

Research Article

Ghrelin Protects Human Lens Epithelial Cells against Oxidative Stress-Induced Damage

Jie Bai,¹ Fan Yang,² Li Dong,¹ and Yi Zheng¹

¹Key Laboratory of Harbin Medical University Eye Center, Eye Hospital, First Affiliated Hospital, Harbin Medical University, Harbin, China

²Public Department, Second Affiliated Hospital, Harbin Medical University, Harbin, China

Correspondence should be addressed to Yi Zheng; yizhengkitty@163.com

Received 23 February 2017; Revised 18 April 2017; Accepted 23 April 2017; Published 10 October 2017

Academic Editor: Grzegorz Bartosz

Copyright © 2017 Jie Bai et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Oxidative stress has been recognized as an important mediator in the pathogenesis of age-related cataracts; using antioxidant supplements is one plausible strategy to protect the antioxidative defense system against oxidative stress. Ghrelin administration is expected to reduce ROS, preventing the onset of different diseases. The role of ghrelin, if any, in protecting against oxidative stress in HLECs has never been examined. In the present study, we investigated the effects of ghrelin against H₂O₂-induced oxidative stress and the associated molecular mechanisms in HLECs and rat lenses. The results showed that pretreatment with ghrelin reduced H₂O₂-induced cellular apoptosis and ROS accumulation, increased the expression levels of SOD and CAT, and decreased the expression level of MDA. The morphological examination showed that the ghrelin-treated lens organ culture maintained transparency. This is the first report to show that ghrelin can protect HLECs from H₂O₂-induced oxidative stress. Our findings suggest that ghrelin may prevent the progression of cataracts, which has treatment value for ophthalmologists.

1. Introduction

Oxidative stress has been recognized as an important mediator in the pathogenesis of age-related cataracts; it is widely recognized as a state of imbalance between prooxidants and antioxidants [1, 2]. Excessive production of reactive oxygen species (ROS) plays an important role in the destruction of normal lens epithelial cell function [3, 4]. Hydrogen peroxide (H₂O₂), a nonradical member of the active oxygen family, is a major intracellular ROS and accumulates in substantial amounts in the lens [5]. Several studies have shown that free radicals and ROS can affect the growth and function of human lens epithelial cells (HLECs) [1, 2, 5]. Our previous studies have found that ROS produced by H₂O₂ causes protein degradation and epithelial cell damage—the damage similar to the damage found in human cataracts [6, 7].

Cataracts, which are the leading cause of visual disability globally, are a protein conformational disease characterized by the aggregation of oxidatively damaged proteins [8]. Oxidation of lens proteins is a major risk factor in cataract formation—any external insult or insufficient degradation of damaged proteins may affect the antioxidant status of the lens and cause opacification [1, 2, 7].

Using antioxidant supplements is one plausible strategy to protect the antioxidative defense system against oxidative stress [9]. Although surgery can remedy cataracts, there are numerous postoperative complications; further, some regions lack the necessary surgical instruments and face shortages of doctors who can meet the medical needs of large numbers of patients. These factors limit the availability of cataract surgery. There are no effective therapeutic agents to halt the formation of a cataractous lens; therefore, it is necessary to develop a pharmacological

intervention to improve lens transparency and delay the progression of cataracts.

The growth hormone-releasing peptide ghrelin, a 28-amino-acid endogenous peptide, is secreted primarily from the gastric mucosa [10]. Its transcripts have also been found in the intestine, pancreas, liver, placenta, heart, lungs, central nervous system (CNS), and kidneys, suggesting its extraendocrine as well as endocrine action. Studies have illustrated that ghrelin exhibits favorable cytoprotective effects against oxidative stress [11]. It can remove ROS and reactive nitrogen species (RNSs) by increasing the expression of antioxidant enzymes and directly scavenging free radicals [12]. Ghrelin administration is expected to reduce ROS, preventing the onset of various diseases. The role of ghrelin, if any, in protecting against oxidative stress in HLECs has never been examined. In the present study, we investigated the effects of ghrelin against H₂O₂-induced oxidative stress and the associated molecular mechanisms in HLECs and rat lenses.

2. Material and Methods

2.1. Reagents and Antibodies. Ghrelin was purchased from Sigma Chemical (St. Louis, MO, USA) and dissolved in dimethylsulfoxide (DMSO) (Sigma, St. Louis, MO). Fetal bovine serum (FBS) and Dulbecco's modified Eagle's medium (DMEM) were obtained from Gibco (Grand Island, NY, USA). Medium 199 was obtained from Sigma-Aldrich (St. Louis, MO). Annexin V-FITC and propidium iodide (PI) were obtained from Becton Dickinson (Mountain View, CA, USA). 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) and H₂DCFDA were obtained from Beyotime (Beyotime Institute of Biotechnology, Shanghai, China). Anti-SOD, anti-CAT, and anti-MDA antibodies were purchased from Santa Cruz Biotechnology Inc. (Santa Cruz, CA, USA). SOD, CAT, and MDA colorimetric kits were purchased from Nanjing Jiancheng Bioengineering Institute (Nanjing, China).

2.2. Cell Culture and Treatment. HLECs (ATCC, America) were cultured in DMEM with heat-inactivated (56°C, 0.5h) FBS (15%), 100 U/mL penicillin, and 100 mg/mL streptomycin in humidified 5% CO₂ at 37°C. The cells were routinely subcultured every 2-3 days. When grown to 70% confluence, they were treated with the indicated concentration of H₂O₂ for 24 h or pretreated with different concentrations of ghrelin for 12 h before the H₂O₂ treatment.

2.3. Cell Viability Assay. The cells were plated at a density of 2×10^4 cells/well in a 96-well culture plate and incubated with 0, 50, 100, 200, 400, and 800 μ M H₂O₂ for 24 h alone or after pretreatment with different concentrations of ghrelin (10^{-9} – 10^{-6} M) for 12 h. The culture medium was removed, and cell viability was measured using the MTT method as previously described [13].

The morphological changes of the cells were observed under an inverted microscope (Olympus CK-30, Tokyo, Japan).

2.4. Cell Apoptosis Assay. Annexin V-FITC/PI staining was used to quantify the amount of cell apoptosis. Cells were plated and incubated on a six-well plate at 1×10^6 cells/well and pretreated with or without different concentrations of ghrelin for 12 h, after which they were treated with 100 μ M H₂O₂ for 24 h. The cells were collected and stained with annexin V-FITC/PI in binding buffer at room temperature in the dark for 20 min. The stained cells were then analyzed using a flow cytometry system.

2.5. Fluorescent Staining of Cells with H₂DCFDA. The levels of intracellular ROS were monitored using H₂DCFDA. The cells were incubated in 10 μ M H₂DCFDA for 20 min at 37°C and then washed twice with PBS. The fluorescence intensity of DCF was detected using a fluorescence microscope (Leica DMI 4000, Germany).

2.6. Lens Organ Culture. Rat eyes (Harbin Medical University, Harbin, China) were removed and placed in mammalian physiological saline prewarmed to 37°C. Freshly extracted transparent lenses were incubated in Medium 199 containing 50 mg/mL gentamicin and 0.1% BSA with 5% CO₂ at 37°C. The lenses were treated with ghrelin at concentrations of 10^{-9} – 10^{-6} M for 24 h at 37°C. The medium was changed, and 100 μ M H₂O₂ was added to the medium for 6 h. The lenses were observed under a stereomicroscope and photographed against a background of black gridlines to record the development of opacity.

2.7. Antioxidant Enzyme Content Assay. Whole lenses were removed from the eyes and sonicated in extract buffer. After centrifugation, the supernatants were used for testing according to the assay manufacturer's instructions. Total SOD content was determined spectrophotometrically at 550 nm, and the results were expressed as U·mg⁻¹ protein [14]. MDA was measured with reference to the MDA assay kit using the thiobarbituric acid method [15]. The results were expressed as U·mg⁻¹, and the intensity of the resulting pink color was read at 532 nm. CAT content was assayed using the ammonium molybdate method according to the instructions of the CAT assay kit [16]. The results were expressed as U·mg⁻¹, and the faint yellow complexes were detected at 405 nm.

2.8. Western Blot Analysis. Lenses from each group were washed with cold saline, dried with filter paper, cut with Vannas scissors, and then ground on ice. The tissue was spun down in a refrigerated centrifuge at 1000 r/min for 10 min, and the supernatants were lysed on ice for 20 min in radioimmunoprecipitation assay (RIPA) buffer containing a protease inhibitor cocktail and centrifuged at 12000g for 20 min. The total protein (30 μ g) was subjected to 10–15% SDS-PAGE and transferred into a polyvinylidene difluoride membrane. The blot was incubated with antibodies against SOD, CAT, and MDA. The enzyme was used with a horseradish peroxidase-conjugated secondary antibody. Enhanced chemiluminescence was used to detect the immunoreactive bands, and ImageJ software (image processing and analysis in Java) was used to quantify the results. After normalizing to the individual actine levels,

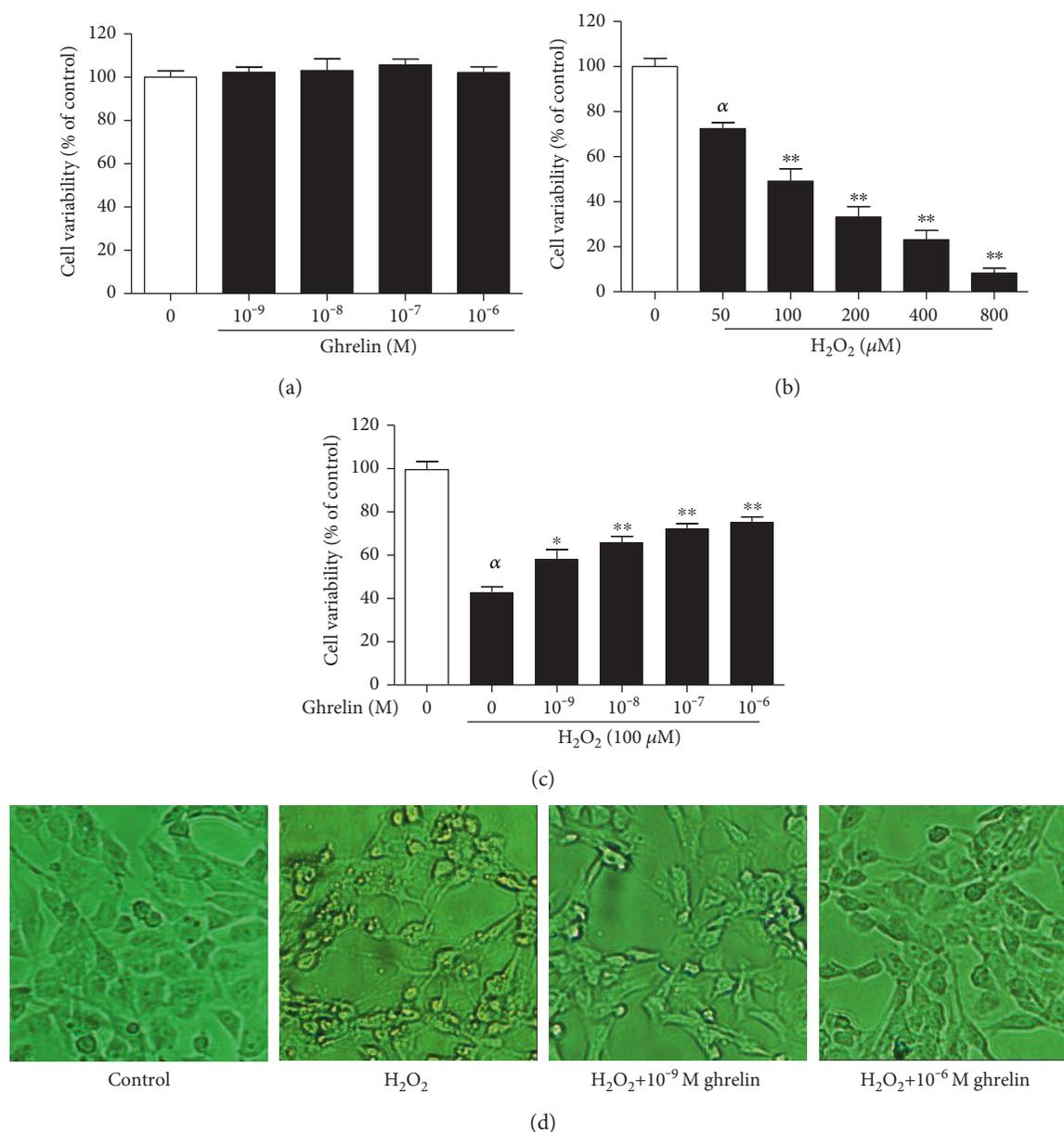


FIGURE 1: The effects of ghrelin on the cell viability of H₂O₂-treated HLECs. (a) HLECs were incubated with different concentrations of ghrelin (10^{-9} – 10^{-6} M) for 24 h. (b) HLECs were incubated with different concentrations of H₂O₂ (50–800 μM) for 24 h. (c) HLECs were preincubated with ghrelin (10^{-9} – 10^{-6} M) for 12 h before being treated with 100 μM H₂O₂ for 24 h. Cell viability was assessed via MTT assay. (d) HLECs were detected by microscopy. The results were represented as the mean ± SEM ($n=3$) from three independent experiments. ^α $P < 0.01$, compared with the untreated control group; ^{*} $P < 0.05$, ^{**} $P < 0.01$, compared with the H₂O₂-treated group.

the ratio of the expression of target proteins was determined. Each experiment was repeated three times.

2.9. Statistical Analysis. Statistical analysis was carried out using GraphPad Prism 5.0 (GraphPad Software, San Diego, CA). All values are expressed as the mean ± standard error of the mean (SEM) from at least three independent experiments. The significance of pairwise group was evaluated using Student's *t*-test. For comparison of more than two groups, one-way ANOVA was used; $P < 0.05$ was considered to be significant.

3. Results

3.1. Effects of H₂O₂ and Ghrelin on HLECs' Viability. As shown in Figure 1(a), ghrelin did not exhibit any cytotoxic effects on HLECs. H₂O₂ impaired cell viability in a dose-dependent manner (Figure 1(b)). Treatment with a concentration of 100 μM H₂O₂ for 24 h was selected for subsequent experiments because it reduced cell viability to approximately 49.12% compared with the control group (cells not treated with H₂O₂). The pretreatment of HLECs with ghrelin showed a dose-dependent protective effect against H₂O₂ damage (Figure 1(c)). Figure 1(d) shows the

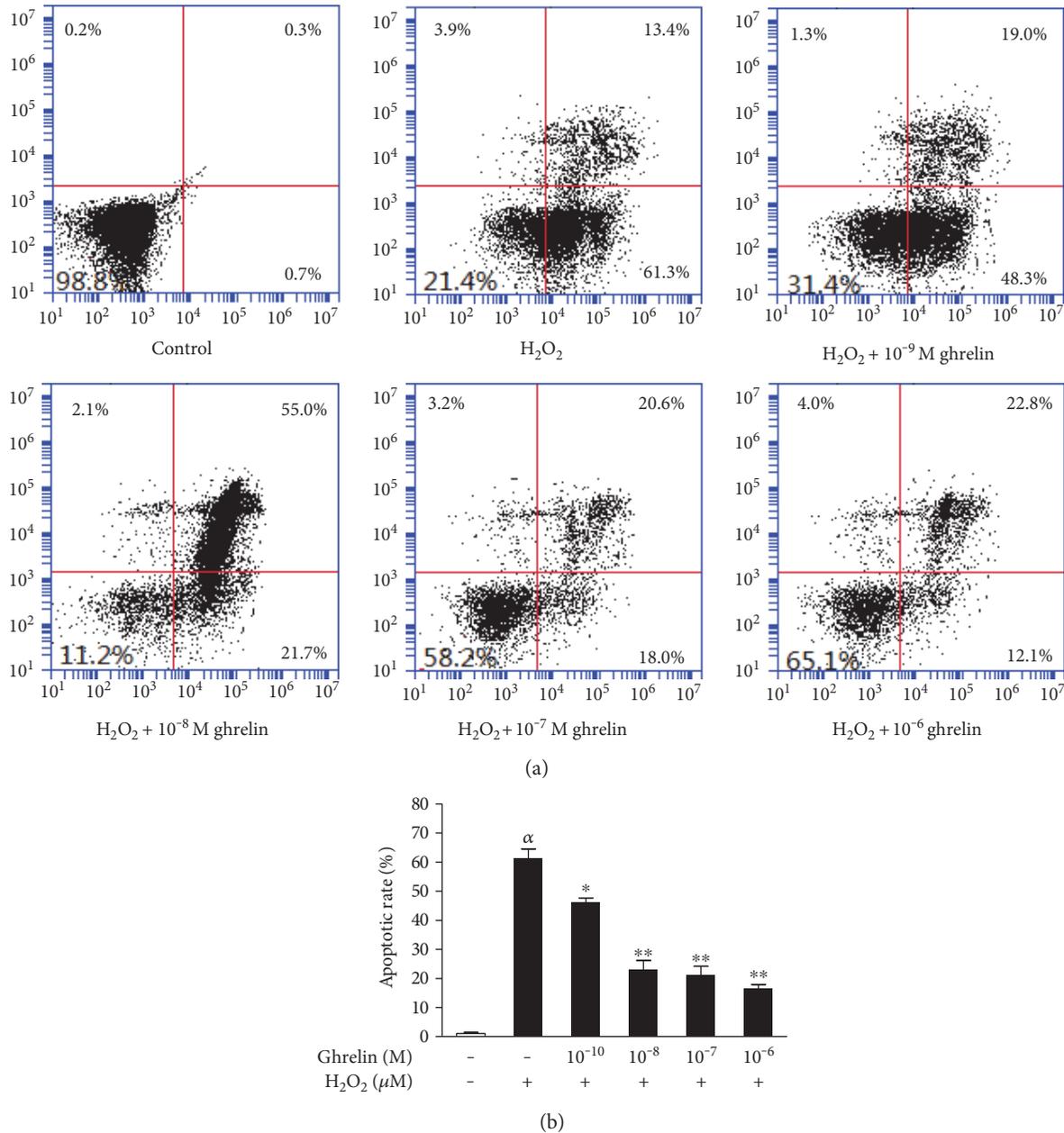


FIGURE 2: Ghrelin prevented H₂O₂-induced cellular apoptosis in HLECs. HLECs were preincubated with ghrelin (10⁻⁹–10⁻⁶ M) for 12 h before being treated with 100 μM H₂O₂ for 24 h. (a) Apoptosis of HLECs as detected by flow cytometry. The results were represented as the mean ± SEM ($n = 3$) from three independent experiments. (b) Ghrelin significantly decreased the apoptosis rate of HLECs. ^α $P < 0.01$, compared with the untreated control group; ^{*} $P < 0.05$, ^{**} $P < 0.01$, compared with the H₂O₂-treated group.

morphological changes of HLECs. The control group showed normal cell morphology and a regular range of cells with intact junctions. The H₂O₂-treated group showed shrinkage of the cells with abnormal shapes, and the distance between cells increased; however, pretreatment with ghrelin inhibited H₂O₂ damage to cell morphology.

3.2. Ghrelin Inhibits Apoptosis Induced by H₂O₂. The rate of cell apoptosis was quantified using flow cytometric analysis by double staining with annexin V-FITC and PI. An increase

of apoptotic cells was observed in the H₂O₂-treated group, while ghrelin pretreatment decreased the apoptosis rate of the cells exposed to H₂O₂ (Figure 2(a)). This positive effect of ghrelin was observed in a concentration-dependent manner (Figure 2(b)).

3.3. Ghrelin Reduced the Generation of ROS in HLECs. As expected, there was a lack of staining in the H₂O₂-free control group (Figure 3). HLECs that were only exposed to H₂O₂ had a light green color, indicating that there was a

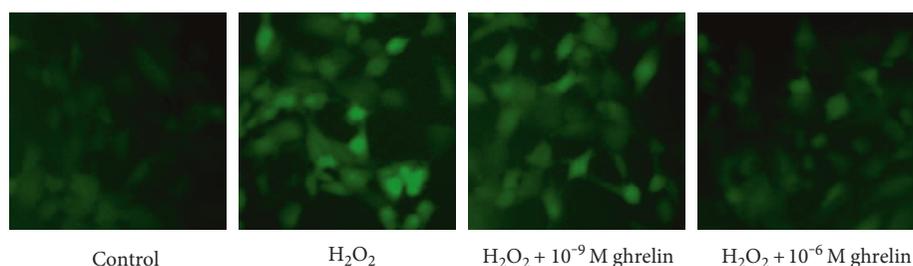


FIGURE 3: Effect of ghrelin on H_2O_2 -induced generation of ROS in HLECs. Cells were treated with $100 \mu M H_2O_2$ for 24 h after incubation in the absence or presence of ghrelin for 12 h. Then, the production of ROS was determined using $10 \mu M H_2DCFDA$. The morphological features of the cells were observed using fluorescence microscopy.

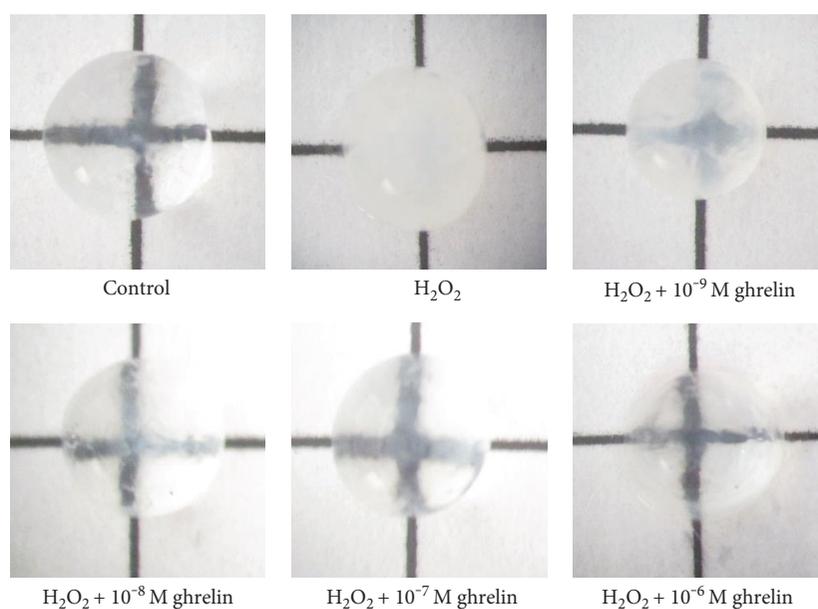


FIGURE 4: Morphological observations confirmed the reduction in opacity when the lenses were incubated in ghrelin. Whole lenses were pretreated with ghrelin at concentrations of 10^{-9} – 10^{-6} M for 24 h and then cultured for 6 h with $100 \mu M H_2O_2$. The control group received no treatment, and the H_2O_2 group was treated only with $100 \mu M H_2O$. Photographs were taken against a background of black gridlines. The higher the concentrations of ghrelin, the less opacity was observed in the lenses.

marked increase in ROS levels. This increase in intracellular ROS was prevented in a dose-dependent manner by pretreatment with ghrelin. This finding indicates that ghrelin can prevent the generation of intracellular ROS in HLECs challenged with H_2O_2 .

3.4. Grading the Lenses. We developed an organ culture experiment to examine the effects of ghrelin on the lenses. Morphological observations confirmed that the lenses incubated in ghrelin showed a reduction in opacity (Figure 4). Lenses in the control group had an absence of opacification, and the gridlines were clearly visible; however, lens opacity measurements showed that $100 \mu M H_2O_2$ induced obvious cataract formation in the lenses. Ghrelin blocked the effect of H_2O_2 in a dose-dependent manner.

3.5. Effects of Ghrelin on the Protein Expression of SOD, CAT, and MDA in Lenses. Western blot was used to measure the levels of SOD, CAT, and MDA in H_2O_2 -stimulated HLECs to detect the antioxidative capability of

ghrelin. Exposure to H_2O_2 markedly decreased the activity of SOD and CAT and increased the MDA content compared with the control group. Pretreatment with ghrelin significantly increased the expression of SOD and CAT, and decreased MDA compared with the H_2O_2 group (Figure 5).

3.6. Effect of Ghrelin on SOD, CAT Activity, and MDA Content in Lenses. The biological activity assay showed that $100 \mu M H_2O_2$ significantly decreased SOD and CAT activity in the lenses, and pretreatment with ghrelin blocked the effect of H_2O_2 . As shown in Table 1, $100 \mu M H_2O_2$ significantly increased MDA content in the lenses. Finally, ghrelin blocked the effect of H_2O_2 in a dose-dependent manner.

4. Discussion and Conclusion

Cataracts are the leading cause of legal blindness worldwide. Until now, there has not been an effective pharmacological agent that can inhibit or reverse the progression of cataracts,

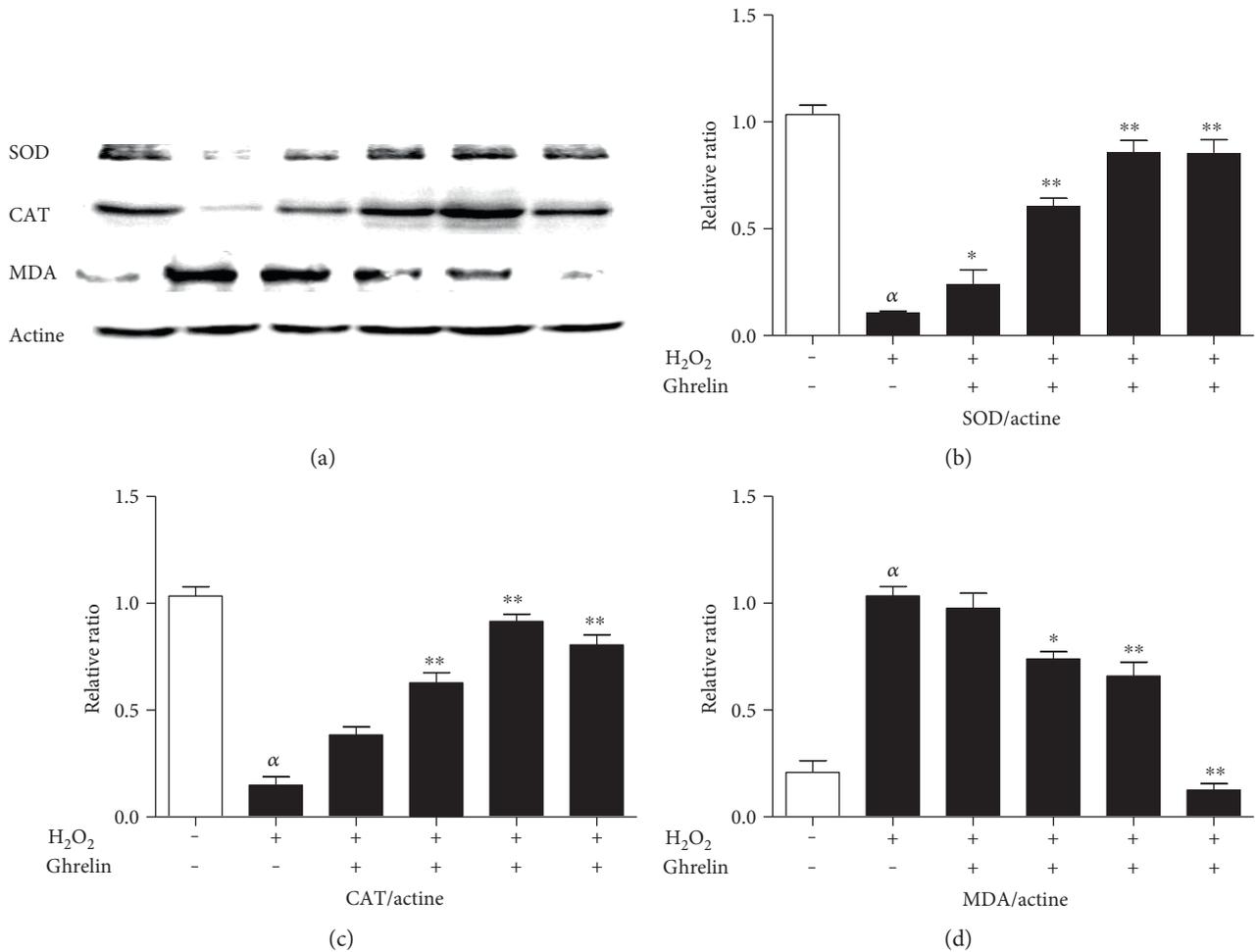


FIGURE 5: Effects of ghrelin on expression of SOD, CAT, and MDA in lenses. (a) The protein levels of SOD, CAT, and MDA were measured using western blot analyses. (b, c, d) The relative protein expression levels of SOD, CAT, and MDA. $\alpha P < 0.05$, compared with the untreated control group; $*P < 0.05$, $**P < 0.01$, compared with the H_2O_2 -treated group.

TABLE 1: Effect of ghrelin on SOD, CAT activity, and MDA content in lenses.

Group	SOD (U·mg ⁻¹)	CAT (U·mg ⁻¹)	MDA (nmol·mg ⁻¹)
Control	27.87 ± 1.57	7.47 ± 1.43	0.44 ± 0.02
H_2O_2	10.27 ± 1.03 ^a	3.34 ± 1.15 ^a	1.83 ± 0.09 ^a
H_2O_2 + 10 ⁻⁹ M ghrelin	15.29 ± 2.11 ^b	4.99 ± 1.23 ^b	0.93 ± 0.04 ^b
H_2O_2 + 10 ⁻⁸ M ghrelin	15.43 ± 2.00 ^b	4.87 ± 1.01 ^b	0.86 ± 0.05 ^b
H_2O_2 + 10 ⁻⁷ M ghrelin	17.03 ± 1.59 ^b	5.34 ± 0.74 ^b	0.77 ± 0.04 ^b
H_2O_2 + 10 ⁻⁶ M ghrelin	19.03 ± 2.08 ^b	5.65 ± 0.81 ^b	0.67 ± 0.07 ^b

Compared with control group ^a $P < 0.05$; compared with the H_2O_2 -treated group ^b $P < 0.05$.

and the search for affordable and nonsurgical pharmacological treatment is necessary to delay the progression of lens opacification [17, 18]. The effects of ghrelin on HLECs and the regulatory mechanism of those effects have never been reported. Therefore, the present study is the first to investigate the effect of ghrelin on H_2O_2 -induced cell injury in HLECs.

Accumulating evidence shows that oxidative stress is a major contributor to the initiation and progression of cataracts. Oxidative stress refers to a state of elevated levels of ROS, with the latter being affected by an intracellular oxidative and antioxidant defense mechanism [19–21]. An increasing level of ROS is a crucial determinant of oxidative damage and impaired cellular function. During oxidative stress, the activity of antioxidant enzymes is inhibited, while the concentration of reactive nitrogen species (RNSs) or ROS increases dramatically. Oxidative stress is involved in the pathogenesis of eye diseases, including age-related cataracts and macular degeneration [19, 22]. Considering that oxidative stress plays an important role in the pathogenesis of cataracts, reducing oxidative stress is a plausible potential therapeutic target for cataracts.

Ghrelin has several biological actions, including regulating cell survival and proliferation, inhibiting inflammation, and exerting antioxidative effects [23, 24]. Previous studies have indicated that the antioxidative effect of ghrelin is based on increasing the activity of endogenous antioxidant enzymes [25]. For example, ghrelin has been reported to alleviate SAH-induced oxidative brain damage by maintaining a

balance in oxidant-antioxidant status [26]. Ghrelin has also been found to significantly reduce the protein expression of iNOS and increase the expression of CuZnSOD, MnSOD, CAT, and GPx in the liver [27]. Ghrelin is an amino acid peptide—it is safe for humans, as has been shown in many studies [28]. Our study also showed that ghrelin exhibited no obvious cytotoxicity in HLECs and pretreatment with ghrelin inhibited H₂O₂ damage to cell morphology. Considering its lack of toxicity and its excellent antioxidative effect, we propose that ghrelin should be considered as a prophylaxis for preserving visual function and could be used to treat age-related cataracts.

HLECs are a single layer of epithelial cells on the lens' anterior surface. The normal construction and function of HLECs is crucial for the maintenance of the transparency and metabolic homeostasis of the entire lens. Once damaged, they cannot self-renew, and they become permanently impaired. Oxidative stress could increase the permeability of HLECs, causing dysfunction. Studies have indicated that oxidants, especially H₂O₂, could trigger lens epithelial cell apoptosis and initiate early cataract formation [6, 7, 29]. Once the lens' defense system is weakened, cataracts begin to form.

The antioxidant systems protect cells against oxidative damage during normal metabolism and after an oxidative insult; they contain numerous antioxidant enzymes such as SOD, CAT, and glutathione peroxidase (GPx). These antioxidative enzymes can protect the lens from oxidative stress and maintain lens clarity [30]. H₂O₂-induced cataracts were associated with decreases in SOD and CAT activity and increase in MDA activity in the lens. These three enzymes are essential in oxidative stress protection and normal lens metabolism (SOD, CAT, and MDA). The antioxidant enzymes are able to catalytically remove free radicals and other reactive species. SOD can prevent lipid peroxidation, scavenge ROS, and protect cells from the damaging effects of toxic oxygen radicals [31]. CAT reduces H₂O₂ to water; a decline in the level of CAT weakens the antioxidant capacity of the lens epithelial cells and induces their apoptosis, which causes cataracts [32]. MDA is one of the metabolic products of lipid peroxides (LPO) and is well known as a widely used marker for oxidative stress [33].

Our *in vitro* test has verified the effect of ghrelin in preventing oxidative stress-induced cell dysfunction; however, can ghrelin also be effective for lens tissue? To answer this, we present the results of an *ex vivo* study to assess the anticataract potential of ghrelin in H₂O₂-induced isolated rat lenses through observation of lens transparency and estimation of some biochemical parameters such as SOD, CAT, and MDA contents.

Western blot results showed increased protein expression of SOD and CAT and a decreased expression of MDA under ghrelin treatment. Antioxidant enzyme content assays showed similar results; the mean content of SOD and CAT significantly decreased in the H₂O₂-treated lenses compared with the control lenses. In the ghrelin-treated group, the mean content of antioxidant enzymes was restored compared with the lenses in the H₂O₂-treated group. The elevated content of MDA in the H₂O₂-treated lenses may account for the disruption of membrane lipids. In addition, the reduction of

the MDA level in ghrelin-treated group suggests that ghrelin may have prevented the disruption of lenticular membrane lipids, thereby impeding opacification of the lens.

In conclusion, the data from our experiments demonstrated that ghrelin effectively retarded H₂O₂-induced cataract formation. As an antioxidant agent, ghrelin increased the levels of SOD and CAT and decreased the level of MDA, thus sustaining lens transparency. Furthermore, ghrelin is an amino acid and should therefore be safe for use in humans. Due to its safety and efficacy, ghrelin has potentially important implications for the prevention of cataractogenesis.

Abbreviations

ROS:	Reactive oxygen species
HLECs:	Human lens epithelial cells
H ₂ O ₂ :	Hydrogen peroxide
SOD:	Superoxide dismutase
CAT:	Catalase
MDA:	Malondialdehyde
MTT:	3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide.

Conflicts of Interest

The authors declare that they have no financial competing interests.

Acknowledgments

This work was supported by the grants from Heilongjiang Postdoctoral Fund (LBH-Z14161), scientific research funds of the First Affiliated Hospital of Harbin Medical University (2016B001), Heilongjiang Postdoctoral Scientific Launch Research (LBH-Q16177), and scientific research innovation projects of Harbin Medical University (2016LCZ39).

References

- [1] D. Škiljić, S. Nilsson, A. Petersen et al., "Oestradiol levels and superoxide dismutase activity in age-related cataract: a case-control study," *BMC Ophthalmology*, vol. 16, no. 1, pp. 210–215, 2016.
- [2] N. Pescosolido, A. Barbato, R. Giannotti, C. Komaiha, and F. Lenarduzzi, "Age-related changes in the kinetics of human lenses: prevention of the cataract," *International Journal of Ophthalmology*, vol. 9, no. 10, pp. 1506–1517, 2016.
- [3] P. Rwei, C. S. Alex Gong, L. J. Luo, M. B. Lin, J. Y. Lai, and H. L. Liu, "In vitro investigation of ultrasound-induced oxidative stress on human lens epithelial cells," *Biochemical and Biophysical Research Communications*, vol. 482, no. 4, pp. 954–960, 2017.
- [4] Y. Yu, H. Jiang, H. Li, W. Song, and X. Xia, "Alpha-A-crystallin protects lens epithelial cell-derived iPSC-like cells against apoptosis induced by oxidative stress," *Cellular Reprogramming*, vol. 18, no. 5, pp. 327–332, 2016.
- [5] T. Ma, T. Chen, P. Li et al., "Heme oxygenase-1 (HO-1) protects human lens epithelial cells (SRA01/04) against hydrogen peroxide (H₂O₂)-induced oxidative stress and apoptosis," *Experimental Eye Research*, vol. 146, pp. 318–329, 2016.

- [6] Y. Zheng, Y. Liu, J. Ge et al., "Resveratrol protects HLECs against H₂O₂-induced oxidative stress by increasing catalase, SOD-1, and HO-1 expression," *Molecular Vision*, vol. 16, pp. 1467–1474, 2010.
- [7] Z. Jia, Z. Song, Y. Zhao, X. Wang, and P. Liu, "Grape seed proanthocyanidin extract protects human lens epithelial cells from oxidative stress via reducing NF- κ B and MAPK protein expression," *Molecular Vision*, vol. 17, pp. 210–217, 2011.
- [8] C. Xie, X. Li, J. Tong, G. Yangshun, and Y. Shen, "Effects of white light-emitting diode (LED) light exposure with different correlated color temperatures (CCTs) on human lens epithelial cells in culture," *Photochemistry and Photobiology*, vol. 90, no. 4, pp. 853–859, 2014.
- [9] K. Yao, L. Zhang, Y. D. Zhang, P. P. Ye, and N. Zhu, "The flavonoid, fisetin, inhibits UV radiation-induced oxidative stress and the activation of NF- κ B and MAPK signaling in human lens epithelial cells," *Molecular Vision*, vol. 14, pp. 1865–1871, 2008.
- [10] H. H. Huang, C. H. Ting, Y. F. Syu, S. C. Chang, and C. Y. Chen, "Correlation between colonic secretion and colonic motility in rats: role of ghrelin," *World Journal of Gastroenterology*, vol. 22, no. 46, pp. 10140–10147, 2016.
- [11] N. Sun, H. Wang, and L. Wang, "Protective effects of ghrelin against oxidative stress, inducible nitric oxide synthase and inflammation in a mouse model of myocardial ischemia/reperfusion injury via the HMGB1 and TLR4/NF- κ B pathway," *Molecular Medicine Reports*, vol. 14, no. 3, pp. 2764–2770, 2016.
- [12] Y. Hao, C. Liu, J. Huang et al., "Ghrelin protects against depleted uranium-induced apoptosis of MC3T3-E1 cells through oxidative stress-mediated p38-mitogen-activated protein kinase pathway," *Toxicology and Applied Pharmacology*, vol. 1, no. 290, pp. 116–125, 2016.
- [13] J. Bai, Y. Zheng, G. Wang, and P. Liu, "Protective effect of D-limonene against oxidative stress-induced cell damage in human lens epithelial cells via the p38 pathway," *Oxidative Medicine and Cellular Longevity*, vol. 2016, Article ID 5962832, 12 pages, 2016.
- [14] J. Pedraza-Chaverri, P. D. Maldonado, O. N. Medina-Campos et al., "Garlic ameliorates gentamicin nephrotoxicity: relation to antioxidant enzymes," *Free Radical Biology and Medicine*, vol. 29, no. 7, pp. 602–611, 2016.
- [15] M. C. Lima, G. Marks, I. S. Silva, B. A. Silva, L. Z. C nsolo, and G. B. Nogueira, "Evaluation of oxidative stress in mice subjected to aerobic exercise," *Acta Cir rgica Brasileira*, vol. 27, no. 8, pp. 544–551, 2012.
- [16] J. Pedraza-Chaverri, M. D. Granados-Silvestre, O. N. Medina-Campos, P. D. Maldonado, I. M. Olivares-Corichi, and M. E. Ibarra-Rubio, "Post-transcriptional control of catalase expression in garlic-treated rats," *Molecular and Cellular Biochemistry*, vol. 216, no. 1-2, pp. 9–19, 2001.
- [17] B. X. Lim, C. H. Lim, D. K. Lim, J. R. Evans, C. Bunce, and R. Wormald, "Prophylactic non-steroidal anti-inflammatory drugs for the prevention of macular oedema after cataract surgery," *Cochrane Database of Systematic Reviews*, vol. 11, article CD006683, 2016.
- [18] C. Skinner and V. Miraldi Utz, "Pharmacological approaches to restoring lens transparency: real world applications," *Ophthalmic Genetics*, vol. 38, no. 3, pp. 201–205, 2017.
- [19] M. A. Babizhayev, "Generation of reactive oxygen species in the anterior eye segment. Synergistic codrugs of N-acetylcarnosine lubricant eye drops and mitochondria-targeted antioxidant act as a powerful therapeutic platform for the treatment of cataracts and primary open-angle glaucoma," *BBA Clinical*, vol. 19, no. 6, pp. 49–68, 2016.
- [20] A. Spector, G. M. Wang, R. R. Wang, W. C. Li, and J. R. Kuszak, "A brief photochemically induced oxidative insult causes irreversible lens damage and cataract. I. Transparency and epithelial cell lacer," *Experimental Eye Research*, vol. 60, no. 5, pp. 471–481, 1995.
- [21] N. Nagai, Y. Ito, and N. Takeuchi, "Correlation between hyper-sensitivity to hydrogen peroxide and low defense against Ca²⁺ influx in cataractogenic lens of Ihara cataract rats," *Biological & Pharmaceutical Bulletin*, vol. 34, no. 7, pp. 1005–1010, 2011.
- [22] H. Cho, M. Kwon, H. Jang, J. B. Lee, K. C. Yoon, and S. C. Jun, "Herbal extracts that reduce ocular oxidative stress may enhance attentive performance in humans," *Computational Intelligence and Neuroscience*, vol. 2016, Article ID 4292145, 13 pages, 2016.
- [23] N. Sun, H. Wang, L. Ma, P. Lei, and Q. Zhang, "Ghrelin attenuates brain injury in septic mice via PI3K/Akt signaling activation," *Brain Research Bulletin*, vol. 124, pp. 278–285, 2016.
- [24] M. Koyuturk, O. Sacan, S. Karabulut et al., "The role of ghrelin on apoptosis, cell proliferation and oxidant-antioxidant system in the liver of neonatal diabetic rats," *Cell Biology International*, vol. 39, no. 7, pp. 834–841, 2015.
- [25] X. X. Tong, D. Wu, X. Wang et al., "Ghrelin protects against cobalt chloride-induced hypoxic injury in cardiac H9c2 cells by inhibiting oxidative stress and inducing autophagy," *Pepptides*, vol. 38, no. 2, pp. 217–227, 2012.
- [26] M. Er ahin, H. Z. Toklu, C. Erzik et al., "The anti-inflammatory and neuroprotective effects of ghrelin in subarachnoid hemorrhage-induced oxidative brain damage in rats," *Journal of Neurotrauma*, vol. 27, no. 6, pp. 1143–1155, 2010.
- [27] B. Dobutovic, E. Sudar, S. Tepavcevic et al., "Effects of ghrelin on protein expression of antioxidative enzymes and iNOS in the rat liver," *Archives of Medical Science*, vol. 10, no. 4, pp. 806–816, 2014.
- [28] Z. C. Luo, J. F. Bilodeau, A. M. Nuyt et al., "Perinatal oxidative stress may affect fetal ghrelin levels in humans," *Scientific Reports*, vol. 8, no. 5, article 17881, 2015.
- [29] Y. Tamada, C. Fukiage, Y. Nakamura, M. Azuma, Y. H. Kim, and T. R. Shearer, "Evidence for apoptosis in the selenite rat model of cataract," *Biochemical and Biophysical Research Communications*, vol. 275, no. 2, pp. 300–306, 2000.
- [30] D. Chang, X. Zhang, S. Rong et al., "Serum antioxidative enzymes levels and oxidative stress products in age-related cataract patients," *Oxidative Medicine and Cellular Longevity*, vol. 2013, Article ID 587826, 7 pages, 2013.
- [31] C. X. Yang, H. Yan, and T. B. Ding, "Hydrogen saline prevents selenite-induced cataract in rats," *Molecular Vision*, vol. 19, pp. 1684–1693, 2013.
- [32] X. Wang, J. Sun, G. F. Dang, Y. Gao, L. Duan, and X. Y. Wu, "Antioxidant content and cytological examination of aqueous fluid from patients with age-related cataracts at different stages," *Genetics and Molecular Research*, vol. 14, no. 2, pp. 6251–6255, 2015.
- [33] H. Omrani, M. R. Alipour, and G. Mohaddes, "Ghrelin improves antioxidant defense in blood and brain in normobaric hypoxia in adult male rats," *Advanced Pharmaceutical Bulletin*, vol. 5, no. 2, pp. 283–288, 2015.



Hindawi
Submit your manuscripts at
<https://www.hindawi.com>

