

## Research Article

# Imbalance of Circulatory T Follicular Helper and T Follicular Regulatory Cells in Patients with ANCA-Associated Vasculitis

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Antineutrophil cytoplasmic antibody- (ANCA-) associated vasculitis (AAV) is characterized by small-vessel inflammation in association with autoantibodies. Balance between T follicular helper (Tfh) cells and T follicular regulatory (Tfr) cells is critical for humoral immune responses. Accumulating evidence supports that Tfh and Tfr are involved in autoimmune diseases; however, their roles in AAV are unclear. In this study, we tested the changes of circulatory Tfh and Tfr in patients with AAV. Twenty patients with AAV and twenty healthy controls were enrolled. Sixteen AAV patients had kidney involvement. We found that the AAV patients had increased circulating Tfh cells ( $CD4^+CXCR5^+CD25^-CD127^{interm-hi}$ ), decreased Tfr cells ( $CD4^+CXCR5^+CD25^+CD127^{lo-interm}$ ), and elevated Tfh/Tfr ratios compared with healthy controls ( $P < 0.01$ ). The Tfh percentage and Tfh/Tfr ratio, but not Tfr percentage, were positively correlated to proteinuria levels and BVAS scores in patients with AAV ( $P < 0.01$ ). In addition, AAV patients had decreased circulating Tfh1 ( $CCR6^-CXCR3^+$ ), but increased Tfh2 cells ( $CCR6^+CXCR3^-$ ), compared with healthy controls ( $P < 0.01$ ), indicating a Tfh1-to-Tfh2 shift. Furthermore, remission achieved by immunosuppressive treatment markedly attenuated the increase of total Tfh ( $P < 0.01$ ) and Tfh2 cells ( $P < 0.05$ ), promoted the Tfh1 response ( $P < 0.05$ ), and recovered the balance between Tfh/Tfr cells ( $P < 0.05$ ) and between Tfh1/Tfh2 cells ( $P < 0.05$ ) in patients with AAV. Plasma levels of IL-21, a cytokine secreted by Tfh cells, were elevated in AAV patients compared with healthy controls ( $P < 0.01$ ), which was attenuated by immunosuppressive treatment ( $P < 0.05$ ). Taken together, our findings indicate that circulatory Tfh/Tfr ratios, Tfh2/Tfh1 shift, and plasma IL-21 levels are associated with AAV and disease activity.

## 1. Introduction

Antineutrophil cytoplasmic antibody- (ANCA-) associated vasculitis (AAV) is a group of potentially life-threatening autoimmune diseases in which the kidney is frequently involved [1]. Pathogenesis of AAV is not fully understood by far [2]. Majority of patients with AAV can achieve temporary disease remission with immunosuppressive induction therapy. However, acute relapses occur in more than 50% of patients during follow-up [3]. Therefore, prediction of

AAV relapse has become a critical and unsolved topic. Biomarkers that are correlated with the disease activity of AAV should be useful for relapse prediction; however, they are still unavailable [4]. ANCA are autoantibodies that are abnormally produced to target and attack cytoplasmic granules of neutrophils and monocytes, leading to necrotizing vascular inflammation [5]. Although ANCA has been considered the initiator of AAV and used for AAV diagnosis for decades, ANCA titers are not associated with disease activity and are not able to predict relapse in patients with AAV [6–8]. Thus,

it is important to investigate novel biomarker candidates that are associated with ANCA and disease activity.

Although production of ANCA autoantibodies is naturally dependent on B cells, certain subsets of T cells play a regulatory role in B cell response and autoantibody synthesis [9, 10]. T follicular helper (Tfh) cells promote germinal center formation and support B cell producing antibodies, while T follicular regulatory (Tfr) cells suppress antibody production [10, 11]. Both Tfh and Tfr cells are subsets of CXCR5<sup>+</sup>CD4<sup>+</sup> T cells. One of their distinctions is that Tfh cells are CD25<sup>-</sup>CD127<sup>interm-hi</sup> whereas Tfr cells are CD25<sup>+</sup>CD127<sup>lo-interm</sup>. As Tfh and Tfr cells have opposing roles in regulating humoral immune responses, their balance is important for immune homeostasis [9, 11, 12]. Imbalance between Tfh and Tfr cells promotes defective antibody production and contributes to the development of autoimmunity [13–15]. Recently, Tfh cells are subdivided into Tfh1 (CCR6<sup>-</sup> CXCR3<sup>+</sup>) and Tfh2 (CCR6<sup>-</sup> CXCR3<sup>-</sup>) subtypes, and the Tfh1/Tfh2 ratio can be altered under certain pathological conditions. However, the roles of Tfh/Tfr balance and Tfh2/Tfh1 shift in AAV are unclear.

Following immune responses, a small number of Tfh and Tfr cells from lymph nodes are released into the circulation. Circulatory Tfh and Tfr cells serve like memory cells and are able to react quickly during subsequent immune responses [16]. In addition, circulatory Tfh and Tfr cells are convenient to test, and their counts could represent the balance between Tfh and Tfr cells [17]. In this study, we investigated the relationship between Tfh/Tfr balance and disease activity in patients with AAV. We found that, compared with healthy controls, AAV patients had increased circulatory Tfh/Tfr and Tfh2/Tfh1 ratios, which were attenuated during disease remission.

## 2. Materials and Methods

**2.1. Patients.** The study was approved by the ethical committee of the First Hospital of Jilin University. Written informed consent was obtained from all individual participants. We enrolled twenty AAV patients with kidney involvement and twenty age-matched healthy controls from the outpatients and inpatients in the First Hospital of Jilin University during July 2016 to June 2017. Demographic, clinical, and laboratory parameters of the patients and healthy controls were collected. Remission was achieved by treatment with a combination of glucocorticoids and cyclophosphamide following the current guideline for clinical practice [18]. AAV is defined as necrotizing vasculitis associated with ANCA and was diagnosed according to the 1994 Chapel Hill Consensus Conference Criteria for Vasculitis [19]. Disease activity of AAV was assessed using the Birmingham Vasculitis Activity Score (BVAS) system. A BVAS of >0 was considered to represent active disease, while BVAS = 0 was defined as remission [19]. The AAV subtypes of microscopic polyangiitis and granulomatosis with polyangiitis were defined based on the Chapel Hill Consensus Criteria [20]. Kidney involvement was defined by a kidney biopsy showing pauci-immune glomerulonephritis or active urine sediment with or without worsening renal dysfunction along with a diagnostic biopsy

TABLE 1: Demographic and clinical characteristics of the patients.

	Healthy controls <i>n</i> = 20	Patients with AAV <i>n</i> = 20
Age (years)	64 ± 3	63 ± 2
Sex (females, %)	10 (50.0%)	9 (45.0%)
p-ANCA <sup>+</sup> ( <i>n</i> , %)	N/A	16 (80.0%)
c-ANCA <sup>+</sup> ( <i>n</i> , %)	N/A	3 (15.0%)
Proteinuria (g/24 h)	N/A	1.56 ± 0.27
Serum creatinine (μmol/L)	72.2 ± 20.3	368.9 ± 60.7**
Blood urea nitrogen (mmol/L)	4.1 ± 0.8	17.0 ± 2.6**
C-reactive protein (mg/L)		62.1 ± 49.3
Erythrocyte sedimentation rate (mm/h)		65.0 ± 38.1
Diagnosis ( <i>n</i> , %)		
Granulomatosis with polyangiitis		6 (30%)
Microscopic polyangiitis		14 (70%)
Lung complications ( <i>n</i> , %)		16 (80%)

\*\**P* < 0.01 vs. healthy control.

result of small-vessel necrotizing vasculitis in an extrarenal tissue [21]. Lung complications were defined as described previously [22].

**2.2. Flow Cytometry.** Fresh venous blood samples were collected from healthy controls and the patients with AAV at the time of diagnosis and when the patients achieved disease remission. Peripheral blood mononuclear cells were isolated by density-gradient centrifugation using the Ficoll-Paque. The isolated cells were stained with the following antibodies: Alexa Fluor 700-conjugated anti-CD4 (Cat#557922, BD Biosciences, San Jose, CA, USA), PerCP-Cy5.5-conjugated CXCR5 (Cat#562781, BD Biosciences), Horizon V500-conjugated CD127 (Cat#563086, BD Biosciences), Alexa Fluor 647-conjugated anti-CD25 (Cat#563598, BD Biosciences), APC-Cy7-conjugated CXCR3 (Cat#353721, BioLegend, San Diego, CA, USA), Alexa Fluor 488-conjugated anti-forkhead box P3 (FOXP3, Cat#561181, BD Biosciences), and PE-Cy7-conjugated CCR6 (Cat#560620, BD Biosciences). Flow cytometry was analyzed with an LSR II instrument (BD Biosciences). The data were analyzed with FlowJo software (Tree Star, Ashland, OR, USA).

**2.3. Enzyme-Linked Immunosorbent Assays (ELISA).** Plasma was collected from the whole blood. Plasma interleukin-21 (IL-21) concentrations were measured by an ELISA kit (Abcam, Cambridge, MA, USA).

**2.4. Statistical Analysis.** Statistical analysis was performed with GraphPad Prism 6.0 software (GraphPad Software Inc., San Diego, CA, USA). Normally distributed continuous variables were presented as the mean ± standard deviation and analyzed by Student's *t*-test for comparing two groups or one-way ANOVA with the post hoc Tukey honestly significant difference test for comparing multiple groups.

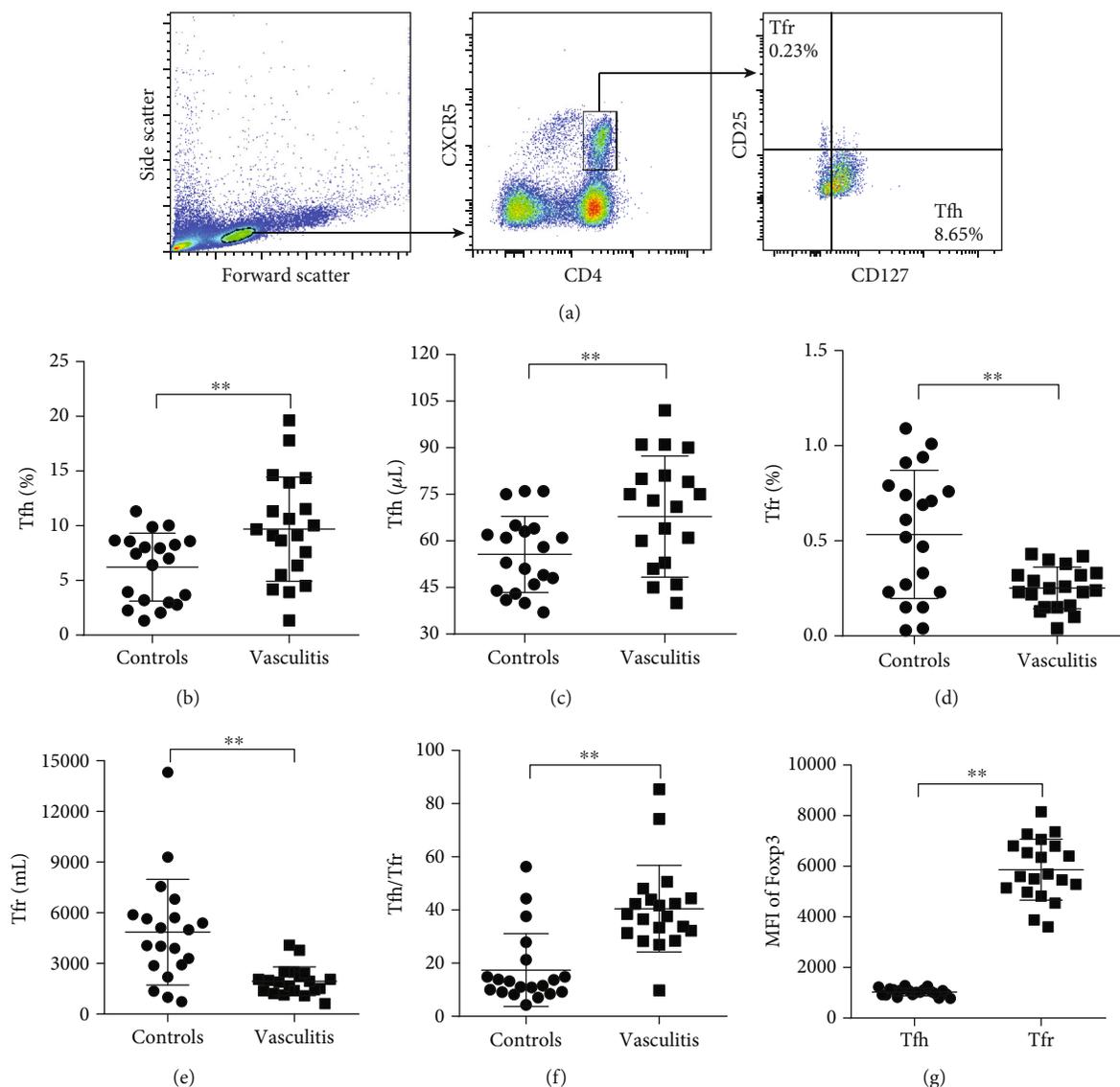


FIGURE 1: Circulatory Tfh and Tfr cells in patients with antineutrophil cytoplasmic antibody- (ANCA-) associated vasculitis (AAV). (a) The gate strategy for circulating Tfh and Tfr cells. The percentage and count of circulating Tfh (b, c) and Tfr (d, e) cells and Tfh/Tfr ratios (f) in patients with AAV and healthy controls. (g) Mean fluorescence intensity (MFI) of FOXP3 in Tfh and Tfr cells. \*\* $P < 0.01$ . Tfh: T follicular helper cells; Tfr: T follicular regulatory cells.

Pearson's correlation and linear regression were performed to examine the relationship between two parameters. Differences were considered significant at a probability level of  $P < 0.05$ .

### 3. Results

**3.1. Patient Characteristics.** The demographic, clinical, and laboratory characteristics of the patients with AAV and healthy controls are shown in Table 1. Sixteen of the twenty AAV patients had kidney involvement and dysfunction (Table 1).

**3.2. Increased Circulatory Tfh but Decreased Tfr Cells in Patients with AAV.** Circulatory CD4<sup>+</sup> T cell percentages were similar between patients with AAV and healthy controls

( $51.49 \pm 12.49\%$  vs.  $46.21 \pm 14.89\%$ ,  $P > 0.05$ ). Circulatory Tfh and Tfr cells were analyzed using a previously reported gating strategy [17]. Figure 1(a) demonstrates circulating Tfh ( $CD4^+CXCR5^+CD25^-CD127^{interm-hi}$ ) and Tfr ( $CD4^+CXCR5^+CD25^+CD127^{lo-interm}$ ) cell populations that were defined by the expression of CD4 and CXCR5 and further separated by the expression of CD25 and CD127. Patients with AAV had a higher number of circulating Tfh cells (Figures 1(b) and 1(c)) while less circulating Tfr cells (Figures 1(d) and 1(e)) than healthy controls (all  $P < 0.01$ ), and the Tfh/Tfr ratio was increased in patients with AAV compared to healthy controls ( $P < 0.01$ , Figure 1(f)). Expression of FOXP3, a T regulatory (Treg) cell marker was abundant in Tfr cells but was much less and minimal in Tfh cells ( $P < 0.01$ , Figure 1(g)). These results indicate that patients with

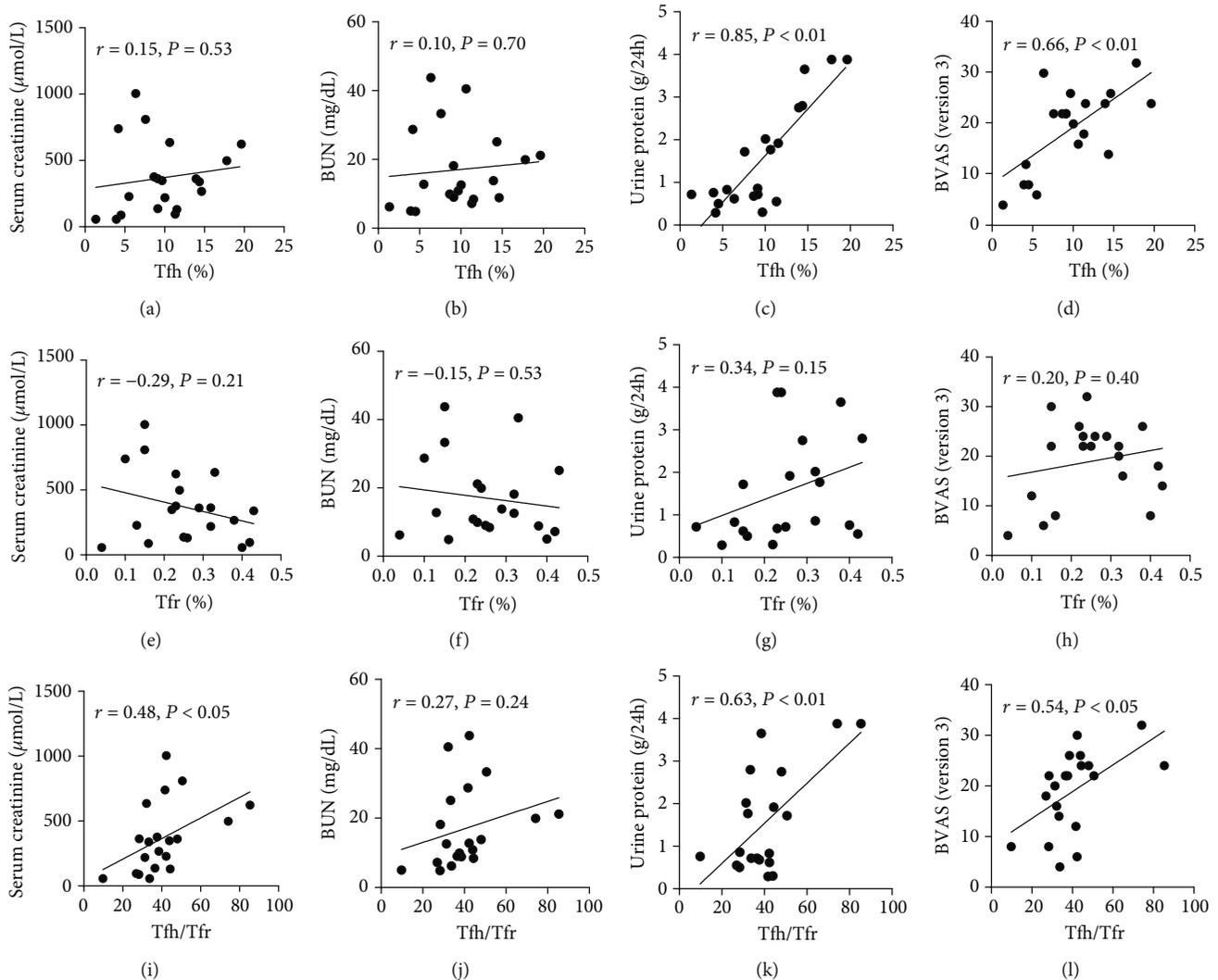


FIGURE 2: Correlation between circulatory Tfh/Tfr cells and renal injury. Correlation of the percentages of circulatory Tfh cells (a–d), Tfr cells (e–h), and Tfh/Tfr ratios (i–l) with serum creatinine, blood urea nitrogen, 24 h urinary protein levels, and BVAS scores in patients with AAV.

AAV have an imbalance between circulatory Tfh and Tfr cell populations.

**3.3. Tfh/Tfr Ratios Correlate with Disease Activity and Renal Injury.** We next evaluated the correlation between Tfh/Tfr ratios and disease activity or renal injury. In patients with AAV, the percentage of circulatory Tfh cells was not significantly correlated with serum creatinine ( $r = 0.15, P = 0.53$ , Figure 2(a)) and blood urea nitrogen ( $r = 0.10, P = 0.70$ ; Figure 2(b)) concentrations, but positively correlated with 24h urinary protein levels ( $r = 0.85, P < 0.01$ ; Figure 2(c)) and BVAS scores ( $r = 0.66, P < 0.01$ ; Figure 2(d)). The percentage of circulatory Tfr population was significantly correlated with neither serum creatinine ( $r = 0.29, P = 0.21$ ; Figure 2(e)), blood urea nitrogen ( $r = -0.15, P = 0.53$ ; Figure 2(f)), 24h urinary protein levels ( $r = 0.34, P = 0.15$ ; Figure 2(g)), nor BVAS scores ( $r = 0.20, P = 0.40$ ; Figure 2(h)). Interestingly, the Tfh/Tfr ratios were positively correlated with serum creatinine ( $r = 0.48, P < 0.05$ ;

Figure 2(i)), 24h urinary protein levels ( $r = 0.63, P < 0.01$ ; Figure 2(k)), and BVAS scores ( $r = 0.54, P < 0.05$ ; Figure 2(l)), but not significantly correlated with blood urea nitrogen ( $r = 0.27, P = 0.24$ ; Figure 2(j)).

**3.4. Increased Circulatory Tfh2 and Decreased Tfh1 Subsets in Patients with AAV.** Tfh cells ( $\text{CD4}^+\text{CXCR5}^+\text{CD25}^-\text{CD127}^{\text{interm-hi}}$ ) were further divided into Tfh1 ( $\text{CCR6}^-\text{CXCR3}^+$ ) and Tfh2 ( $\text{CCR6}^-\text{CXCR3}^-$ ) based on a documented gating strategy [23]. As shown in Figure 3(a), most Tfh cell populations were negative for CCR6, which were defined as Tfh1 or Tfh2. Tfh17 ( $\text{CCR6}^+\text{CXCR3}^-$ ) and Tfh1/17 ( $\text{CCR6}^+\text{CXCR3}^+$ ) were too low to be calculated in both patients with AAV and healthy controls. We included the positive control for CCR6 staining in the Supplementary Materials (Figure S1). We found that patients with AAV had a lower percentage and count of circulatory Tfh1 cells while more circulatory Tfh2 cells than healthy controls ( $P < 0.01$  or  $P < 0.05$ , Figures 3(b)–3(e)).

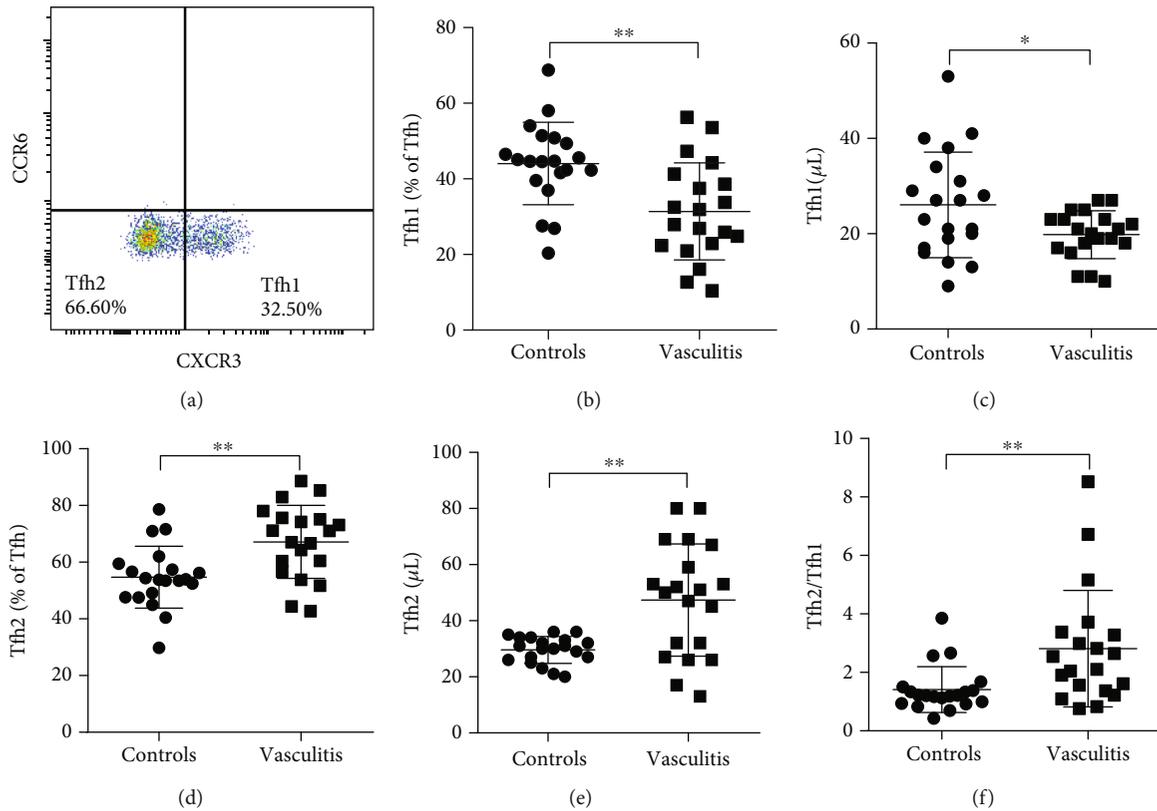


FIGURE 3: Circulatory Tfh1 and Tfh2 cells in patients with AAV. (a) The gate strategy for circulatory Tfh1 and Tfh2 cells. The percentage and count of circulatory Tfh1 (b, c) and Tfh2 (d, e) cells and Tfh2/Tfh1 ratios (f) in patients with AAV and healthy controls. \* $P < 0.05$ ; \*\* $P < 0.01$ . Tfh: T follicular helper cells.

In addition, the Tfh2/Tfh1 ratio was increased in patients with AAV when compared to healthy controls ( $P < 0.01$ , Figure 3(f)). These results suggest that patients with AAV have a shift from Tfh1 to Tfh2 cell populations.

**3.5. Remission Is Associated with Decreased Tfh Cells.** Disease remission was reached in all twenty patients with AAV after immunosuppressive treatment with a combination of glucocorticoids and cyclophosphamide. We found that the Tfh cell percentage and count were significantly decreased after the treatment compared to baseline ( $P < 0.01$  or  $P < 0.05$ , Figures 4(a) and 4(b)). However, the Tfr cell percentage and count were not significantly altered by the treatment (Figures 4(c) and 4(d)). Accordingly, the Tfh/Tfr ratios were decreased in AAV patients with remission compared to baseline ( $P < 0.05$ , Figure 4(e)). In addition, immunosuppressive treatment significantly increased circulatory Tfh1 cell percentage and count (both  $P < 0.05$ , Figures 4(f) and 4(g)), decreased Tfh2 cells (both  $P < 0.05$ , Figures 4(h) and 4(i)), and lowered the Tfh2/Tfh1 ratios ( $P < 0.05$ , Figure 4(j)) in patients with AAV.

**3.6. Increased Plasma IL-21 Levels in Patients with AAV.** IL-21 is produced by Tfh cells and is critical for germinal center formation [24]. Plasma IL-21 levels were higher in patients with AAV than in healthy controls ( $P < 0.01$ , Figure 5). As expected, plasma IL-21 levels were significantly decreased after remission ( $P < 0.05$ , Figure 5).

## 4. Discussion

Accumulating evidence has shown that dysregulation of Tfh or Tfr cells contributes to the development of autoimmune diseases. Here, we showed that patients with AAV had increased circulatory Tfh/Tfr and Tfh2/Tfh1 ratios and elevated plasma IL-21 levels compared with healthy controls, which were attenuated when disease remission was successfully achieved by immunosuppressive treatment.

Tfh cells are a novel T cell subset in the germinal center [11]. B cells in the germinal center are responsible for producing autoantibodies which are the culprit of a variety of autoimmune diseases, including AAV [5]. Tfh cells play a critical role in autoantibody production through helping B cells form germinal centers and driving B cells to differentiate into antibody-producing plasma cells [25]. Circulatory Tfh cell population is increased in myasthenia gravis, systemic lupus erythematosus (SLE), and multiple sclerosis [26, 27]. The present study identified increased Tfh cells in patients with AAV, which is consistent with previous studies showing that Tfh cells were increased in patients with granulomatosis with polyangiitis and in patients with immunoglobulin A vasculitis [23, 28]. These findings suggest that increased Tfh cells may contribute to the development of AAV.

Tfr cells are a specialized subset of Treg cells [29]. Like Tfh cells, Tfr cells also express CXCR5 which help direct

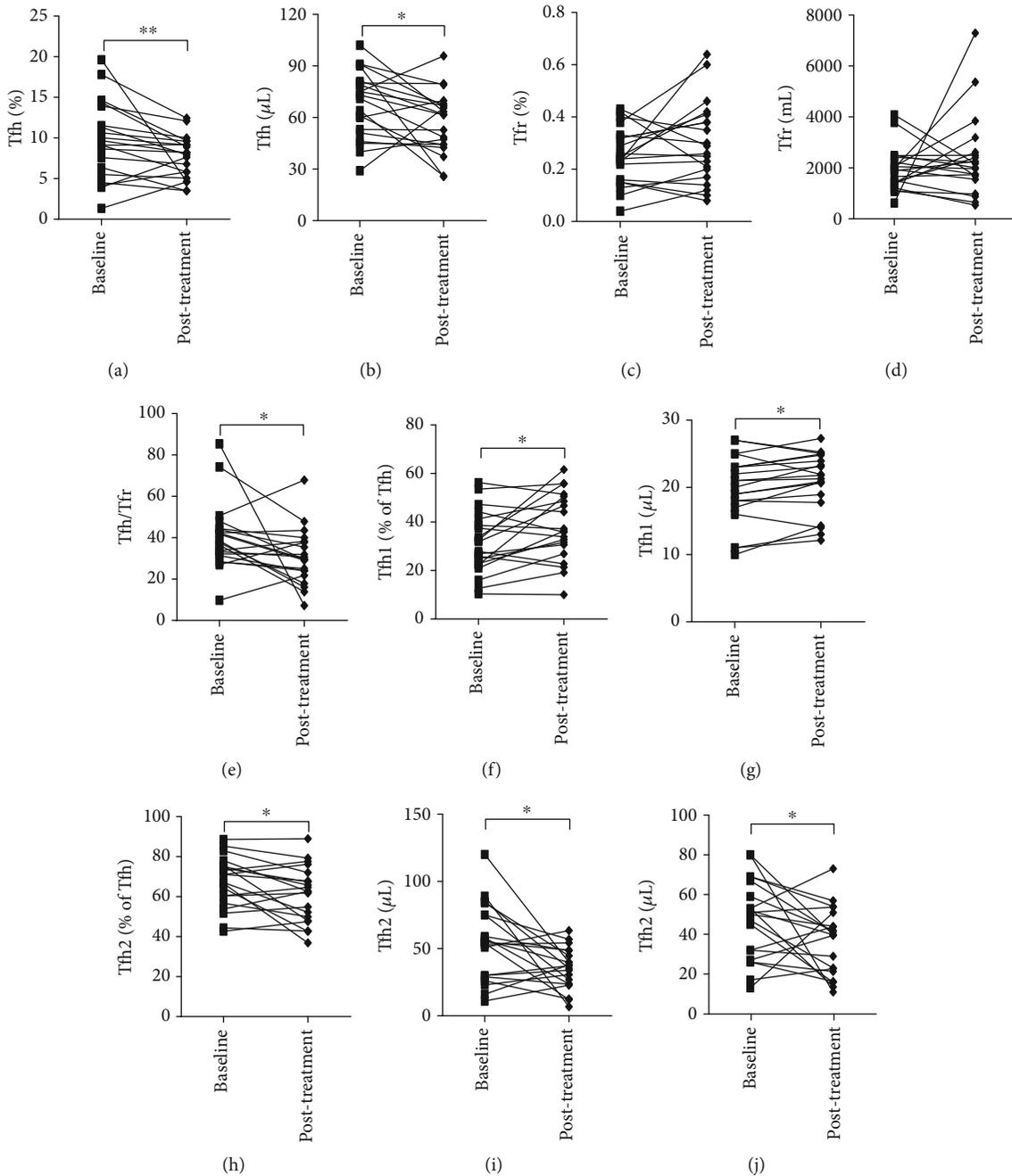


FIGURE 4: Effects of immunosuppressive treatment on circulating Tfh/Tfr and Tfh2/Tfh1 ratios. The percentage and count of circulating Tfh (a, b) and Tfr (c, d) cells and Tfh/Tfr ratios (e) in patients with AAV at baseline and after remission were achieved by immunosuppressive treatment. The percentage of circulating Tfh1 (f, g) and Tfh2 (h, i) cells and Tfh2/Tfh1 ratios (j) in patients with AAV at baseline and after remission was achieved by immunosuppressive treatment. \* $P < 0.05$ ; \*\* $P < 0.01$ .

them into the germinal center [14]. Tfr cells play a suppressive role in germinal center reactions [9, 10, 29]. Given that Tfh and Tfr cells have opposite effects on humoral immune responses, the balance between them is critical for immune homeostasis. The uncontrolled decrease of Tfr activity or breakdown of Tfh/Tfr balance can cause autoimmunity, leading to autoimmune diseases [13, 14, 26, 30–32]. For instance, decreased Tfr cells have been found in myasthenia gravis, SLE, and multiple sclerosis [17, 33, 34]. Evidence

shows that AAV patients have defective Treg cell function [35]. However, the role of Tfr in AAV has not been reported. The present study demonstrated that patients with AAV had decreased Tfr cells and increased Tfh/Tfr ratios compared with healthy controls. These results suggest that Tfr and Treg cells play a similar role in AAV. Collectively, the findings indicate that imbalance between Tfh and Tfr may contribute to the development of AAV. There are also reports showing increased Treg or Tfr cells in autoimmune

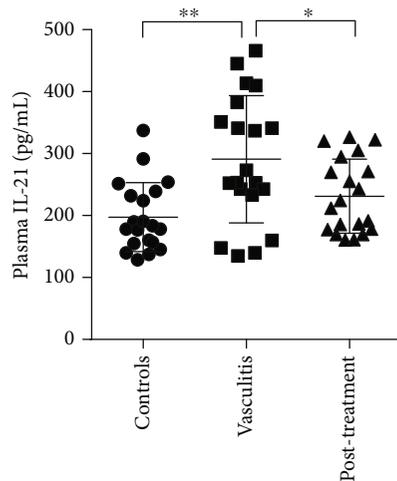


FIGURE 5: Plasma levels of cytokine IL-21. Plasma IL-21 levels were detected in healthy controls and patients with AAV at baseline and after remission achieved by immunosuppressive treatment. \* $P < 0.05$ ; \*\* $P < 0.01$ .

diseases, including Sjögren syndrome, SLE, and rheumatoid arthritis [30, 36–40]. These differences may reflect different pathogenic mechanisms between AAV and other autoimmune diseases.

Kidney involvement frequently occurs in patients with AAV, and delayed diagnosis of ANCA-associated renal injury is potentially organ- and life-threatening [41, 42]. In the present study, we found that the percentage of Tfh cells was associated with proteinuria but not serum creatinine and blood urea nitrogen, which suggests that Tfh cells may be related to kidney injury rather than the status of renal function. The percentage of Tfr cells was not associated with either proteinuria or renal function parameters, indicating that Tfr cells may not be a sensitive biomarker for detecting renal injury. Interestingly, the Tfh/Tfr ratio was correlated to both proteinuria and renal function, indicating that the Tfh/Tfr ratio could be a better parameter for evaluating kidney involvement in patients with AAV.

As Tfh cells may play an important role in AAV, we further investigate the distribution of its subtypes. The present study demonstrated that AAV patients had decreased Tfh1 cells, increased Tfh2 cells, and Tfh2/Tfh1 ratios. The results are consistent with those of a previous study showing that Tfh2 cells were increased in child patients with immunoglobulin A vasculitis [23]. However, they found that Tfh17 cells were decreased while Tfh1 and Tfh1/17 cells were not altered when compared to healthy controls [23]. The percentages of Tfh17 and Tfh1/17 were quite low in the subjects of the present study, so these two subsets were not analyzed. Some previous studies also demonstrated that the percentage of Tfh17 was the least subset and was minimal even in healthy controls [34]. The changes and function of Tfh17 cells in autoimmune disease warrant further investigations. Currently, the function of Tfh1 and Tfh2 subpopulations is not well documented. A previous study demonstrated that

Tfh1 cells lack the capacity to help naïve B cells and that patients with juvenile dermatomyositis, an autoimmune disease, had decreased Tfh1 and increased Tfh2 cells [43, 44]. These findings suggest that a Tfh1-to-Tfh2 shift might be involved in the development of autoimmune disease. However, a correlation between Tfh1 cells and autoantibody titers was not observed in the present study (data not shown). Taken together, the Tfh2/Tfh1 ratio may be used as a biomarker for AAV, but the role of these subsets in AAV requires further experimental investigation.

We found that immunosuppressive treatment attenuated the increases in Tfh cells and Tfh/Tfr ratios in patients with AAV. These results indicate that Tfh cells and Tfh/Tfr ratios are associated with remission of AAV. These two parameters may be useful for predicting relapse, however, which warrants further study. Previous studies demonstrated that immunosuppressive treatment with sirolimus increased Treg cells [45]. In contrast, the present study showed that remission induction failed to change the decreased Tfr cells, suggesting that immunosuppressive treatment may not target the Tfr subset.

IL-21 is a specific cytokine secreted by Tfh cells. It was not surprising that changes in plasma IL-21 concentrations were consistent with the percentage of Tfh cells. Percentages of IL-21 producing Tfh cells were significantly increased in patients with granulomatosis with polyangiitis [46]. However, plasma IL-21 concentrations in patients with AAV were not reported previously. Given the more feasible measurement of IL-21 than Tfh cells, plasma IL-21 may be a promising biomarker for patients with AAV.

The relationship between Tfh and Tfr cells was not fully understood. Based on the current evidence, Tfh cells may suppress the generation of Tfr cells through secreting IL-21, and IL-21 may play a critical role in the maintenance of Tfh and Tfr cell balance [47]. It was reported that IL-21 was able to suppress Tfr cell-mediated suppressive effects on B cells [48]. In turn, Tfr cells can impair Tfh cells [49]. Taken together, the direct relationship between Tfh and Tfr cells is still unclear. Serial measurements of Tfh and Tfr cells in patients with AAV or *in vitro* experimental studies may help uncover the causative correlation of Tfh and Tfr cells.

There are some limitations in the present study. The sample size was relatively small. The differences in terms of Tfh, Tfr, Tfh1, and Tfh2 subsets between seronegative and seropositive patients could be analyzed with a larger number of cases. The changes of the Tfh/Tfr ratio that we found in the present study may be not specific to patients with AAV. Changes of the Tfh/Tfr ratio may also be present in other autoimmune diseases such as lupus nephritis, which needs to be tested in future studies. Another limitation is that we only selected patients who achieved remission with standard therapy. The comparison between patients with remission and refractory patients should be helpful to define the utility of Tfh/Tfr as a prognostic factor in patients with AAV. In addition, analyzing the relationship between AAV relapse and Tfh/Tfr or IL-21 levels should be helpful in the prediction of AAV relapse.

## 5. Conclusion

In conclusion, circulatory Tfh/Tfr ratios, Tfh2/Tfh1 ratios, and plasma IL-21 levels are increased in patients with AAV and may be associated with disease activity, renal injury, and remission in patients with AAV.

## Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

## Disclosure

Ying Xu and Hongmei Xu contributed equally as the first authors.

## Conflicts of Interest

The authors declare that there is no conflict of interest regarding the publication of this article.

## Authors' Contributions

YX, HX, and HY designed the study. YX, HX, YZ, XS, HW, and CH are responsible for the patient diagnosis, follow-up, and data collection. YX, HX, ZM, and MY are assigned to the data interpretation and analysis. YX, HX, and HY drafted the manuscript and revised critically the important intellectual content. All authors read and approved the final version to be published.

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## Supplementary Materials

Figure S1: positive controls for CCR6 staining. (*Supplementary Materials*)

## References

- [1] S. Moiseev, P. Novikov, D. Jayne, and N. Mukhin, "End-stage renal disease in ANCA-associated vasculitis," *Nephrology, Dialysis, Transplantation*, vol. 32, pp. gfw046–gfw053, 2017.
- [2] E. J. Gapud, P. Seo, and B. Antiochos, "ANCA-associated vasculitis pathogenesis: a commentary," *Current Rheumatology Reports*, vol. 19, no. 4, p. 15, 2017.
- [3] E. M. Miloslavsky, U. Specks, P. A. Merkel et al., "Outcomes of nonsevere relapses in antineutrophil cytoplasmic antibody-associated vasculitis treated with glucocorticoids," *Arthritis & Rheumatology*, vol. 67, no. 6, pp. 1629–1636, 2015.
- [4] U. Specks, "Accurate relapse prediction in ANCA-associated vasculitis—the search for the Holy Grail," *Journal of the American Society of Nephrology*, vol. 26, no. 3, pp. 505–507, 2015.
- [5] C. G. Kallenberg, C. A. Stegeman, W. H. Abdulahad, and P. Heeringa, "Pathogenesis of ANCA-associated vasculitis: new possibilities for intervention," *American Journal of Kidney Diseases*, vol. 62, no. 6, pp. 1176–1187, 2013.
- [6] R. Nowack, I. Grab, L. F. Flores-Suarez, P. Schnulle, B. Yard, and F. J. van der Woude, "ANCA titres, even of IgG subclasses, and soluble CD14 fail to predict relapses in patients with ANCA-associated vasculitis," *Nephrology, Dialysis, Transplantation*, vol. 16, no. 8, pp. 1631–1637, 2001.
- [7] J. Ara, E. Mirapeix, R. Rodriguez, A. Saurina, and A. Darnell, "Relationship between ANCA and disease activity in small vessel vasculitis patients with anti-MPO ANCA," *Nephrology, Dialysis, Transplantation*, vol. 14, no. 7, pp. 1667–1672, 1999.
- [8] G. Tomasson, P. C. Grayson, A. D. Mahr, M. Lavalley, and P. A. Merkel, "Value of ANCA measurements during remission to predict a relapse of ANCA-associated vasculitis—a meta-analysis," *Rheumatology*, vol. 51, no. 1, pp. 100–109, 2012.
- [9] M. A. Linterman, W. Pierson, S. K. Lee et al., "Foxp3<sup>+</sup> follicular regulatory T cells control the germinal center response," *Nature Medicine*, vol. 17, no. 8, pp. 975–982, 2011.
- [10] I. Wollenberg, A. Agua-Doce, A. Hernandez et al., "Regulation of the germinal center reaction by Foxp3<sup>+</sup> follicular regulatory T cells," *The Journal of Immunology*, vol. 187, no. 9, pp. 4553–4560, 2011.
- [11] J. S. Weinstein, E. I. Herman, B. Lainez et al., "TFH cells progressively differentiate to regulate the germinal center response," *Nature Immunology*, vol. 17, no. 10, pp. 1197–1205, 2016.
- [12] P. T. Sage, L. M. Francisco, C. V. Carman, and A. H. Sharpe, "The receptor PD-1 controls follicular regulatory T cells in the lymph nodes and blood," *Nature Immunology*, vol. 14, no. 2, pp. 152–161, 2013.
- [13] P. T. Sage, C. L. Tan, G. J. Freeman, M. Haigis, and A. H. Sharpe, "Defective TFH cell function and increased TFR cells contribute to defective antibody production in aging," *Cell Reports*, vol. 12, no. 2, pp. 163–171, 2015.
- [14] P. T. Sage and A. H. Sharpe, "T follicular regulatory cells in the regulation of B cell responses," *Trends in Immunology*, vol. 36, no. 7, pp. 410–418, 2015.
- [15] Q. Niu, Z. C. Huang, X. J. Wu et al., "Enhanced IL-6/phosphorylated STAT3 signaling is related to the imbalance of circulating T follicular helper/T follicular regulatory cells in patients with rheumatoid arthritis," *Arthritis Research & Therapy*, vol. 20, no. 1, p. 200, 2018.
- [16] P. T. Sage, D. Alvarez, J. Godec, U. H. von Andrian, and A. H. Sharpe, "Circulating T follicular regulatory and helper cells have memory-like properties," *The Journal of Clinical Investigation*, vol. 124, no. 12, pp. 5191–5204, 2014.
- [17] B. Xu, S. Wang, M. Zhou et al., "The ratio of circulating follicular T helper cell to follicular T regulatory cell is correlated with disease activity in systemic lupus erythematosus," *Clinical Immunology*, vol. 183, pp. 46–53, 2017.
- [18] M. Yates, R. A. Watts, I. M. Bajema et al., "EULAR/ERA-EDTA recommendations for the management of ANCA-associated vasculitis," *Annals of the Rheumatic Diseases*, vol. 75, no. 9, pp. 1583–1594, 2016.
- [19] S. Gopaluni, O. Flossmann, M. A. Little et al., "Effect of disease activity at three and six months after diagnosis on long-term outcomes in antineutrophil cytoplasmic antibody-associated vasculitis," *Arthritis & Rheumatology*, vol. 71, no. 5, pp. 784–791, 2018.

- [20] J. C. Jennette, R. J. Falk, P. A. Bacon et al., “2012 revised International Chapel Hill Consensus Conference Nomenclature of Vasculitides,” *Arthritis & Rheumatism*, vol. 65, no. 1, pp. 1–11, 2013.
- [21] R. L. Rhee, S. L. Hogan, C. J. Poulton et al., “Trends in long-term outcomes among patients with antineutrophil cytoplasmic antibody-associated vasculitis with renal disease,” *Arthritis & Rheumatology*, vol. 68, no. 7, pp. 1711–1720, 2016.
- [22] D. R. Thickett, A. G. Richter, N. Nathani, G. D. Perkins, and L. Harper, “Pulmonary manifestations of anti-neutrophil cytoplasmic antibody (ANCA)-positive vasculitis,” *Rheumatology*, vol. 45, no. 3, pp. 261–268, 2006.
- [23] D. Liu, J. Liu, J. Wang et al., “Distribution of circulating T follicular helper cell subsets is altered in immunoglobulin A vasculitis in children,” *PLoS One*, vol. 12, no. 12, article e0189133, 2017.
- [24] R. Spolski and W. J. Leonard, “IL-21 and T follicular helper cells,” *International Immunology*, vol. 22, no. 1, pp. 7–12, 2010.
- [25] N. Gensous, M. Charrier, D. Duluc et al., “T follicular helper cells in autoimmune disorders,” *Frontiers in Immunology*, vol. 9, p. 1637, 2018.
- [26] Y. Gong, J. Tong, and S. Wang, “Are follicular regulatory T cells involved in autoimmune diseases?,” *Frontiers in Immunology*, vol. 8, p. 1790, 2017.
- [27] J. Y. Choi, J. H. Ho, S. G. Pasoto et al., “Circulating follicular helper-like T cells in systemic lupus erythematosus: association with disease activity,” *Arthritis & Rheumatology*, vol. 67, no. 4, pp. 988–999, 2015.
- [28] Y. Zhao, P. M. Lutalo, J. E. Thomas et al., “Circulating T follicular helper cell and regulatory T cell frequencies are influenced by B cell depletion in patients with granulomatosis with polyangiitis,” *Rheumatology*, vol. 53, no. 4, pp. 621–630, 2014.
- [29] Y. Chung, S. Tanaka, F. Chu et al., “Follicular regulatory T cells expressing Foxp3 and Bcl-6 suppress germinal center reactions,” *Nature Medicine*, vol. 17, no. 8, pp. 983–988, 2011.
- [30] V. R. Fonseca, A. Agua-Doce, A. R. Maceiras et al., “Human blood Tfh cells are indicators of ongoing humoral activity not fully licensed with suppressive function,” *Science Immunology*, vol. 2, no. 14, article eaan1487, 2017.
- [31] M. Puthenparampil, A. Zito, G. Pantano et al., “Peripheral imbalanced TFH/TFR ratio correlates with intrathecal IgG synthesis in multiple sclerosis at clinical onset,” *Multiple Sclerosis Journal*, vol. 25, no. 7, pp. 918–926, 2018.
- [32] W. Fu, X. Liu, X. Lin et al., “Deficiency in T follicular regulatory cells promotes autoimmunity,” *The Journal of Experimental Medicine*, vol. 215, no. 3, pp. 815–825, 2018.
- [33] Y. Wen, B. Yang, J. Lu, J. Zhang, H. Yang, and J. Li, “Imbalance of circulating CD4<sup>+</sup>CXCR5<sup>+</sup>FOXP3<sup>+</sup> Tfr-like cells and CD4<sup>+</sup>CXCR5<sup>+</sup>FOXP3<sup>-</sup> Tfh-like cells in myasthenia gravis,” *Neuroscience Letters*, vol. 630, pp. 176–182, 2016.
- [34] T. Dhaeze, E. Peelen, A. Hombrouck et al., “Circulating follicular regulatory T cells are defective in multiple sclerosis,” *The Journal of Immunology*, vol. 195, no. 3, pp. 832–840, 2015.
- [35] M. E. Free, D. O. Bunch, J. A. McGregor et al., “Patients with antineutrophil cytoplasmic antibody-associated vasculitis have defective Treg cell function exacerbated by the presence of a suppression-resistant effector cell population,” *Arthritis & Rheumatism*, vol. 65, no. 7, pp. 1922–1933, 2013.
- [36] M. Ivanchenko, L. A. Aqrawi, A. Bjork, M. Wahren-Herlenius, and K. Chemin, “FoxP3<sup>+</sup>CXCR5<sup>+</sup>CD4<sup>+</sup>T cell frequencies are increased in peripheral blood of patients with primary Sjögren’s syndrome,” *Clinical and Experimental Immunology*, vol. 195, no. 3, pp. 305–309, 2019.
- [37] C. Liu, D. Wang, Y. Song, S. Lu, J. Zhao, and H. Wang, “Increased circulating CD4<sup>+</sup>CXCR5<sup>+</sup>FoxP3<sup>+</sup> follicular regulatory T cells correlated with severity of systemic lupus erythematosus patients,” *International Immunopharmacology*, vol. 56, pp. 261–268, 2018.
- [38] V. R. Fonseca, V. C. Romao, A. Agua-Doce et al., “The ratio of blood T follicular regulatory cells to T follicular helper cells marks ectopic lymphoid structure formation while activated follicular helper T cells indicate disease activity in primary Sjögren’s syndrome,” *Arthritis & Rheumatology*, vol. 70, no. 5, pp. 774–784, 2018.
- [39] C. Liu, D. Wang, S. Lu et al., “Increased circulating follicular Treg cells are associated with lower levels of autoantibodies in patients with rheumatoid arthritis in stable remission,” *Arthritis & Rheumatology*, vol. 70, no. 5, pp. 711–721, 2018.
- [40] V. C. Romao, J. E. Fonseca, A. Agua-Doce, and L. Graca, “T follicular regulatory cells are decreased in patients with established treated rheumatoid arthritis with active disease: comment on the article by Liu et al.,” *Arthritis & Rheumatology*, vol. 70, no. 11, pp. 1893–1895, 2018.
- [41] V. Binda, G. Moroni, and P. Messa, “ANCA-associated vasculitis with renal involvement,” *Journal of Nephrology*, vol. 31, no. 2, pp. 197–208, 2018.
- [42] E. Houben, J. W. van der Heijden, B. van Dam, W. A. Bax, A. E. Voskuyl, and E. L. Penne, “Screening for renal involvement in ANCA-associated vasculitis: room for improvement?,” *The Netherlands Journal of Medicine*, vol. 75, no. 1, pp. 21–26, 2017.
- [43] N. Schmitt, S. E. Bentebibel, and H. Ueno, “Phenotype and functions of memory Tfh cells in human blood,” *Trends in Immunology*, vol. 35, no. 9, pp. 436–442, 2014.
- [44] R. Morita, N. Schmitt, S. E. Bentebibel et al., “Human Blood CXCR5<sup>+</sup>CD4<sup>+</sup> T Cells Are Counterparts of T Follicular Cells and Contain Specific Subsets that Differentially Support Antibody Secretion,” *Immunity*, vol. 34, no. 1, pp. 108–121, 2011.
- [45] Y. Li, Y. Shi, Y. Liao, L. Yan, Q. Zhang, and L. Wang, “Differential regulation of Tregs and Th17/Th1 cells by a sirolimus-based regimen might be dependent on STAT-signaling in renal transplant recipients,” *International Immunopharmacology*, vol. 28, no. 1, pp. 435–443, 2015.
- [46] W. H. Abdulahad, N. Lepse, C. A. Stegeman et al., “Increased frequency of circulating IL-21 producing Th-cells in patients with granulomatosis with polyangiitis (GPA),” *Arthritis Research & Therapy*, vol. 15, no. 3, p. R70, 2013.
- [47] Y. Ding, J. Li, P. Yang et al., “Interleukin-21 promotes germinal center reaction by skewing the follicular regulatory T cell to follicular helper T cell balance in autoimmune BXD2 mice,” *Arthritis & Rheumatology*, vol. 66, no. 9, pp. 2601–2612, 2014.
- [48] P. T. Sage, N. Ron-Harel, V. R. Juneja et al., “Suppression by T<sub>FR</sub> cells leads to durable and selective inhibition of B cell effector function,” *Nature Immunology*, vol. 17, no. 12, pp. 1436–1446, 2016.
- [49] B. Miles, S. M. Miller, J. M. Folkvord et al., “Follicular regulatory T cells impair follicular T helper cells in HIV and SIV infection,” *Nature Communications*, vol. 6, no. 1, p. 8608, 2015.



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