Sustained delivery of nicotinamide limits cortical injury and improves functional recovery following traumatic brain injury

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Introduction

Traumatic brain injury (TBI) is a major public health crisis. An estimated 1.4 million people incur a TBI annually; a rate of 1 injury every 23 seconds.1 The resulting incidence rate is approximately 100 per 100,000 US citizens and is respectively 8 and 34 times higher than breast cancer and HIV/AIDS diagnoses each year.2 Military personnel, particularly those serving in Afghanistan and Iraq are a special population highly susceptible to TBI as blasts are the primary cause of injury for this group.3 Sports related head injuries are also becoming more common and it is estimated that 300,000 such injuries occur annually.4 Despite these staggering statistics, no clinically effective pharmacological treatments exist to aid recovery during the acute or secondary stages of injury.4-6

Previously, we have demonstrated that nicotinamide (NAM), a neuroprotective soluble B-group vitamin, improves recovery of function following traumatic brain injury (TBI). However, no prior studies have examined whether NAM is beneficial following continuous infusions over 7 days post-TBI. The purpose of this study was to investigate the preclinical efficacy of NAM treatment as it might be delivered clinically; over several days by slow infusion. Rats were prepared with either unilateral controlled cortical impact (CCI) injuries or sham procedures and divided into three groups: CCI-NAM, CCI-vehicle and sham. Thirty minutes following CCI, Alzet osmotic mini-pumps were implanted subcutaneously. NAM was delivered at a rate of 50 mg/kg/day for 7 days immediately post-CCI. On day 7 following injury, the pumps were removed and blood draws were collected for serum NAM and nicotinamide adenine dinucleotide (NAD+) analyses. Starting on day 2 post-CCI, animals were tested on a battery of sensorimotor tests (bilateral tactile adhesive removal, locomotor placing and limb-use asymmetry). Continuous infusion of NAM resulted in a significant serum elevation in NAM, but not NAD+. Statistical analyses of the tactile removal and locomotor placing data revealed that continuous administration of NAM significantly reduced the initial magnitude of the injury deficit and improved overall recovery compared to the vehicle-treated animals. NAM treatment also significantly decreased limb-use asymmetries compared to vehicle-treated animals. The overall extent of the cortical damage was also reduced by NAM treatment. No detrimental effects were seen following continuous infusion. The present results suggest that NAM delivered via a clinically relevant therapeutic regimen may truncate behavioral damage following TBI. Thus our results offer strong support for translation into the clinical population.

Key Abbreviations: TBI, traumatic brain injury; CCI, cortical contusion injury; NAM, nicotinamide; MWM, morris water maze; PARP, poly-ADP-ribose polymerase; NAD+, nicotinamide adenine dinucleotide; BBB, blood-brain barrier; ATP, adenosine triphosphate; FPI, fluid percussion injury; ANOVA, analysis of variance; ad lib, ad libitum

promising, but then do not translate clinically. It has been stated that there is a gap between the bench to the bedside.5 In other words, inconsistencies exist between preclinical testing in the laboratory and the clinic. The gaps most relevant to experimentalists are: treatment window and duration.4 For animal models the treatment window (in which the pharmacotherapy begins) is approximately less than an hour while clinically it is usually four to eight hours; while treatment durations can be over several days. Surprisingly few studies have examined continuous dosing regimens in TBI. Subcutaneous osmotic mini-pumps were used in order to deliver the immunosuppressant cyclosporine A (CsA) following cortical contusion injury (CCI).7 In a second study, a single bolus intraperitoneal (i.p.) injection, as a loading dose, was administered 15 mins post-CCI followed by chronic infusion of CsA. In another study, nicotine was infused continuously following TBI and it was shown to attenuate nicotinic receptor deficits

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following injury. In addition, continuous delivery of progesterone by silastic capsules following TBI has been shown to reduce anxiety. All of these studies demonstrated significant treatment effects; however, few studied the effect on recovery of function following injury. These studies used subcutaneous infusions as a clinically relevant route of administration since continuous intravenous infusion in rodents is problematic. Therefore, these studies could not conclude whether continuous infusions result in prolonged and enhanced recovery of function following TBI.

A promising preclinical treatment for TBI is nicotinamide (NAM), a neuroprotective soluble B-group vitamin. The administration of an i.p. dose of NAM (500 mg/kg) administered 15 minutes and 24 hours post-TBI, resulted in significant behavioral and anatomical recovery following bilateral frontal injury. Using the same dosing regimen, NAM was able to prevent blood-brain barrier (BBB) breakdown and reduce apoptosis at 5, 24 and 72 hours post injury following unilateral CCI. In a model of fluid percussion injury (FPI), 2 doses of 50 or 500 mg/kg NAM administered within 24 hrs of injury, induced sensorimotor and cognitive recovery up to thirty-five days post injury and thus need to be examined preclinically. Since continuous i.v. infusion into awake, behaving rodents is problematic at best, the alternative is the use of subcutaneous osmotic mini-pumps. Therefore, the main purpose of this study was to investigate the preclinical efficacy of NAM treatment at a targeted delivery rate of 0.625 mg/hr (to achieve 50 mg/kg), as it would be delivered clinically; over several days by slow, continuous infusion.

**Results**

NAM and NAD+ analysis. Two separate one-way ANOVAs were conducted to analyze NAM and NAD+ blood serum levels (µg/mL). Three levels were used in this analysis (NAM-treated, vehicle-treated and sham). There was a statistically significant difference between the three groups in terms of NAM blood levels, F(2,20) = 3.81, p = 0.04 (see Fig. 1). Pairwise comparisons revealed significant differences in serum NAM levels [LSD(15) = 5.00, p = 0.02] between the NAM (M = 16.63, SEM = 2.24) and vehicle-treated groups (M = 11.62, SEM = 0.62). Therefore, injured animals treated with chronic infusions of NAM had higher serum NAM levels than the injured animals treated with vehicle. The NAD blood serum level analysis revealed no overall significant differences between groups F(2,20) = 0.46, p = 0.64; however, there was a differences between the NAM (M = 9.63, SEM = 3.82) and vehicle-treated groups (M = 6.10, SEM = 1.60 (see Fig. 1).

**Bilateral tactile removal.** Latency to remove the tactile stimuli (adhesive patches) was analyzed in a 3 x 6 mixed ANOVA. The factors were group (NAM-treated, vehicle-treated and sham) x day (1–6). Latency to remove the contralateral patch was not significantly affected by days F(4.44,88.73) = 1.61, p = 0.17. The group by day interaction was also not statistically significant F(8.87,88.73) = 0.96, p = 0.48. However, the main effect of group was statistically significant, F(2,20) = 4.55, p = 0.02. The chronic infusion of NAM significantly reduced the removal latencies in lesioned animals (see Fig. 2). Fischer’s LSD pairwise comparisons revealed significant differences in removal latencies between the NAM-treated group and the vehicle-treated group on days 2 (LSD(15) = 44.50, p = 0.006), 4 (LSD(15) = 32.63, p = 0.04) and 6 (LSD(15) = 36.44, p = 0.01). No significant injury-induced impairments in patch removal were seen on the left forelimb p > 0.05.

**Locomotor placing.** Foot-faults on the grid floor were analyzed in a 3 x 6 mixed ANOVA. The factors were group (NAM-treated, vehicle-treated and sham) x day (1–6). The number of foot-faults was significantly affected by days F(3.76, 75.11) = 3.22, p = 0.001 and group, F(2, 20) = 63.17, p = 0.001 (see Fig. 3). However, the interaction was not statistically significant F(7.51, 75.11) = 0.57, p = 0.79. Pairwise comparisons revealed significant differences in foot-faults between the NAM-treated group and the vehicle-treated group on days 2 (LSD(15) = 44.50, p = 0.006), 4 (LSD(15) = 32.63, p = 0.04) and 6 (LSD(15) = 36.44, p = 0.01). No significant injury-induced impairments in patch removal were seen on the left forelimb p > 0.05.

**Limb-use asymmetry.** The percentage of bias towards use of the contralateral forelimb was analyzed in a 3 x 3 mixed
ANOVA. The factors were group (NAM-treated, vehicle-treated and sham) x day (1–3). The percentage of contralateral forelimb bias was not statistically significantly affected by day, $F(2,40) = 0.75, p = 0.48$. The interaction between day and group was also not statistically significant, $F(4,20) = 1.04, p = 0.40$. However, the group factor was statistically significant $F(2,20) = 32.90, p = 0.001$ (see Fig. 4). Pairwise comparisons revealed significant differences in contralateral biases between the NAM-treated and vehicle-treated groups on day 4 [LSD(15) = 11.32, $p = 0.04$], 15 [LSD(15) = 15.78, $p = 0.007$] and 28 [LSD(15) = 11.33, $p = 0.05$]. Therefore, chronic administration of NAM significantly reduced the bias towards the contralateral forelimb.

Lesion analysis. An initial one-way ANOVA was performed on the volume of the intact contralateral cortex. Three groups were used in this analysis (NAM-treated, vehicle-treated and sham). There was not a significant difference in contralateral cortical volume between the groups, $F(2,20) = 2.36, p = 0.12$. A second, one-way ANOVA was used to analyze the percentage of cortex volume reduction in the contralateral hemisphere compared to the ipsilateral hemisphere. There was a significant lesion effect between the groups, $F(2,20) = 80.84, p = 0.001$ (see Fig. 5). Pairwise comparisons revealed significant differences in cortex reduction (LSD(15) = 6.38, $p = 0.01$) between the NAM (M = 21.91, SEM = 1.45) and vehicle-treated groups (M = 28.29, SEM = 2.33). Therefore, lesioned animals treated with chronic infusions of NAM had significantly lower percentage of reduction in the contralateral cortex than the lesioned animals treated with vehicle. Representative histology sections are shown in Figure 6.

Discussion

The purpose of this study was to examine the preclinical efficacy of continuous treatment with NAM following TBI. The preclinical efficacy of NAM following TBI has been well established for bolus administrations; however, it is of clinical interest to determine if sustained, continuous infusion of NAM also demonstrates preclinical efficacy.

The sensorimotor testing on the bilateral tactile adhesive removal test following unilateral CCI of the SMC resulted in strong initial injury impairments in the vehicle-treated animals. In contrast, continuous infusion of NAM resulted in a significant reduction in the initial magnitude of the behavioral deficit and facilitation in recovery of function. On the locomotor placing test, unilateral CCI produced enduring deficits in the vehicle-treated animals. Infusion of NAM significantly reduced the magnitude of the initial placing deficit; in fact, NAM-treated animals were significantly less impaired on every test day compared to the vehicle-treated animals. On the forelimb asymmetry test CCI produced very enduring deficits in the vehicle-treated animals. On all test days (4, 15 and 28 days) post-injury the vehicle-treated animals showed a forelimb bias favoring the unimpaired forelimb that was greater than 80%. In contrast, the sham controls showed virtually no bias and used both the ipsilateral and contralateral forelimbs equally. The animals treated with infusion of NAM demonstrated a significant reduction in their bias toward the unimpaired limb on each test day compared to

Figure 2. Continuous infusion of nicotinamide (NAM) significantly improved recovery on the bilateral adhesive removal test following cortical contusion injury (CCI). The graph shows the plotted mean (+SEM) of the tactile stimuli removal latencies for all three experimental groups (NAM-treated, vehicle-treated and sham) following unilateral CCI. Seven days of continuous infusion of NAM significantly improved performance compared to vehicle (*$p < 0.05$). There were no statistically significant differences between the NAM-treated and sham animals.

Figure 3. Continuous infusion of nicotinamide (NAM) significantly improved recovery on the locomotor placing test following cortical contusion injury (CCI). The graph shows the plotted mean (+SEM) of the percentage of foot-faults on the grid for all three experimental groups (NAM-treated, vehicle-treated, and sham) following unilateral CCI. Seven days of continuous infusion of NAM significantly improved locomotor placing performance compared to vehicle (*$p < 0.05$). Although, a significant treatment effect was seen with NAM compared to vehicle in the injured animals.
the vehicle-treated animals. Together, all of the behavioral data in this study suggest that a 7-day regimen of low dose NAM significantly improved recovery of function following TBI.

The behavioral data as a whole in this study replicate and extended the beneficial effects we have seen with bolus administrations of NAM following CCI and fluid percussion injury (FPI) on a multitude of behavioral measures, including motor, sensorimotor and cognitive tests. In the present study direct comparison between bolus administrations of NAM and continuous administration of NAM were not performed, so experimentally, it cannot be determined if the continuous infusions were better than the bolus route. However, some speculations can be made. The current continuous infusion findings are consistent with a previous study in which bolus administrations of NAM (50 mg/kg) were performed daily for 5 days post-CCI. On the bilateral tactile removal test, animals receiving injections at 15 minutes or 4 and 8 hours post-CCI showed recovery over time, with the best recovery shown in the 15 minute group. However, in the current study, the NAM-treated group demonstrated a greater degree of initial injury reduction on day 2 post-CCI. A strong initial reduction in magnitude of the tactile removal deficit has also been shown following 12 days of bolus injections of 50 mg/kg NAM. However, in that paper animals did not start to perform at sham control levels until day 14 post-CCI; whereas, in the current study NAM-treated animals were performing at sham levels on day 2. A similar effect was also seen on the loco-motor placing test. Bolus injections of NAM administered for 12 day starting 4 hours post-CCI resulted in the animals performing at sham control levels. When treatment was initiated 8 and 24 hours after injury, the animals did not perform at sham levels, but were lower than vehicle-treated animals. In the present study, animals treated with NAM favored the use of their contralateral forelimb significantly less than vehicle-treated animals. A similar finding showing that NAM treatment when started at 4 or 8 hours post-CCI demonstrated a significant reduction in contralateral forelimb bias has been reported. Thus, the results from the current study continue to support the preclinical efficacy of NAM for TBI. However, in our experience the beneficial behavioral effects we saw in the current study are some of the strongest we have observed in these animals models.

The data from the lesion analysis also revealed a significant improvement in tissue integrity. Infusion of NAM significantly reduced the size of the injury, as measured by a cortical volume analysis. The vehicle-treated group had an average reduction in frontal cortical volume of 28.3% compared to the NAM-treated group that showed a reduction of 21.9%. This cortical protective effect is similar to those effects we have seen in previous studies. Furthermore, previous studies in our laboratory have shown NAM to significantly reduce apoptosis, GFAP expression, edema and neuronal death. Thus, it is likely that by reducing the pathophysiological events associated with CCI in previous studies that NAM resulted in the beneficial behavioral effects observed in the current study.

We have previously demonstrated that systemic administration of NAM elevated serum NAM concentrations. The data from the current study confirmed that sustained, low-level delivery of

![Figure 4. Continuous infusion of nicotinamide (NAM) significantly improved recovery on the forelimb asymmetry test following cortical contusion injury (CCI). The graph shows the plotted mean (+SEM) of the percent forelimb bias for all three experimental groups (NAM-treated, vehicle-treated and sham) following unilateral CCI. The no bias line indicates the point at which the ipsilateral and contralateral forelimbs are being used equally. Seven days of continuous infusion of NAM significantly improved performance compared to vehicle. Although, a significant treatment effect was seen with NAM compared to vehicle in the injured animals, there were still enduring deficits compared to the sham controls.](image1)

![Figure 5. Continuous infusion of nicotinamide (NAM) significantly limited cortical injury following cortical contusion injury (CCI). Plotted are the mean (+SEM) percentages of cortical volume reductions in the NAM-treated animals, vehicle-treated animals, and sham animals. NAM significantly reduced the extent of the cortical injury compared to vehicle (*p < 0.05).](image2)
Interestingly, there were lower levels of NAD⁺ at 7 days following injury in the vehicle-treated animals compared to the sham controls; furthermore, treatment with NAM appeared to have

NAM for 7 days also increased serum NAM levels significantly. Serum NAD⁺ levels were also analyzed and no such significant elevation was seen in the blood following NAM administration.

Figure 6. Representative histology showing that continuous infusion of nicotinamide (NAM) significantly improved recovery on the forelimb asymmetry test following cortical contusion injury (CCI). Unilateral CCI produced marked cortical cavitation. Shown are representative images of the effect of NAM infusion and vehicle treatment through the extent of the injury (0.44x) at coordinates +1.00, 0.00 and -1.00 mm relative to bregma. Measurement bar = 2.0 mm.
Our ability to see significant changes in serum NAD⁺ levels is partially offset this reduction. One factor that may have limited ways include forkhead transcription factors and the sirtuins.¹⁹,²⁰ NAD⁺ levels as well.²⁴,²⁶ NAM also has a role as an essential precursor for the coenzyme NAD⁺.²⁷ NAM is able to maintain levels of PARP thereby restoring ATP levels and to elevate NAD⁺ thereby restoring ATP levels and to serve as a PARP inhibitor, also restoring ATP.²² Using NAM to elevate NAD⁺ levels in conjunction with PARP inhibition are additive in respect to ATP restoration and has proven to be more beneficial.²⁹ Additionally, NAM is critical for synthesizing nicotinamide adenine dinucleotide phosphate (NADP⁺), modulating cellular metabolism, plasticity, longevity, inflammatory microglial responses, and maintenance of the internal and external cellular environment.²⁷,³⁰ Recently, new cellular pathways have emerged that appear to govern the beneficial actions of NAM in disease conditions and have recently been reviewed.²⁰ These pathways include forkhead transcription factors and the sirtuins.¹⁹,²⁰ NAM also activates protein kinase B and inhibits transcription of proapoptotic genes. A downstream effect of protein kinase B is closing of the mitochondrial transition pores opened by excessive Ca²⁺ influx. The mitochondrion is able to regain ionic homeostasis and bring cytochrome C back to physiological conditions thereby inhibiting caspase 9 mediated cell death.²⁰,²⁷

In our experience with NAM therapy for experimental TBI we have not observed any detrimental effects of treatment at 50 or 500 mg/kg.²⁰,²⁶,²⁷ These studies have included daily dosing regimens of NAM (50 mg/kg) for 7 and 14 days.¹³,¹⁴ In the current study, continuous infusion of NAM did not result in any overtly observable detrimental effects. Currently, NAM is used clinically in the treatment of pellagra (nicotinamide deficiency) and diabetes. Doses less than 600 mg/kg have been used daily for up to several weeks in studies using rats without complications, but the at higher doses liver and kidney damage have been reported.²⁶ For treatment of various diseases in the human population, daily doses of 1 to 12 grams have been used.³¹ Drug safety tests indicate that daily doses up to 3 grams can be used long-term safely and without side effects.³¹ However, a recent reports suggests that there is some concern that NAM therapy may be a double-edged sword when treating oxidative or metabolic conditions, especially in the aged populations.¹⁹,²⁰ Current studies in our laboratory are examining the ability of NAM to be neuroprotective in aged animals.

In conclusion, the current study showed that unilateral SMC injury produced relatively enduring behavioral deficits on a variety of forelimb sensorimotor tasks. Sustained delivery of NAM throughout the first 7 days post-injury resulted in significant improvements in these behavioral deficits. Future studies will be aimed at maximizing NAM administration by providing sustained, steady-state levels of treatment with NAM post-TBI.

Methods

Animals. Twenty-three male Sprague-Dawley rats were used for this study and weighed an average of 369.60 g (SEM 7.42) at the time of injury and pump implantation. A single animal in the sham group died during recovery and was not replaced. The experimental procedures conducted within this study were reviewed and approved by the Institutional Animal Care and Use Committee and the study was conducted in a facility certified by the American Association for the Accreditation of Laboratory Animal Care. Animals were individually housed in standard cages and allowed ad lib access to food and water. A twelve hour light:dark cycle was used. Rats were handled daily for approximately one week before surgery. Behavioral testing took place during the light cycle.

CCI surgery. Surgical procedures were based on previous studies and performed under aseptic conditions. Animals were anesthetized with a combination of Isofluorane (2–4%) and oxygen (0.8 L/minute) and placed on a heated surgical stage (37°C) in a stereotaxic frame. A unilateral CCI over the left sensorimotor cortex or sham procedure was then performed. A midline incision was made through the skin and underlying fascia. A craniotomy measuring 5.0 mm in diameter was created using a motor tool. A specially designed drill bit was used to prevent damage to the meninges and cortex. The craniotomy was positioned at Ap = 0.0, ML = +2.5. The cortical region containing the sensorimotor cortex (forelimb region) was then exposed. A sterile, stainless steel impactor tip with a 4.0 mm diameter was used to create the injury. The tip was attached to an electromagnetic impactor (www.myneurolab.com) and situated at a 10 degree angle. The impactor measuring 5.0 mm in diameter was created using a motor tool. A specially designed drill bit was used to prevent damage to the meninges and cortex. The craniotomy was positioned at Ap = 0.0, ML = +2.5. The cortical region containing the sensorimotor cortex (forelimb region) was then exposed. A sterile, stainless steel impactor tip with a 4.0 mm diameter was used to create the injury. The tip was attached to an electromagnetic impactor (www.myneurolab.com) and situated at a 10 degree angle. The tip made impact with the cortex at a velocity of 2.75 m/s for 0.5 seconds. A 2.0 mm compression of the cortex occurred. After injury, bleeding was controlled and incisions were sutured closed. Afterwards, animals were placed in a heated recovery cage until locomotion returned. Sham procedures were the same, except cortical impact did not occur.

Drug administration. Twelve hours prior to surgery, Alzet osmotic mini-pumps (model 2001; delivery rate of 1 µL per hour for 7 days) were primed and loaded with a concentration of NAM (625 mg of NAM dissolved per 1 mL of 0.9% PBS) or 0.9% PBS. The pumps were then incubated overnight. Approximately thirty minutes after CCI, the osmotic mini-
pumps were implanted (subcutaneous, s.c.). Briefly, under gas anesthesia an incision was made just posterior to the shoulder blades. After the incision was made, the underlying fascia and connective tissue was separated from the skin and the osmotic mini-pump was placed within the “pocket.” Afterwards, the incision was sutured closed. The osmotic mini-pumps delivered the NAM or vehicle at a rate of 50 mg/kg per day for seven days following CCI, based on the weight of the animal at the time of injury. On day seven the animals were again anesthetized and the pumps were removed and blood was collected from the tail vein for NAM and NADþ serum analysis. There were three groups. Group 1 received CCI and was treated with an osmotic mini-pump containing NAM (NAM-treated; n = 8). Group 2 also received CCI and was treated with an osmotic mini-pump containing the vehicle solution (0.9% PBS; vehicle-treated; n = 8). Group 3 received sham procedures and an osmotic mini-pump containing vehicle solution (sham; n = 7). See Table 1.

**Bilateral tactile removal.** In order to test somatosensory function this test was administered on pre-injury day 2 and post-CCI days 2, 4, 6, 8, 14 and 28.13,14 Small adhesive patches (approximately 113.0 mm²) were applied to the radial aspects of each forelimb. The animal was then returned to their home cage. The latency for removal of the patch was recorded for each forelimb and was recorded with a stopwatch. A trial ended after both patches were completely removed or 120 seconds elapsed. Animals received two trials per test day. The latency to remove the adhesive patch from the contralateral forelimb was the dependent variable of interest.

**Locomotor placing.** In order to assess recovery of coordinated locomotor forelimb movement, this test was administered on pre-injury day 2 and post-CCI days 2, 4, 6, 8, 14 and 28.13,14 On each test day, rats were placed on an elevated grid floor (56.0 x 54.0 cm) and allowed to explore for three minutes. There were openings measuring 3.5 cm² in size on the grid floor. A “foot-fault” occurred when a rat inaccurately placed the forelimb through these openings. Rats were administered one trial per test day. The number of contralateral and ipsilateral forelimb “foot-faults” was the dependent variable. The following equation was used to calculate the percentage of contralateral foot-faults: [(Contralateral foot-faults-ipsilateral foot-faults)/number of steps during the trial]×100.

**Ipsilateral paw placements.** To assess recovery of impaired (contralateral) forelimb use, this test was administered on post-CCI days 4, 15 and 20.13,14 Rats were placed in a clear observation chamber (20 cm x 20 cm x 30 cm) with four vertical surfaces. Animals were given a single five minute trial on each test day and allowed to place their paws on the vertical surface. The number of ipsilateral and contralateral paw placements on the vertical surface was the dependent variable. The following equation was used to calculate the percent forelimb bias: [(ipsilateral paw placements + (both paw placements at once/2))/[(ipsilateral + contralateral + both paw placements)]×100].

**NAM and NADþ analysis.** These methods were based on previous studies.22–23 Immediately following removal of the pumps, blood samples were collected from the animals’ tail vein. The samples were separated with a microcentrifuge and stored frozen at -80°C. NAM was analyzed in serum using HPLC with UV detection at 254 nm on a Varian Pro Star 210 HPLC system using a modification of a previously developed assay used to measure NADþ.24 Briefly, 60 μL serum samples and the added internal standard, nicotinamide hypoxanthine dinucleotide sodium salt (de-amino NAD), were deproteinized with 1 M perchloric acid, buffered with 1 M potassium phosphate pH 7.5, and centrifuged. The supernatant was injected onto an Adsorbsil C-18, 5 um, 250 x 4.6 mm column (Grace Davison Discovery Sciences, Deerfield, IL) and separated using a mobile phase gradient of 6% to 20% methanol over 10 mins in 0.2 M ammonium phosphate at pH 5.25. The de-amino NAD eluted at 6.3 minutes and NAM eluted at 6.3 minutes and NAM eluted at 13.5 mins.

**Histology.** Twenty-eight days after CCI, animals were anesthetized with a lethal dose of urethane (3.0 g/kg, i.p.). Once an absence of the pedal reflex response occurred, animals followed by 10% phosphate buffered formalin (PBF). Once the brain was removed from the skull, they were post-fixed in PBF for 24 hours. Brains were then cryoprotected in a 30% sucrose solution for three days prior to frozen sectioning. Serial coronal sections (40 μm thick) were taken using a sliding microtome and a freezing stage. The sections were then collected into a cryopreservative solution, stored and then frozen.

**Lesion analysis.** The coronal sections were mounted on gelatin-subbed microscope slides and stained with cresyl violet to determine the extent of lesion cavity formation. An Olympus microscope (BX-51) with an Olympus 13.5 megapixel digital camera (DP-70) attached to it was used to capture images of four sections thoroughly traversing the lesion cavity (1.60, 0.70, -0.40, -1.60 relative to bregma).35 The Camvalieri method was used to calculate the volume of the ipsilateral and contralateral cortex.36 The number of sections analyzed and their respective thickness (40 μm) were multiplied by the mean area of the lesion cavity. The extent of cortical injury was calculated using the formula [100 - (ipsilateral cortex volume/contralateral cortex volume)]×100. This formula calculated the percent reduction in the ipsilateral cortex.

**Data analysis.** All behavioral testing and histological analyses were performed without knowledge of treatment parameters.

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**Table 1. Experimental groups and timeline**

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<tr>
<th>Groups</th>
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<tr>
<td>Group 2</td>
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<tr>
<td>Group 3</td>
<td>Sham + Vehicle</td>
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**Timeline**

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<td>4, 15, 20</td>
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<td>Perfusion</td>
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Means and standard error of the means (SEM) were calculated for all data. The General Linear Model (GLM) ANOVA procedure with the repeated measures option was used to evaluate the effects of chronic NAM treatment across days. The groups included in the analysis were NAM-treated, vehicle-treated and sham. The between groups factor was treatment group assignment and the within groups factor was day of testing. The Huynh-Feldt correction was used to correct for repeated measures. Fischer’s Least Significant Different (LSD) test was the pairwise comparison procedure used. One-way ANOVAs were calculated on the serum levels and anatomical data and were followed by appropriate post-hoc (LSD) analyses. Statistical significance was determined with a p value less than 0.05.

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