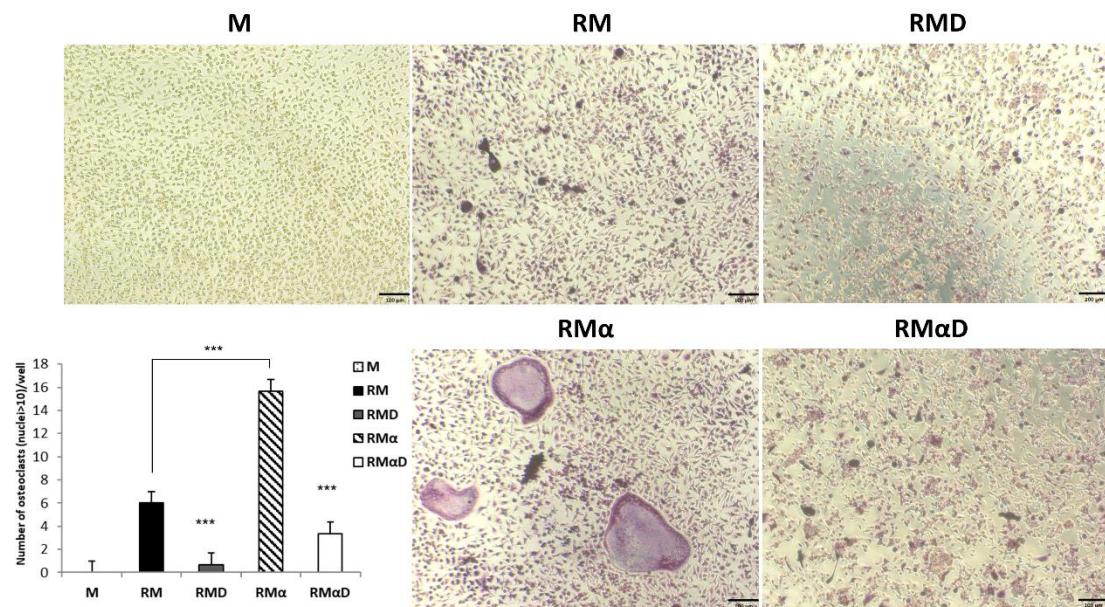


Supplementary figures

DCR3 affects osteoclast differentiation in BMMs

Figure 1



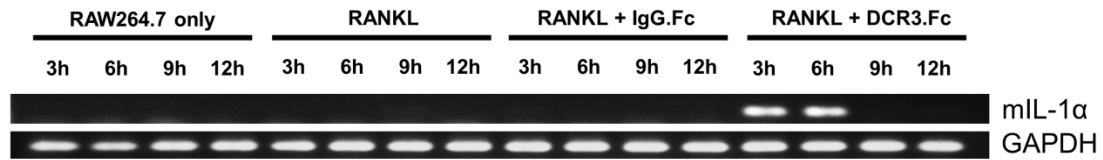
sFig. 1 Effects of DCR3 on RANKL plus IL-1 α induced osteoclast differentiation in BMMs.

BMMs were treated with DCR3 in the presence of RANKL (50 ng/ml) and M-CSF (30 ng/ml) or RANKL, IL-1 α , and M-CSF for 5 days. After incubation, the cells were fixed and stained for TRAP and TRAP⁺ multinucleated cells containing more than five nuclei in RAW264.7 or ten nuclei in BMMs were counted as multinucleated osteoclasts. (M: BMM cells + MCSF; RM: RANKL + MCSF; RMD: RANKL + MCSF + DCR3; RM α : RANKL + MCSF + IL-1 α ; RM α D: RANKL + MCSF + IL-1 α + DCR3) The data represent the means \pm S.D. of more than three cultures. (***) $P < 0.001$)

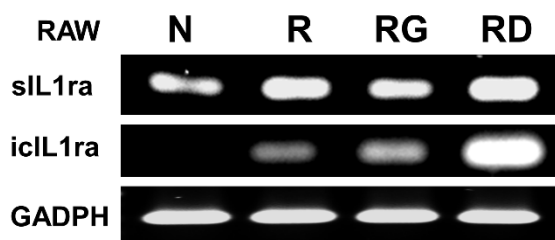
mRNA expression pattern of IL-1 α , icIL-1ra, and sIL-1ra in DCR3 treated OC

Figure 2

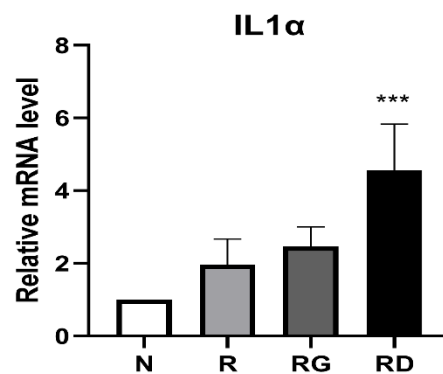
A



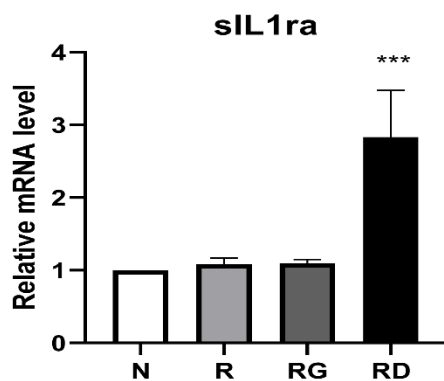
B



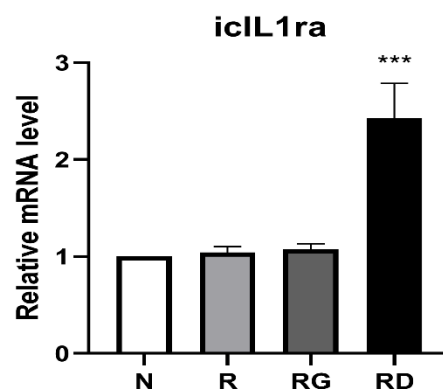
C



D



E

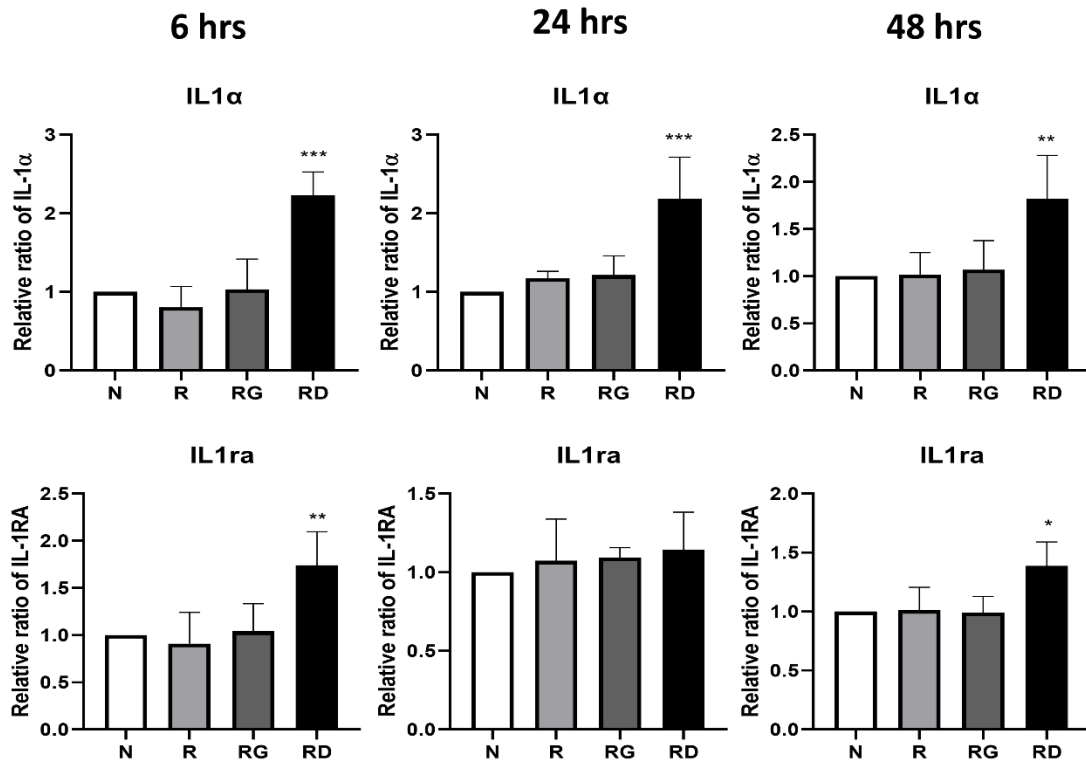


sFig. 2 Effects of DCR3 on IL-1 α and IL-1ra mRNA regulation in RANKL-induced osteoclast differentiation. RAW 264.7 cells were seeded in density of 2×10^5 cells/well at 24 well plate and treated with 10 μ g/ml of DCR3 or IgG control in the presence of RANKL. After 3, 6, 9, and 12 hours treating with 10 μ g/ml DCR3 or IgG control in the present of RANKL, total RNA was isolated and 1 μ g of total RNA was used to transcribe cDNA. Mouse specific IL-1 α , sIL-1ra, and icIL-1ra were detected by RT-PCR (A, B). According to the peak expression of IL-1 α at 6 hours, IL-1 α , sIL-1ra, and icIL-1ra were detected and quantified at 6 hours by QPCR.

A representative result of at least three independent experiments is shown. (N: RAW264.7 cells;
R: RANKL; RG: RANKL + IgG; RD: RANKL + DCR3; ***P < 0.001)

Quantification of IL-1 α and IL-1ra protein expression in DCR3 treated OC

Figure 3



sFig. 3 Effects of DCR3 on IL-1 α and IL-1ra protein expression on RANKL-induced osteoclast differentiation.

RAW264.7 cells were treated with 10 μ g/ml DCR3 or IgG in the presence of RANKL (50 ng/ml) stimulation for 6, 24 or 48 hours. Cell extracts were analysed by immunoblotting assay. Equal amounts of protein were loaded in each lane as demonstrated by the level of GAPDH. A representative result of at least three independent experiments is shown. (N: RAW264.7 cells; R: RANKL; RG: RANKL + IgG; RD: RANKL + DCR3; *P < 0.05; **P < 0.01; ***P < 0.001)

List of murine PCR primers

Supplementary table

Table 1

Genes	Nucleotide sequences
IL-1 α	5'- CGCTTGAGTCGGCAAAGAAA -3' (forward)
	5'- CTTCCCGTTGCTTGACGTTG -3' (reverse)
sIL-1ra	5'- CCTCGGGATGGAAATCTG -3' (forward)
	5'- CTGGTTGTTTCTCAGGTAAAAGG -3' (reverse)
icIL-1ra	5'- GCTCCTTTATACACAGCAAGTCTCT -3' (forward)
	As antisense sIL-1ra
GAPDH	5'- GTGAGGCCGGTGCTGAGTATGT -3' (forward)
	5'- ACAGTCTTCTGGGTGGCAGTGAT -3' (reverse)