Therapeutic Potential of Curcumin for the Treatment of Brain Tumors

Neil V. Klinger1 and Sandeep Mittal1,2,3
1Department of Neurosurgery, Wayne State University, Detroit, MI, USA
2Department of Oncology, Wayne State University, Detroit, MI, USA
3Karmanos Cancer Institute, Wayne State University, Detroit, MI, USA

Correspondence should be addressed to Sandeep Mittal; smittal@med.wayne.edu

Received 23 July 2016; Accepted 7 September 2016

Academic Editor: Mohamed Essa

Copyright © 2016 N. V. Klinger and S. Mittal. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Brain malignancies currently carry a poor prognosis despite the current multimodal standard of care that includes surgical resection and adjuvant chemotherapy and radiation. As new therapies are desperately needed, naturally occurring chemical compounds have been studied for their potential chemotherapeutic benefits and low toxicity profile. Curcumin, found in the rhizome of turmeric, has extensive therapeutic promise via its antioxidant, anti-inflammatory, and antiproliferative properties. Preclinical in vitro and in vivo data have shown it to be an effective treatment for brain tumors including glioblastoma multiforme. These effects are potentiated by curcumin’s ability to induce G2/M cell cycle arrest, activation of apoptotic pathways, induction of autophagy, disruption of molecular signaling, inhibition of invasion, and metastasis and by increasing the efficacy of existing chemotherapeutics. Further, clinical data suggest that it has low toxicity in humans even at large doses. Curcumin is a promising nutraceutical compound that should be evaluated in clinical trials for the treatment of human brain tumors.

1. Introduction

Primary central nervous system tumors have an incidence of 27.86 per 100,000 [1]. Glioblastoma (GBM) is the most common malignant primary CNS tumor with an annual incidence of 3.19 per 100,000. GBM accounts for 15.4% of all primary brain tumors and 45.6% of primary malignant brain tumors [1, 2]. The current standard of care for GBM consists of surgical resection followed by the Stupp regimen (adjuvant chemotherapy and radiation) and it has been used in practice for over a decade without substantial improvements. This aggressive multimodal treatment prolongs median overall survival to 15 months [3–5]. Other therapies approved for use in GBM include alternating electrical fields produced by Optune Therapy (Novocure Inc.), further prolonging median survival to 20 months [6]. With such poor prognoses, additional therapeutics are needed to improve survival and quality of life for patients with malignant gliomas. Complementary medicine and nutraceuticals are of particular interest and have been studied extensively as anticancer agents because they are usually associated with low toxicity profiles. This allows them to be safely used at high doses or added to existing chemotherapy regimens as adjuvant treatment.

Curcumin is found in rhizome of turmeric (Curcuma longa), which is a member of the ginger family (Figure 1) [7]. Its traditional uses, long known by Ayurvedic medicine from India and traditional Chinese medicine, include treating infections, liver and skin disorders, dressing wounds, burns, and decreasing inflammation [7]. In fact, powdered Curcuma longa has been used in Asian medicine, cosmetics, and fabric dying for more than 2000 years [8]. Curcumin possesses multiple beneficial chemical properties, including antioxidant, anti-inflammatory, and chemotherapeutic potential both against cultured cells and in treatment studies using animal models. Current data suggests that curcumin may be useful in a wide spectrum of human disorders including Alzheimer’s disease, Parkinson’s disease, diabetes, cardiovascular disease, arthritis, and various neoplasms including brain tumors [9, 10].
2. Mechanism and Preclinical Data

Curcumin's effects on glioma cells in vitro and in vivo have been well studied. Curcumin has many molecular targets (Figure 2) and therefore diverse and complex mechanisms of action. The antitumoral effects of curcumin are thought to act through many different signaling pathways such as cellular proliferation [11–13], apoptosis [11, 12, 14], autophagy [15–17], angiogenesis [18–20], immunomodulation [21], invasion [14, 22, 23], and metastasis [24–26]. These pathways have been reviewed comprehensively elsewhere [27–31]. In addition to its effectiveness as an antineoplastic, curcumin has been found to be protective against reactive oxygen species (ROS). A study by Rathore et al. [32] examined the effects of curcumin oil on a transient ischemia model in Sprague-Dawley rats. The middle cerebral artery was occluded using a nylon filament for two hours, followed by a 24-hour reflow period. Treatment with curcuma oil reduced the size of infarcted cerebral tissue, improved neurologic function, and reduced oxidative stress caused during the reperfusion period. The authors reported that these protective effects were obtained by inhibition of ROS, reducing peroxynitrite levels and caspase-3 activity and preventing destabilization of the mitochondrial membrane potential [32]. In an in vitro model of ischemia-reperfusion using rat cortical neurons, curcumin was found to protect neurons from death caused by oxygen and glucose deprivation. The authors of this study suggested that curcumin activates an antioxidant protein, thioredoxin, in the Nrf2 pathway [33]. In addition, curcumin may inhibit cellular glyoxalases leading to decreased ATP and glutathione, which can impact cellular metabolism and may account for some of its anti-inflammatory and antitumor effects [34].

2.1. Curcumin Induces G2/M Cell Cycle Arrest. There is a bevy of data to suggest that curcumin is able to induce G2/M cell cycle arrest and apoptosis. More recent work has tried to explain how this cell cycle arrest can occur. Wu et al. published a study in 2013 [35] that suggests that this might be due to increased DAPK1 expression. The authors treated the U251 GBM cell line with curcumin and found a dose-dependent increase in DAPK1 mRNA by real-time RT-PCR and verified a corresponding increase in protein expression by western blot analysis. Further, they used siRNA (si-DAPK1-1 and si-DAPK1-2) transfection to suppress DAPK1 and found that curcumin had an attenuated ability to suppress STAT3 and NF-κB phosphorylation. They also showed that their knockdown of DAPK1 inhibited curcumin-mediated caspase-3 activation and led to decreased apoptosis (33.0% apoptotic cells versus 58.3% in their control) [35]. These findings suggest that DAPK1 plays an important role in curcumin-mediated cell death. Other studies in U251 GBM cells show that p53 expression is upregulated by curcumin treatment, as were CDK1 p21Waf1/Cip1 (p21) and ING4—a tumor suppressor gene that has been found to be suppressed in gliomas [36, 37]. Curcumin owes part of its antiproliferative effects to suppression of cyclin D1 and to induction of p21. In a study that treated U87 GBM cells with curcumin, transcription factor Egr-1 was found to activate transcription of p21 independent of p53 activation. Egr-1 expression was reportedly induced by curcumin via ERK.
and JNK [13]. An in vitro study using U87 human glioma cell line showed that 5–10 μM curcumin could inhibit proliferation and that treated cells were arrested in G2/M stage by increased expression of DUSP-2 and inhibition of ERK and JNK phosphorylation. Interestingly, trapping cells in the G2/M phase appears to enhance their sensitivity to radiation [38, 39]. Curcumin’s radiosensitizing effects have also been examined in neuroblastoma (SK-N-MC) cells. Pretreating cells with 100 nM curcumin suppressed radiation induced NF-κB, enhanced radiation induced caspase activation, and inhibited antiapoptotic molecules [40].

2.2. Curcumin Activates Apoptotic Pathways. An in vitro study using human GBM (A172, KNS60, U251MG(KO)) and medulloblastoma (ONS76) cell lines showed that while curcumin was able to inhibit growth of these cell lines, only KNS60 and ONS76 were arrested at G2/M. These findings suggest that the mechanism for growth inhibition is not exclusively due to cell cycle arrest. They also found a significant increase in caspase 3/7 activity, curcumin induced DNA damage, and apoptosis triggered by overexpression of BAX and downregulating Bcl2 and survivin, as well as inhibition of telomerase and downregulation of hTERT [41]. An in vitro study using 8401 GBM cell line found that curcumin decreased cell proliferation, reduced mitochondrial membrane potential, induced DNA fragmentation, induced apoptosis via a caspase-dependent pathway (caspase-3, caspase-8, and caspase-9), and inhibited NF-κB transcription factor activity [42]. In U87MG GBM cell line, curcumin can induce apoptosis by suppressing antiapoptotic signals, by promoting activation of caspase-8, and through an increased BAX/Bcl-2 ratio [43]. Solubilized curcumin (667 μM in PBS, 3% DMSO) can increase caspase 3/7 activity in both mouse (B16F10 melanoma, GL261 glioma, and N18...
neuroblastoma) and human (HOG oligodendroglioma and A549 lung cancer) cell lines and it decreased tumor cell viability as measured by MTT assay. Further, LDH release in these experiments was not increased, suggesting apoptotic cell death [19]. The authors of this study also report that cyclin D1, NF-κB, AKT, ERK, and Bcl-xL were suppressed when B16F10 cells were treated with curcumin [19]. In an in vivo arm of this experiment, the authors injected 10,000 B16F10 mouse melanoma cells in the brain of C57BL/6 mice. They then received either daily tail vein injections of 200 μL of their solubilized curcumin (estimated final plasma concentration of 35 μM in PBS, 0.15% DMSO) or a control injection (PBS, 0.15% DMSO) for 18 days. Additionally, this experiment was repeated with intracerebral delivery of the curcumin solution via a stainless steel cannula. While all of the control animals developed intracranial tumors, only one of the 5 treated mice developed detectable tumor [19].

2.3. CurcuminInducesAutophagy. Autophagy is a cellular process for disassembling unnecessary or dysfunctional cellular components and is essential for maintaining energy during states of nutritional stress and during programmed cell death. It also appears to have an important role in regulation of glioma-initiating cells, which are poorly differentiated and share features of neural stem cells. An in vitro study published in 2012 showed that curcumin can induce differentiation and halt growth of glioma-initiating cells (SU-2 and SU-3) from surgically resected human GBM by activating autophagy. In the same study, glioma-initiating cells were implanted intracranially in athymic nude mice and randomized to a treatment group consisting of intraperitoneal (IP) injection of curcumin (300 mg/kg) every 3 days or control group (n = 14 each). Those mice who received treatment had increased survival (study halted after 120 days) versus non-treated animals [16]. In a separate study, curcumin activated the ERK1/2 pathway and inhibited the AKT/mTOR/p70S6K pathway, resulting in autophagy both in vitro and in vivo using a subcutaneous xenograft model in nude mice with U87 cells. The authors report that curcumin induced G2/M cell cycle arrest in addition to autophagy in U87 and U373 GBM cells [17].

2.4. CurcuminDisruptsMolecularSignaling. The NF-κB pathway is upregulated in GBM, and inhibitors of NF-κB exhibit relatively low toxicity to normal tissues. A preclinical study of NF-κB inhibitors found that NF-κB activity correlated with percentage cell viability in C6 and U138 GBM lines [45]. The authors report that inhibition of NF-κB leads to mitochondrial dysfunction with arrest in the G2/M phase of the cell cycle. Interestingly, NF-κB is found to be overstimulated in cisplatin-resistant C6 cells and NF-κB inhibitors were able to overcome cisplatin resistance [45]. Similarly, progranulin has been found to be overexpressed in many GBM cell lines (U87, GBM8904, and SIRI) and tumor samples [46]. Progranulin overexpression contributes to tumorigenesis and treatment resistance by upregulating DNA repair and stemness genes using the transcription factor AP-1. Curcumin is an AP-1 inhibitor, and it was found to downregulate progranulin promoter activity and expression [46]. Other studies have shown that curcumin can reduce GBM cell survival through inhibition of AP-1 and NF-κB by preventing constitutive activation of JNK and AKT [47]. An in vitro study examined the role of glucose-6-phosphate translocase (G6PT) in U87 GBM cell signal transduction. In this experiment, suppression of gene expression was accomplished using siRNAs and led to apoptosis and necrosis. Curcumin (35 μM) also inhibited G6PT gene expression >90%. However, the authors also report transfection with an expression vector for G6PT rescued cells from curcumin-mediated cell death. This suggests that G6PT may be a novel chemotherapeutic target and that G6PT overexpression may lead to treatment resistance [48]. In a separate set of experiments, murine glioma cell lines (Tu-2449, Tu-9648, and Tu-251) with constitutively expressed STAT3 treated with curcumin showed a dose-dependent decrease in the activity of phosphorylated JAK1 and JAK2 and led to downstream inactivation of STAT3. In an in vivo arm of this study, C6B3F1 mice were fed a diet rich in fat and cholesterol with or without fortification with curcumin for 7 days. Subsequently, they received intracranial implantation of either Tu-2449 or Tu-9648 glioma cells and maintained their diet. Fifteen percent of Tu-2449 implanted mice who received curcumin-enriched diets had tumor-free long-term survival versus 0% of the control dieted animals. Curcumin fed Tu-9648 implanted mice experienced a 38% increase in tumor-free long-term survival [49].

The sonic hedgehog (SHH) signaling pathways are important in the carcinogenesis of medulloblastoma. It is a major regulator of cell proliferation and death. Curcumin has been shown to downregulate SHH, leading to decreased downstream targets GLI and PTCH and cytotoxicity in cell lines MED-4, MED-5, and DAOY [50]. Targeting SHH and GLI with curcumin treatment has also been done in glioma cells (U87 and T98G). Similar to medulloblastoma, mRNA and protein levels of SHH and GLI were downregulated with curcumin treatment. IP injection of curcumin in U87-implanted nude mice also reduced tumor volume, prolonged survival, and decreased GLI1 expression [51]. In the medulloblastoma cell line DAOY, curcumin has been shown to inhibit Wnt/β-catenin signaling and suppress proliferation with IC50 of 35 μM after 48 hours [52]. Mechanistically, curcumin may decrease histone deacetylase 4 expression leading to increased tubulin acetylation and mitotic catastrophe in addition to its effects on G2/M cell cycle arrest in DAOY medulloblastoma cells [53].

2.5. CurcuminDecreasesInvasionandMetastasis. Abnormal expression of matrix metalloproteinases (MMPs), membrane-associated or secreted enzymes used to digest extracellular matrix proteins, is one method by which glioma cells are able to invade normal brain tissue. Data has shown that curcumin is able to suppress expression of MMP-1, MMP-3, MMP-9, and MMP-14 in GBM cell lines U87MG and U373MG via the common upstream AP-1 and MAP kinases [22, 23, 54]. Urokinase-type plasminogen activator (uPA) is a serine protease that starts a degradative cascade by converting extracellular plasminogen to plasmin and eventually degrades extracellular matrix collagen and
activates other MMPs to aid in invasion [55]. Curcumin can prevent nuclear translocation of RelA/NF-κB, which prevents upregulation of uPA [55]. Curcumin has also been found to dramatically reduce MMP-9 in murine glioma cell lines Tu-2449, Tu-9648, and Tu-251 [49]. An important contributor to tumors propensity to invade and metastasize is angiogenesis. Cell lines derived from surgically resected pituitary adenomas treated with curcumin have decreased HIF1A and VEGFA, both of which are involved in tumoral angiogenesis [56].

2.6. Curcumin Increases the Efficacy of Existing Chemotherapeutics. Data also exists to suggest that curcumin acts synergistically with chemotherapeutics already approved for the treatment of brain tumors. A preclinical study examined the efficacy of curcumin for the treatment of GBM in vitro and in vivo [57]. Cell lines U138MG, U87, U373, and C6 were treated with curcumin and found to have IC_{50} values of 29, 19, 21, and 25 μM, respectively, whereas IC_{50} for astrocyes was 135 μM. Chemotherapy synergism was subsequently tested using U138 and C6 cell lines. Curcumin alone resulted in 55% and 75% viability, respectively. When cisplatin was added in combination with 25 μM curcumin, the viability dropped to 30% and 10% for U138 and C6 cell lines, respectively. When doxorubicin was added in combination with 25 μM curcumin, viability was reported to be 36% and 46%. Mechanistically, the authors reported that curcumin decreased activation of PI3K/AKT and NF-κB pathways, decreased expression of Bcl-xL, and caused mitochondrial dysfunction and apoptosis. The authors also implanted Wistar rats with C6 GBM cells intracranially and treated them with 50 mg/kg/day curcumin via IP injection within days 10–20 after implantation. Sixty-four percent of curcumin treated animals developed tumors as compared to 100% of the DMSO treated controls. Of the treated animals who developed tumors, average tumor volume was 73% less than that of the controls [57]. Curcumin in combination with temozolomide (TMZ) appears to have additive cytotoxic benefit in GBM cells [15]. Further, both drugs appear to cause G2/M arrest by activating proteins such as Wee, Cdc2, CHK1, and Cdc25c. Decreased phosphorylation of cyclin B1 and cyclin D1 was also observed [15]. The authors found that treatment with either TMZ or curcumin appears to induce autophagy that was dependent on ERK1/2 prior to apoptosis. Curcumin was also found to inhibit STAT3, NF-κB, and PI3K/AKT [15]. A study by Ramachandran et al. [58] investigated potentiation of etoposide (ETP) and TMZ by curcumin and a trade preparation of turmeric called Tumeric Force™ (TF) in human U87 GBM and D283 medulloblastoma cell lines. U87 IC_{50} values for ETP, TMZ, curcumin, and TF were 6.5, >2000, 373, and 30.8 μg/mL, respectively. For D283 cells, those IC_{50} values were 0.19, 147, 28.7, and 1.6 μg/mL. Combination index values in U87 glioma cells for ETP + curcumin, TMZ + curcumin, and ETP + TMZ + curcumin were 0.55, 2.07, and 0.39 μg/mL, respectively. The combination index values with TF instead of curcumin were 0.38, 0.57, and 0.05 μg/mL [58]. Curcumin has also been studied for efficacy in combination with paclitaxel (PTX) for the treatment of GBM. LN18 and U138MG cell lines treated with 20 μM curcumin and 10 nM PTX were found to have a combination index of 0.1 and 0.09, respectively, indicating a synergistic effect. This combination activated caspase-3, caspase-8, and calpain, increased BAX, and reduced Bcl-2 initiating apoptosis. Combination therapy also decreased the ability of LN18 and U138MG cells to invade matrigel [59]. In addition to acting synergistically, curcumin may prevent chemotherapy resistance. A study reported in 2008 on Sprague-Dawley rats showed curcumin can inhibit a protein linked with multidrug resistance in the luminal membrane of capillaries at the blood-brain barrier- (BBB-) ATP-binding cassette transporter ABCG2 [60]. A separate, intriguing method by which GBM may resist hydrophobic therapeutic agents is by accumulating lipid droplets that sequester these drugs. Curcumin is hydrophobic and has been shown to concentrate near the cell membrane and in cytoplasmic droplets of U251 human GBM cells [61]. Zhang et al. sought to overcome this mechanism of resistance in vitro using pyrrolidine-2, a cytoplasmic phospholipase A2α inhibitor, in combination with curcumin. When the cells were pretreated with pyrrolidine-2 24 hours before curcumin administration, cell viability was found to approach 0% [61].

2.7. Curcuminoids and Curcumin Derivatives. Demethoxycurcumin (C2) and bisdemethoxycurcumin (C3) (Figure 1) are two curcuminoids often found in small percentages of curcumin extractions. An in silico study published in 2009 [62] examined the docking of curcuminoids with the Bcl-2 apoptotic proteins (Protein Data Bank accession numbers 1G5M and 1GJH). The authors calculated free energies and inhibition constants for these molecules and found that C2 binds more favorably than curcumin (ΔG = −6.97 versus −4.53 kcal/mol; K_{i} 0.56 versus 2.21 nm). In vitro treatment of U87 GBM cells showed marked decrease in Bcl-2 expression with all three compounds after 48 hours and that C2 showed significant progression of percent apoptosis compared to curcumin or C3. An in vitro binding assay utilizing circular dichroism spectroscopy showed that C2, like the natural Bcl-2 inhibitor Bak, caused a conformational change different from curcumin or C3 [62]. These findings suggest that C2 induces Bcl-2 mediated apoptosis more effectively than curcumin or C3. A study published by Huang et al. [63] also examined the effects of C2. They showed that C2 had IC_{50} of 22.7 μM in GBM 8401 cells and that apoptosis was induced by decreasing the mitochondrial membrane potential and caspase-dependent pathways similar to other studies of curcumin [63]. Other curcumin analogs have been developed in an attempt to improve upon the compound’s therapeutic promise. For example, lead compounds identified by Campos et al. show comparable IC_{50} values against GBM (U87MG) and neuroblastoma (SK-N-SH and SK-N-FI) cell lines [64].

3. Delivery Mechanisms

The therapeutic benefits of curcumin are limited by its poor absorption, rapid metabolism, and poor water solubility [65]. Curcumin is broken down by multiple enzymatic pathways,
including glucuronidation, sulfation, alcohol dehydrogenase, and p450 system leading to rapid metabolism and excretion [66]. However, its lack of toxicity allows for administration of large doses. Twenty-four subjects with mean age of 34 were provided capsules with 0.5–12 g of curcumin and serum analysis was conducted prior to testing at 1, 2, and 4 hours after dosing. Peak serum concentration was <2 μM even at 10- and 12-gram doses [67, 68]. These clinical data also show that doses up to 12 grams per day do not provoke serious side effects, though trouble with tablet bulk, diarrhea, and yellow stool were noted [67, 68]. Piperine, an inhibitor of hepatic and intestinal glucuronidation, has long been suggested to be administered as an adjuvant to prevent the degradation of medications including curcumin [69–71]. Ten subjects with mean age of 60 years participated in a randomized crossover trial to compare the effects of oral curcumin (2 g) alone or with the addition of piperine (20 mg). A two-week washout period was used before the crossover. They found that, when administered with piperine, peak serum concentrations of curcumin increased 30-fold and the relative bioavailability was 2000%. The Vₚ₅ for curcumin administered with piperine was 203 L/kg. No adverse events were reported in this study [69].

Though current literature contains mixed data, curcumin distribution to the brain might be hindered by limited BBB permeability [65, 72]. Certain nanoparticle formulations, such as poly(lactic-co-glycolic acid), have been shown to increase distribution to brain tissue [65]. The half-life of curcumin-loaded poly(lactic-co-glycolic acid) in brain tissues from Sprague-Dawley rats for this study was reported to increase from 9 minutes to 15 minutes. When brain regions were examined separately, the compound’s half-life was found to be significantly extended in the cerebral cortex (20 versus 2 minutes) and hippocampus (17 versus 8 minutes) but not in the cerebellum, brainstem, striatum, or other brain regions [65]. Curcumin has also been loaded into poly(butyl)cyanoacrylate (PBCA) nanoparticles using apolipoprotein E3 to increase transport across the BBB [73]. This delivery system was found to induce apoptosis in SH-SY5Y neuroblastoma cells more efficaciously than curcumin in IC₅₀ values of 14.3 and 7.5 μM for curcumin alone against WEHI-164 cells [73]. WEHI-164 cells were injected subcutaneously in the flank in an in vivo experiment using BALB/c mice, and the results showed that treatment with dendrosomal curcumin led to significant reductions in tumor burden compared to curcumin treated animals or control [76]. Dendrosomal curcumin has also shown efficacy in the treatment of metastatic breast cancer cells (4T1) in vitro and in vivo with 11% of animals having metastases at necropsy compared to 89% of control. The authors also reported that treatment with dendrosomal curcumin led to downregulation of VEGF, COX-2, and MMP expression [80].

Curcumin has also been conjugated as a poly(glycerol-sebacate-curcumin) polymer similar to carmustine polymers used for local treatment with gliomas [81]. In vitro studies show IC₅₀ values in response to this polymer of 23.2 μg/mL with U87 cells and 20.2 μg/mL with T98 cells [81]. Poly-caprolactone implants have been studied as another delivery method for test agents with poor bioavailability. An in vitro assay with 9 L rat glioma cells reported that curcumin-loaded poly(e-caprolactone)-poly(ethylene glycol)-poly(ε-caprolactone) nanofibers caused dose-dependent growth inhibition [82]. Gupta et al. reported a preclinical study [83] in August-Copenhagen Irish rats who received four polycaprolactone implants that were loaded with 40 mg of...
curcumin. Analysis of liver, brain, and plasma by high-performance liquid chromatography (HPLC) with fluorescence spectroscopy showed an increase in curcumin levels versus animals receiving sham implant and a diet supplemented with 1000 ppm curcumin [83]. They also demonstrated that these implants can reduce benzo[a]pyrene induced lung DNA adducts by 62% in Sprague-Dawley rats [83].

Curcumin microparticles in a poly(D,L-lactide-co-glycolide) polymer has been formulated in an attempt to improve delivery of the drug and sustain its release. The investigators reported that a single subcutaneous injection was able to sustain release in the blood and body tissues of BALB/c mice for 4 weeks. Levels of curcumin in the lungs and brain show 10–30-fold greater distribution than the blood. Further, animals implanted with MDA-MB-231 human breast adenocarcinoma cells were effectively treated with these microparticles (49% decreased tumor volume versus empty microparticle), whereas repeated systemic injections of curcumin alone were no different than the vehicle treatment [84]. Curcumin nanoparticles (NanoCurc™) [21, 85] have also been used as a method to overcome the delivery and bioavailability barriers of curcumin. These particles, unlike free curcumin, are able to disperse in an aqueous environment [21]. Studies using xenograft models of human pancreatic cancer in athymic mice show that NanoCurc™ can lead to approximately 50% reduction in tumor volume. This inhibition was enhanced with the addition of gemcitabine.

The authors reported that these effects were observed through decreased NF-κB activation, MMP-9, and cyclin D1 [85]. These nanoparticles have also been shown to suppress in vitro growth by MTS cell proliferation assay of embryonal tumor lines (DAOY and D283Med), U87 GBM cells, and glioblastoma neurosphere lines (JHH-GBM14 and HSR-GBM1) in a dose-dependent manner [86]. The authors suggested that, in this experiment, the effects could be due to a combination of G2/M arrest and apoptotic induction [86].

Additional benefits of using delivery systems include the ability to deliver multiple compounds simultaneously. For example, Dilnawaz and Sahoo used magnetic nanoparticles (MNP) to deliver both curcumin and TMZ to monolayer and spheroid T-98G GBM cultures. In monolayer, IC50 values (μg/mL) for TMZ, curcumin, TMZ and curcumin were 1.1, 6.6, and 0.8 without MNP delivery and 0.2, 0.6, and 0.1 with MNP delivery. These same values in spheroid cultures were 23, 28, and 10 without MNP and 10, 11, and 5.2 with MNP delivery. Combination index analysis showed that the drug combination functioned synergistically in both cultures [87]. Another study using magnetic core nanocapsules reported the ability to deliver drugs with differing hydrophobicity [88]. They were able to accommodate doxorubicin (hydrophilic) and curcumin (hydrophobic) in the same particles for delivery to RG2 rat glioma cells [88].

Curcumin has also been coupled to an antibody as a delivery vehicle. Langone et al. [89] demonstrated that curcumin coupled to Muc18, a melanoma specific antibody, increased its efficacy against B16F10 melanoma. The in vitro IC50 value was reduced from 22 μM to 0.09 μM in cells treated with the curcumin-antibody adduct versus curcumin alone. In addition, C57BL/6 mice implanted with 1,000 B16F10 cells in their right forebrain showed a 10-fold decrease in tumor burden when treated with and curcumin-antibody adduct [89]. The authors reported that this effect was achieved by suppressing NF-κB [89]. In 2014, Langone et al. published another study that used an antibody delivery system for curcumin. In this experiment, a GBM-specific CD68 antibody was used to target GL261 (mouse), T98G (human), and U87MG (human) GBM cells [18]. GL261 cells were treated in vitro with 50 μM of curcumin for 24 hours, and western blot analysis showed suppression of NF-κB. In addition, curcumin was found to inhibit AKT-1 (neural survival and antiapoptotic), Bcl-xL (antiapoptotic), cyclin D1 (cell cycle promoter), and VEGF (promoting angiogenesis) [18]. Curcumin alone yielded IC50 15 μM, whereas treatment with the curcumin-antibody adduct had IC50 of 0.125 μM. IC50 values for U87MG and T98G cells were 25 and 8 μM for curcumin alone and 0.400 and 0.225 μM, respectively, for treatment with the CD68 antibody-linked curcumin. GL261 cells were also implanted in the right forebrain of C57BL6 mice. These mice were treated with intracranial infusions of 16 pmol CD68 antibody-linked curcumin followed by tail vein infusion of curcumin and resulted in a reduction of intracranial tumor burden [18].

4. Toxicity/Safety

Natural products are of interest as anticancer agents because they have been associated with low toxicity profiles. This allows them to be safely used at high doses or added to existing regimens. Indeed, curcumin has been administered to human subjects at large doses without major side effects [67–69]. However, some data exists that serves to caution against its use. Despite being an antioxidant, curcumin may lead to a temporary increase in ROS and a decrease in cell viability by depletion of glutathione [90]. Curcumin might cause DNA base damage and fragmentation through cytochrome p450 enzyme generation of a curcumin radical and lead to apoptotic cell death in healthy as well as tumoral tissues [91]. Interestingly, this damage was attenuated when curcumin was present at larger concentrations [92]. Similar DNA damage findings have been noted during in vivo studies [93, 94]. Curcumin may inhibit the tumor suppressor p53 function in colon cancer cells and contribute to tumorogenesis [95]. Administration of curcumin with camptothecin or cyclophosphamide may inhibit their effectiveness; thus careful study of curcumin is warranted prior to its addition to an existing chemotherapy [96].

5. Clinical Data

There are over one hundred clinical trials examining the potential therapeutic effects of curcumin (Table 1, https://clinicaltrials.gov/). Many of these trials examine the efficacy of curcumin for the treatment of gastrointestinal diseases (e.g., Cohn’s disease, ulcerative colitis, and inflammatory bowel disease), endocrine disorders (e.g., diabetes mellitus), neoplasms (breast, gastrointestinal, cervical, lymphoma, and pancreatic), immune system diseases (e.g., atopy, multiple
<table>
<thead>
<tr>
<th>Trial Status</th>
<th>Site</th>
<th>Disease</th>
<th>Measure</th>
<th>Trial ID*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vascular</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Curcumin or placebo Ongoing</td>
<td>Lawson Health Research Institute, CA</td>
<td>AAA</td>
<td>Serum creatinine</td>
<td>NCT01225094</td>
</tr>
<tr>
<td>Curcumin or placebo Ongoing</td>
<td>University of Colorado Boulder, USA</td>
<td>Vascular aging</td>
<td>Arterial stiffness, arterial pulse-wave velocity, NO dilation</td>
<td>NCT01968564</td>
</tr>
<tr>
<td>Curcumin or placebo Ongoing</td>
<td>UPEI, CA</td>
<td>Vascular stiffness</td>
<td>Arterial stiffness by tonometry, IL-6, CRP, CK</td>
<td>NCT02281981</td>
</tr>
<tr>
<td>Neurologic/psychiatric</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Curcumin with or without yoga Recruiting</td>
<td>VA Los Angeles, USA</td>
<td>AD</td>
<td>Biomarkers for MCI</td>
<td>NCT01811381</td>
</tr>
<tr>
<td>Curcumin Recruiting</td>
<td>SHSC, CA</td>
<td>Adolescent bipolar disorder</td>
<td>Mood by CDRS-R, biomarkers</td>
<td>NCT01928043</td>
</tr>
<tr>
<td>Curcumin with other supplements or placebo Ongoing</td>
<td>HCL, France</td>
<td>Fibromyalgia</td>
<td>QOL by GIQLI</td>
<td>NCT01469936</td>
</tr>
<tr>
<td>Curcumin or placebo Ongoing</td>
<td>UCLA, USA</td>
<td>MCI, normal aging</td>
<td>Cognitive changes by neuropsych assessment</td>
<td>NCT01383161</td>
</tr>
<tr>
<td>Curcumin or placebo with omega-3 fatty acid Invitation only</td>
<td>TUMS, Iran</td>
<td>Migraine</td>
<td>HA, endothelial factors, inflammation</td>
<td>NCT02532023</td>
</tr>
<tr>
<td>Curcumin or placebo with IFN-β1A Ongoing</td>
<td>Merck Serono, Italy</td>
<td>Multiple sclerosis</td>
<td>Proportion of subjects with active T2 lesions</td>
<td>NCT01514370</td>
</tr>
<tr>
<td>Curcumin or placebo Recruiting</td>
<td>Beersheva Mental Health Center, Israel</td>
<td>Schizophrenia</td>
<td>Psychotic symptoms (PANSS)</td>
<td>NCT02298985</td>
</tr>
<tr>
<td>Curcumin Recruiting</td>
<td>VA Los Angeles, USA</td>
<td>Schizophrenia</td>
<td>MCCB</td>
<td>NCT02104752</td>
</tr>
<tr>
<td>Curcumin or placebo Not yet recruiting</td>
<td>Yale, USA</td>
<td>Schizophrenia, schizoaffective</td>
<td>MCCB</td>
<td>NCT02476708</td>
</tr>
<tr>
<td>Gastrointestinal</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Curcumin or placebo with thiopurines Recruiting</td>
<td>CHU, France</td>
<td>Crohn's disease</td>
<td>Rutgeerts endoscopic score</td>
<td>NCT02255370</td>
</tr>
<tr>
<td>Curcumin with triple therapy or triple therapy alone Not yet recruiting</td>
<td>Rabin Medical Center, Israel</td>
<td>Helicobacter pylori infection</td>
<td>Eradication by urea breath test</td>
<td>NCT02018328</td>
</tr>
<tr>
<td>Curcumin with selenium and green tea or placebo Recruiting</td>
<td>Meir Medical Center, Israel</td>
<td>Irritable bowel syndrome</td>
<td>QOL by questionnaires</td>
<td>NCT01167673</td>
</tr>
<tr>
<td>Curcumin or placebo Not yet recruiting</td>
<td>NCI, USA</td>
<td>MAG and/or GIM</td>
<td>Changes in IL-1β, safety, tolerability, histologic grade</td>
<td>NCT02782949</td>
</tr>
<tr>
<td>Curcumin or placebo Not yet recruiting</td>
<td>Schneider Children’s Medical Center, Israel</td>
<td>Pediatric ulcerative colitis</td>
<td>Disease activity by PUCAI</td>
<td>NCT02277223</td>
</tr>
<tr>
<td>Curcumin or placebo with 5-ASA Recruiting</td>
<td>Asian Institute of Gastroenterology, India</td>
<td>Ulcerative colitis</td>
<td>Time to clinical and endoscopic remission</td>
<td>NCT02683733</td>
</tr>
<tr>
<td>Curcumin or placebo with 5-ASA Recruiting</td>
<td>Asian Institute of Gastroenterology, India</td>
<td>Ulcerative colitis in remission</td>
<td>Percentage of patients in remission</td>
<td>NCT02683759</td>
</tr>
<tr>
<td>Oncologic</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Curcumin or placebo Recruiting</td>
<td>Emory University, USA</td>
<td>Breast cancer</td>
<td>NF-κB by ELISA</td>
<td>NCT01740323</td>
</tr>
<tr>
<td>Curcumin at two different doses Recruiting</td>
<td>OSU, USA</td>
<td>Breast cancer, obesity</td>
<td>Adherence, tolerability, safety</td>
<td>NCT01975363</td>
</tr>
<tr>
<td>Curcumin or HPR Plus or placebo Recruiting</td>
<td>University of Rochester, USA</td>
<td>Noninflammatory breast cancer</td>
<td>Mean radiation dermatitis severity</td>
<td>NCT02556632</td>
</tr>
<tr>
<td>Curcumin with standard treatment Recruiting</td>
<td>UZ Leuven, Belgium</td>
<td>Endometrial carcinoma</td>
<td>Peripheral blood inflammatory markers</td>
<td>NCT02017353</td>
</tr>
<tr>
<td>Trial</td>
<td>Status</td>
<td>Site</td>
<td>Disease</td>
<td>Measure</td>
</tr>
<tr>
<td>-------</td>
<td>--------------</td>
<td>-------------------------------------------</td>
<td>--------------------------------------</td>
<td>-------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Curcumin</td>
<td>Recruiting</td>
<td>Baylor Research Institute, USA</td>
<td>Squamous CIN3</td>
<td>Safety, feasibility, overall and pathologic response</td>
</tr>
<tr>
<td>Curcumin and piperine</td>
<td>Recruiting</td>
<td>Mayo, USA</td>
<td>Cancer with inflamed ureteral stent</td>
<td>AE, max tolerable dose, optimal dose</td>
</tr>
<tr>
<td>Curcumin with cholecalciferol</td>
<td>Recruiting</td>
<td>Case CCC, USA</td>
<td>CLL or SLL, stages 0-II</td>
<td>Overall response rate by NCI-WG (CLL) or Cheson (SLL)</td>
</tr>
<tr>
<td>Curcumin</td>
<td>Recruiting</td>
<td>University Hospital Salzburg, Austria</td>
<td>Locally advanced or metastatic cancer</td>
<td>Safety, max tolerable dose, tumor response</td>
</tr>
<tr>
<td>Curcumin or placebo</td>
<td>Recruiting</td>
<td>UPR, Puerto Rico</td>
<td>FAP</td>
<td>Tolerability and efficacy by polyp number and size</td>
</tr>
<tr>
<td>Curcumin or placebo</td>
<td>Ongoing</td>
<td>NCI, USA</td>
<td>FAP</td>
<td>Laboratory biomarker analysis</td>
</tr>
<tr>
<td>Curcumin with anthocyanin extract</td>
<td>Recruiting</td>
<td>The Hospital Galliera, Italy</td>
<td>Colorectal adenoma</td>
<td>β-Catenin, NFκB, Ki-67, p53 by IHC</td>
</tr>
<tr>
<td>Curcumin with 5-FU</td>
<td>Recruiting</td>
<td>Baylor Research Institute, USA</td>
<td>Metastatic colon cancer resistant to 5-FU</td>
<td>Safety, toxicity, response, biomarkers</td>
</tr>
<tr>
<td>Curcumin with FOLFOX or FOLFOX alone</td>
<td>Ongoing</td>
<td>University of Leicester, UK</td>
<td>Metastatic colorectal cancer</td>
<td>Tolerable long-term dose, safety</td>
</tr>
<tr>
<td>Curcumin with Avastin/FOLFIRI</td>
<td>Not yet recruiting</td>
<td>Gachon University Gil Medical Center, Korea</td>
<td>Metastatic colorectal cancer</td>
<td>PFS</td>
</tr>
<tr>
<td>Curcumin with irinotecan</td>
<td>Recruiting</td>
<td>UNC Lineberger, USA</td>
<td>Metastatic colorectal cancer</td>
<td>Max tolerated dose, pharmacokinetics</td>
</tr>
<tr>
<td>Curcumin or placebo with capecitabine and radiation</td>
<td>Ongoing</td>
<td>MD Anderson, USA</td>
<td>Rectal cancer</td>
<td>Pathologic complete response rate</td>
</tr>
<tr>
<td>Curcumin or placebo with docetaxel</td>
<td>Recruiting</td>
<td>Centre Jean Perrin, France</td>
<td>Metastatic prostate cancer</td>
<td>Time to progression, PSA response</td>
</tr>
<tr>
<td>Curcumin or placebo</td>
<td>Recruiting</td>
<td>UT Southwestern, USA</td>
<td>Prostate cancer</td>
<td>PSA, recurrence free survival</td>
</tr>
<tr>
<td>Curcumin or placebo with RT</td>
<td>Recruiting</td>
<td>SBUMS, Iran</td>
<td>Prostate cancer</td>
<td>Proctitis and cystitis by CTCAE, PSA</td>
</tr>
<tr>
<td>Curcumin with gemcitabine, metformin, and paclitaxel</td>
<td>Recruiting</td>
<td>City of Hope Medical Center, USA</td>
<td>Metastatic pancreatic cancer</td>
<td>Feasibility, compliance, toxicity, survival</td>
</tr>
<tr>
<td>Curcumin with EGFR-TKI</td>
<td>Recruiting</td>
<td>Lady Davis Institute, CA</td>
<td>Nonresectable mutant EGFR NSCLC</td>
<td>Feasibility, adherence, AE, QOL (FACT-L), CRP</td>
</tr>
</tbody>
</table>

**Other**

<table>
<thead>
<tr>
<th>Trial</th>
<th>Status</th>
<th>Site</th>
<th>Measure</th>
<th>Trial ID*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Curcumin or placebo</td>
<td>Recruiting</td>
<td>University of Colorado Denver, USA</td>
<td>ADPKD</td>
<td>NCT02494141</td>
</tr>
<tr>
<td>Curcumin mouthwash</td>
<td>Recruiting</td>
<td>Aurora BayCare Medical Center, USA</td>
<td>Chemotherapy induced mucositis</td>
<td>NCT02300727</td>
</tr>
<tr>
<td>Curcumin or placebo</td>
<td>Recruiting</td>
<td>Lawson Health Research Institute, CA</td>
<td>CKD</td>
<td>NCT02369549</td>
</tr>
<tr>
<td>Curcumin or placebo</td>
<td>Recruiting</td>
<td>NNFTI, Iran</td>
<td>DMII</td>
<td>NCT02529969</td>
</tr>
<tr>
<td>Curcumin or placebo</td>
<td>Recruiting</td>
<td>NNFTI, Iran</td>
<td>DMII</td>
<td>NCT02529982</td>
</tr>
<tr>
<td>Curcumin with other nutraceuticals or placebo</td>
<td>Not yet recruiting</td>
<td>IRCCS Neuromed, Italy</td>
<td>NAFLD</td>
<td>NCT02369536</td>
</tr>
</tbody>
</table>
Table 1: Continued.

<table>
<thead>
<tr>
<th>Trial</th>
<th>Status</th>
<th>Site</th>
<th>Disease</th>
<th>Measure</th>
<th>Trial ID*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Curcumin in Orabase</td>
<td>Ongoing</td>
<td>SVSIDS, India</td>
<td>Oral submucous fibrosis</td>
<td>Reduction of lesion, number of bands</td>
<td>NCT02645656</td>
</tr>
<tr>
<td>Curcumin</td>
<td>Recruiting</td>
<td>University of Arizona, USA</td>
<td>Rheumatoid arthritis</td>
<td>AE, pharmacokinetics, ESR, CRP</td>
<td>NCT02543931</td>
</tr>
</tbody>
</table>

* Source: https://www.clinicaltrials.gov/. A search performed using the keyword “curcumin” revealed 129 studies. Only active studies (48) were included in the table. Clinical trials that were complete (58), were terminated (3), or have unknown status (12) were excluded.

CA: Canada; CCC: Comprehensive Cancer Center; CHU: Clermont-Ferrand University Hospital; HCL: Hospices Civiles de Lyon; NCI: National Cancer Institute; NNFTI: National Nutrition and Food Technology Institute; OSU: Ohio State University; SBUMS: Shahid Beheshti University of Medical Sciences; SHSC: Sunnybrook Health Sciences Centre; UPEI: University of Prince Edward Island; UCLA: University of California, Los Angeles; UT: University of Texas; USA: United States of America.

Nutritional agents (e.g., 5-ASA, 5-FU, 6-mercaptopurine, and bevacizumab) have been used in conjunction with curcumin to identify new therapeutic agents. As in the case with GBM, it is unfortunate that large clinical trials have been undertaken without identifying improvements to the standard of care that was formulated over 10 years ago [100–104]. Clinical trials should be undertaken to corroborate the benefits of curcumin seen in preclinical studies and improve the prognoses of individuals with brain tumors.

Competing Interests

The authors declare that there are no competing interests regarding the publication of this paper.

References


