Research Article

Curcumin Induces Nrf2 Nuclear Translocation and Prevents Glomerular Hypertension, Hyperfiltration, Oxidant Stress, and the Decrease in Antioxidant Enzymes in 5/6 Nephrectomized Rats

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Renal injury resulting from renal ablation induced by 5/6 nephrectomy (5/6NX) is associated with oxidant stress, glomerular hypertension, hyperfiltration, and impaired Nrf2-Keap1 pathway. The purpose of this work was to know if the bifunctional antioxidant curcumin may induce nuclear translocation of Nrf2 and prevents 5/6NX-induced oxidant stress, renal injury, decrease in antioxidant enzymes, and glomerular hypertension and hyperfiltration. Four groups of rats were studied: (1) control, (2) 5/6NX, (3) 5/6NX+CUR, and (4) CUR (n = 8–10). Curcumin was given by gavage to NX5/6+CUR and CUR groups (60 mg/kg/day) starting seven days before surgery. Rats were studied 30 days after NX5/6 or sham surgery. Curcumin attenuated 5/6NX-induced proteinuria, systemic and glomerular hypertension, hyperfiltration, glomerular sclerosis, interstitial fibrosis, interstitial inflammation, and increase in plasma creatinine and blood urea nitrogen. This protective effect was associated with enhanced nuclear translocation of Nrf2 and with prevention of 5/6NX-induced oxidant stress and decrease in the activity of antioxidant enzymes. It is concluded that the protective effect of curcumin against 5/6NX-induced glomerular and systemic hypertension, hyperfiltration, renal dysfunction, and renal injury was associated with the nuclear translocation of Nrf2 and the prevention of both oxidant stress and the decrease of antioxidant enzymes.

1. Introduction

Curcumin is the major active component of turmeric, a yellow compound isolated from the plant Curcuma longa and has been used for centuries in traditional medicines [1]. Extensive research over the past 30 years has indicated that this molecule has therapeutic potential against a wide range of diseases, such as cancer, lung diseases, renal diseases, neurological diseases, liver diseases, metabolic diseases, cardiovascular diseases, and various other inflammatory diseases [1–3]. Numerous lines of evidence indicate that curcumin is highly pleiotropic with anti-inflammatory [4–6], hypoglycemic [7], antioxidant [8–11], wound healing [12] and antimicrobial activities [13]. Curcumin exerts both direct and indirect antioxidant effects by scavenging reactive oxygen species (ROS) [14, 15] and inducing the expression of cytoprotective proteins in an Nrf2-dependent way [16], respectively. It is considered a bifunctional antioxidant [17]. The nuclear-factor-erythroid-2-related factor 2 (Nrf2), is a cap “n” collar (CNC) basic region leucine zipper transcription protein and a ubiquitous master transcription factor which induces cytoprotective proteins trough binding
to antioxidant response elements (AREs) [18, 19]. Nrf2 is held in the cytoplasm, under basal conditions, as an inactive complex bound to a repressor molecule known as Kelch-like ECH-associated protein 1 (Keap1), which facilitates its ubiquitination. Keap1 is the redox-sensitive protein that on stimulus allows the activation of Nrf2. Nrf2 can be activated by diverse stimuli including oxidants, antioxidants, and chemopreventive agents. Keap1 contains several reactive cysteine residues that serve as sensors of intracellular redox state [20]. Oxidative or covalent modification of thiols in some of these cysteine residues results in dissociation of Nrf2 from Keap1 and its translocation to the nucleus. Previous work of our group has shown that curcumin indeed was able to induce in vivo translocation of Nrf2 to the nucleus which was associated with the renoprotection of this antioxidant against potassium-dichromate-induced nephrotoxicity in rats [8]. On the other hand, it has been shown that curcumin exerts protective effects against renal damage and inflammation induced by 5/6 nephrectomy (5/6NX) in rats [4, 21], a widely used model to study the progression of renal disease [22, 23]. This experimental model is characterized by proteinuria, hypertension, glomerular sclerosis, arteri- olopathy, renal inflammation and fibrosis, and alterations in renal hemodynamics including glomerular hypertension and hyperfiltration [22–25]. It has been shown that glomerular hypertension and hyperfiltration are involved in the progression of renal damage in 5/6NX rats [24–26]. Furthermore, it has been shown that oxidant stress, increase in nicotinamide adenine dinucleotide phosphate (NADPH) oxidase and alterations in antioxidant enzymes are involved in the pathogenesis of 5/6NX [27, 28]. More recently, Kim and Vaziri [28] have shown that the impaired Nrf2-Keap1 pathway contributes to oxidative stress and inflammation in 5/6NX rats. They showed that remnant kidney tissue Nrf2 activity (nuclear translocation) was reduced, whereas the Nrf2 repressor Keap1 was upregulated [28]. At present it is unknown if curcumin is able to ameliorate 5/6NX-induced glomerular hypertension and hyperfiltration and to modulate nuclear translocation of Nrf2 in this model. Based on the above information, the hypothesis was made that curcumin may enhance nuclear Nrf2 translocation which may decrease 5/6NX-induced oxidant stress, alterations in renal hemodynamics, and renal injury. The present study was designed to investigate if curcumin administration results in attenuation of renal hemodynamic alterations in rats with 5/6NX and if this may be associated to Nrf2 nuclear translocation and attenuation of both oxidant stress and decrease in antioxidant enzymes. Furthermore, it was explored if this experimental model is associated with mitochondrial alterations that may be involved in the pathophysiology of the disease as has been shown in other models of renal damage [8, 29–33].

2. Method

2.1. Chemicals. Curcumin, nitroblue tetrazolium (NBT), tetramethoxypropane, glutathione reductase, reduced glutathione (GSH), oxidized glutathione (GSSG), 1-chloro-2, 4-dinitrobenzene (CDNB), ethylenediaminetetraacetic acid (EDTA), Tris-HCl, 1-methyl-2-phenylindole, xanthine, xanthine oxidase, NADPH, aprotinin, leupeptin, pepstatin, N-[2-hydroxyethyl]piperazine-N’-[2-ethanesulfonic acid] (HEPES), butylated hydroxytoluene, bovine serum albumin, dimedone (5, 5-dimethyl-1, 3-cyclohexanedione), boric acid, Brij solution 30% (w/v), O-phenthalddehyde, 2-mercaptoethanol, ethanol, adenosine diphosphate (ADP), potassium succinate, rotenone, sodium glutamate, sodium malate, carbonyl cyanide m-chlorophenyl hydrazone (CCCP), Sudan Black, hematoxylin, eosin, and periodic acid Schiff were purchased from Sigma-Aldrich (St. Louis, MO, USA). Hydrogen peroxide (H2O2) and trichloroacetic acid (TCA) were purchased from J. T. Baker (Xalostoc, Edomex, México). Sodium pentobarbital was from Holland of México (Mexico City). Rabbit IgG anti-Nrf2 polyclonal antibody (Cat #sc722) was purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). Inutest (25% polyfructosan solution) was from Fresenius Kabi Austria GmbH (Linz, Austria). All other reagents and chemicals used were of the highest grade of purity commercially available.

2.2. Experimental Design. Male Wistar rats with an initial body weight of 280–300 g were used. All animal procedures were performed in accordance to the Mexican Federal Regulation for animal experimentation and care (NOM-062-ZOO-2001) and for the disposal of biological residues (NOM-087-ECOL-1995) and were approved by Bioethics and Investigation Committees of Instituto Nacional de Cardiología Ignacio Chávez (Approval No. 12-767). Under anesthesia with sodium pentobarbital, 60 mg/kg intraperitoneally, renal ablation was performed by removal of the right kidney and selective infarction of approximately two thirds of the left kidney by ligation of two or three branches of the renal artery. Sham operation consisted of ventral laparotomy and manipulation of the kidneys and renal pedicle without destruction of renal tissue [22]. Four groups of rats were studied and all animals were sacrificed 30 days after 5/6NX or laparotomy: (1) control rats (n = 9), treated daily via oral gavage with carboxymethylcellulose 0.05% 7 days before and 30 days after laparotomy; (2) 5/6NX-treated rats (5/6NX group, n = 9); (3) 5/6NX + curcumin (5/6NX+CUR group, n = 8); (4) curcumin-treated rats (CUR group, n = 10). Curcumin was dissolved in carboxymethylcellulose 0.05% and was given daily at a dose of 60 mg/kg/day via oral gavage for 7 days before and 30 days after 5/6NX (5/6NX+CUR group) or sham operation (CUR group). The dose of curcumin used (60 mg/kg/day) was chosen based on previous experiments in which lower doses of 15 and 30 mg/kg/day were ineffective to ameliorate proteinuria and hypertension in our rats. In addition, Kuhad et al. [34] observed the higher protection against cisplatin-induced nephrotoxicity with 60 mg of curcumin/kg/day. Systolic blood pressure (SBP) was measured in conscious restrained rats by tail-cuff plethysmography (XBP-1000 Kent Scientific, Connecticut, USA). Rats were preconditioned twice before SBP was measured at basal period, and every two weeks for the rest of the study as described in earlier studies [22]. In addition to tail-cuff plethysmography, blood pressure
was also determined by direct intra-arterial measurement through the experiment by a catheter placed in the femoral artery during the micropuncture experiments (see later). Twenty-four-hour urine collections were obtained placing the rats in metabolic cages at baseline and at every 2 weeks during the study, before the micropuncture experiments. Proteinuria and systolic blood pressures were measured in basal conditions and on days 15 and 30 and plasma creatinine and blood urea nitrogen (BUN) were measured on day 30.

2.3. Micropuncture Studies to Evaluate Renal Hemodynamics. Micropuncture studies were performed 30 days after the surgical procedure (5/6NX or sham) under sodium pentobarbital anesthesia (30 mg/kg body weight intraperitoneally) with supplementary doses as required. Micropuncture methodology has been previously described [22] briefly; the rats were placed on a temperature-regulated table, at 37°C. Polyethylene tubing was used to catheterize the trachea jugular veins, femoral arteries, and the left ureter. The left kidney was exposed, through a lumbar incision, placed in a Lucite holder, and sealed, covering the kidney surface with 0.9% saline solution. One femoral artery catheter was used for blood sampling and the other for monitoring mean arterial pressure (MAP) with a pressure transducer (Model p23 Db, Gould, Puerto Rico, USA) and recorded on a polygraph (Grass Instruments, Quincy, MA, USA). During the surgery, rats received an albumin (6%) infusion (1% of body weight), through a jugular catheter. Immediately after a bolus injection of 100 mg of polyfructosan, an infusion of 5% polyfructosan in Ringer solution was started at a rate of 2.2 mL/h. Sixty minutes were allowed for equilibration before the studies were done. Sampling blood was simultaneously replaced by an equal volume obtained from a normal donor rat. At the end of the experiment the kidneys were removed and weighed. Samples of proximal tubule fluid were obtained from seven different nephrons after inserting an oil block with a micropipette for determination of flow rate and polyfructosan concentration to calculate single-nephron glomerular filtration rate (SNGFR). Polyfructosan was measured in plasma and urine samples to calculate whole-kidney glomerular filtration rate (GFR). Using a continuous-recording servo-null micropipette transducer (Servo Nulling Pressure System, Instrumentation for Physiology and Medicine, Inc., California, CA, USA), intratubular hydrostatic pressure (FF), was measured in additional proximal tubules under free-flow conditions and after stopping tubular flow with an oil block (stop-flow pressure); hydrostatic pressure was also measured in peritubular capillaries. Colloid osmotic pressure (π) in glomerular capillaries was estimated from the protein concentration in blood taken from the femoral artery (Ca) and in blood obtained by puncturing surface efferent arterioles (Ce) [35]. The following variables were measured: MAP, stop-flow pressure (SFP), transcapillary hydraulic pressure (ΔP), hematocrit (Hct), free flow pressure (FFP), capillary pressure (Pc), and afferent (CA) and efferent (CE) protein concentration.

2.4. Analytical Procedures. Urea and creatinine were measured with an autoanalyzer (I lab 300 Plus Instrumentation Laboratory, Bedford, MA, USA). Proteinuria was measured by a turbidimetric method at 420 nm using 12.5% TCA and data were expressed as mg/24h [36]. Polyfructosan concentrations in plasma and urine were determined by the anthrone method [37]. The volume of fluid collected from individual proximal tubules was estimated from the length of the fluid column in a constant-bore capillary tube of known internal diameter. The concentration of polyfructosan in the tubular fluid was measured in triplicate by a microfluorometric method [38]. Protein concentrations were determined in efferent samples and femoral arterial blood plasma using a fluorometric method[39]. GFR, SNGFR, glomerular capillary pressure (P_{GC}), single-nephron plasma flow (Qa), single-nephron filtration fraction (SNFF), afferent (AR) an efferent (ER) resistances, afferent (πA) and efferent (πE) onctic pressure, and ultrafiltration coefficient (Kf) were calculated according by Baylis et al. [35].

2.5. Histological Analysis. Paraffin-embedded sections stained with trichrome and periodic acid-Schiff reagent (PAS) were examined in a blinded fashion. Tubulointerstitial fibrosis was evaluated in Masson’s trichrome sections. Thirty nonoverlapping fields of cortex (640 × 477 mM at 10X) per biopsy were analyzed by light microscopy (Olympus BX51, Olympus American, New York, USA) and captured with a digital video camera (Evolution VF, Media Cybernetics, Madison, USA). Positive blue-color areas (excluding glomeruli and vessels) were analyzed using Image-Pro-Plus 7.0 (Media Cybernetics). The extension of positive areas (30 microscopic fields per biopsy) was expressed as a fraction of the total tubulointerstitial area examined. Tubulointerstitial cellular infiltration was studied in PAS sections. The number of cells by field was quantified and expressed as positive cells per field. Glomerulosclerosis was evaluated in Masson’s trichrome stained sections. The degree of sclerosis was scored as follows: in each biopsy, the number of glomeruli with segmental, mesangial, and global sclerosis, as well as normal glomerular tufts, was assessed. Sclerosis was defined as a peripheral segmental or global increase in mesangial matrix with obliteration of capillary loops, far from the hilus. The resulting index in each animal was expressed as the percent of sclerosed glomeruli [24].

2.6. Nrf2 Immunohistochemistry. Sections (2 μm) were deparaffinized and rehydrated through submersion in graded alcohols (xylene: 1:1 xylene-alcohol, alcohol, and 70% alcohol for 10 min each, and finally rinsed in distilled water). Antigen retrieval was performed with 10 mM citrate buffer pH 6, for 5 min in a microwave oven. Samples were treated with H2O2 (3%) to quench endogenous peroxidase. Sections were incubated at 4°C overnight with a dilution 1:100 of anti-Nrf2 antibody. To detect the specific binding of the primary antibody, an immunocytochemical staining kit was used in which tissues were incubated sequentially with blocking serum, a specific secondary antibody and streptavidin/peroxidase complex. Finally, dianinobenzidine.
was used as chromogen. Sections were counterstained with haematoxylin. The percentage of cells with immunostained nuclei was determined and results were expressed as positive nuclei/field.

2.7. Lipid Peroxidation. Lipid peroxidation was assessed by measuring malondialdehyde (MDA), and 4-hydroxy-2-nonenal (4-HNE) in the kidney tissue using a standard curve of tetramethoxypropane. A solution of 1-methyl-2-phenyllindole in a mixture of acetonitrile/methanol (3:1) was added to the renal homogenates and the reaction was started by adding 37% HCl. Optical density was measured at 586 nm after 1 h of incubation at 45°C as previously described [8]. Data were expressed as nmol MDA and 4-HNE/mg protein.

2.8. Activity of Antioxidant Enzymes. Catalase (CAT) activity was assayed by a method based on the disappearance of 30 mM H2O2 at 240 nm [8] and the data were expressed as k/mg protein. GR activity was assayed using GSSG as substrate and measuring the disappearance of NADPH at 340 nm [8]. One unit of GR was defined as the amount of enzyme that oxidizes 1 μmol of NADPH/min. Data were expressed as U/mg protein. Glutathione peroxidase (GPx) activity was measured at 340 nm using GR and NADPH in a coupled reaction [8]. One unit of GPx was defined as the amount of enzyme that oxidizes 1 μmol of NADPH/min. Data were expressed as U/mg protein. Glutathione-S-transferase (GST) activity was assayed in a mixture containing GSH and CDNB as previously described [8]. One unit of GST was defined as the amount of enzyme that conjugates 1 μmol of CDNB with GSH per minute [8]. Data were expressed as U/mg protein. Total superoxide dismutase (SOD) activity in renal homogenates was assayed spectrophotometrically at 560 nm by a previously reported method using NBT as the indicator reagent [8]. The amount of protein that inhibited NBT reduction to 50% of maximum was defined as one unit of SOD activity. Results were expressed as U/mg protein.

2.9. Isolation of Renal Mitochondria. Mitochondria were isolated from separate groups of rats prepared exclusively for this purpose. Kidneys were removed from rats, washed, cleaned of fatty and conjunctive tissue, and placed in cold isolation buffer containing 250 mM sucrose, 10 mM Tris-HCl, 1 mM EDTA, and pH 7.3. Kidneys were minced in isolation buffer before being homogenized. Mitochondria were obtained by differential centrifugation as previously described [40].

2.10. Mitochondrial Oxygen Consumption. Mitochondrial oxygen consumption was measured using a Clark-type oxygen electrode (Yellow Springs Instruments, Yellow Spring, OH, USA). The experiments were carried out in 1.5 mL of basic medium containing 125 mM KCl, 10 mM HEPES, and 3 mM inorganic phosphate (Pi), pH 7.3. State 4 respiration was evaluated in the presence of 10 mM succinate plus 1 μg/mL rotenone, or with 10 mM sodium glutamate and 10 mM sodium maleate. State 3 respiration was stimulated by the addition of 200 μM ADP. Respiratory rates are expressed as ng-atoms oxygen/min/mg protein (ngAO/min/mg). Respiratory control index (RC) was calculated as the ratio state 3/state 4. Uncoupled respiration was measured by adding 1 μM CCCP; phosphorylation efficiency (ADP/O ratio) was calculated from the added amount of ADP and total amount of oxygen consumed during state 3 [8].

2.11. Statistical Analysis. Data are presented as mean ± standard error of the mean (SEM). Differences between groups were evaluated by two-way analysis of variance (ANOVA) followed by multiple comparisons according to Bonferroni using a commercially available statistical package (GraphPad Prism 4.0, California, USA). Two-tailed P < 0.05 was considered significant.

3. Results

3.1. Plasma Creatinine, BUN, Proteinuria, and Systolic Blood Pressure. Rats in all the experimental groups had comparable body weight, blood pressure, and urinary protein excretion in the baseline studies and had comparable body weights at the end of the experiment (data not shown). At the end of the study, it was found that curcumin attenuates proteinuria and the increase in plasma creatinine, blood urea nitrogen, and systolic blood pressure (Figure 1). BUN decreased from 100 mg/dL in the 5/6NX group to 30 mg/dL in the 5/6NX+CUR group. Plasma creatinine decreased from 1.7 mg/dL in the 5/6NX group to 1.0 mg/dL in the 5/6NX+CUR group. Rats with renal ablation developed severe systemic hypertension. Subtotal renal ablation was followed by the progressive increase in proteinuria. As shown in Figure 1(c), proteinuria in the 5/6NX+CUR group was reduced by approximately two-thirds (P < 0.05) and remained at essentially steady levels 30 days after 5/6NX. Systolic blood pressure rose from 120 ± 3 at day 0 to 178 ± 6 mm Hg at 30 days. Treatment with curcumin was associated with a less-pronounced increment in blood pressure (152 ± 4 mm Hg, Figure 1). In addition the increase in kidney weight (KW) in 5/6NX rats was not significant (Table 1) but curcumin decreased significantly KW about 25% in the 5/6NX+CUR group (Table 1).

3.2. Micropuncture Studies. Findings in the micropuncture studies 30 days after 5/6NX are shown in Figure 2. It was found that the increase in (a) MAP, (b) SNGFR, (c) Qa, (d) PGC, (e) SFP, and (f) ΔP observed in 5/6NX rats was ameliorated by curcumin (5/6NX+CUR group). Curcumin prevented glomerular hypertension (the increase in PGC) and hyperfiltration (the increase in SNGFR). In addition it was found that the decrease in (g) AR, (h) ER, and (i) GFR observed in 5/6NX rats was ameliorated by curcumin (5/6NX+CUR group). The most relevant changes observed in the 5/6NX+CUR group were the normalization of PGC and SNGFR resulting from a significant increase in the preglomerular tone evidenced by the rise in AR. No significant differences were observed in Hct, SNFF, FFP, Pc, Kf, CA, πA, CE, and πE (Table 1).
3.3. Histological Analysis. Figure 3 shows that 5/6NX rats developed glomerular sclerosis. Representative images are shown in the upper panel and quantitative data are shown in the lower panel. Curcumin treatment ameliorated significantly glomerular sclerosis from 25% in the 5/6NX groups to 12% in the 5/6NX+CUR group (P < 0.05). No sclerosis was observed in control and CUR groups (Figure 3).

Figure 4 shows that 5/6NX rats developed interstitial fibrosis. Representative images are shown in the upper panel and quantitative data are shown in the lower panel. Curcumin treatment ameliorated significantly interstitial fibrosis from 9% in the 5/6NX group to 4% in the 5/6NX+CUR group (P < 0.05). These values were lower than 1% in control and CUR groups (Figure 4).

Figure 5 shows that 5/6NX rats developed interstitial inflammation. Representative images are shown in the upper panel and quantitative data are shown in the lower panel. Curcumin treatment ameliorated significantly interstitial inflammation from 50 cells/field in the 5/6NX groups to 20 cells/field in the 5/6NX+CUR group (P < 0.05). These values were lower than 15 cells/field in control and CUR groups (Figure 5).
The increase was higher in the 5/6NX+CUR group (about 6 positive cells/field in control and CUR groups (Figure 6). It has been shown that oxidant stress plays a major role in animal models [27, 42–45]. In the 5/6NX model oxidant stress may be explained from an increase in ROS production and a decrease in the expression/activity of antioxidant enzymes [28, 46]. In a recent study, Kim and Vaziri [28] showed that the Nrf2–Keap1/ARE pathway is altered in rats with 5/6NX. The cytoplasmic concentration of Keap1 was increased and the nuclear Nrf2 translocation was decreased. These changes were accompanied by a decrease in the renal content of GSH and of the several enzymes dependent of Nrf2 as well as an increase in the NADPH oxidase expression [28]. In this context, the use of antioxidant compounds and of inducers of Nrf2 may be useful to attenuate the respiration (state 3, state 4, RC, uncoupled respiration, and ADP/O ratio using either malate/glutamate or succinate as substrates, Table 2). No significant alterations were found among all the studied groups (Table 2).

### 4. Discussion

It has been shown that oxidant stress plays a major role in the progression of renal damage in patients [41] and in animal models [27, 42–45]. In the 5/6NX model oxidant stress may be explained from an increase in ROS production and a decrease in the expression/activity of antioxidant enzymes [28, 46]. In a recent study, Kim and Vaziri [28] showed that the Nrf2–Keap1/ARE pathway is altered in rats with 5/6NX. The cytoplasmic concentration of Keap1 was increased and the nuclear Nrf2 translocation was decreased. These changes were accompanied by a decrease in the renal content of GSH and of the several enzymes dependent of Nrf2 as well as an increase in the NADPH oxidase expression [28]. In this context, the use of antioxidant compounds and of inducers of Nrf2 may be useful to attenuate the

### Table 1: Glomerular hemodynamic findings 30 days after 5/6 nephrectomy.

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>5/6NX</th>
<th>5/6NX + CUR</th>
<th>CUR</th>
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<tr>
<td>KW (g)</td>
<td>1.58 ± 0.05</td>
<td>1.72 ± 0.12</td>
<td>1.29 ± 0.05**</td>
<td>1.66 ± 0.08</td>
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<td>Hct (%)</td>
<td>0.48 ± 0.01</td>
<td>0.47 ± 0.01</td>
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<td>SNFF (%)</td>
<td>0.30 ± 0.02</td>
<td>0.30 ± 0.02</td>
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<td>FFP (mm Hg)</td>
<td>11.54 ± 0.93</td>
<td>11.02 ± 0.47</td>
<td>10.24 ± 1.49</td>
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<td>Pc (mm Hg)</td>
<td>11.68 ± 0.62</td>
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<tr>
<td>Kf (nl/sec/mm Hg)</td>
<td>0.0338 ± 0.0016</td>
<td>0.0296 ± 0.0008</td>
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<tr>
<td>Malate/glutamate</td>
<td>State 3 (ngAO/min/mg)</td>
<td>193.9 ± 19.2</td>
<td>197.3 ± 7.3</td>
<td>187.9 ± 12.8</td>
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<td></td>
<td>State 4 (ngAO/min/mg)</td>
<td>72.6 ± 8.7</td>
<td>77.7 ± 6.4</td>
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<td></td>
<td>RC</td>
<td>2.8 ± 0.2</td>
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<td></td>
<td>Uncoupled respiration (ngAO/min/mg)</td>
<td>233.1 ± 28.0</td>
<td>238.3 ± 22.7</td>
<td>225.3 ± 19.5</td>
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<td></td>
<td>ADP/O</td>
<td>2.2 ± 0.4</td>
<td>1.8 ± 0.1</td>
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<table>
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<tr>
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<th>5/6NX + CUR</th>
<th>CUR</th>
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<tr>
<td>Succinate</td>
<td>State 3 (ngAO/min/mg)</td>
<td>277.1 ± 31.3</td>
<td>276.1 ± 20.7</td>
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<td>State 4 (ngAO/min/mg)</td>
<td>136.4 ± 14.0</td>
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<td></td>
<td>RC</td>
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<td>Uncoupled respiration (ngAO/min/mg)</td>
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<td>340.7 ± 53.3</td>
<td>399.7 ± 26.8</td>
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<td>ADP/O</td>
<td>1.3 ± 0.1</td>
<td>1.2 ± 0.0</td>
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**Table 2: Effect of CUR and 5/6NX on mitochondrial oxygen consumption using malate/glutamate or succinate as substrates.**

**Table 3: Mitochondrial function.** Mitochondrial function was evaluated in all groups in order to explore the possible role of this organelle in the observed renal dysfunction induced by 5/6NX. On day 30, it was found that 5/6NX or curcumin was unable to induce alterations in renal mitochondrial function (state 3, state 4, RC, uncoupled respiration, and ADP/O ratio using either malate/glutamate or succinate as substrates, Table 2). No significant alterations were found among all the studied groups (Table 2).
progression of renal damage in rats with 5/6NX. In fact it has been shown in rats with 5/6NX the beneficial effect of the antioxidants such as vitamin E [45], N-acetylcysteine [47], melatonin [48], S-allylcysteine [27], and curcumin [4]. The data obtained in the present paper about the protective effect of curcumin on 5/6NX-induced proteinuria, systemic hypertension, histological alterations, and inflammation are consistent with the findings of Ghosh et al. [4, 21]. In addition, here is shown clearly that curcumin significantly ameliorated glomerular hypertension and hyperfiltration, which may limit the extension of renal injury [24–26]. In fact, it has been shown that renal ablation results in hemodynamic changes in the remnant nephrons that engage the participation of proinflammatory and profibrotic mechanisms that amplify nephron loss [49]. The renoprotective effect of curcumin was clearly associated with enhanced translocation of Nrf2, attenuation of oxidant stress, and preservation of the activity of several antioxidant enzymes. Based on the fact that curcumin was given daily along of the study and to the fact that Nrf2 nuclear translocation was enhanced, it is possible that the curcumin-induced decrease of oxidant stress and preservation of the activity of antioxidant enzymes may be secondary to the direct and indirect effect of this bifunctional antioxidant. With this administration protocol is not possible to distinguish between direct and indirect antioxidant effects of curcumin. Interestingly, the protective effect of curcumin against quinolinic acid-induced neurotoxicity is associated to the induction of Nrf2 in striatal tissue [50]. It has been reported that curcumin possesses nephroprotective properties in the experimental models of renal damage induced by cisplatin [34], potassium-dichromate [8], gentamicin [51], sodium fluoride [52], vancomycin [53], cyclosporine [54], acetaminophen [55], ferric nitrilotriacetate [56], and adriamycin [57]. Interestingly, no alterations

Figure 2: Curcumin (60 mg/kg/day) ameliorated 5/6NX-induced alterations in renal hemodynamics at the end of the study (day 30): (a) mean arterial pressure (MAP), (b) single-nephron glomerular filtration rate (SNGFR), (c) single-nephron plasma flow (Qa), (d) glomerular capillary pressure (Pgc), (e) stop-flow pressure (SFP) and (f) hydrostatic pressure gradient (ΔP), (g) afferent resistance (AR), and (h) efferent resistance (ER), and (i) glomerular filtration rate (GFR) observed in 5/6NX rats was prevented by curcumin (5/6NX+CUR group). Data are mean ± SEM, n = 8 – 10. Two-way ANOVA followed by Bonferroni. *P < 0.05 versus Control, **P < 0.05 versus 5/6NX.
were found in mitochondrial respiration in 5/6NX rats 30 days after surgery. State 3, state 4, RC, uncoupled respiration, and ADP/O ratio were unchanged in the four groups of rats studied. To our knowledge, this is the first study of mitochondrial respiration performed in rats with 5/6NX. Alterations in mitochondrial respiration have been found in other models of renal damage induced by potassium dichromate [8], ischemia and reperfusion [29], mercury
Figure 4: Curcumin (60 mg/kg/day) ameliorated 5/6NX-induced interstitial fibrosis at the end of the study (day 30). Upper panels shows representative images of the groups studied. 40X. Lower panel show quantitative data. Data are mean ± SEM of 8–10 rats. *P < 0.05 versus control, **P < 0.05 versus 5/6NX.

It is possible that alterations mitochondrial respiration may be found in this experimental model at longer periods of study (more than 30 days). In conclusion, curcumin treatment results in prevention of the hemodynamic changes in glomerular microcirculation, renal inflammatory injury, and functional deterioration associated with renal mass reduction. The protective effect of curcumin in the remnant kidney was associated with the nuclear translocation of Nrf2 and the prevention of both oxidant stress and the decrease of antioxidant enzymes. It is possible that both direct and indirect antioxidant effects of curcumin may be involved in
Figure 5: Curcumin (60 mg/kg/day) ameliorated 5/6NX-induced Interstitial inflammation at the end of the study (day 30). Upper panels shows representative images of the groups studied. 40X. Lower panel show quantitative data. Data are mean ± SEM of 8–10 rats. *P < 0.05 versus control, **P < 0.05 versus 5/6NX.
**Figure 6:** Curcumin (60 mg/kg/day) induced nuclear translocation of Nrf2 at the end of the study (day 30). Arrows indicate immunostained nuclei. Upper panels show representative images of the groups studied. 100X. Lower panel show quantitative data. Data are mean ± SEM, *P < 0.01 versus control, **P < 0.05 versus 5/6NX.
Figure 7: Curcumin (60 mg/kg/day) on 5/6NX-induced lipid peroxidation and decrease in the activity of antioxidant enzymes in kidney homogenates. CAT: catalase, GR: glutathione reductase, GPx: glutathione peroxidase, GST: glutathione S-transferase, SOD: superoxide dismutase, MDA: malondialdehyde, 4-HNE: 4-hydroxy-2-nonenal, k: constant first-order reaction, U: international units, mg: milligram of protein. Data represent mean ± SEM, n = 6–8. *P < 0.05 versus control; **P < 0.01 versus 5/6NX.
the protective effect observed. Potential clinical benefits of curcumin therapy deserve further study.

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