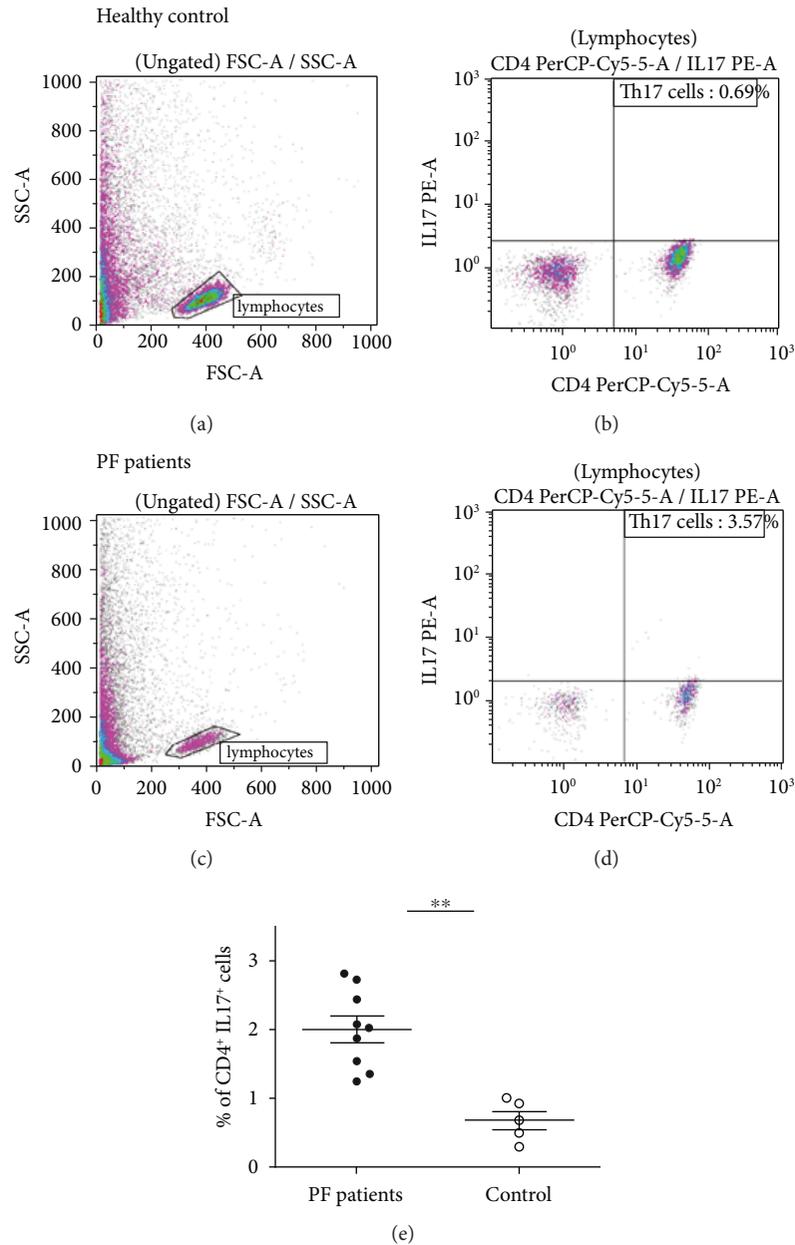


FIGURE 4: Frequency of IL17⁺ cells in the peripheral blood mononuclear cells of PF patients and healthy controls. Th17 cells identified using specific antibodies CD4-PerCP-Cy5 and IL17A-PE. Representative flow cytometry analysis of Th17 cells in healthy controls (a, b) and in PF patients (c, d). (e) Percentage of Th17 cells in control and patients' PBMCs.

5. Conclusion

In this case-control study, a high frequency of circulating Th17 cells was revealed in patients' blood compared to healthy controls. This increased level was confirmed by the mRNA expression analysis of IL23R and ROR γ t, specific markers of these cells. Moreover, we reported for the first time, to our knowledge, the genetic contribution of the IL23/Th17 pathway gene's variants in PF pathogenesis. Thus, these results could provide an argument for the contribution of the IL23/Th17 pathway in the pathogenesis of PF. However, much deeper investigation on larger patient groups should be done to support these findings.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that there is no conflict of interest regarding the publication of this paper.

Acknowledgments

The authors thank Ms Fekria Abdennadher and Ms Emna Hentati, proficient in the English language, for proofreading

this article. This study was supported by the Ministry of High Education and Scientific Research of Tunisia, the Habib Bourguiba University hospital of Sfax, and the “Comité de Recherche de l’hôpital Habib Bourguiba, Sfax, Tunisia”.

References

- [1] M. Kasperkiewicz, C. T. Ellebrecht, H. Takahashi et al., “Pemphigus,” *Nature Reviews Disease Primers*, vol. 3, p. 17026, 2017.
- [2] J. T. Roscoe, L. Diaz, S. A. P. Sampaio et al., “Brazilian pemphigus foliaceus autoantibodies are pathogenic to BALB/c mice by passive transfer,” *Journal of Investigative Dermatology*, vol. 85, no. 6, pp. 538–541, 1985.
- [3] E. H. Beutner, L. S. Prigenzi, W. Hale, C. D. A. Leme, and O. G. Bier, “Immunofluorescent studies of autoantibodies to intercellular areas of epithelia in Brazilian pemphigus foliaceus,” *Proceedings of the Society for Experimental Biology and Medicine*, vol. 127, no. 1, pp. 81–86, 1968.
- [4] A. P. Kowalczyk, J. E. Anderson, J. E. Borgwardt, T. Hashimoto, J. R. Stanley, and K. J. Green, “Pemphigus sera recognize conformationally sensitive epitopes in the amino-terminal region of desmoglein-1,” *The Journal of Investigative Dermatology*, vol. 105, no. 2, pp. 147–152, 1995.
- [5] M. K. Hacker, M. Janson, J. A. Fairley, and M.-S. Lin, “Isotypes and antigenic profiles of pemphigus foliaceus and pemphigus vulgaris autoantibodies,” *Clinical Immunology*, vol. 105, no. 1, pp. 64–74, 2002.
- [6] S. J. P. Warren, L. A. Arteaga, L. A. Diaz et al., “The role of subclass switching in the pathogenesis of endemic pemphigus foliaceus,” *Journal of Investigative Dermatology*, vol. 120, no. 1, pp. 1–5, 2003.
- [7] Y. Futei, M. Amagai, K. Ishii, K. Kuroda-Kinoshita, K. Ohya, and T. Nishikawa, “Predominant IgG4 subclass in autoantibodies of pemphigus vulgaris and foliaceus,” *Journal of Dermatological Science*, vol. 26, no. 1, pp. 55–61, 2001.
- [8] R. J. Armitage, B. M. Macduff, M. K. Spriggs, and W. C. Fanslow, “Human B cell proliferation and Ig secretion induced by recombinant CD40 ligand are modulated by soluble cytokines,” *The Journal of Immunology*, vol. 150, no. 9, pp. 3671–3680, 1993.
- [9] M. S. Lin, C. L. Fu, V. Aoki et al., “Desmoglein-1-specific T lymphocytes from patients with endemic pemphigus foliaceus (fogo selvagem),” *Journal of Clinical Investigation*, vol. 105, no. 2, pp. 207–213, 2000.
- [10] K. L. Gebhard, C. M. Veldman, R. Wassmuth, E. Schultz, G. Schuler, and M. Hertl, “Ex vivo analysis of desmoglein 1-responsive T-helper (th) 1 and Th2 cells in patients with pemphigus foliaceus and healthy individuals,” *Experimental Dermatology*, vol. 14, no. 8, pp. 586–592, 2005.
- [11] R. C. Xu, H. Q. Zhu, W. P. Li et al., “The imbalance of Th17 and regulatory T cells in pemphigus patients,” *European Journal of Dermatology*, vol. 23, no. 6, pp. 795–802, 2013.
- [12] M. Arakawa, T. Dainichi, S. Yasumoto, and T. Hashimoto, “Lesional Th17 cells in pemphigus vulgaris and pemphigus foliaceus,” *Journal of Dermatological Science*, vol. 53, no. 3, pp. 228–231, 2009.
- [13] L. Cosmi, R. de Palma, V. Santarlasci et al., “Human interleukin 17-producing cells originate from a CD161+CD4+ T cell precursor,” *The Journal of Experimental Medicine*, vol. 205, no. 8, pp. 1903–1916, 2008.
- [14] F. Annunziato, L. Cosmi, F. Liotta, E. Maggi, and S. Romagnani, “Defining the human T helper 17 cell phenotype,” *Trends in Immunology*, vol. 33, no. 10, pp. 505–512, 2012.
- [15] M. S. Maddur, P. Miossec, S. V. Kaveri, and J. Bayry, “Th17 cells: Biology, pathogenesis of autoimmune and inflammatory diseases, and therapeutic strategies,” *American Journal of Pathology*, vol. 181, no. 1, pp. 8–18, 2012.
- [16] M. J. McGeachy, Y. Chen, C. M. Tato et al., “The interleukin 23 receptor is essential for the terminal differentiation of interleukin 17-producing effector T helper cells in vivo,” *Nature Immunology*, vol. 10, no. 3, pp. 314–324, 2009.
- [17] H. Park, Z. Li, X. O. Yang et al., “A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17,” *Nature Immunology*, vol. 6, no. 11, pp. 1133–1141, 2005.
- [18] M. Essakalli, C. Brick, N. Bennani, N. Benseffaj, S. Ouadghiri, and O. Atouf, “Le lymphocyte TH17 dernier-né de la famille des lymphocytes T CD4+,” *Pathologie Biologie*, vol. 58, no. 6, pp. 437–443, 2010.
- [19] D. V. Jovanovic, J. A. di Battista, J. Martel-Pelletier et al., “IL-17 stimulates the production and expression of proinflammatory cytokines, IL-beta and TNF-alpha, by human macrophages,” *The Journal of Immunology*, vol. 160, no. 7, pp. 3513–3521, 1998.
- [20] A. L. Croxford, F. Mair, and B. Becher, “IL-23: one cytokine in control of autoimmunity,” *European Journal of Immunology*, vol. 42, no. 9, pp. 2263–2273, 2012.
- [21] P. Di Meglio, A. Di Cesare, U. Laggner et al., “The IL23R R381Q gene variant protects against immune-mediated diseases by impairing IL-23-induced Th17 effector response in humans,” *PLoS One*, vol. 6, no. 2, article e17160, 2011.
- [22] H. Puwipirom, N. Hirankarn, P. Sodsai, Y. Avihingsanon, J. Wongpiyabovorn, and T. Palaga, “Increased interleukin-23 receptor(+) T cells in peripheral blood mononuclear cells of patients with systemic lupus erythematosus,” *Arthritis Research & Therapy*, vol. 12, no. 6, p. R215, 2010.
- [23] H. Shen, J. C. Goodall, and J. S. Hill Gaston, “Frequency and phenotype of peripheral blood Th17 cells in ankylosing spondylitis and rheumatoid arthritis,” *Arthritis and Rheumatism*, vol. 60, no. 6, pp. 1647–1656, 2009.
- [24] P. Joly and N. Litrowski, “Pemphigus group (vulgaris, vegetans, foliaceus, herpetiformis, brasiliensis),” *Clinics in Dermatology*, vol. 29, no. 4, pp. 432–436, 2011.
- [25] D. J. Cua, J. Sherlock, Y. Chen et al., “Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain,” *Nature*, vol. 421, no. 6924, pp. 744–748, 2003.
- [26] C. A. Murphy, C. L. Langrish, Y. Chen et al., “Divergent pro- and antiinflammatory roles for IL-23 and IL-12 in joint autoimmune inflammation,” *The Journal of Experimental Medicine*, vol. 198, no. 12, pp. 1951–1957, 2003.
- [27] E. Lee, W. L. Trepicchio, J. L. Oestreicher et al., “Increased expression of interleukin 23 p19 and p40 in lesional skin of patients with psoriasis vulgaris,” *The Journal of Experimental Medicine*, vol. 199, no. 1, pp. 125–130, 2004.
- [28] J. Xue, W. Su, Z. Chen, Y. Ke, X. Du, and Q. Zhou, “Overexpression of interleukin-23 and interleukin-17 in the lesion of pemphigus vulgaris: a preliminary study,” *Mediators of Inflammation*, vol. 2014, Article ID 463928, 5 pages, 2014.
- [29] E. Abdollahi, F. Tavasolian, A. A. Momtazi-Borojeni, M. Samadi, and H. Rafatpanah, “Protective role of R381Q

- (rs11209026) polymorphism in IL-23R gene in immune-mediated diseases: A comprehensive review,” *Journal of Immunotoxicology*, vol. 13, no. 3, pp. 286–300, 2016.
- [30] S. Pidasheva, S. Trifari, A. Phillips et al., “Functional studies on the IBD susceptibility gene IL23R implicate reduced receptor function in the protective genetic variant R381Q,” *PLoS One*, vol. 6, no. 10, article e25038, 2011.
- [31] S. Yang, C. Li, X. Li et al., “Relationship of IL-17A and IL-17F genetic variations to cervical cancer risk: a meta-analysis,” *Biomarkers in Medicine*, vol. 11, no. 5, pp. 459–471, 2017.
- [32] Y.-W. Gao, M. Xu, Y. Xu, D. Li, and S. Zhou, “Effect of three common IL-17 single nucleotide polymorphisms on the risk of developing gastric cancer,” *Oncology Letters*, vol. 9, no. 3, pp. 1398–1402, 2015.
- [33] O. S. Marwa, T. Kalthoum, K. Wajih, and H. Kamel, “Association of IL17A and IL17F genes with rheumatoid arthritis disease and the impact of genetic polymorphisms on response to treatment,” *Immunology Letters*, vol. 183, pp. 24–36, 2017.
- [34] J. D. Torzecka, J. Narbutt, A. Sysa-jedrzejowska et al., “Tumour necrosis factor- α polymorphism as one of the complex inherited factors in pemphigus,” *Mediators of Inflammation*, vol. 12, no. 5, 307 pages, 2003.
- [35] Y. M. Mosaad, H. Fathy, Z. Fawzy, and M. A. El-Saied, “Tumor necrosis factor- α -308 G>A and interleukin-6 -174 G>C promoter polymorphisms and pemphigus,” *Human Immunology*, vol. 73, no. 5, pp. 560–565, 2012.
- [36] S. A. Dar, N. Akhter, S. Haque et al., “Tumor necrosis factor (TNF)- α -308G/a (rs1800629) polymorphism distribution in North India and its association with pemphigus: case-control study and meta-analysis,” *Autoimmunity*, vol. 49, no. 3, pp. 179–187, 2016.
- [37] J. Bidwell, L. Keen, G. Gallagher et al., “Cytokine gene polymorphism in human disease: on-line databases,” *Genes and Immunity*, vol. 1, no. 1, pp. 3–19, 1999.
- [38] P. Juszczyński, E. Kalinka, J. Bienvenu et al., “Human leukocyte antigens class II and tumor necrosis factor genetic polymorphisms are independent predictors of non-Hodgkin lymphoma outcome,” *Blood*, vol. 100, no. 8, pp. 3037–3040, 2002.
- [39] F. Pociot, A. G. Wilson, J. Nerup, and G. W. Duff, “No independent association between a tumor necrosis factor- α promoter region polymorphism and insulin-dependent diabetes mellitus,” *European Journal of Immunology*, vol. 23, no. 11, pp. 3050–3053, 1993.
- [40] B. Brinkman, D. Zuijdeest, E. Kaijzel, F. Breedveld, and C. Verweij, “Relevance of the tumor necrosis factor alpha (TNF alpha) -308 promoter polymorphism in TNF alpha gene regulation,” *Journal of Inflammation*, vol. 46, no. 1, pp. 32–41, 1995.
- [41] O. Abida, N. Mahfoudh, A. Kammoun et al., “Polymorphisms of HLA microsatellite marker in Tunisian pemphigus foliaceus,” *Human Immunology*, vol. 74, no. 1, pp. 104–109, 2013.



Hindawi

Submit your manuscripts at
www.hindawi.com

