

Review Article

Writers and Readers of DNA Methylation/Hydroxymethylation in Physiological Aging and Its Impact on Cognitive Function

Rodrigo F. Torres ^{1,2} Ricardo Kouro,^{1,3} and Bredford Kerr ^{1,4}

¹Centro de Estudios Científicos, Valdivia 5110466, Chile

²Fundación Cultura Científica, Valdivia 5112119, Chile

³Programa de Honor en Investigación, Facultad de Medicina, Universidad Austral de Chile, Valdivia 5110566, Chile

⁴Centro de Biología Celular y Biomedicina-CEBICEM, Facultad de Medicina y Ciencia, Universidad San Sebastián, Santiago 7510157, Chile

Correspondence should be addressed to Rodrigo F. Torres; rtorres@cecs.cl and Bredford Kerr; bredford.kerr@uss.cl

Received 22 February 2019; Revised 25 April 2019; Accepted 26 May 2019; Published 14 July 2019

Guest Editor: Vanessa Nieratschker

Copyright © 2019 Rodrigo F. Torres et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The chromatin landscape has acquired deep attention from several fields ranging from cell biology to neurological and psychiatric diseases. The role that DNA modifications have on gene expression regulation has become apparent in several physiological processes, and numerous efforts have been performed to establish a relationship between DNA modifications and physiological conditions, such as cognitive performance and aging. DNA modifications are incorporated by specific sets of enzymes—the writers—and the modified DNA-interacting partners—the readers—are ultimately responsible for maintaining a functional epigenetic landscape. Therefore, understanding how these epigenetic mediators—writers and readers—are modulated in physiological aging will contribute to unraveling how aging-associated neuronal disturbances arise and contribute to the cognitive decline associated with this period of life. In this review, we focused on DNA modifications, writers and readers, highlighting that despite some methodological disparities, the evidence suggests a critical role for epigenetic mediators in the aging-associated neuronal dysfunction.

1. Introduction

Changes in gene expression allow the dynamic nature of the neuronal function. The role of gene expression regulation in directing neuronal states has been highlighted in synaptic plasticity and learning and memory processes [1]. However, its relevance has acquired attention from several biological processes relevant to neuronal function, such as energy metabolism, redox balance, unfolded protein response, neurological and psychiatric diseases, and physiological aging [2–4].

One of the mechanisms through which gene expression is controlled in a long-lasting manner is by the remodeling of the chromatin landscape induced by epigenetic changes, which are chemical modifications in histone proteins and DNA bases. Modifications of the chromatin landscape are functionally relevant, as it settles the accessibility of protein complexes to regulatory regions of promoters and thus direct gene expression. The epigenetic modifications are signatures

through which environmental factors chemically modify the genome to leave a record and form, a many times permanent, genetic-molecular memory. In this regard, epigenetic modifications have emerged as a relevant mechanism underlying dynamic transcriptional regulation contributing to neuronal function [5], and several efforts have been made to establish a relationship between aging and epigenetic modifications [6, 7]. DNA modifications, such as DNA methylation and DNA hydroxymethylation, are dynamically incorporated by specific sets of enzymes or writers. DNA-modified interacting proteins, or readers, allow the translation of these modifications into functional transcriptional signals that modulate gene expression contributing to neuronal function.

Besides those changes in histone and DNA methylation/hydroxymethylation, aging-associated changes in epigenetic mediators—writers and readers—could be involved in the age-related changes in gene expression that underlie one of the most singular characteristics associated to this period of

life, the cognitive decline. Moreover, the processes leading to epigenetic mark erasure could also contribute to aging-associated disturbances and should be considered. Therefore, a clear understanding of the age-associated expression dynamics of epigenetic writers and readers will contribute to unraveling the aging-associated disturbances of neuronal function. In this review, we focused on DNA methylation and DNA hydroxymethylation writers and readers to highlight our current knowledge of how physiological aging modifies the expression of epigenetic mediators and the relevance of these changes for the maintenance of cognitive abilities.

2. DNA Methylation and DNA Hydroxymethylation Writers

The covalent binding of a methyl group to carbon 5 of a cytosine (5meC) is an epigenetic mark that is subjected to dynamic modifications. 5meC occurs at a higher frequency in cytosines of CpG dinucleotides; however, 5meC in non-CpG dinucleotides has acquired an increased relevance [8]. 5meC is catalyzed by DNA methyltransferases (DNMTs), a family of proteins that bind to DNA and write the epigenome. There are two classes of DNMTs: *maintenance* DNMTs and *de novo* DNMTs. Both groups share the use of S-adenosyl-L-methionine (SAM) as a methyl donor, and both groups of DNMTs are expressed in the brain [9, 10]. DNMT1 is the main *maintenance* DNMT and is proposed to maintain DNA methylation after DNA replication [11]. On the other hand, DNMT3a and DNMT3b are the main *de novo* DNMTs and incorporate a previously nonexistent 5meC to the genome. The *DNMT3a* locus gives rise to 2 protein isoforms named DNMT3a1 and a DNMT3a2. The DNMT3a2 lacks 219 aminoterminal residues in comparison to DNMT3a1, and it has been shown to be sensitive to an increase in neuronal activity [12]. In addition, non-CpG methylation accumulates in the genome through development, reaching levels similar to that of CpG in the adult brain [8]. Non-CpG accumulation is mediated by a transient recruitment of DNMT3a during early postnatal development [13, 14]. Interestingly, DNMT3a recruitment is modulated by early-life experiences, affecting non-CpG levels in later life [14], increasing the complexity of DNA methylation signal. Interestingly, despite its postmitotic nature, DNA methylation in neurons is far from being stable and is dynamically regulated by natural and artificial stimuli [15–17].

5meC can undergo an active series of chemical modifications that lead to cytosine demethylation. This pathway involves the oxidation of methylcytosine to hydroxymethylcytosine (5hmeC), 5-formylcytosine (fC), and 5-carboxylcytosine (caC) followed by excision of fC or caC mediated by thymine DNA glycosylase coupled with base excision repair [18]. The first of these chemical reactions is mediated by the ten-eleven translocator protein family (TETs). The TET family of proteins has three members in mammals (TET1-3), and all of them catalyze the modification of 5meC to 5hmeC [18]. The TET-mediated oxidation of 5meC to 5hmeC is α -ketoglutarate and Fe (II)-dependent reaction, generating an interesting link to cellular metabolism [19]. TET enzymes and 5hmeC have also been shown

to have a pivotal participation in learning and memory processes and are dynamically regulated by environmental stimulation, suggesting a role for this epigenetic modification in activity-dependent gene expression [20–22].

3. DNA Methylation and DNA Hydroxymethylation Readers

5meC regulates gene expression either directly, by inhibiting the association of transcription factors to its binding site, or indirectly via a family of proteins containing a methyl-binding domain (MBD) [23]. The MBD was first characterized in the amino acid sequence of the methyl CpG binding protein-2 (MeCP2) and is shared by the proteins MBD1, MBD2, MBD3, and MBD4. These proteins, excluding MBD3, bind to 5meC and recruit several proteins associated to chromatin remodeling [24]. The relevance of MBD proteins in the central nervous system was highlighted by the association of MECP2 mutations as the major cause for Rett syndrome (RTT, MIM 312750), a devastating neurodevelopmental disorder [25]. Interestingly, restoring *Mecp2* expression in postmitotic neurons recovers RTT in mice [26], and *Mecp2* ablation in later age windows is incompatible with life [27], highlighting how important is the proper reading of epigenetic signals for the maintenance of postmitotic neuronal function. *Mecp2* is the most abundantly expressed member of the MBD family in the brain, and it is coded by a 4-exon locus in the X chromosome, giving rise to 2 MECP2 isoforms. In addition to its MBD, MECP2 has a transcriptional repressor domain (TRD) and a carboxy-terminal domain. The MBD is necessary and sufficient for methylcytosine binding in a DNA sequence-independent manner [28], and the TRD domain has been shown to interact with HDACs among other proteins [29]. In spite that *Mecp2* was initially described as a transcriptional repressor, it was latter shown that some of *Mecp2* target genes are downregulated in the brain of a mouse model lacking the expression of *Mecp2*, suggesting that *Mecp2* is also able to activate gene expression [30]. Currently, several evidences suggest that *Mecp2* acts as a chromatin structure regulator, and therefore, the effect of *Mecp2* mutations or posttranslational modifications will be dependent upon the chromatin context [31, 32].

The mechanism through which *Mecp2* activates the expression of its target genes is still a matter of study. It has been described that *Mecp2* interacts with the cAMP response element-binding protein (CREB) [30], a widely known activator of gene expression. Considering that CREB phosphorylation and CBP recruiting are induced by neuronal activity [33], it is not risky to propose this interaction as a molecular linker between environmental factors and gene expression control.

It has also been shown that *Mecp2* binds 5hmeC in the central nervous system [34]; therefore, *Mecp2* represents an intersectional link between DNA methylation and hydroxymethylation. The affinity of *Mecp2* for 5meC vs. 5hmeC is a matter of study that will contribute to understand how epigenetic modifications relate to the tuning of gene expression. Recent evidences suggest that MBD1 has a higher affinity for 5meC than *Mecp2*; on the other hand, *Mecp2* seems to have higher affinity for 5hmeC than MBD1 [35]. Hence, the

binding of MBD1 or Mecp2 to a certain promoter region could act to switch gene expression [35]. Thus, the control of TET-mediated oxidation activity over discrete genomic regions could have profound effects on the chromatin landscape and gene regulation, highlighting the dynamism of epigenetic changes.

Interestingly, neuronal activity modulates Mecp2 phosphorylation, decreasing phosphoserine 80 [36] and increasing phosphoserine 421 [37, 38], suggesting that Mecp2 posttranscriptional modifications are relevant to synaptic plasticity and activity-dependent gene expression [39], further highlighting the role of Mecp2 in the adult brain.

As exemplified by Mecp2 phosphorylation, there is a vast range of posttranslational modifications regulating the function of Mecp2 [39]. Understanding how these posttranscriptional modifications contribute to Mecp2 functionality in aging will be crucial to understand the physiological aging-associated neuronal dysfunction.

The information regarding Mecp2 contribution to cognitive function comes mainly from studies using mouse models of RTT. For instance, a mouse model containing a truncated form of Mecp2 shows impaired synaptic plasticity and learning and memory [40]. Similarly, the Mecp2-null mice showed a disruption of synaptic plasticity and deficits in cerebellar and amygdala-based learning [41, 42]. Of relevance to aging-associated cognitive decline, it has been shown that Mecp2-null mice show an impairment of place cell function, resulting in the incapacity for maintaining contextual memory in long-time scales [43]. Moreover, by using a knock-down approach to delete the expression of Mecp2 from the hippocampus of adult mice, Gulmez Karaca et al. reported that Mecp2 is required for long-term memory formation and consolidation [44].

4. Writers, Readers, and Aging

The cognitive decline associated with aging is linked to a change in gene expression that drives several neuronal functions [45, 46]. As DNA methylation and hydroxymethylation are dynamically regulated and sensitive to neuronal activity, the observation that these modifications also vary across the lifespan suggested a role for DNA modifications during physiological aging [6, 47–49]. The reader is directed to specialized works and reviews to address the aging-associated genome modifications of 5mC and 5hmC [50–54]. However, systematic studies assessing the dynamics of the expression of epigenetic writers and readers across the lifespan are lacking and the available information raises technical hitches which contribute to variability between laboratories, such as the age of “aged mice” sampling and the age of the “young” mice used as controls. However, it is remarkable that despite these differences, similar findings have been reported which are summarized in Table 1.

Although few efforts have been made to elucidate the role of epigenetic mediators in aging, the results obtained have been highly interesting and call for further attention. Based on the general consensus that global DNA methylation decreases in the aged brain [52], Oliveira et al. investigated a possible causal role for DNMTs. They observed that the

expression of DNMT3a1 and DNMT3a2 is decreased in the hippocampus of 18-month-old mice. Restoring the expression of DNMT3a2 recovered the declined cognitive performance in fear conditioning and the object-place recognition test. Inversely, decreasing DNMT3a2 levels were detrimental in young 3-month-old adult mice [12]. Concordantly, decreased DNMT1 and DNMT3a expression, but not DNMT3b, was observed in postmortem human samples and in the hippocampus of 30-month-old rats [55]. DNMT1 haploinsufficiency, evaluated at 12 months of age, has also been associated with an age-dependent cognitive impairment [56]. Altogether, this evidence suggests a pivotal role for DNMT dosage in aging.

A reduction in hippocampal TET1 and TET2 expression was observed in 18-month-old mice [57]. Concordantly, TET2 expression also showed a premature and age-dependent decreased expression in the hippocampus of mice starting at 6 months of age [58]. Diminished TET2 expression was associated to a reduction in the levels of hippocampal 5hmC and to reduced neurogenesis in the dentate gyrus. Moreover, increasing TET2 expression in 6-month-old mice restored neurogenesis to the level observed for young 3-month-old adult mice and enhanced associative fear memory [58]. This evidence suggests a critical role for this enzyme in aging-associated cognitive decline.

As was mentioned above, MECP2 binds to either 5mC or 5hmC and mutation in its coding gene is the main cause of RTT. Since RTT symptoms arise early in development, most of MECP2 literature overlooks the role for this protein in aging. However, tissue microarray analyses showed that MECP2 expression increases along aging in humans [59, 60] and similar results were observed at mRNA levels in 28-month-old mice [61]. Moreover, Mecp2 deletion in 7–10-month-old mice is incompatible with life [27] and increased Mecp2 protein levels have been shown to be detrimental, leading to a progressive neurological disorder [62]. This evidence suggests a relevant role for Mecp2 in aging that has not been explored and remains a key piece of information to understand the role of epigenetic mediators in the aging-associated cognitive decline. Furthermore, the evidence suggests that Mecp2 is target to miRNAs [63, 64] and the MECP2 gene contains a CpG island that could be associated with its transcriptional control [65]; however, such mechanism for the regulation of Mecp2 expression in aging has not been explored.

5. Environmental Stimulation

Some environmental conditions attenuate or delay characteristics associated to physiological aging. Among those conditions, physical exercise (PE), caloric restriction (CR), and environmental enrichment (EE) have been shown to modulate the epigenome and contribute to improve cognitive function [66–68]. For instance, CR was shown to delay the aging-associated modifications of DNA methylation [69] and PE has widely documented effects on the epigenetic landscape (for a review, see Fernandes et al., [66]). EE is an experimental paradigm widely used to increase neural plasticity in an experience-dependent manner [68]. It was shown that EE

TABLE 1: Reported changes in DNMTs, Mecp2, and TETs expression in aging.

	Species	Age (months)	Reported change	Brain region	Reference
Dnmt1	m	18	=	Hippocampus	[12]
	m	24	↓	Frontal cortex	[82]
	r	30	↓	Hippocampus	[55]
	h	*	↓	Hippocampus	[55]
Dnmt3a1	m	18	↓	Hippocampus	[12]
	r	30	↓	Hippocampus	[55]
	h	*	↓	Hippocampus	[55]
Dnmt3a2	m	18	↓	Hippocampus	[12]
Dnmt3b	m	18	=	Hippocampus	[12]
	r	30	=	Hippocampus	[55]
	h	*	=	Hippocampus	[55]
Mecp2	m	31	↑	Whole brain	[61]
	h	*	↑	Cerebral cortex	[59, 60]
Tet1	m	18	↓	Hippocampus	[57]
	m	24	=	Frontal cortex	[82]
	m	31	↓	Whole brain	[61]
Tet2	m	18	↓	Hippocampus	[57]
	m	18	↓	Hippocampus	[58]
	m	31	↑	Whole brain	[61]
Tet3	m	18	=	Hippocampus	[58]
	m	24	=	Frontal cortex	[82]
	m	31	↓	Whole brain	[61]

Species: h: human, m: mouse, r: rat. Age is presented in months. *Human postmortem samples include varied ages.

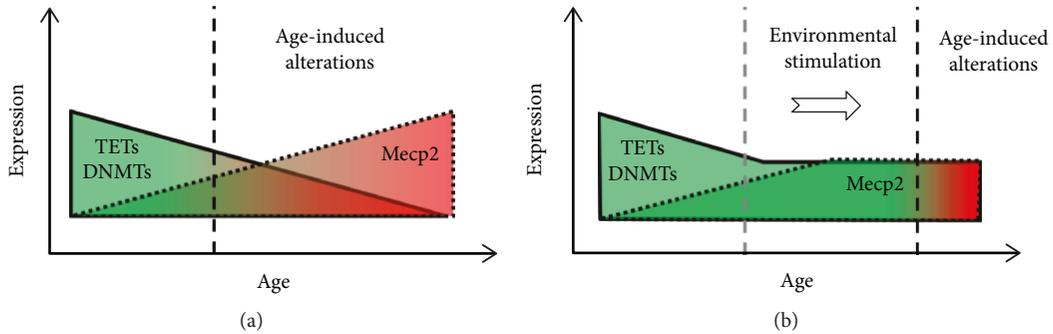


FIGURE 1: Aging-induced changes in the expression of epigenetic mediators. (a) Aging is associated with decreased DNMT1, DNMT3a1, DNMT3a2, TET1, and TET2 expression. Conversely, Mecp2 expression is increased in postmortem human samples. (b) The exposure to environmental stimulation could diminish or revert the aging-associated alterations in the expression of epigenetic mediators, as it has been observed for TET2 and physical exercise and DNMT1 activity in glucose restriction.

modulates the dynamics of hippocampal DNA hydroxy-methylation, with changes affecting mainly genes related to axonal guidance [21]. Moreover, it has been reported that short-term EE (3 weeks) in 21-month-old rats restores synaptic and visual plasticity, suggesting that environmental stimulation recovers, at least in part, the transcriptional changes associated to aging [70, 71]. A similar conclusion can be drawn by comparing the gene expression of 18-month-old mice with or without access to PE by a running wheel [72]. Mechanism associated with the effect of environmental stimulation over aging ranges across several neuronal

processes, including diminished synaptic protein loss, metabotropic glutamatergic signaling enhancement, and increased BDNF maturation, among others [73–75]. However, it is relevant to elucidate if environmental stimulation directs changes in the expression of epigenetic writers and readers in aging.

Although intrinsic age-associated disruption of neuronal circuits occurs during aging [76], the cognitive decline observed is also associated with a perceptual decline that could determine a reduction of neuronal stimulation due to the impoverished perceptual input, a hypothesis known as

“information degradation” [77, 78]. As neuronal activity arising from environmental stimuli is required to direct the changes in DNA methylation and hydroxymethylation, the impoverished perceptual input might contribute to the aging-related changes in epigenetic mediators. Under this view, it could be hypothesized that an increase in environmental stimulation would restore, prevent, or diminish the aging-associated changes in the expression of epigenetic mediators (Figure 1). In this line, it was reported that the age-associated reduction in hippocampal TET2 expression observed in 18-month-old mice was restored to the level observed in 3-month-old mice by exposure to voluntary PE in a running wheel for 4 weeks [57]. Similarly, glucose restriction increased DNMT1 activity *in vitro*, counteracting the diminished DNMT1 expression reported for aging [79]. Interestingly, *Mecp2* expression is downregulated by neuronal activity *in vitro* and it has been shown that EE reduced *Mecp2* mRNA expression in 8-week-old mice [64, 80], which raises the possibility that an aging-associated reduction in neuronal activity could underlie the increase in *Mecp2* expression observed in both aged humans and mice [59–61]. Supporting this notion, it was reported that in 2-month-old mice voluntary running (7 days) reduced *Mecp2* expression in the cerebellum [81]. Therefore, it would be interesting to assess if EE, PE, and CR modify the level of epigenetic mediators in the brain of aged mice and the consequences at the level of gene expression and cognition. Moreover, alteration of *Mecp2* protein levels could impact *Mecp2* posttranslational modifications in the aged brain, contributing to aging-associated neuronal disturbances.

The aging-induced reduction in neuronal stimulation adds a layer of complexity to the interpretation of aging-related changes in gene expression. Notwithstanding, the relevance of epigenetic mediators in aging has been strikingly highlighted by some of the works reviewed, raising an urge to fulfill the voids in our understanding.

6. Conclusions and Prospects

As it was discussed above, physiological aging is accompanied by a cognitive performance decline associated with changes in the expression of epigenetic writers and readers that modify chromatin landscape and thus impact on gene expression. However, more evidence is required to fulfill the gaps among all these processes and take our knowledge beyond correlations between these aging-associated events. To achieve that, it is needed to unveil the mechanism underlying the changes in the expression of DNA readers and writers and the physiological factors associated with the signaling that leads to these modifications. Since the transcriptional changes that accompany aging seem to be gene-specific and do not affect the entire genome, it is critical to identify the mechanism through which DNA writers select their target regions to write on it. Then, with that knowledge, we could understand how through environmental factors these changes can be prevented or mitigated to maintain a better cognitive performance during aging, which is critical nowadays in which the life expectancy has raised.

Disclosure

In this review, we expressed our own opinion which is not based on an official position of the institution or funding agency.

Conflicts of Interest

We confirm that this review was conducted with no conflict of interest.

Authors' Contributions

This review was initially conceived by RFT and BK. The text was drafted by RFT. RFT, RK, and BK contributed to the collection and sorting of evidence.

Acknowledgments

This review was supported by the Fondo Nacional de Desarrollo Científico y Tecnológico (FONDECYT) (3180518 to Rodrigo F. Torres and 1181574 to Bredford Kerr). RK is part of the Research Honor Program for medical student of the Faculty of Medicine (PHIM), Universidad Austral de Chile. The Centro de Estudios Científicos is funded by the Chilean Government through the Centers of Excellence Basal Financing Program of CONICYT.

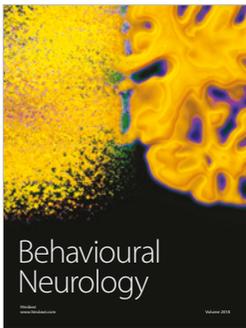
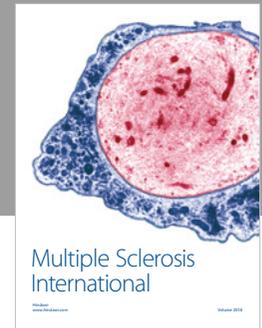
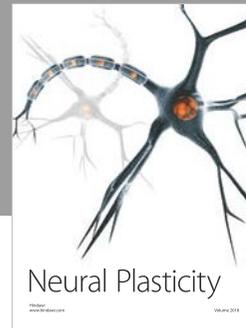
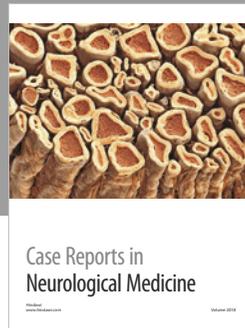
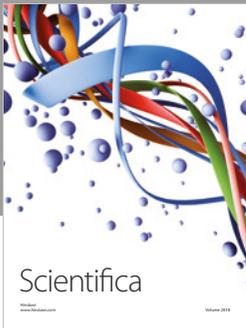
References

- [1] F. Telese, Q. Ma, P. M. Perez et al., “LRP8-Reelin-regulated neuronal enhancer signature underlying learning and memory formation,” *Neuron*, vol. 86, no. 3, pp. 696–710, 2015.
- [2] C. Bas-Orth, Y.-W. Tan, D. Lau, and H. Bading, “Synaptic activity drives a genomic program that promotes a neuronal Warburg effect,” *Journal of Biological Chemistry*, vol. 292, no. 13, pp. 5183–5194, 2017.
- [3] E. J. Ryu, H. P. Harding, J. M. Angelastro, O. V. Vitolo, D. Ron, and L. A. Greene, “Endoplasmic reticulum stress and the unfolded protein response in cellular models of Parkinson’s disease,” *The Journal of Neuroscience*, vol. 22, no. 24, pp. 10690–10698, 2002.
- [4] J. Yang, E. Ruchti, J.-M. Petit et al., “Lactate promotes plasticity gene expression by potentiating NMDA signaling in neurons,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 111, no. 33, pp. 12228–12233, 2014.
- [5] J. Cortés-Mendoza, S. Díaz de León-Guerrero, G. Pedraza-Alva, and L. Pérez-Martínez, “Shaping synaptic plasticity: the role of activity-mediated epigenetic regulation on gene transcription,” *International Journal of Developmental Neuroscience*, vol. 31, no. 6, pp. 359–369, 2013.
- [6] L. Ianov, A. Riva, A. Kumar, and T. C. Foster, “DNA methylation of synaptic genes in the prefrontal cortex is associated with aging and age-related cognitive impairment,” *Frontiers in Aging Neuroscience*, vol. 9, 2017.
- [7] H. A. Irier and P. Jin, “Dynamics of DNA methylation in aging and Alzheimer’s disease,” *DNA and Cell Biology*, vol. 31, Supplement 1, pp. S-42–S-48, 2012.
- [8] J. U. Guo, Y. Su, J. H. Shin et al., “Distribution, recognition and regulation of non-CpG methylation in the adult

- mammalian brain," *Nature Neuroscience*, vol. 17, no. 2, pp. 215–222, 2014.
- [9] T. H. Bestor, "The DNA methyltransferases of mammals," *Human Molecular Genetics*, vol. 9, no. 16, pp. 2395–2402, 2000.
- [10] R. J. Klose and A. P. Bird, "Genomic DNA methylation: the mark and its mediators," *Trends in Biochemical Sciences*, vol. 31, no. 2, pp. 89–97, 2006.
- [11] H. Leonhardt, A. W. Page, H.-U. Weier, and T. H. Bestor, "A targeting sequence directs DNA methyltransferase to sites of DNA replication in mammalian nuclei," *Cell*, vol. 71, no. 5, pp. 865–873, 1992.
- [12] A. M. M. Oliveira, T. J. Hemstedt, and H. Bading, "Rescue of aging-associated decline in Dnmt3a2 expression restores cognitive abilities," *Nature Neuroscience*, vol. 15, no. 8, pp. 1111–1113, 2012.
- [13] B. H. Ramsahoye, D. Biniszkiwicz, F. Lyko, V. Clark, A. P. Bird, and R. Jaenisch, "Non-CpG methylation is prevalent in embryonic stem cells and may be mediated by DNA methyltransferase 3a," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 10, pp. 5237–5242, 2000.
- [14] H. Stroud, S. C. Su, S. Hrvatin et al., "Early-life gene expression in neurons modulates lasting epigenetic states," *Cell*, vol. 171, no. 5, pp. 1151–1164.e16, 2017.
- [15] J. U. Guo, D. K. Ma, H. Mo et al., "Neuronal activity modifies the DNA methylation landscape in the adult brain," *Nature Neuroscience*, vol. 14, no. 10, pp. 1345–1351, 2011.
- [16] R. Halder, M. Hennion, R. O. Vidal et al., "DNA methylation changes in plasticity genes accompany the formation and maintenance of memory," *Nature Neuroscience*, vol. 19, no. 1, pp. 102–110, 2016.
- [17] C. G. Duke, A. J. Kennedy, C. F. Gavin, J. J. Day, and J. D. Sweatt, "Experience-dependent epigenomic reorganization in the hippocampus," *Learning & Memory*, vol. 24, no. 7, pp. 278–288, 2017.
- [18] X. Wu and Y. Zhang, "TET-mediated active DNA demethylation: mechanism, function and beyond," *Nature Reviews Genetics*, vol. 18, no. 9, pp. 517–534, 2017.
- [19] J.-P. Etchegaray and R. Mostoslavsky, "Interplay between metabolism and epigenetics: a nuclear adaptation to environmental changes," *Molecular Cell*, vol. 62, no. 5, pp. 695–711, 2016.
- [20] Y. Alagband, T. W. Bredy, and M. A. Wood, "The role of active DNA demethylation and Tet enzyme function in memory formation and cocaine action," *Neuroscience Letters*, vol. 625, pp. 40–46, 2016.
- [21] H. Irier, R. C. Street, R. Dave et al., "Environmental enrichment modulates 5-hydroxymethylcytosine dynamics in hippocampus," *Genomics*, vol. 104, no. 5, pp. 376–382, 2014.
- [22] A. Rudenko, M. M. Dawlaty, J. Seo et al., "Tet1 is critical for neuronal activity-regulated gene expression and memory extinction," *Neuron*, vol. 79, no. 6, pp. 1109–1122, 2013.
- [23] P. A. Jones and D. Takai, "The role of DNA methylation in mammalian epigenetics," *Science*, vol. 293, no. 5532, pp. 1068–1070, 2001.
- [24] X. Zou, W. Ma, I. A. Solov'yov, C. Chipot, and K. Schulten, "Recognition of methylated DNA through methyl-CpG binding domain proteins," *Nucleic Acids Research*, vol. 40, no. 6, pp. 2747–2758, 2012.
- [25] R. E. Amir, I. B. Van den Veyver, M. Wan, C. Q. Tran, U. Francke, and H. Y. Zoghbi, "Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2," *Nature Genetics*, vol. 23, no. 2, pp. 185–188, 1999.
- [26] S. Luikenhuis, E. Giacometti, C. F. Beard, and R. Jaenisch, "Expression of MeCP2 in postmitotic neurons rescues Rett syndrome in mice," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 16, pp. 6033–6038, 2004.
- [27] H. Cheval, J. Guy, C. Merusi, D. De Sousa, J. Selfridge, and A. Bird, "Postnatal inactivation reveals enhanced requirement for MeCP2 at distinct age windows," *Human Molecular Genetics*, vol. 21, no. 17, pp. 3806–3814, 2012.
- [28] A. Free, R. I. D. Wakefield, B. O. Smith, D. T. F. Dryden, P. N. Barlow, and A. P. Bird, "DNA recognition by the methyl-CpG binding domain of MeCP2," *Journal of Biological Chemistry*, vol. 276, no. 5, pp. 3353–3360, 2001.
- [29] X. Nan, H.-H. Ng, C. A. Johnson et al., "Transcriptional repression by the methyl-CpG-binding protein MeCP2 involves a histone deacetylase complex," *Nature*, vol. 393, no. 6683, pp. 386–389, 1998.
- [30] M. Chahrour, S. Y. Jung, C. Shaw et al., "MeCP2, a key contributor to neurological disease, activates and represses transcription," *Science*, vol. 320, no. 5880, pp. 1224–1229, 2008.
- [31] A. Becker, P. Zhang, L. Allmann et al., "Poly(ADP-ribosyl)ation of methyl CpG binding domain protein 2 regulates chromatin structure," *Journal of Biological Chemistry*, vol. 291, no. 10, pp. 4873–4881, 2016.
- [32] C. S. Casas-Delucchi, A. Becker, J. J. Boliuis, and M. C. Cardoso, "Targeted manipulation of heterochromatin rescues MeCP2 Rett mutants and re-establishes higher order chromatin organization," *Nucleic Acids Research*, vol. 40, no. 22, article e176, 2012.
- [33] E. Korzus, M. G. Rosenfeld, and M. Mayford, "CBP histone acetyltransferase activity is a critical component of memory consolidation," *Neuron*, vol. 42, no. 6, pp. 961–972, 2004.
- [34] M. Mellén, P. Ayata, S. Dewell, S. Kriaucionis, and N. Heintz, "MeCP2 binds to 5hmC enriched within active genes and accessible chromatin in the nervous system," *Cell*, vol. 151, no. 7, pp. 1417–1430, 2012.
- [35] A. Marco, T. Kisliouk, T. Tabachnik, A. Weller, and N. Meiri, "DNA CpG methylation (5-methylcytosine) and its derivative (5-hydroxymethylcytosine) alter histone posttranslational modifications at the Pomc promoter, affecting the impact of perinatal diet on leanness and obesity of the offspring," *Diabetes*, vol. 65, no. 8, pp. 2258–2267, 2016.
- [36] J. Tao, K. Hu, Q. Chang et al., "Phosphorylation of MeCP2 at serine 80 regulates its chromatin association and neurological function," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 12, pp. 4882–4887, 2009.
- [37] B. Buchthal, D. Lau, U. Weiss, J.-M. Weislogel, and H. Bading, "Nuclear calcium signaling controls methyl-CpG-binding protein 2 (MeCP2) phosphorylation on serine 421 following synaptic activity," *Journal of Biological Chemistry*, vol. 287, no. 37, pp. 30967–30974, 2012.
- [38] Z. Zhou, E. J. Hong, S. Cohen et al., "Brain-specific phosphorylation of MeCP2 regulates activity-dependent Bdnf transcription, dendritic growth, and spine maturation," *Neuron*, vol. 52, no. 2, pp. 255–269, 2006.

- [39] E. Bellini, G. Pavesi, I. Barbiero et al., “MeCP2 post-translational modifications: a mechanism to control its involvement in synaptic plasticity and homeostasis?,” *Frontiers in Cellular Neuroscience*, vol. 8, p. 236, 2014.
- [40] P. Moretti, J. M. Levenson, F. Battaglia et al., “Learning and memory and synaptic plasticity are impaired in a mouse model of Rett syndrome,” *The Journal of Neuroscience*, vol. 26, no. 1, pp. 319–327, 2006.
- [41] G. J. Pelka, C. M. Watson, T. Radziewicz et al., “*Mecp2* deficiency is associated with learning and cognitive deficits and altered gene activity in the hippocampal region of mice,” *Brain*, vol. 129, no. 4, pp. 887–898, 2006.
- [42] Y. Asaka, D. G. M. Jugloff, L. Zhang, J. H. Eubanks, and R. M. Fitzsimonds, “Hippocampal synaptic plasticity is impaired in the *Mecp2*-null mouse model of Rett syndrome,” *Neurobiology of Disease*, vol. 21, no. 1, pp. 217–227, 2006.
- [43] S. E. Kee, X. Mou, H. Y. Zoghbi, and D. Ji, “Impaired spatial memory codes in a mouse model of Rett syndrome,” *eLife*, vol. 7, 2018.
- [44] K. Gulmez Karaca, D. V. C. Brito, B. Zeuch, and A. M. M. Oliveira, “Adult hippocampal MeCP2 preserves the genomic responsiveness to learning required for long-term memory formation,” *Neurobiology of Learning and Memory*, vol. 149, pp. 84–97, 2018.
- [45] H. B. Fraser, P. Khaitovich, J. B. Plotkin, S. Pääbo, and M. B. Eisen, “Aging and gene expression in the primate brain,” *PLoS Biology*, vol. 3, no. 9, article e274, 2005.
- [46] D. Glass, A. Viñuela, M. N. Davies et al., “Gene expression changes with age in skin, adipose tissue, blood and brain,” *Genome Biology*, vol. 14, no. 7, article R75, 2013.
- [47] H. Chen, S. Dzitoyeva, and H. Manev, “Effect of aging on 5-hydroxymethylcytosine in the mouse hippocampus,” *Restorative Neurology and Neuroscience*, vol. 30, no. 3, pp. 237–245, 2012.
- [48] M. Fasolino, S. Liu, Y. Wang, and Z. Zhou, “Distinct cellular and molecular environments support aging-related DNA methylation changes in the substantia nigra,” *Epigenomics*, vol. 9, no. 1, pp. 21–31, 2017.
- [49] E. Palomer, A. Martín-Segura, S. Baliyan et al., “Aging triggers a repressive chromatin state at *Bdnf* promoters in hippocampal neurons,” *Cell Reports*, vol. 16, no. 11, pp. 2889–2900, 2016.
- [50] B. C. Christensen, E. A. Houseman, C. J. Marsit et al., “Aging and environmental exposures Aater tissue-specific DNA methylation dependent upon CpG island context,” *PLOS Genetics*, vol. 5, no. 8, article e1000602, 2009.
- [51] F. Ciccarone, S. Tagliatesta, P. Caiafa, and M. Zampieri, “DNA methylation dynamics in aging: how far are we from understanding the mechanisms?,” *Mechanisms of Ageing and Development*, vol. 174, pp. 3–17, 2018.
- [52] M. Jung and G. P. Pfeifer, “Aging and DNA methylation,” *BMC Biology*, vol. 13, no. 1, 2015.
- [53] S. Pal and J. K. Tyler, “Epigenetics and aging,” *Science Advances*, vol. 2, no. 7, article e1600584, 2016.
- [54] M. Zampieri, F. Ciccarone, R. Calabrese, C. Franceschi, A. Bürkle, and P. Caiafa, “Reconfiguration of DNA methylation in aging,” *Mechanisms of Ageing and Development*, vol. 151, pp. 60–70, 2015.
- [55] Z. Tong, C. Han, M. Qiang et al., “Age-related formaldehyde interferes with DNA methyltransferase function, causing memory loss in Alzheimer’s disease,” *Neurobiology of Aging*, vol. 36, no. 1, pp. 100–110, 2015.
- [56] L. Liu, T. van Groen, I. Kadish et al., “Insufficient DNA methylation affects healthy aging and promotes age-related health problems,” *Clinical Epigenetics*, vol. 2, no. 2, pp. 349–360, 2011.
- [57] P. Jessop and M. Toledo-Rodriguez, “Hippocampal TET1 and TET2 expression and DNA hydroxymethylation are affected by physical exercise in aged mice,” *Frontiers in Cell and Developmental Biology*, vol. 6, p. 45, 2018.
- [58] G. Gontier, M. Iyer, J. M. Shea et al., “Tet2 rescues age-related regenerative decline and enhances cognitive function in the adult mouse brain,” *Cell Reports*, vol. 22, no. 8, pp. 1974–1981, 2018.
- [59] D. Balmer, J. Goldstine, Y. M. Rao, and J. M. LaSalle, “Elevated methyl-CpG-binding protein 2 expression is acquired during postnatal human brain development and is correlated with alternative polyadenylation,” *Journal of Molecular Medicine*, vol. 81, no. 1, pp. 61–68, 2003.
- [60] R. C. Samaco, R. P. Nagarajan, D. Braunschweig, and J. M. LaSalle, “Multiple pathways regulate MeCP2 expression in normal brain development and exhibit defects in autism-spectrum disorders,” *Human Molecular Genetics*, vol. 13, no. 6, pp. 629–639, 2004.
- [61] S. A. S. Langie, K. M. Cameron, G. Ficiz et al., “The ageing brain: effects on DNA repair and DNA methylation in mice,” *Genes*, vol. 8, no. 2, p. 75, 2017.
- [62] A. L. Collins, J. M. Levenson, A. P. Vilaythong et al., “Mild overexpression of MeCP2 causes a progressive neurological disorder in mice,” *Human Molecular Genetics*, vol. 13, no. 21, pp. 2679–2689, 2004.
- [63] R. Bijkerk, C. Trimpert, C. van Solingen et al., “MicroRNA-132 controls water homeostasis through regulating MECP2-mediated vasopressin synthesis,” *American Journal of Physiology-Renal Physiology*, vol. 315, no. 4, pp. F1129–F1138, 2018.
- [64] R. F. Torres, C. Hidalgo, and B. Kerr, “*Mecp2* mediates experience-dependent transcriptional upregulation of ryanodine receptor type-3,” *Frontiers in Molecular Neuroscience*, vol. 10, p. 188, 2017.
- [65] C. O. Olson, R. M. Zachariah, C. D. Ezeonwuka, V. R. B. Liyanage, and M. Rastegar, “Brain region-specific expression of MeCP2 isoforms correlates with DNA methylation within *Mecp2* regulatory elements,” *PLoS One*, vol. 9, no. 3, article e90645, 2014.
- [66] J. Fernandes, R. M. Arida, and F. Gomez-Pinilla, “Physical exercise as an epigenetic modulator of brain plasticity and cognition,” *Neuroscience and Biobehavioral Reviews*, vol. 80, pp. 443–456, 2017.
- [67] S. Fusco, C. Ripoli, M. V. Podda et al., “A role for neuronal cAMP responsive-element binding (CREB)-1 in brain responses to calorie restriction,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 109, no. 2, pp. 621–626, 2012.
- [68] J. Nithianantharajah and A. J. Hannan, “Enriched environments, experience-dependent plasticity and disorders of the nervous system,” *Nature Reviews Neuroscience*, vol. 7, no. 9, pp. 697–709, 2006.
- [69] S. Maegawa, Y. Lu, T. Tahara et al., “Caloric restriction delays age-related methylation drift,” *Nature Communications*, vol. 8, no. 1, p. 539, 2017.
- [70] M. Scali, L. Baroncelli, M. C. Cenni, A. Sale, and L. Maffei, “A rich environmental experience reactivates visual cortex plasticity in aged rats,” *Experimental Gerontology*, vol. 47, no. 4, pp. 337–341, 2012.

- [71] L. R. Stein, K. A. O'Dell, M. Funatsu, C. F. Zorumski, and Y. Izumi, "Short-term environmental enrichment enhances synaptic plasticity in hippocampal slices from aged rats," *Neuroscience*, vol. 329, pp. 294–305, 2016.
- [72] A. M. Stranahan, K. Lee, K. G. Becker et al., "Hippocampal gene expression patterns underlying the enhancement of memory by running in aged mice," *Neurobiology of Aging*, vol. 31, no. 11, pp. 1937–1949, 2010.
- [73] G. P. Cortese, A. Olin, K. O'Riordan, R. Hullinger, and C. Burger, "Environmental enrichment improves hippocampal function in aged rats by enhancing learning and memory, LTP, and mGluR5-Homer1c activity," *Neurobiology of Aging*, vol. 63, pp. 1–11, 2018.
- [74] P. Obiang, E. Maubert, I. Bardou et al., "Enriched housing reverses age-associated impairment of cognitive functions and tPA-dependent maturation of BDNF," *Neurobiology of Learning and Memory*, vol. 96, no. 2, pp. 121–129, 2011.
- [75] L. Wang, M. Cao, T. Pu, H. Huang, C. Marshall, and M. Xiao, "Enriched physical environment attenuates spatial and social memory impairments of aged socially isolated mice," *The International Journal of Neuropsychopharmacology*, vol. 21, no. 12, pp. 1114–1127, 2018.
- [76] P. R. Patrylo and A. Williamson, "The effects of aging on dentate circuitry and function," *Progress in Brain Research*, vol. 163, pp. 679–696, 2007.
- [77] Z. A. Monge and D. J. Madden, "Linking cognitive and visual perceptual decline in healthy aging: the information degradation hypothesis," *Neuroscience and Biobehavioral Reviews*, vol. 69, pp. 166–173, 2016.
- [78] K. L. Roberts and H. A. Allen, "Perception and cognition in the ageing brain: a brief review of the short- and long-term links between perceptual and cognitive decline," *Frontiers in Aging Neuroscience*, vol. 8, 2016.
- [79] Y. Li, L. Liu, and T. O. Tollefsbol, "Glucose restriction can extend normal cell lifespan and impair precancerous cell growth through epigenetic control of hTERT and p16 expression," *The FASEB Journal*, vol. 24, no. 5, pp. 1442–1453, 2010.
- [80] D. Tropea, N. Mortimer, S. Bellini et al., "Expression of nuclear methyl-CpG binding protein 2 (Mecp2) is dependent on neuronal stimulation and application of insulin-like growth factor 1," *Neuroscience Letters*, vol. 621, pp. 111–116, 2016.
- [81] J. L. Abel and E. F. Rissman, "Running-induced epigenetic and gene expression changes in the adolescent brain," *International Journal of Developmental Neuroscience*, vol. 31, no. 6, pp. 382–390, 2013.
- [82] S. Dzitoyeva, H. Chen, and H. Manev, "Effect of aging on 5-hydroxymethylcytosine in brain mitochondria," *Neurobiology of Aging*, vol. 33, no. 12, pp. 2881–2891, 2012.



Hindawi

Submit your manuscripts at
www.hindawi.com

