

Review Article

Function of Connexins in the Interaction between Glial and Vascular Cells in the Central Nervous System and Related Neurological Diseases

Yinan Zhao,¹ Yanguo Xin,² Zhiyi He ,¹ and Wenyu Hu ³

¹Department of Neurology, The First Affiliated Hospital of China Medical University, Shenyang, Liaoning 110001, China

²Department of Cardiology, West China Hospital of Sichuan University, Chengdu, Sichuan 610041, China

³Department of Cardiology, The First Affiliated Hospital of China Medical University, Shenyang, Liaoning 110001, China

Correspondence should be addressed to Zhiyi He; hezhiyi0320@yahoo.co.jp and Wenyu Hu; huwen0320@sohu.com

Received 9 March 2018; Revised 6 May 2018; Accepted 14 May 2018; Published 10 June 2018

Academic Editor: Jean-Pierre Mothet

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Neuronal signaling together with synapse activity in the central nervous system requires a precisely regulated microenvironment. Recently, the blood-brain barrier is considered as a “neuro-glia-vascular unit,” a structural and functional compound composed of capillary endothelial cells, glial cells, pericytes, and neurons, which plays a pivotal role in maintaining the balance of the microenvironment in and out of the brain. Tight junctions and adherens junctions, which function as barriers of the blood-brain barrier, are two well-known kinds in the endothelial cell junctions. In this review, we focus on the less-concerned contribution of gap junction proteins, connexins in blood-brain barrier integrity under physio-/pathology conditions. In the neuro-glia-vascular unit, connexins are expressed in the capillary endothelial cells and prominent in astrocyte endfeet around and associated with maturation and function of the blood-brain barrier through a unique signaling pathway and an interaction with tight junction proteins. Connexin hemichannels and connexin gap junction channels contribute to the physiological or pathological progress of the blood-brain barrier; in addition, the interaction with other cell-cell-adhesive proteins is also associated with the maintenance of the blood-brain barrier. Lastly, we explore the connexins and connexin channels involved in the blood-brain barrier in neurological diseases and any programme for drug discovery or delivery to target or avoid the blood-brain barrier.

1. Introduction

In the 1900s, the term “blood-brain barrier (BBB)” was firstly proposed to explain the reason that certain medicines failed to treat brain conditions [1]. The previous concept of BBB is a pure endothelial barrier of the brain, a physical barrier comprised of tight junctions (TJs) and adherens junctions (AJs) that limit paracellular diffusion of solutes between the blood and the brain [2, 3]. Currently, BBB is more likely to be considered as “a neuro-glia-vascular unit” [4, 5], a functional unit composed of capillary endothelial cells (ECs), glial cells, pericytes, and neurons and which interacts with local segments of blood vessels. Functionally, in healthy conditions, BBB serves as a higher selective barrier, protecting and separating the central nervous system (CNS) from toxic

and damaging blood-borne compounds and peripheral neurotransmitter pools, protecting the synaptic and axonal signaling from ionic fluctuations, permitting the entry of required chemical molecules, and clearing larger molecules and brain metabolites, maintaining the balance of the brain microenvironment [6, 7]. In pathological conditions, brain-targeted diseases may be the origins, followed by vasculopathy, change of blood flow, and altered secretion of matrix molecules. Microglia activated and recruited by signals from astrocytes and neurons secrete more inflammatory cytokines, ultimately causing neuronal injury and synaptic dysfunction [5, 8–10].

According to previous studies, it is well known that the two kinds of vascular endothelial cell-cell junction, TJs and AJs, play key roles in the barrier function of BBB [2, 11];

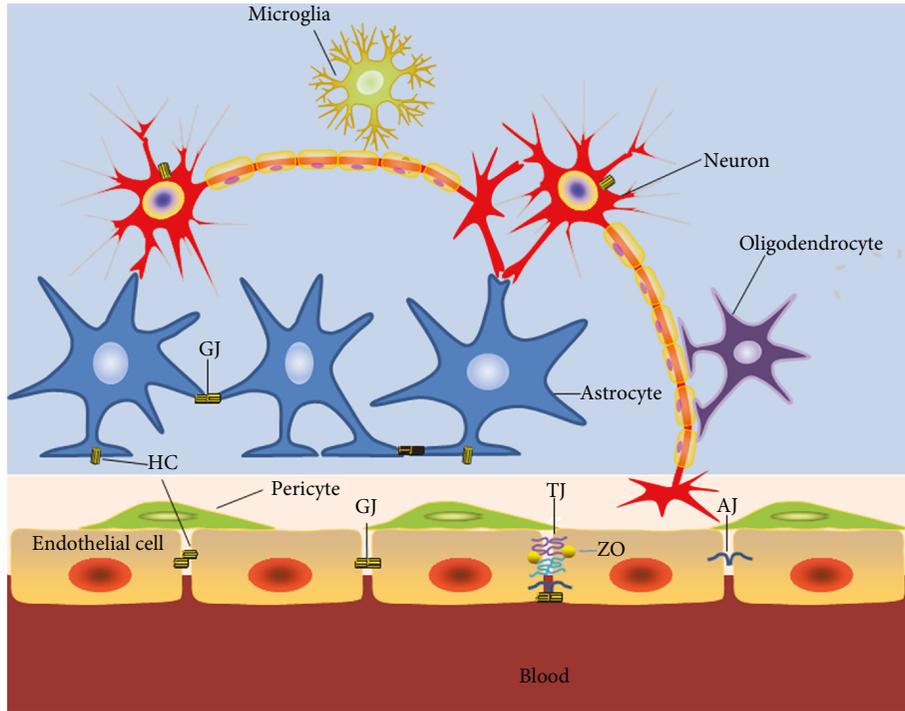


FIGURE 1: Connexin-based channels in BBB. A functional unit composed of capillary endothelial cells (ECs), glial cells, pericytes, neurons, and interaction with local segments of blood vessels.

nevertheless, very recently, researchers discovered a third category, gap junction (GJ) channels, and the cross work with other cell-cell junctions in maintaining the barrier function [12]. Gap junction intercellular communication (GJIC) is formed by two hemichannels (HCs) located on the opposing cell membranes; each one is composed of six transmembrane proteins termed as connexins (Cxs) [13, 14]. Cx channels contribute to the intercellular communication between different cellular compartments. In addition, crapping around the capillary endothelial cells in CNS, dysfunction of vascular barriers caused by deficiency of Cxs on the membrane of astrocytes has been observed [15], indicating that Cx channels on astrocyte endfeet also play a pivotal role in the maintenance of function in BBB. However, it is still unclear how Cx channels work on astrocyte endfeet and contribute to the regulation of BBB. In addition, some believe that Cxs on the membrane of ECs rather than astrocytes are involved in the permeability status of BBB [16]. Accordingly, there are still debates about where and how Cxs regulate the barrier function and interact with other junctional proteins in CNS, hence the problem. This review focuses on the pivotal roles of Cxs and Cx channels in regulating the permeability of CNS/PNS barriers and related diseases caused by dysfunction of the “neuro-glia-vascular unit.”

2. BBB Structure

BBB is considered as a functional unit, and glial cell is one of the important constituent structures, which forms a continuous membranous network around the vessels,

where molecular signaling through pericytes and ECs is organized (Figure 1). This “highly selective permeability barrier” maintains brain homeostasis through permitting the entry of required nutrients and excluding potentially harmful compounds into the CNS [17, 18]. Thus, BBB plays a pivotal role in protecting the specific neuron function from biochemical attack in the systemic circulation.

Capillary ECs, another constituent structure of BBB, supply a junctional complex comprising of AJs and TJs and control paracellular diffusion of solute between the blood and the brain [2, 4, 11, 19] (Figure 2). TJs contain a complex of proteins spanning the intercellular cleft (occludin and claudin) and junctional adhesion molecules [2, 20]. The zona occludin protein family (ZO-1, ZO-2, and ZO-3) plays a key role in anchoring the occludin and claudin to the cytoskeleton via interaction with actin [21]. The AJ protein, cadherin (α , β , and γ), spans the intercellular cleft and provides structural support [2]. Recently, more focus is on the significant role of Cx proteins in the junctional complex [22, 23]. Unlike TJ and AJ proteins, Cx proteins do not form a tight seal between the connecting cytoplasm and adjacent cells, despite adhesive properties indispensable for the adjacent cells.

3. Cxs in Glial Cells and Neurons

GJs constitute connected exchange channels between adjacent cells for the interchange of cytoplasm components, small molecules, ions, and so on, and maintain metabolic cooperation [24–26], completeness of intercellular signaling pathways, liquid buffer, and electric coupling [27, 28]. GJs

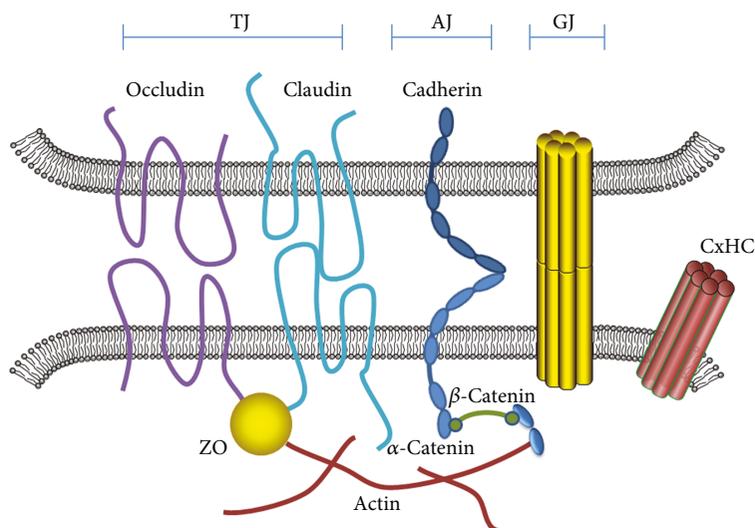


FIGURE 2: Cx channels form the intercellular junctional complex. TJ contains a complex of proteins spanning the intercellular cleft (occludin and claudin). The ZO protein family plays a role in anchoring the occludin and claudin to the cytoskeleton via interaction with actin. The AJ protein, cadherin, spans the intercellular cleft and provides structural support. Transmembrane Cx proteins constitute a third partner in the intercellular junctional complex. GJ is formed by two HCs located on the opposing cell membranes, which are composed of six transmembrane Cx proteins.

are formed by two HCs located on the opposing cell membranes (Figure 2), which are composed of six transmembrane proteins termed as Cxs. The Cx superfamily is comprised of 21 isoforms in humans and 20 isoforms in mice in a tissue-specific expression pattern and named according to their molecular weight (Figure 3) [29, 30].

Eleven kinds of Cxs have been detected in the rodent brain [31]; Cx30 and Cx43 are two major categories expressed in astrocytes. The expression of astrocytic Cxs varies by brain region and stage of development [31]. For instance, the time pattern of the postnatal expression of Cx30 (from P12) and Cx43 (from P2) in astrocyte end-feet correlates with the maturation of the BBB [15, 32]. White matter astrocytes express minimal or no Cx30 [33]. In contrast, expression of Cx30 is higher compared with that in white matter, even more than Cx43, in the thalamus [34]. Lack of Cx43 leads to the reduction of coupling and absence of both Cx43 and Cx30 abolishment of interastrocyte coupling [35–37], indicating that Cx43 and Cx30 are the major components of a direct intercellular communication between astrocytes. Cx36, together with a site-specific expression of Cx45 and Cx57, is the principal connexin expressed in neurons. Cx36 was firstly reported to be expressed specifically in mammalian neurons [38]. The expression of Cx36 is sharply reduced during the course of postnatal maturation and is sparsely expressed in the neocortex; however, it remains enriched in subsets of interneurons in the hippocampus, olfactory bulb, and thalamus [38–40]. There was evidence indicating the absence of GJs between neurons and glial cells in the mammalian CNS [31, 41, 42]; however, both electrical and dye coupling between Bergmann glial cells and Purkinje neurons has been observed in adult rats [43].

4. Cxs in Different Regions of Vascular Tree (Arteries, Arterioles/Venous System, and Lymphatic System)

The expression regions of Cx40, Cx37, and Cx43 are mainly restricted on the cytomembrane of vascular ECs [44]. Cx37 and Cx40 are widely expressed in arteries and arterioles, while Cx43 in the regions is consistent with turbulent blood flow [45–47]. Cx37 expression is prominent in the straight portion of carotid arteries, but reduced from the carotid bifurcation [48]. Cx43 is robustly expressed at branching vessels; turbulent shear stress caused by disturbed blood flow makes the endothelium of these small vessels prone to developing atherosclerotic lesions [46, 49].

Very limited studies focused on the expression pattern of Cxs in the venous/lymphatic system. Cxs expression (Cx37, Cx40, and Cx43) in the venous system also exhibited the same manner as in the arterial systems, which is a regional variation [50].

Kanady et al. observed Cx37, Cx43, and Cx47, especially the first two which exhibited a regional and dynamic variation of expression manner in mice. Interestingly, coexpression of Cx47 and Cx43 was observed in the region upstream of the adult valve. A knockout mouse model showed that Cx37 and Cx43 but not Cx47 affected lymphatic morphology [51–53].

Microvasculature is the region where the exchange of nutrients and materials between tissue and blood takes place. As mentioned above, the expression of Cx43 is higher in microvasculature compared with that in macrovasculature [54], indicating that Cx43 is a major regulator of vascular permeability. Upregulation of Cx43 in ECs, associated with the vascular hyperpermeability, has been verified to be consistent with the above conclusion [54–57].

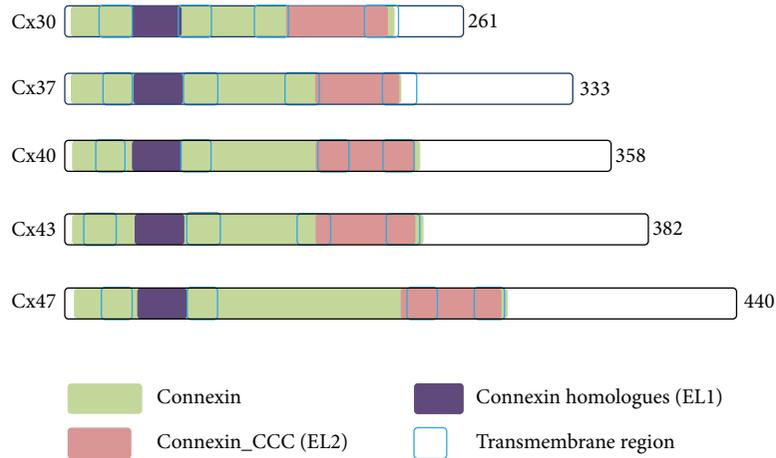


FIGURE 3: Schematic representation of connexins. Each Cx is a four-pass transmembrane (red frame) protein comprising two extracellular loops (EL1, purple region; EL2, green region) and a C-terminal cytoplasmic tail. The domains and motifs of connexins are depicted according to the size.

5. Function of Endothelial Cxs in BBB

Endothelial cells individually or jointly express Cx37, Cx40, and Cx43, in a vessel type-dependent manner as mentioned above. Several evidences point to the possible contribution of endothelial Cxs in maintaining the permeability status of the BBB. Cx40 and Cx43 are associated with TJ proteins (occludin and claudin-5) in porcine brain ECs through cytoplasmic scaffold proteins, such as ZO-1. GJ blocker 18 β -glycyrrhetic acid and oleamide did weaken the barrier function of TJs, but did not influence the expression or distribution of Cxs or TJ proteins in porcine BBB ECs [12] (Table 1). Damaged BBB permeability was considered to be caused by loss of function of GJIC and TJs without morphological change; however, the mechanism needs further research. In contrast with pathological conditions, the expression of Cx43 in the endothelial wall of healthy blood vessels appears low and increases after inflammation occurred, especially in the endothelial cells of smaller blood vessels including those of the BBB [55, 58]. Cx43 antisense treatment further reduces vascular permeability and invasion of neutrophils in spinal cord injury (SCI) [55] (Table 2). Very recently, increased expression of Cx43 and permeability of BBB were observed in familial cerebral cavernous malformation type III (FCCM3) lesions of mouse model brains [59], partly consistent with the observation in *ccm3* knockdown brain microvascular ECs, which was accompanied by dislocation of ZO-1, limitation to formation, and transinteraction of TJs *in vitro*. Gap27, a Cx43 GJ inhibitor, rescued CCM3KD hyperpermeability through inhibiting Cx43 GJs, restoring ZO-1 to TJ structures, and reducing plaque accumulation in Cx43 GJs, indicating that Cx43 GJs are increased in Fccm3 and regulate barrier permeability in a TJ-dependent manner [59] (Table 1). It is unclear by what mechanism CCM3 regulates Cx43 and whether Cx43 is exclusive to participate in the formation of TJs.

6. Involvement of Glial and Neuronal Connexins and BBB Function

In the nervous system, astrocytic Cx is unique and contributes pivotally to brain metabolism and processing [60]. Cx30 and Cx43 are two major Cx proteins expressed by astrocytes and that form Cx-based channels to mediate intercellular signaling, especially that involving Ca^{2+} and maintaining the BBB integrity [15, 32]. A knockout model was induced to clarify the role of astroglial Cxs in the morphology and function of BBB. The combination of global Cx30 and astrocyte-specific Cx43 deletion (conditional knockout from glial fibrillary acidic protein-expressing cells) downregulates aquaporin 4 (AQP4), which is located in astrocytes throughout the CNS in the brain, spinal cord, and optic nerve and is particularly concentrated in foot processes adjacent to microvessels at BBB [61] (Table 2). In contrast, TJ proteins, occludin, and ZO-1 show no difference in expression or morphology, compared with the WT mice [15]. Consequently, double knockout of astrocytic Cx43 and Cx30 leads to systemic astrocyte endfeet swell and increased BBB permeability with the increase in vascular and shear stress [15]. However, absence of Cx30 did not affect organization or permeability of BBB according to a further analysis of the separate role of Cx30 [62] (Table 2). Absence of astroglial Cx43 compromises the ability of the brain to maintain immune quiescence [63]; recruitment of T cells, B cells, macrophages, and neutrophils; enhanced antigen presentation; and autoimmune reaction [64–66] (Table 2). Immune cell recruitment is not the consequence of BBB breakdown in Cx43-KO mouse brain, despite progressively weakened BBB permeability under shear and pressure [64]. Loss of immune-quiescent, immune-competent cells abnormally crossing the BBB and developing autoimmune mechanisms could be harmful as suggested in pathological conditions linked to a decreased expression of Cx43 in multiple sclerosis (MS) and neuromyelitis optica (NMO) patients [67, 68]. Deficiency of Cx43

TABLE 1: Connexin inhibitors link to the BBB permeability *in vitro*.

Cell type	Stimulus	Treatment	Main outcome	Reference
Porcine BBB ECs		18 β -Glycyrrhetic acid and oleamide	Inhibits the barrier function of TJs and GJIC without morphological change	[12]
Mouse brain CCM3KD ECs		Gap27	Inhibits BBB hyperpermeability through blocking Cx43 GJs, restoring ZO-1 to TJ structures	[59]
Rat brain RBE4 cells	Bradykinin	Gap27, Cx37-siRNA, and Cx43-siRNA	Inhibits endothelial hyperpermeability through reducing intracellular calcium oscillations	[57]
Rat brain RBE4 cells	A reduction in extracellular Ca ²⁺ concentration	Gap27	Inhibits endothelial hyperpermeability through blocking the intercellular Ca ²⁺ waves and repressing PKC, CaMKII, and actomyosin contraction	[77]
Rat brain (RBE4, GP8 cells)	TNF- α	Gap26	No effect on the elevated baseline ATP release from rat brain endothelial cells treated by TNF- α	[82]
Rat brain GP8 cells		Gap26 and Gap27	Inhibits intracellular ATP release through blocking Cx43-HCs which contribute to the intercellular propagation of calcium signals	[81]
Organotypic hippocampal slices from 7-day-old Wistar rats	Albumin	Carbenoxolone, Gap27, and SLS peptide	In developing hippocampal networks, the generation and initiation of spontaneous recurrent seizure-like activity depend on the opening of glial GJs	[119]
Astrocytes from the cortex and spinal cord of the SCI mouse model	TNF- α	Gap26, Gap27, and Cx43-siRNA	TNF- α activated Cx43-HCs, rather than GJIC, to release CXCL1 which contribute to neuropathic pain	[94]
Astrocytes from cortex of Sprague-Dawley rat	A β -peptide	Octanol (an uncoupler of gap junctions)	Functional GJs are not required for calcium-wave propagation; they play a role in the enhancement of calcium waves induced by A β .	[110]

ECs: endothelial cells; SCI: spinal cord injury; TJs: tight junctions; GJs: gap junctions; ZO: zona occludin; PKC: protein kinase C; CaMKII: Ca²⁺/calmodulin-dependent kinase II; HCs: hemichannels; GJIC: gap junction intercellular communication; A β : amyloid β .

TABLE 2: Connexin inhibitors link to the BBB permeability *in vivo*.

	Disease model	Interference	Main outcome	Reference
Spinal cord	SCI	Cx43-asODN	Reduce vascular permeability and invasion of neutrophils in spinal cord injury	[55]
BBB	Neuroinflammation	Gap27	Gap27 coadministered together with bradykinin reduced leakage of reporter dye from the vascular lumen into the tissue	[57]
BBB		Global Cx30 and astrocytic-specific Cx43 deletion	Systemic astrocyte endfeet swell and increased BBB permeability with the increase in vascular and shear stress	[61]
BBB		Global Cx30 deletion	Not affecting organization or permeability of BBB	[62]
BBB		Astrocytic-specific Cx43 deletion	Compromises the ability of the brain to maintain immune quiescence and recruitment of immune cells without BBB breakdown	[63–66]
Embryo		Cx43 deletion	Migration impairment of neurons through adhesive interaction with radial fibers	[69, 70]
Brain	AD	INI-1602 (microglia HC blocker)	Improve memory deficits	[114]

SCI: spinal cord injury; HCs: hemichannels; BBB: brain-blood barrier.

nonchannel function is associated with migration impairment of neurons through adhesive interaction with radial fibers during embryogenesis [69, 70] (Table 2). However, the mechanism by which knockdown of Cx43 leads to the migration of immune-competent cells abnormally crossing the BBB and the Cx-channel-dependent or nondependent function important to mediate the migration process needs to be clarified further.

In an ultrasound-induced BBB opening Wistar rat model [71], reorganization of neuronal Cx36-containing GJs with an increase in gap-junctional plaque size was found, similar to the results in another study [72]; neuronal Cx36 interacts with a ZO-1 PDZ domain, the same as in astrocytic Cx43, thus indicating that ultrasound-induced disruption of ZO-1 might lead to an increase in Cx36 plaque size. However, in the same BBB opening model [71], changes in Cx36 plaque size were not as striking as in astrocytic Cx43 plaques, suggesting the key role of astrocytes for the maintenance of homeostasis of BBB.

7. The Roles of Cx Channels in BBB Function

Several factors, such as inflammatory cytokines and growth factors, contribute to the regulation of Cx-HC and GJ channels in physiological or pathological progress and modulatory mechanism of BBB. Accumulating evidence indicated that Ca^{2+} transients in astrocyte endfeet may induce dilation and constrictions in the adjacent arterioles and may further precede changes in blood flow and disturbance of the healthy brain function [73, 74]. One research showed an increased BBB and bovine brain capillary EC permeability through triggering calcium oscillations involved in Cx-HC opening, indicating that HCs may play a vital role in the process of calcium signaling and propagation of ICW [57]. Divergent functional properties of Cx43 channels have been set forth in inflammatory conditions. Retamal et al. found that the proinflammatory cytokines IL-1 β and TNF- α reduced the

intercellular communication via Cx43 GJIC, whereas it increased the cellular exchange with the extracellular milieu via Cx43 HCs [75]. Inhibition of GJIC that is observed under the condition of inflammation indicates that the increased membrane permeability may be attributable to active HCs [75, 76]. On the other hand, the effect of proinflammatory cytokines enhanced the uptake and reduced the intercellular diffusion of glucose, which might explain the change of metabolic status in astrocytes under inflammatory conditions. Accordingly, such opposite regulation effects of Cx channels may impact glucose trafficking and modify the metabolic status of astrocytes involved in brain inflammation [75]. Connexin channels provide a target to manipulate brain endothelial calcium dynamics and BBB permeability [57]. Bradykinin, a typical inflammatory messenger, increases BBB and rat brain endothelial (RBE4) cell permeability through triggering calcium oscillations involved in Cx-HC opening. Gap27, a GJ mimetic peptide identical to part of the second extracellular loop of Cx37 and Cx43 structurally, reduced intracellular oscillations in RBE4 cells in response to bradykinin, in accordance with the Cx37/Cx43-targeted siRNA [57] (Table 1). Another group indicated that intercellular Ca^{2+} waves (ICWs) increased the endothelial permeability by lowering the extracellular Ca^{2+} . The endothelial permeability increase was furthermore inhibited by Gap27, which also blocked the ICWs, and with inhibition of protein kinase, Ca^{2+} /calmodulin-dependent kinase II, and actomyosin contraction. In conclusion, ICWs significantly increase endothelial permeability and the Cxs underlying Ca^{2+} wave propagation form a target to limit BBB alteration [77] (Table 1).

GJs mimetic peptides, Gap26 and Gap27, which specifically target Cx channels, are widely used to close the HCs and, in certain conditions, gap junction communication [78]. ATP is the best known signaling factor linked to Cx-HC activity and released from astrocytes directly through Cx43-HCs [79]. Gap26 binds to the first extracellular loop

TABLE 3: Cxs/Cx-channels in BBB in CNS pathology.

CNS pathology	Cxs/Cx channel remodeling	BBB dysfunction	Reference
MOG-induced EAE mouse model	Deletion of both <i>Cx30</i> and <i>Cx43</i>	None	[126]
Carrageenan-induced inflammatory pain rat model	Not mentioned	Increased expression of ZO-1 in TJs and permeability of the BBB	[92]
MCAO mouse model	<i>Cx43</i> ^{+/-} , GJIC reduction	Weakened barrier function of BBB and exacerbated ischemic injury	[98]
Age-related neurodegenerative disease mouse model	Increased expression and relocalization of <i>Cx43</i> in ECs	An increase in the permeability of the BBB without change of ZO-1	[108]
4-AP-induced epilepsy rat model	Gradually elevated expression of <i>Cx43</i> and GJIC remodeling	Not mentioned	[118]
HIV-infected neuroAIDS macaque model	HIV amplified by GJs	Compromised BBB integrity	[123]

Cx: connexin; TJs: tight junctions; GJs: gap junctions; ZO: Zona occludin; GJIC: gap junction intercellular communication; BBB: brain-blood barrier; 4-AP: 4-aminopyridine.

of *Cx43* and blocks *Cx43*-HCs in ECs, demonstrating that intracellular ATP is released through Cx-HCs in a Cx-dependent manner, which contributes to the intercellular propagation of calcium signals, thereby affecting the postulated spatial spread of BBB opening [80, 81] (Table 1). Nevertheless, one *in vitro* study showed that *Gap26* has no effect on the elevated baseline ATP from rat brain ECs treated by $\text{TNF-}\alpha$, suggesting that ATP release is non-Cx-dependent under proinflammatory cytokine-stimulated conditions [82] (Table 1). *Gap27* targets the second extracellular loop of *Cx37* and *Cx43*, and exposure to *Gap27* upon short time (minutes) inhibits Cx-HC-mediated ATP release and dye uptake [32, 81, 83], while longer exposure duration (hours) blocks junctional coupling, as well [81, 84]. *Gap27* limits BBB permeability induced by bradykinin *in vivo* through reducing intracellular calcium oscillations without modifications of TJs and cytoskeletal proteins such as occludin or ZO-1 [57] (Table 2). Some problems remain to be solved. First, some GJ mimetic peptides are widely used to inhibit the HC and GJ channels; however, what role GJ channels and HCs play in glial endfeet and how they contribute to regulation of BBB permeability need to be explained in detail. Secondly, Wnt, together with the Sonic hedgehog (Shh) signaling pathway, regulates the maturation of the BBB during development [85–87]; however, which signaling molecules that passed through Cx channels contribute to astrocyte endfeet structure adhesion and affect the integrity of BBB also needs to be clarified.

8. Cxs and Cx Channels in Nervous System Diseases

8.1. Multiple Sclerosis. Downregulation of oligodendrocytic *Cx32*, *Cx47*, and astrocytic *Cx43* had been identified in the active lesions of CNS in MS patients and EAE mice [88–90]. Lutz et al. reported that deletion of both *Cx30* and *Cx43* in the EAE model did not aggravate clinical signs or pathological progress; moreover, no difference was observed in BBB permeability [91], indicating that absence of astrocytic *Cx43* and *Cx30* does not contribute to the

pathology progress of the nerve tissue or further weaken BBB intensity under inflammatory condition (Table 3).

8.2. Neuropathic Pain. Inflammatory-mediated pain alters in TJs; ZO-1 expression was significantly increased, increasing the permeability of the BBB of the carrageenan-induced inflammatory pain rat model [92] (Table 3). An *in vivo* evidence suggests that carrageenan-induced spinal $\text{IL-1}\beta$ inhibits astrocyte activation during the early phase of inflammation through the suppression of *Cx43* expression [93]. *Cx43* has been reported to induce chemokine CXCL1 (keratinocyte-derived cytokine) release to contribute to neuropathic pain. $\text{TNF-}\alpha$ activated *Cx43*-HCs, rather than GJIC, to release CXCL1 [94], which could be blocked by *Gap26*, *Gap27*, and *Cx43*-siRNA in cultured astrocytes from the cortex and spinal cord of the SCI mouse model, a mouse model for chronic neuropathic pain (Table 1). *Cx43* is increased in cervical level 6 to 8 dorsal root ganglia, and specific peptide inhibitors of *Cx43* ameliorated established tactile allodynia after severe SCI. Notably, SCI-induced mechanical allodynia was prevented in *Cx43/Cx30* double-knockout mice, highlighting the importance of glial intercommunication in chronic neuropathic pain [95, 96].

8.3. Stroke. *Cx43* deficiency not only creates intercellular communication reduction and weakened barrier function of BBB but also exacerbates ischemic injury [15, 97, 98] (Table 3). After ischemia-reperfusion, endothelial Cx expression is upregulated and Cx-HCs open [57, 99, 100], leading to glutamate excitotoxicity and inflammatory mediator release including nitric oxide [101] and further leading to EC edema and rupture [56, 102], astrocyte activity, T cell and monocyte migration, and increased inflammatory reaction, ultimately inducing tissue damage [103–105]. Inhibition of astrocytic GJs by octanol, for instance, could restrict the flow of neurotoxins, which exacerbate neuronal damage of cerebral ischemia [106]. In addition, *Cx43* mimetic peptide, a *Cx43* inhibitor, exhibited protective effects after cerebral and retinal ischemia [58, 107], which proves the point in reverse.

8.4. Alzheimer's Disease (AD). An age-related decline in BBB integrity correlates with the loss of sex steroids and increase in gonadotropins with menopause [108]. Ovariectomy of young female mice induces an increased expression and dislocation of Cx43 in ECs accompanied by a rise in the permeability of the BBB without change of TJ protein, ZO-1 [108] (Table 3). It implicates Cx43 in regulating changes in BBB permeability and serum gonadotropins in the cerebral pathophysiology of neurodegenerative diseases, such as AD. One study reported that amyloid β ($A\beta$) peptide, a self-aggregating 40–42-amino-acid protein that is crucially involved in AD as the main component of the amyloid plaques, did harm to neurons resulting in their dysfunction and death [109], contributing to AD pathology through dramatically enhancing GJC/HC-mediated Ca^{2+} wave propagation in astrocytes [110] (Table 1). Recently, several studies [111–113] indicated that expression of astroglial Cxs is increased at amyloid plaques and that there is an increase in HC activity in reactive astrocytes that should certainly have an impact on gliovascular interaction. Until recently, INI-0602, a putative HC blocker, was verified to inhibit LPS-induced glutamate release from microglia *in vivo* and improve memory deficits in double transgenic mice expressing human amyloid precursor protein with K595N and M596L mutations and presenilin 1 with A264E mutation as an AD mouse model [114] (Table 2). An important next step underlying Cx32 conditional knockout in microglia will be needed to demonstrate the unique regulation of microglial HCs in the progress of AD.

8.5. Epilepsy. TGF- β signaling is a key trigger of albumin-induced epileptogenesis, following BBB breakdown, which can be inhibited by the TGF- β pathway blocker and can manifest the TGF- β pathway as a novel therapeutic target for preventing injury-related epileptogenesis [115, 116]. Downregulation of GJ proteins Cx43 and Cx30, inwardly rectifying K^+ channels (Kir4.1), the glial excitatory amino acid transporters (EAAT1 and EAAT2), and dislocation of aquaporins (AQP4), represents astrocyte activation induced by BBB disruption, as well as by focal application of albumin or TGF- β . These data indicate that activation of astrocytes is a critical step in epileptogenesis preceding alteration in neuronal function [115, 117]. Although many of those signaling cascades are altered in activated astrocytes, very little is known about neurovascular coupling in epilepsy. In the immature rat brain, GJIC is involved in rhythm genesis and synchronization of cortical activity and may enhance the epileptogenicity of the 4-aminopyridine- (4-AP-) induced epilepsy model [118]. In addition, repeated seizures can induce changes in different Cx gene (Cx26, 32, and 43) expression during the postnatal development, especially gradually elevated expression of Cx43 and GJIC remodeling [118] (Table 3). Assuming a selective inhibition of a Cx43-dependent process by the mimetic peptides and preferential localization of this Cx isoform in astrocytes suggests that in developing hippocampal networks, the generation and initiation of spontaneous recurrent seizure-like activity may depend in large part upon the opening of glial GJs [119] (Table 1).

8.6. NeuroAIDS. The main cells infected with HIV within the CNS are macrophages, microglia, and a small fraction of astrocytes [120–122]. Both *in vitro* and *in vivo* evidence uncovered a novel mechanism of bystander BBB toxicity mediated by low numbers of HIV-infected astrocytes and amplified by GJs [123] (Table 3). Few HIV-infected astrocytes compromise BBB integrity through blocking GJ channels, indicating that HIV affects the CNS by a GJ-dependent manner [123].

9. Discussion

Homeostasis of metabolites and ions is strictly regulated by the barrier between blood and nerve tissue. The BBB shields the direct contact between systemic circulation and the brain, as well as the spinal cord and peripheral nerve. TJs and AJs, two pivotal junctional complexes physically located in the ECs, are considered as the main structural components of the barriers. In this review, we introduce a third junctional component, Cx proteins, as an important player in the interactions with other junctional complexes that occur at the neuro-glia-vascular interface. In a cerebral microenvironment, Cx-forming channels of both astrocytes and ECs are critical in the control of BBB function. Interestingly, GJ channels and HC channels of BBB appear to play opposite roles in the regulation of BBB permeability. GJIC contributes to maintaining BBB integrity, while HCs are associated with ATP signaling release and necessary to generate the Ca^{2+} oscillations linked to BBB disruption induced by proinflammatory signals. A cluster of papers have provided strong evidence in favor of a Cx channel-dependent manner contributing to the regulation of BBB function; however, few researchers focus on a Cx channel-independent manner, adhesive contact, and gene expression, for instance [16, 124, 125]. It remains to be demonstrated how nonchannel Cxs interact with other cell-cell-adhesive proteins to maintain the barrier. Cxs have been considered as novel therapeutic targets for neurologic diseases in connection with BBB dysfunctions. Astrocytic and endothelial Cx channels are vital to BBB permeability change caused by neurologic pathology, especially inflammation. Microglial cells also play an important role in activating immune cells; however, the role of Cxs in the intercellular cross talk between BBB endothelial cells and microglia, and microglia in the BBB development in physiological or pathologic conditions, remains unclear. More studies should be focused on the mechanism of microglia-endothelial Cx channels in BBB permeability regulation.

Conflicts of Interest

The authors declare no competing interests.

Acknowledgments

The authors gratefully acknowledge financial support from the China Scholarship Council.

References

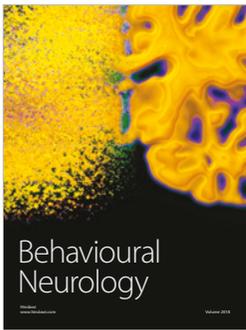
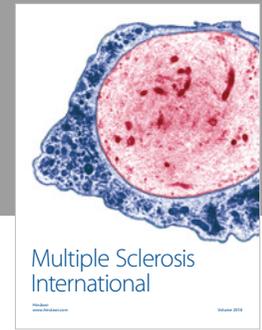
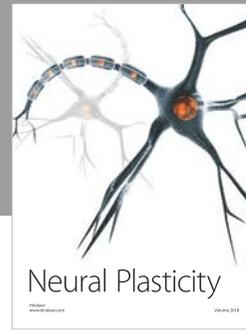
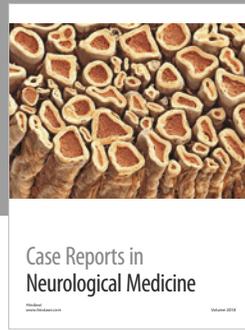
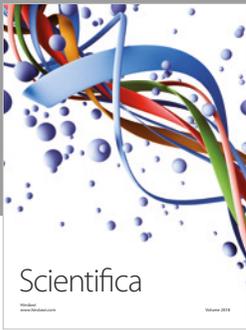
- [1] S. A. Liddelow, "Fluids and barriers of the CNS: a historical viewpoint," *Fluids and Barriers of the CNS*, vol. 8, no. 1, p. 2, 2011.
- [2] H. Wolburg and A. Lippoldt, "Tight junctions of the blood-brain barrier: Development, composition and regulation," *Vascular Pharmacology*, vol. 38, no. 6, pp. 323–337, 2002.
- [3] M. M. Tenreiro, R. Ferreira, L. Bernardino, and M. A. Brito, "Cellular response of the blood-brain barrier to injury: potential biomarkers and therapeutic targets for brain regeneration," *Neurobiology of Disease*, vol. 91, pp. 262–273, 2016.
- [4] N. J. Abbott, A. A. K. Patabendige, D. E. M. Dolman, S. R. Yusof, and D. J. Begley, "Structure and function of the blood-brain barrier," *Neurobiology of Disease*, vol. 37, no. 1, pp. 13–25, 2010.
- [5] J. Lok, P. Gupta, S. Guo et al., "Cell-cell signaling in the neurovascular unit," *Neurochemical Research*, vol. 32, no. 12, pp. 2032–2045, 2007.
- [6] B. V. Zlokovic, "The blood-brain barrier in health and chronic neurodegenerative disorders," *Neuron*, vol. 57, no. 2, pp. 178–201, 2008.
- [7] N. J. Abbott, L. Ronnback, and E. Hansson, "Astrocyte-endothelial interactions at the blood-brain barrier," *Nature Reviews Neuroscience*, vol. 7, no. 1, pp. 41–53, 2006.
- [8] S. Boillee, C. Vande Velde, and D. W. Cleveland, "ALS: a disease of motor neurons and their nonneuronal neighbors," *Neuron*, vol. 52, no. 1, pp. 39–59, 2006.
- [9] B. V. Zlokovic, "Neurovascular mechanisms of Alzheimer's neurodegeneration," *Trends in Neurosciences*, vol. 28, no. 4, pp. 202–208, 2005.
- [10] C. Iadecola, "Neurovascular regulation in the normal brain and in Alzheimer's disease," *Nature Reviews Neuroscience*, vol. 5, no. 5, pp. 347–360, 2004.
- [11] I. Bechmann, I. Galea, and V. H. Perry, "What is the blood-brain barrier (not)?," *Trends in Immunology*, vol. 28, no. 1, pp. 5–11, 2007.
- [12] K. Nagasawa, H. Chiba, H. Fujita et al., "Possible involvement of gap junctions in the barrier function of tight junctions of brain and lung endothelial cells," *Journal of Cellular Physiology*, vol. 208, no. 1, pp. 123–132, 2006.
- [13] R. Bruzzone, T. W. White, and D. L. Paul, "Connections with connexins: the molecular basis of direct intercellular signaling," *European Journal of Biochemistry*, vol. 238, no. 1, pp. 1–27, 1996.
- [14] D. W. Laird, "Life cycle of connexins in health and disease," *Biochemical Journal*, vol. 394, no. 3, pp. 527–543, 2006.
- [15] P. Ezan, P. André, S. Cisternino et al., "Deletion of astroglial connexins weakens the blood-brain barrier," *Journal of Cerebral Blood Flow & Metabolism*, vol. 32, no. 8, pp. 1457–1467, 2012.
- [16] N. Prochnow and R. Dermietzel, "Connexons and cell adhesion: a romantic phase," *Histochemistry and Cell Biology*, vol. 130, no. 1, pp. 71–77, 2008.
- [17] L. L. Rubin and J. M. Staddon, "The cell biology of the blood-brain barrier," *Annual Review of Neuroscience*, vol. 22, no. 1, pp. 11–28, 1999.
- [18] R. Daneman and A. Prat, "The blood-brain barrier," *Cold Spring Harbor Perspectives in Biology*, vol. 7, no. 1, article a020412, 2015.
- [19] S. Ge, L. Song, and J. S. Pachter, "Where is the blood-brain barrier ... really?," *Journal of Neuroscience Research*, vol. 79, no. 4, pp. 421–427, 2005.
- [20] H. Wolburg, S. Noell, A. Mack, K. Wolburg-Buchholz, and P. Fallier-Becker, "Brain endothelial cells and the gliovascular complex," *Cell and Tissue Research*, vol. 335, no. 1, pp. 75–96, 2009.
- [21] E. Dejana, "Endothelial cell-cell junctions: happy together," *Nature Reviews Molecular Cell Biology*, vol. 5, no. 4, pp. 261–270, 2004.
- [22] M. Derangeon, D. C. Spray, N. Bourmeyster, D. Sarrouilhe, and J. C. Herve, "Reciprocal influence of connexins and apical junction proteins on their expressions and functions," *Biochimica et Biophysica Acta (BBA) - Biomembranes*, vol. 1788, no. 4, pp. 768–778, 2009.
- [23] R. A. Meyer, D. W. Laird, J. P. Revel, and R. G. Johnson, "Inhibition of gap junction and adherens junction assembly by connexin and A-CAM antibodies," *The Journal of Cell Biology*, vol. 119, no. 1, pp. 179–189, 1992.
- [24] G. S. Goldberg, P. D. Lampe, and B. J. Nicholson, "Selective transfer of endogenous metabolites through gap junctions composed of different connexins," *Nature Cell Biology*, vol. 1, no. 7, pp. 457–459, 1999.
- [25] G. S. Goldberg, A. P. Moreno, and P. D. Lampe, "Gap junctions between cells expressing connexin 43 or 32 show inverse permselectivity to adenosine and ATP," *The Journal of Biological Chemistry*, vol. 277, no. 39, pp. 36725–36730, 2002.
- [26] Y. Qu and G. Dahl, "Function of the voltage gate of gap junction channels: selective exclusion of molecules," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 2, pp. 697–702, 2002.
- [27] S. M. Lee and M. G. Clemens, "Subacinar distribution of hepatocyte membrane potential response to stimulation of gluconeogenesis," *The American Journal of Physiology-Gastrointestinal and Liver Physiology*, vol. 263, no. 3, pp. G319–G326, 1992.
- [28] S. Rohr, "Role of gap junctions in the propagation of the cardiac action potential," *Cardiovascular Research*, vol. 62, no. 2, pp. 309–322, 2004.
- [29] R. Dobrowolski and K. Willecke, "Connexin-caused genetic diseases and corresponding mouse models," *Antioxidants & Redox Signaling*, vol. 11, no. 2, pp. 283–296, 2009.
- [30] A. Pfenniger, A. Wohlwend, and B. R. Kwak, "Mutations in connexin genes and disease," *European Journal of Clinical Investigation*, vol. 41, no. 1, pp. 103–116, 2011.
- [31] J. I. Nagy, F. E. Dudek, and J. E. Rash, "Update on connexins and gap junctions in neurons and glia in the mammalian nervous system," *Brain Research Reviews*, vol. 47, no. 1–3, pp. 191–215, 2004.
- [32] W. H. Evans, E. De Vuyst, and L. Leybaert, "The gap junction cellular internet: connexin hemichannels enter the signalling limelight," *Biochemical Journal*, vol. 397, no. 1, pp. 1–14, 2006.
- [33] J. I. Nagy, D. Patel, P. A. Y. Ochalski, and G. L. Stelmack, "Connexin30 in rodent, cat and human brain: selective expression in gray matter astrocytes, co-localization with connexin43 at gap junctions and late developmental appearance," *Neuroscience*, vol. 88, no. 2, pp. 447–468, 1999.
- [34] S. Griemsmann, S. P. Höft, P. Bedner et al., "Characterization of panglial gap junction networks in the thalamus, neocortex,

- and hippocampus reveals a unique population of glial cells," *Cerebral Cortex*, vol. 25, no. 10, pp. 3420–3433, 2015.
- [35] N. Rouach, A. Koulakoff, V. Abudara, K. Willecke, and C. Giaume, "Astroglial metabolic networks sustain hippocampal synaptic transmission," *Science*, vol. 322, no. 5907, pp. 1551–1555, 2008.
- [36] L. Roux, K. Benchenane, J. D. Rothstein, G. Bonvento, and C. Giaume, "Plasticity of astroglial networks in olfactory glomeruli," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 45, pp. 18442–18446, 2011.
- [37] A. Wallraff, R. Köhling, U. Heinemann, M. Theis, K. Willecke, and C. Steinhäuser, "The impact of astrocytic gap junctional coupling on potassium buffering in the hippocampus," *The Journal of Neuroscience*, vol. 26, no. 20, pp. 5438–5447, 2006.
- [38] D. F. Condorelli, R. Parenti, F. Spinella et al., "Cloning of a new gap junction gene (Cx36) highly expressed in mammalian brain neurons," *European Journal of Neuroscience*, vol. 10, no. 3, pp. 1202–1208, 1998.
- [39] D. F. Condorelli, A. Trovato-Salinaro, G. Mudo, M. B. Mirone, and N. Belluardo, "Cellular expression of connexins in the rat brain: neuronal localization, effects of kainate-induced seizures and expression in apoptotic neuronal cells," *European Journal of Neuroscience*, vol. 18, no. 7, pp. 1807–1827, 2003.
- [40] M. R. Deans, J. R. Gibson, C. Sellitto, B. W. Connors, and D. L. Paul, "Synchronous activity of inhibitory networks in neocortex requires electrical synapses containing connexin36," *Neuron*, vol. 31, no. 3, pp. 477–485, 2001.
- [41] J. E. Rash, T. Yasumura, F. E. Dudek, and J. I. Nagy, "Cell-specific expression of connexins and evidence of restricted gap junctional coupling between glial cells and between neurons," *Journal of Neuroscience*, vol. 21, no. 6, pp. 1983–2000, 2001.
- [42] J. E. Rash, T. Yasumura, K. G. V. Davidson, C. S. Furman, F. E. Dudek, and J. I. Nagy, "Identification of cells expressing Cx43, Cx30, Cx26, Cx32 and Cx36 in gap junctions of rat brain and spinal cord," *Cell Communication & Adhesion*, vol. 8, no. 4–6, pp. 315–320, 2001.
- [43] P. Pakhotin and A. Verkhratsky, "Electrical synapses between Bergmann glial cells and Purkinje neurones in rat cerebellar slices," *Molecular and Cellular Neurosciences*, vol. 28, no. 1, pp. 79–84, 2005.
- [44] S. Johnstone, B. Isakson, and D. Locke, "Chapter 2 biological and biophysical properties of vascular connexin channels," *International Review of Cell and Molecular Biology*, vol. 278, pp. 69–118, 2009.
- [45] R. Bruzzone, J. A. Haefliger, R. L. Gimlich, and D. L. Paul, "Connexin40, a component of gap junctions in vascular endothelium, is restricted in its ability to interact with other connexins," *Molecular Biology of the Cell*, vol. 4, no. 1, pp. 7–20, 1993.
- [46] J. E. Gabriels and D. L. Paul, "Connexin43 is highly localized to sites of disturbed flow in rat aortic endothelium but Connexin37 and Connexin40 are more uniformly distributed," *Circulation Research*, vol. 83, no. 6, pp. 636–643, 1998.
- [47] J. A. Haefliger, P. Nicod, and P. Meda, "Contribution of connexins to the function of the vascular wall," *Cardiovascular Research*, vol. 62, no. 2, pp. 345–356, 2004.
- [48] A. Pfenniger, C. Wong, E. Sutter et al., "Shear stress modulates the expression of the atheroprotective protein Cx37 in endothelial cells," *Journal of Molecular and Cellular Cardiology*, vol. 53, no. 2, pp. 299–309, 2012.
- [49] B. R. Kwak, P. Silacci, N. Stergiopoulos, D. Hayoz, and P. Meda, "Shear stress and cyclic circumferential stretch, but not pressure, alter connexin43 expression in endothelial cells," *Cell Communication & Adhesion*, vol. 12, no. 5–6, pp. 261–270, 2005.
- [50] T. Inai and Y. Shibata, "Heterogeneous expression of endothelial connexin (Cx) 37, Cx40, and Cx43 in rat large veins," *Anatomical Science International*, vol. 84, no. 3, pp. 237–245, 2009.
- [51] J. D. Kanady, M. T. Dellinger, S. J. Munger, M. H. Witte, and A. M. Simon, "Connexin37 and Connexin43 deficiencies in mice disrupt lymphatic valve development and result in lymphatic disorders including lymphedema and chylothorax," *Developmental Biology*, vol. 354, no. 2, pp. 253–266, 2011.
- [52] A. Sabine, Y. Agalarov, H. Maby-el Hajjami et al., "Mechanotransduction, PROX1, and FOXC2 cooperate to control connexin37 and calcineurin during lymphatic-valve formation," *Developmental Cell*, vol. 22, no. 2, pp. 430–445, 2012.
- [53] M. J. Meens, I. Kutkut, V. Rochemont et al., "Cx47 fine-tunes the handling of serum lipids but is dispensable for lymphatic vascular function," *PLoS One*, vol. 12, no. 7, article e0181476, 2017.
- [54] H. H. Wang, C. H. Su, Y. J. Wu et al., "Reduction of connexin43 in human endothelial progenitor cells impairs the angiogenic potential," *Angiogenesis*, vol. 16, no. 3, pp. 553–560, 2013.
- [55] M. Cronin, P. N. Anderson, J. E. Cook, C. R. Green, and D. L. Becker, "Blocking connexin43 expression reduces inflammation and improves functional recovery after spinal cord injury," *Molecular and Cellular Neuroscience*, vol. 39, no. 2, pp. 152–160, 2008.
- [56] H. V. Danesh-Meyer, R. Huang, L. F. B. Nicholson, and C. R. Green, "Connexin43 antisense oligodeoxynucleotide treatment down-regulates the inflammatory response in an *in vitro* interphase organotypic culture model of optic nerve ischaemia," *Journal of Clinical Neuroscience*, vol. 15, no. 11, pp. 1253–1263, 2008.
- [57] M. De Bock, M. Culot, N. Wang et al., "Connexin channels provide a target to manipulate brain endothelial calcium dynamics and blood-brain barrier permeability," *Journal of Cerebral Blood Flow & Metabolism*, vol. 31, no. 9, pp. 1942–1957, 2011.
- [58] H. V. Danesh-Meyer, N. M. Kerr, J. Zhang et al., "Connexin43 mimetic peptide reduces vascular leak and retinal ganglion cell death following retinal ischaemia," *Brain*, vol. 135, no. 2, pp. 506–520, 2012.
- [59] A. M. Johnson, J. P. Roach, A. Hu et al., "Connexin 43 gap junctions contribute to brain endothelial barrier hyperpermeability in familial cerebral cavernous malformations type III by modulating tight junction structure," *The FASEB Journal*, vol. 32, no. 5, pp. 2615–2629, 2018.
- [60] C. Giaume, A. Koulakoff, L. Roux, D. Holcman, and N. Rouach, "Astroglial networks: a step further in neuroglial and gliovascular interactions," *Nature Reviews Neuroscience*, vol. 11, no. 2, pp. 87–99, 2010.
- [61] S. Nielsen, E. Arnulf Nagelhus, M. Amiry-Moghaddam, C. Bourque, P. Agre, and O. Petter Ottersen, "Specialized membrane domains for water transport in glial cells: high-

- resolution immunogold cytochemistry of aquaporin-4 in rat brain,” *Journal of Neuroscience*, vol. 17, no. 1, pp. 171–180, 1997.
- [62] A.-C. Boulay, B. Saubaméa, S. Cisternino et al., “The Sarcoglycan complex is expressed in the cerebrovascular system and is specifically regulated by astroglial Cx30 channels,” *Frontiers in Cellular Neuroscience*, vol. 9, 2015.
- [63] A. Lampron, A. Elali, and S. Rivest, “Innate immunity in the CNS: redefining the relationship between the CNS and its environment,” *Neuron*, vol. 78, no. 2, pp. 214–232, 2013.
- [64] A. C. Boulay, A. Mazeraud, S. Cisternino et al., “Immune quiescence of the brain is set by astroglial connexin 43,” *Journal of Neuroscience*, vol. 35, no. 10, pp. 4427–4439, 2015.
- [65] C. A. Whittaker and R. O. Hynes, “Distribution and evolution of von Willebrand/integrin A domains: widely dispersed domains with roles in cell adhesion and elsewhere,” *Molecular Biology of the Cell*, vol. 13, no. 10, pp. 3369–3387, 2002.
- [66] N. Naba, K. R. Clauser, S. Hoersch, H. Liu, S. A. Carr, and R. O. Hynes, “The matrisome: *in silico* definition and *in vivo* characterization by proteomics of normal and tumor extracellular matrices,” *Molecular & Cellular Proteomics*, vol. 11, no. 4, article M111 014647, 2012.
- [67] K. Markoullis, I. Sargiannidou, N. Schiza et al., “Gap junction pathology in multiple sclerosis lesions and normal-appearing white matter,” *Acta Neuropathologica*, vol. 123, no. 6, pp. 873–886, 2012.
- [68] K. Masaki, S. O. Suzuki, T. Matsushita et al., “Connexin 43 astrocytopathy linked to rapidly progressive multiple sclerosis and neuromyelitis optica,” *PLoS One*, vol. 8, no. 8, article e72919, 2013.
- [69] L. A. B. Elias, D. D. Wang, and A. R. Kriegstein, “Gap junction adhesion is necessary for radial migration in the neocortex,” *Nature*, vol. 448, no. 7156, pp. 901–907, 2007.
- [70] L. A. B. Elias, M. Turmaine, J. G. Parnavelas, and A. R. Kriegstein, “Connexin 43 mediates the tangential to radial migratory switch in ventrally derived cortical interneurons,” *Journal of Neuroscience*, vol. 30, no. 20, pp. 7072–7077, 2010.
- [71] A. Alonso, E. Reinze, J. W. Jenne et al., “Reorganization of gap junctions after focused ultrasound blood-brain barrier opening in the rat brain,” *Journal of Cerebral Blood Flow & Metabolism*, vol. 30, no. 7, pp. 1394–1402, 2010.
- [72] X. Li, C. Olson, S. Lu et al., “Neuronal connexin36 association with zonula occludens-1 protein (ZO-1) in mouse brain and interaction with the first PDZ domain of ZO-1,” *European Journal of Neuroscience*, vol. 19, no. 8, pp. 2132–2146, 2004.
- [73] S. J. Mulligan and B. A. MacVicar, “Calcium transients in astrocyte endfeet cause cerebrovascular constrictions,” *Nature*, vol. 431, no. 7005, pp. 195–199, 2004.
- [74] M. Zonta, M. C. Angulo, S. Gobbo et al., “Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation,” *Nature Neuroscience*, vol. 6, no. 1, pp. 43–50, 2003.
- [75] M. A. Retamal, N. Froger, N. Palacios-Prado et al., “Cx43 hemichannels and gap junction channels in astrocytes are regulated oppositely by proinflammatory cytokines released from activated microglia,” *Journal of Neuroscience*, vol. 27, no. 50, pp. 13781–13792, 2007.
- [76] M. A. Retamal, K. A. Schalper, K. F. Shoji, M. V. L. Bennett, and J. C. Saez, “Opening of connexin 43 hemichannels is increased by lowering intracellular redox potential,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 104, no. 20, pp. 8322–8327, 2007.
- [77] M. De Bocka, M. Culot, N. Wang et al., “Low extracellular Ca^{2+} conditions induce an increase in brain endothelial permeability that involves intercellular Ca^{2+} waves,” *Brain Research*, vol. 1487, pp. 78–87, 2012.
- [78] W. H. Evans and L. Leybaert, “Mimetic peptides as blockers of connexin channel-facilitated intercellular communication,” *Cell Communication & Adhesion*, vol. 14, no. 6, pp. 265–273, 2007.
- [79] J. Kang, N. Kang, D. Lovatt et al., “Connexin 43 hemichannels are permeable to ATP,” *Journal of Neuroscience*, vol. 28, no. 18, pp. 4702–4711, 2008.
- [80] K. Braet, W. Vandamme, P. E. Martin, W. H. Evans, and L. Leybaert, “Photoliberating inositol-1,4,5-trisphosphate triggers ATP release that is blocked by the connexin mimetic peptide gap 26,” *Cell Calcium*, vol. 33, no. 1, pp. 37–48, 2003.
- [81] K. Braet, S. Aspeslagh, W. Vandamme et al., “Pharmacological sensitivity of ATP release triggered by photoliberation of inositol-1,4,5-trisphosphate and zero extracellular calcium in brain endothelial cells,” *Journal of Cellular Physiology*, vol. 197, no. 2, pp. 205–213, 2003.
- [82] W. Vandamme, K. Braet, L. Cabooter, and L. Leybaert, “Tumour necrosis factor alpha inhibits purinergic calcium signalling in blood-brain barrier endothelial cells,” *Journal of Neurochemistry*, vol. 88, no. 2, pp. 411–421, 2004.
- [83] H. K. Eltzschig, T. Eckle, A. Mager et al., “ATP release from activated neutrophils occurs via connexin 43 and modulates adenosine-dependent endothelial cell function,” *Circulation Research*, vol. 99, no. 10, pp. 1100–1108, 2006.
- [84] E. Decrock, E. de Vuyst, M. Vinken et al., “Connexin 43 hemichannels contribute to the propagation of apoptotic cell death in a rat C6 glioma cell model,” *Cell Death and Differentiation*, vol. 16, no. 1, pp. 151–163, 2009.
- [85] T. Constantinou, F. Baumann, M. D. Lacher, S. Saurer, R. Friis, and A. Dharmarajan, “SFRP-4 abrogates Wnt-3a-induced β -catenin and Akt/PKB signalling and reverses a Wnt-3a-imposed inhibition of *in vitro* mammary differentiation,” *Journal of Molecular Signaling*, vol. 3, p. 10, 2008.
- [86] L. Liu, W. Wan, S. Xia, B. Kalionis, and Y. Li, “Dysfunctional Wnt/ β -catenin signaling contributes to blood-brain barrier breakdown in Alzheimer’s disease,” *Neurochemistry International*, vol. 75, pp. 19–25, 2014.
- [87] B. Obermeier, R. Daneman, and R. M. Ransohoff, “Development, maintenance and disruption of the blood-brain barrier,” *Nature Medicine*, vol. 19, no. 12, pp. 1584–1596, 2013.
- [88] E. Brand-Schieber, P. Werner, D. A. Jacobas et al., “Connexin43, the major gap junction protein of astrocytes, is down-regulated in inflamed white matter in an animal model of multiple sclerosis,” *Journal of Neuroscience Research*, vol. 80, no. 6, pp. 798–808, 2005.
- [89] E. A. Eugenin, D. Basilio, J. C. Sáez et al., “The role of gap junction channels during physiologic and pathologic conditions of the human central nervous system,” *Journal of Neuroimmune Pharmacology*, vol. 7, no. 3, pp. 499–518, 2012.
- [90] K. Markoullis, I. Sargiannidou, C. Gardner, A. Hadjisavvas, R. Reynolds, and K. A. Kleopa, “Disruption of oligodendrocyte gap junctions in experimental autoimmune encephalomyelitis,” *Glia*, vol. 60, no. 7, pp. 1053–1066, 2012.
- [91] S. E. Lutz, C. S. Raine, and C. F. Brosnan, “Loss of astrocyte connexins 43 and 30 does not significantly alter susceptibility

- or severity of acute experimental autoimmune encephalomyelitis in mice," *Journal of Neuroimmunology*, vol. 245, no. 1-2, pp. 8-14, 2012.
- [92] J. D. Huber, K. A. Witt, S. Hom, R. D. Egleton, K. S. Mark, and T. P. Davis, "Inflammatory pain alters blood-brain barrier permeability and tight junctional protein expression," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 280, no. 3, pp. H1241-H1248, 2001.
- [93] H.-S. Choi, D. H. Roh, S. Y. Yoon et al., "The role of spinal interleukin-1 β and astrocyte connexin 43 in the development of mirror-image pain in an inflammatory pain model," *Experimental Neurology*, vol. 287, pp. 1-13, 2017.
- [94] G. Chen, C. K. Park, R. G. Xie, T. Berta, M. Nedergaard, and R. R. Ji, "Connexin-43 induces chemokine release from spinal cord astrocytes to maintain late-phase neuropathic pain in mice," *Brain*, vol. 137, no. 8, pp. 2193-2209, 2014.
- [95] C. A. Lee-Kubli, M. Ingves, K. W. Henry et al., "Analysis of the behavioral, cellular and molecular characteristics of pain in severe rodent spinal cord injury," *Experimental Neurology*, vol. 278, pp. 91-104, 2016.
- [96] M. J. Chen, B. Kress, X. Han et al., "Astrocytic CX43 hemichannels and gap junctions play a crucial role in development of chronic neuropathic pain following spinal cord injury," *Glia*, vol. 60, no. 11, pp. 1660-1670, 2012.
- [97] Z. Zhou, X. Wei, J. Xiang et al., "Protection of erythropoietin against ischemic neurovascular unit injuries through the effects of connexin43," *Biochemical and Biophysical Research Communications*, vol. 458, no. 3, pp. 656-662, 2015.
- [98] T. Nakase, S. Fushiki, and C. C. G. Naus, "Astrocytic gap junctions composed of connexin 43 reduce apoptotic neuronal damage in cerebral ischemia," *Stroke*, vol. 34, no. 8, pp. 1987-1993, 2003.
- [99] N. M. Kerr, C. S. Johnson, C. F. de Souza et al., "Immunolocalization of gap junction protein connexin43 (GJA1) in the human retina and optic nerve," *Investigative Ophthalmology & Visual Science*, vol. 51, no. 8, pp. 4028-4034, 2010.
- [100] J. A. Orellana, P. J. Sáez, K. F. Shoji et al., "Modulation of brain hemichannels and gap junction channels by pro-inflammatory agents and their possible role in neurodegeneration," *Antioxidants & Redox Signaling*, vol. 11, no. 2, pp. 369-399, 2009.
- [101] N. N. Osborne, R. J. Casson, J. P. M. Wood, G. Chidlow, M. Graham, and J. Melena, "Retinal ischemia: mechanisms of damage and potential therapeutic strategies," *Progress in Retinal and Eye Research*, vol. 23, no. 1, pp. 91-147, 2004.
- [102] A. Rodriguez-Sinovas, A. Cabestrero, D. Lopez et al., "The modulatory effects of connexin 43 on cell death/survival beyond cell coupling," *Progress in Biophysics and Molecular Biology*, vol. 94, no. 1-2, pp. 219-232, 2007.
- [103] C. Kaur, V. Sivakumar, Y. Zhang, and E. A. Ling, "Hypoxia-induced astrocytic reaction and increased vascular permeability in the rat cerebellum," *Glia*, vol. 54, no. 8, pp. 826-839, 2006.
- [104] W. D. Dietrich, O. Alonso, and M. Halley, "Early microvascular and neuronal consequences of traumatic brain injury: a light and electron microscopic study in rats," *Journal of Neurotrauma*, vol. 11, no. 3, pp. 289-301, 1994.
- [105] A. Hirano, T. Kawanami, and J. F. Llana, "Electron microscopy of the blood-brain barrier in disease," *Microscopy Research & Technique*, vol. 27, no. 6, pp. 543-556, 1994.
- [106] A. Rami, T. Volkman, and J. Winckler, "Effective reduction of neuronal death by inhibiting gap junctional intercellular communication in a rodent model of global transient cerebral ischemia," *Experimental Neurology*, vol. 170, no. 2, pp. 297-304, 2001.
- [107] J. O. Davidson, C. R. Green, L. F. B. Nicholson et al., "Connexin hemichannel blockade improves outcomes in a model of fetal ischemia," *Annals of Neurology*, vol. 71, no. 1, pp. 121-132, 2012.
- [108] A. C. Wilson, L. Clemente, T. Liu, R. L. Bowen, S. V. Meethal, and C. S. Atwood, "Reproductive hormones regulate the selective permeability of the blood-brain barrier," *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, vol. 1782, no. 6, pp. 401-407, 2008.
- [109] M. P. Mattson, "Cellular actions of beta-amyloid precursor protein and its soluble and fibrillogenic derivatives," *Physiological Reviews*, vol. 77, no. 4, pp. 1081-1132, 1997.
- [110] N. J. Haughey and M. P. Mattson, "Alzheimer's amyloid β -peptide enhances ATP/gap junction-mediated calcium-wave propagation in astrocytes," *NeuroMolecular Medicine*, vol. 3, no. 3, pp. 173-180, 2003.
- [111] C. Yi, P. Ezan, P. Fernández et al., "Inhibition of glial hemichannels by boldine treatment reduces neuronal suffering in a murine model of Alzheimer's disease," *Glia*, vol. 65, no. 10, pp. 1607-1625, 2017.
- [112] C. Yi, X. Mei, P. Ezan et al., "Astroglial connexin43 contributes to neuronal suffering in a mouse model of Alzheimer's disease," *Cell Death & Differentiation*, vol. 23, no. 10, pp. 1691-1701, 2016.
- [113] X. Mei, P. Ezan, C. Giaume, and A. Koulakoff, "Astroglial connexin immunoreactivity is specifically altered at β -amyloid plaques in β -amyloid precursor protein/presenilin1 mice," *Neuroscience*, vol. 171, no. 1, pp. 92-105, 2010.
- [114] H. Takeuchi, H. Mizoguchi, Y. Doi et al., "Blockade of gap junction hemichannel suppresses disease progression in mouse models of amyotrophic lateral sclerosis and Alzheimer's disease," *PLoS One*, vol. 6, no. 6, article e21108, 2011.
- [115] L. P. Cacheaux, S. Ivens, Y. David et al., "Transcriptome profiling reveals TGF- β signaling involvement in epileptogenesis," *Journal of Neuroscience*, vol. 29, no. 28, pp. 8927-8935, 2009.
- [116] R. Kovacs, U. Heinemann, and C. Steinhäuser, "Mechanisms underlying blood-brain barrier dysfunction in brain pathology and epileptogenesis: role of astroglia," *Epilepsia*, vol. 53, Supplement 6, pp. 53-59, 2012.
- [117] Y. David, L. P. Cacheaux, S. Ivens et al., "Astrocytic dysfunction in epileptogenesis: consequence of altered potassium and glutamate homeostasis?," *The Journal of Neuroscience*, vol. 29, no. 34, pp. 10588-10599, 2009.
- [118] Z. Gajda, E. Hermes, E. Gyengesi, Z. Szupera, and M. Szenté, "The functional significance of gap junction channels in the epileptogenicity and seizure susceptibility of juvenile rats," *Epilepsia*, vol. 47, no. 6, pp. 1009-1022, 2006.
- [119] M. Samoilova, K. Wentlandt, Y. Adamchik, A. A. Velumian, and P. L. Carlen, "Connexin 43 mimetic peptides inhibit spontaneous epileptiform activity in organotypic hippocampal slice cultures," *Experimental Neurology*, vol. 210, no. 2, pp. 762-775, 2008.
- [120] M. A. Cosenza, M.-L. Zhao, Q. Si, and S. C. Lee, "Human brain parenchymal microglia express CD14 and CD45 and

- are productively infected by HIV-1 in HIV-1 encephalitis,” *Brain Pathology*, vol. 12, no. 4, pp. 442–455, 2002.
- [121] E. A. Eugenin and J. W. Berman, “Gap junctions mediate human immunodeficiency virus-bystander killing in astrocytes,” *The Journal of Neuroscience*, vol. 27, no. 47, pp. 12844–12850, 2007.
- [122] M. J. Churchill, S. L. Wesselingh, D. Cowley et al., “Extensive astrocyte infection is prominent in human immunodeficiency virus-associated dementia,” *Annals of Neurology*, vol. 66, no. 2, pp. 253–258, 2009.
- [123] E. A. Eugenin, J. E. Clements, M. C. Zink, and J. W. Berman, “Human immunodeficiency virus infection of human astrocytes disrupts blood-brain barrier integrity by a gap junction-dependent mechanism,” *Journal of Neuroscience*, vol. 31, no. 26, pp. 9456–9465, 2011.
- [124] J. Z. Zhou and J. X. Jiang, “Gap junction and hemichannel-independent actions of connexins on cell and tissue functions—an update,” *FEBS Letters*, vol. 588, no. 8, pp. 1186–1192, 2014.
- [125] M. Vinken, E. Decrock, L. Leybaert et al., “Non-channel functions of connexins in cell growth and cell death,” *Biochimica et Biophysica Acta (BBA) - Biomembranes*, vol. 1818, no. 8, pp. 2002–2008, 2012.
- [126] R. Yamasaki, H. Lu, O. Butovsky et al., “Differential roles of microglia and monocytes in the inflamed central nervous system,” *Journal of Experimental Medicine*, vol. 211, no. 8, pp. 1533–1549, 2014.



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