Review Article
Salvianolic Acid Exerts Cardioprotection through Promoting Angiogenesis in Animal Models of Acute Myocardial Infarction: Preclinical Evidence

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Received 27 February 2017; Revised 9 April 2017; Accepted 24 April 2017; Published 21 June 2017

Academic Editor: Pei Luo

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Radix Salviae miltiorrhize, danshen root (danshen), is one of the widely used Chinese herbal medicines in clinics, containing rich phenolic compounds. Salvianolic acid is the main active compound responsible for the pharmacologic effects of danshen. Here, we aimed to evaluate the effects of salvianolic acid on cardioprotection through promoting angiogenesis in experimental myocardial infarction. Studies of salvianolic acid in animal models of myocardial infarction were obtained from 6 databases until April 2016. The outcome measures were vascular endothelium growth factor (VEGF), blood vessel density (BVD), and myocardial infarct size. All the data were analyzed using Rev-Man 5.3 software. Ultimately, 14 studies were identified involving 226 animals. The quality score of studies ranged from 3 to 6. The meta-analysis of six studies showed significant effects of salvianolic acid on increasing VEGF expression compared with the control group (P < 0.01). The meta-analysis of the two salvianolic acid A studies and three salvianolic acid B studies showed significantly improving BVD compared with the control group (P < 0.01). The meta-analysis of five studies showed significant effects of salvianolic acid for decreasing myocardial infarct size compared with the control group (P < 0.01). In conclusion, these findings demonstrated that salvianolic acid can exert cardioprotection through promoting angiogenesis in animal models of myocardial infarction.

1. Introduction

Ischemic heart disease (IHD) remains the leading cause of death worldwide [1]. According to the World Health Organization report, 740 million people die of IHD annually all around the world, accounting for the death of 1.32% of the total population [2]. Myocardial infarction (MI) is one of the main manifestation of IHD, which makes myocardial necrosis or apoptosis in a short time [3], leading to heart failure with a poor prognosis [4]. It has been ranked as the leading cause of death in IHD [5].

In recent years, there are many types of treatments for MI, such as reducing incidence of coronary atherosclerosis [6], antithrombotic therapy including vitamin K antagonists [7], antiplatelet therapy with low-dose aspirin [8], and clopidogrel [9]. In addition, invasive vascular reconstruction is widely used, which improves coronary perfusion, such as percutaneous coronary intervention (PCI) and coronary artery bypass grafting (CABG) [10]. In the short term, clinical interventions and treatments of MI have achieved positive effectiveness [11]. However, the side effect of drugs such as lipid-lowering drugs leading to skeletal muscle, metabolic and neurological adverse events [12], antithrombotic therapy [13] and/or anti-platelet therapy [14] leading to bleeding, and the high incidence rate of restenosis or stent thrombosis limits the long-term success of treatment [15]. Thus, the promotion of therapeutic angiogenesis as a new treatment strategy has been proposed. Angiogenesis appears in all
vascularized organs during the whole embryonic development stage, formatting of new blood vessels from pre-existing ones [16]. Although ischemia leads to endogenous myocardial angiogenesis, it cannot reach the effect to maintain normal capillary density [17]. Therefore, therapeutic stimulation of angiogenesis has been regarded as an effective treatment for myocardial ischemia [18].

Radix *Salviae miltiorrhizae*, danshen root (danshen), the dried root of *Salvia miltiorrhiza* Bge., known as a popular traditional Chinese herbal medicine, has been widely used and well received for the treatment of coronary artery diseases, such as angina pectoris and MI [19]. Salvianolic acid is the main active compound responsible for the pharmacologic effects of danshen [20] and exerts the significant cardiovascular protection [21]. Currently, various studies have indicated its significant function of promoting angiogenesis [22].

The use of preclinical systematic review can more systematically evaluate the efficacy, identify an area for testing in further animal experiments, provide reliable information about the drugs study, and list the base of future clinical research [23]. However, currently, there is no systematic review in this area. Thus, the aim of this study is to evaluate the effects of salvianolic acid on cardioprotection through promoting angiogenesis in animal experiments of MI.

2. Methods

2.1. Search Strategies. We searched studies of salvianolic acid in animal models of acute myocardial infarction from PubMed, EMBASE, Chinese National Knowledge Infrastructure (CNKI), VIP information database, and Wanfang Data information site from inception to April 2016. The search term used was “danshen OR *Salvia miltiorrhiza* OR Salvianolic acid OR Daiclzein” AND “myocardial infarction OR Myocardial Ischemia OR myocardial ischemia OR myocardial infarct OR myocardial stems.” All the research objects were limited to animals.

2.2. Inclusion/Exclusion Criteria. We included studies about the effect of salvianolic acid on animal models with myocardial infarction, in which the outcome measures were vascular endothelium growth factor (VEGF) and/or blood vessel density (BVD). To prevent bias, inclusion criteria were pre-specified as follows: (1) acute myocardial infarction (AMI) experimental model was induced by ligating of the left anterior descending coronary artery (LAD); (2) experimental drug was Salvianolic acid; and VEGF and/or BVD (3) is the primary outcome measurement and (4) is compared with control animal models receiving saline or no treatment. Prespecified exclusion criteria were treatment with single danshen or danshen-based prescription, a nonmyocardial infarct model, no control group, and duplicate publications.

2.3. Data Extraction. Two authors independently extracted data as follows: (1) publication year and the first author’s name; (2) the information of experimental animals including number, species, sex, weight, and age; (3) a model of myocardial infarction; (4) the time of giving experimental drug; (5) the type and the administration methods of anesthetic; (6) the characteristics of treatment used in the experimental group containing the types of salvianolic acid, administration method, and duration of treatment; (7) the primary outcome measures, other outcome measures, and timing for outcome assessments; and (8) side effect. If there were many different time point outcomes, only the last was recorded. Likewise, if the experimental animals received different doses of the drug, only the highest dose was recorded. If the primary data were incomplete, further information was retrieved by contacting with authors. For each comparison, we extracted the mean value and standard deviation from each experimental and control group of every study. Discrepancies were resolved after discussion between the two authors.

2.4. Quality Assessment. We evaluated the methodological quality of the included studies using the ten-item scale [24] with minor modification as follows: (a) peer-reviewed publication; (b) control of temperature; (c) random allocation to treatment or control; (d) blinded induction of model; (e) blinded assessment of outcome; (f) use of anesthetic without significant intrinsic vascular protection activity; (g) appropriate animal model (aged, diabetic, or hypertensive); (h) sample size calculation; (i) compliance with animal welfare regulations; and (j) statement of potential conflict of interests. Every item was given one point. Two authors independently evaluated the study quality, and the final result was identified by discussion when countering disagreement.

2.5. Statistical Analysis. All the data of VEGF and BVD were considered as continuous data, and then, we used the standard mean difference (SMD) with the random effect model to assess the comprehensive results, because of the heterogeneity between multistudies. Then we utilized *I*² statistic to estimate heterogeneity. The significance of differences between *n* groups was estimated by partitioning heterogeneity and by using the *x*² distribution with *n*–1 degrees of freedom (df), where *n* equals the number of groups. The publication bias was expressed by a funnel plot. Probability values of 0.05 were considered significant. We utilized RevMan version 5.3 to carry out the meta-analysis.

3. Results

3.1. Study Inclusion. We searched 573 potentially relevant studies, and 315 were excluded because of duplication. After screening titles and abstracts, 73 studies were excluded because of a nonanimal study, clinical trial, case report, comments, or review. By reading the full text of the remaining articles, 135 studies were excluded because of at least one of the following reasons: (1) the outcome measures did not include VEGF and/or BVD; (2) nonmyocardial infarct model; (3) treatment with single danshen or danshen-based prescriptions; (4) no control group; and (5) duplicate publications. Ultimately, 14 eligible studies were included in qualitative synthesis and 11 eligible studies [25–38] in quantitative synthesis (Figure 1).

3.2. Study Characteristics. Fourteen studies with 226 animals were included. All the studies were published between 2004 and 2016, including four studies [25–27, 38] in English and
seven studies [29, 32–37], two online PhD theses [30, 31], and one online Master’s thesis [28] in Chinese. A male/female Sprague Dawley rat, male Wistar rat, and male piglet model were used in 12 studies [25–31, 33, 34, 36–38], 1 study [35], and 1 study [32], respectively. All the myocardial infarction models were produced by ligation of the LAD. Twelve studies used blood vessel density (BVD) [26–35, 37, 38], ten studies [25–29, 31, 33, 34, 36, 38] used VEGF, and eight studies [25–27, 30, 31, 34, 35, 37] utilized myocardial infarct size as outcome measures. Anesthetic was reported in 13 studies, including pentobarbital (n = 3) [30, 34, 35], urethane (n = 2) [26, 28], chloral hydrate (n = 4) [25, 29, 31, 33], ether (n = 2) [36, 37], ketamine and diazepam (n = 1) [32], and hydrochloride (n = 1) [27], whereas one study [38] did not report the anesthetic. Seven studies [25, 26, 30, 34, 35, 37, 38] used salvianolic acid B, two studies [27, 31] used salvianolic acid A, and five studies [28, 29, 32, 33, 36] used mixed salvianolic acids. There were three administration methods, including intragastric administration (n = 7) [26, 28, 30, 33, 35–37], intravenous administration (n = 5) [25, 27, 31, 32, 34], and intraperitoneal administration (n = 1) [29]. The characteristics of the included studies are concluded in Table 1.

3.3. Study Quality. The quality score of studies ranged from 3 to 6. All studies were publications in a peer-reviewed journal or thesis. Five studies reported control of room temperature. All studies described random allocation to the groups. Thirteen studies used anesthetic without significant intrinsic vascular protection activity. No studies described a sample size calculation. Four studies reported a compliance with animal welfare regulations, and five studies mentioned a statement of potential conflict of interests. None of the studies described masked induction of appropriate animal models (aged, diabetic, or hypertensive). The methodological quality is concluded in Table 2.

3.4. Effectiveness

3.4.1. VEGF. Ten studies [25–29, 31, 33, 34, 36, 38] utilized VEGF as the outcome measure. The meta-analysis of six studies [26, 27, 29, 31, 34, 36] showed significant effects of salvianolic acid for increasing VEGF expression compared with the control group (n = 101, SMD 2.02, 95% CI: 1.45–2.59, P < 0.00001; heterogeneity χ² = 5.70, P = 0.34, I² = 12%), Figure 2. The subgroup analysis showed that
<table>
<thead>
<tr>
<th>Study (years)</th>
<th>Species (sex, n = experimental/control group)</th>
<th>Weight</th>
<th>Model (method)</th>
<th>Time drug given</th>
<th>Anesthetic</th>
<th>Treatment group and administration methods</th>
<th>Control group</th>
<th>Angiogenesis outcome index</th>
<th>Intergroup differences (time)</th>
<th>Secondary outcome</th>
<th>Intergroup differences</th>
<th>Side effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chao et al. [25]</td>
<td>Male Sprague Dawley rats (8/8)</td>
<td>200–220 g</td>
<td>AMI</td>
<td>1 day after the surgery</td>
<td>Chloral hydrate [0.3 g/kg, i.p.], Salvinolic acid B 24 mg/kg·4 h, i.v.</td>
<td>Normal saline</td>
<td>(1) VEGF</td>
<td>(1) $P &lt; 0.05$; 8 h</td>
<td>(1) Myocardial infarct size</td>
<td>(1) $P &lt; 0.001$; 8 h</td>
<td>No report</td>
<td></td>
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<tr>
<td>He et al. [26]</td>
<td>Male Sprague Dawley rats (15/15)</td>
<td>200–220 g</td>
<td>AMI</td>
<td>24 h after the surgery</td>
<td>Urethane [1.2 g/kg, intraperitoneally (i.p.)], Salvinolic acid B 100 mg/kg·d, i.g.</td>
<td>Normal saline</td>
<td>(2) BVD</td>
<td>(2) $P &lt; 0.05$; 4 w</td>
<td>(2) LVSP</td>
<td>(2) $P &lt; 0.01$; 4 w</td>
<td>No report</td>
<td></td>
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<tr>
<td>Li et al. [27]</td>
<td>Male Sprague Dawley rats (5/5)</td>
<td>200–220 g</td>
<td>AMI</td>
<td>24 hours after the surgery</td>
<td>3.5% hydrochloride [3.5 mg/100 g, i.p.], 10 mg/kg/d salvianolic acid A i.v.</td>
<td>Normal saline</td>
<td>(1) VBD</td>
<td>(1) $P &lt; 0.05$; 7 d</td>
<td>(1) Myocardial infarct size</td>
<td>(1) $P &lt; 0.01$; 1 w</td>
<td>No report</td>
<td></td>
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<tr>
<td>Yang [28]</td>
<td>Male Sprague Dawley rats (8/8)</td>
<td>180–200 g</td>
<td>AMI</td>
<td>1 week after the surgery</td>
<td>Urethane [1.2 g/kg, i.p.], Salvia extract (salvianolic acid B 20.6%, Danshensu 23.6%, protocatechuic acid 3.9%) 100 mg/kg·d, i.g.</td>
<td>Normal saline</td>
<td>(1) VEGF</td>
<td>(1) $P &lt; 0.05$; 60 d</td>
<td>(1) LVSP</td>
<td>(1) $P &lt; 0.05$; 60 d</td>
<td>No report</td>
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<tr>
<td>Fang [29]</td>
<td>Male Sprague Dawley rats (13/13)</td>
<td>220–260 g</td>
<td>AMI</td>
<td>3 days after the surgery</td>
<td>10% chloral hydrate [4 ml/kg, i.p.], Salvianolate (main composition salvianolic acid B) 30 mg/kg·d, i.p.</td>
<td>Normal saline</td>
<td>(1) VBD</td>
<td>(1) $P &lt; 0.01$; 4 w</td>
<td>(2) VEGF</td>
<td>(2) $P &lt; 0.01$; 4 w</td>
<td>No report</td>
<td></td>
</tr>
<tr>
<td>Ma [30]</td>
<td>Male Sprague Dawley rats (10/10)</td>
<td>Three months</td>
<td>AMI</td>
<td>No report</td>
<td>3% pentobarbital 45 mg/kg, i.p.</td>
<td>Salvianolic acid B 1 g/kg·d, i.g.</td>
<td>Normal saline</td>
<td>(1) BVD</td>
<td>(1) $P &lt; 0.01$; 14 d</td>
<td>(1) Myocardial infarct size</td>
<td>(1) $P &lt; 0.01$; 14 d</td>
<td>No report</td>
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<tr>
<td>Li [31]</td>
<td>Male Sprague Dawley rats (5/5)</td>
<td>200–220 g</td>
<td>AMI</td>
<td>1 day after the surgery</td>
<td>3.5% chloral hydrate [35 g/kg, i.p.], Salvinolic acid A 10 mg/kg, i.v.</td>
<td>Normal saline</td>
<td>(2) VEGF</td>
<td>(2) $P &lt; 0.001$; 1 w</td>
<td>(2) SDF-1</td>
<td>(2) $P &lt; 0.01$; 1 w</td>
<td>No report</td>
<td></td>
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<tr>
<td>Wang [32]</td>
<td>Male piglets (6/6)</td>
<td>28 ± 10 kg</td>
<td>AMI</td>
<td>1 day after the surgery</td>
<td>Ketamine, 20 mg/kg, and diazepam, 0.05 mg/kg, i.m.</td>
<td>Salvianolate (main composition salvianolic acid B) 400 mg, i.v.</td>
<td>Normal saline</td>
<td>(1) BVD</td>
<td>(1) $P &lt; 0.01$; 4 w</td>
<td>(1) LVSP</td>
<td>(1) $P &lt; 0.05$; 4 w</td>
<td>No report</td>
</tr>
<tr>
<td>Nuan-Liu [33]</td>
<td>Male Sprague Dawley rats (8/8)</td>
<td>200–240 g</td>
<td>AMI</td>
<td>2 days after the surgery</td>
<td>10% chloral hydrate i.p.</td>
<td>Salvia extract (main composition salvianolic acid B) 40 mg/kg·d, i.g.</td>
<td>Normal saline</td>
<td>(1) VEGF</td>
<td>(1) $P &lt; 0.01$; 4 w</td>
<td>(2) BVD</td>
<td>(2) $P &lt; 0.01$; 4 w</td>
<td>No report</td>
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<tr>
<td>Study (years)</td>
<td>Species (sex, n = experimental/control group)</td>
<td>Weight</td>
<td>Model (method)</td>
<td>Time drug given</td>
<td>Anesthetic</td>
<td>Treatment group and administration methods</td>
<td>Control group</td>
<td>Angiogenesis outcome index</td>
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<td>Secondary outcome</td>
<td>Intergroup differences</td>
<td>Side effect</td>
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<tr>
<td>Chen [34]</td>
<td>Male/female Sprague Dawley rats (8/8)</td>
<td>180–220 g</td>
<td>AMI</td>
<td>1 day after the surgery</td>
<td>3% pentobarbital 30 mg/kg i.v.</td>
<td>Salvinolic acid B 6.4 mg, i.v.</td>
<td>Normal saline</td>
<td>(1) VEGF</td>
<td>(1) $P &lt; 0.01$; 2 w</td>
<td>(1) Myocardial infarct size</td>
<td>(1) $P &lt; 0.01$; 2 w</td>
<td>(2) NO</td>
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<tr>
<td>Fang [35]</td>
<td>Male/female Wistar rats (10/10)</td>
<td>250 ± 50 g</td>
<td>AMI</td>
<td>6 days before the surgery</td>
<td>3% pentobarbital 30 mg/kg, i.p.</td>
<td>Salvinolic acid B 100 mg/(kg·d), i.g.</td>
<td>Normal saline</td>
<td>(1) BVD</td>
<td>(1) $P &lt; 0.05$; 6 d</td>
<td>(1) Myocardial infarct size</td>
<td>(1) $P &lt; 0.01$; 6 d</td>
<td>(2) Fibroblast</td>
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<td>Ye [36]</td>
<td>Male Sprague Dawley rats (6/7)</td>
<td>240 ± 60 g</td>
<td>AMI</td>
<td>48 h after the surgery</td>
<td>Ether inhaler</td>
<td>Salvia extract (main composition salvinolic acid B (40 mg/kg·d), i.g.</td>
<td>Normal saline</td>
<td>(1) VEGF</td>
<td>(1) $P &lt; 0.01$; 8 w</td>
<td>No</td>
<td>No</td>
<td>No report</td>
</tr>
<tr>
<td>Pang [37]</td>
<td>Male Sprague Dawley rats (10/11)</td>
<td>180–220 g</td>
<td>AMI</td>
<td>24 h after the surgery</td>
<td>Ether inhaler</td>
<td>Salvinolic acid B 120 mg/(kg·d) i.g</td>
<td>Normal saline</td>
<td>(1) BVD</td>
<td>(1) $P &lt; 0.05$; 2 w</td>
<td>(1) Myocardial infarct size</td>
<td>(1) $P &lt; 0.05$; 2 w</td>
<td>(2) Ventricle thickness</td>
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<tr>
<td>Guo et al. [38]</td>
<td>Female SD rats (no report)</td>
<td>No report</td>
<td>AMI</td>
<td>No report</td>
<td>No report</td>
<td>80 μl of phosphate-buffered saline (PBS) alone and 5 × 10⁶ salvinolic acid B pretreated MSC/d</td>
<td>80 μl of phosphate-buffered saline alone and 5 × 10⁶ MSC/d</td>
<td>(1) BVD</td>
<td>(1) $P &lt; 0.01$; 4 w</td>
<td>(1) $P &lt; 0.01$; 4 w</td>
<td>(2) VEGF</td>
<td>(2) $P &lt; 0.05$; 4 w</td>
</tr>
</tbody>
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Note: LAD: the left anterior descending coronary artery; VEGF: vascular endothelium growth factor; LVSP: left ventricular systolic pressure; LVEDP: left ventricular end-diastolic pressure; IL: infarct length; LVEF: left ventricular ejection fraction.
salvianolic acid A [27, 31] (n = 20, SMD 3.31, 95% CI: 1.72–4.90, \( P < 0.0001 \); heterogeneity \( \chi^2 = 0.21, P = 0.65, I^2 = 0% \)), salvianolic acid B [26, 34] (n = 42, SMD 1.51, 95% CI: 0.81–2.21, \( P < 0.0001 \); heterogeneity \( \chi^2 = 0.04, P = 0.84, I^2 = 0% \)), and a mixture of salvianolic acids [29, 36] (n = 39, SMD 2.28, 95% CI: 1.43–3.14, \( P < 0.0001 \); heterogeneity \( \chi^2 = 0.55, P = 0.46, I^2 = 0% \)) were significantly increasing VEGF expression compared with the control group, respectively (Figure 3). After removing one study [34] that used female animals, the meta-analysis of five studies [26, 27, 29, 31, 36] that used male animals showed significantly increasing VEGF expression compared with the control group (n = 89, SMD 2.17, 95% CI: 1.52–2.82, \( P < 0.0001 \); heterogeneity \( \chi^2 = 4.81, P = 0.31, I^2 = 17\% \)), Figure 4. The remaining 4 studies [25, 28, 33, 38] failed to pool the analysis because of the absence of primary data, but all of them reported significant effects of salvianolic acid for increasing VEGF expression compared with the control group (\( P < 0.05 \) or \( P < 0.01 \)).

3.4.2. BVD. Twelve studies [26–35, 37, 38] utilized BVD as the outcome measure, including salvianolic acid A [27, 31], salvianolic acid B [26, 30, 34, 35, 37, 38], and salvianolic acid mixture [28, 29, 32, 33]. The meta-analysis of the two salvianolic acid A studies [27, 31] showed significantly improving BVD compared with the control group (n = 20, SMD 3.56, 95% CI: 1.89–5.23, \( P < 0.0001 \); heterogeneity \( \chi^2 = 0, P = 0.97, I^2 = 0\% \)), Figure 5. The meta-analysis of the five salvianolic acid B studies [26, 30, 34, 35, 37] showed significantly improving BVD compared with the control group (n = 107, SMD 1.9, 95% CI: 0.9–2.9, \( P = 0.0002 \); heterogeneity \( \chi^2 = 16.47, P = 0.002, I^2 = 76\% \)). Owing to the significant statistical heterogeneity, we utilized subgroup analyses to explore the sources of the heterogeneity. Among the five included studies, three studies [30, 34, 35] used pentobarbital as anesthetic, one study [26] used urethane, and the last one [37] used ether. The meta-analysis of three studies [30, 34, 35] showed significant effects of salvianolic acid B for improving BVD compared with the control group (n = 56, SMD 2.83, 95% CI: 2.04–3.62, \( P < 0.0001 \); heterogeneity \( \chi^2 = 0, P = 1, I^2 = 0\% \)), Figure 6, suggesting that anesthetic was the potential cause of the heterogeneity. Additionally, after removing two studies [34, 35] which used female animals, the meta-analysis of three studies [26, 30, 37] showed significant effects of salvianolic acid B for improving BVD compared with the control group (n = 71, SMD 1.38, 95% CI: 0.26–2.49, \( P = 0.02 \); heterogeneity \( \chi^2 = 8.04, P = 0.02, I^2 = 75\% \)). The reason of the high heterogeneity was possibly different anesthetics used. The remaining one salvianolic acid B study [38] failed to analyze because of the absence of primary data, but it also reported significant increasing BVD compared with the control group (\( P < 0.05 \)). The meta-analysis of the 3 mixtures of salvianolic acid studies [29, 32, 33] showed significant effects for improving BVD compared with the control group (n = 54, SMD 8.46, 95% CI: 1.40–15.53, \( P < 0.0001 \); heterogeneity \( \chi^2 = 19.78, P = 0.02, I^2 = 90\% \)). The reason of the high heterogeneity might be that each of them had different types of salvianolic acid. The remaining one mixture of a salvianolic acid study [28] failed to pool the analysis because of the absence of primary data. However, all of them reported a positive effect on increasing BVD compared with the control group (\( P < 0.05 \) or \( P < 0.01 \)).

3.4.3. Myocardial Infarct Size. Eight studies [25–27, 30, 31, 34, 35, 37] utilized myocardial infarct size as outcome measure. The meta-analysis of five studies [25, 27, 34, 35, 37] showed significant effects of salvianolic acid for decreasing myocardial infarct size compared with the control group (n = 79, SMD −2.16, 95% CI: −2.81–−1.51, \( P < 0.0001 \); heterogeneity \( \chi^2 = 4.53, P = 0.34, I^2 = 12\% \)), Figure 7. After removing one salvianolic acid A study [27], the meta-analysis of four salvianolic acid B studies [25, 34, 35, 37] showed significantly decreasing myocardial infarct size compared with the control group (n = 69, SMD −2.02, 95% CI: −2.63–−1.41, \( P < 0.0001 \); heterogeneity \( \chi^2 = 0.63, P = 0.89, I^2 = 0\% \)), Figure 8. After removing two studies [34, 35] that used female animals, the meta-analysis of three studies [25, 27, 37] showed significantly decreasing myocardial infarct size compared with the control group (n = 47, SMD −2.34, 95% CI: −3.67–−1.02, \( P = 0.0005 \); heterogeneity \( \chi^2 = 4.36, P = 0.11, I^2 = 54\% \)), Figure 9. The reason of the high heterogeneity was possibly the use of different types of salvianolic acid. The remaining three studies [26, 30, 31] failed to pool the analysis because of the absence of primary data, but all of them reported the significant effects of salvianolic acid for decreasing myocardial infarct size compared with the control group (\( P < 0.05 \) or \( P < 0.01 \)).

4. Discussion

4.1. Summary of Evidences. To our knowledge, this is the first systematic review to estimate the effects of salvianolic acid for
3: The forest plot: subgroup analysis of salvianolic acid A, salvianolic acid B, and a mixture of salvianolic acids for improving VEGF expression compared with the control group.

**Figure 2:** The forest plot: effects of salvianolic acid for increasing VEGF expression compared with the control group.

**Figure 3:** The forest plot: subgroup analysis of salvianolic acid A, salvianolic acid B, and a mixture of salvianolic acids for improving VEGF compared with the control group.

**Figure 4:** The forest plot: effects of salvianolic acid in male animals for increasing VEGF expression compared with the control group.
**Figure 5:** The forest plot: effects of salvianolic acid A for improving BVD compared with the control group.

<table>
<thead>
<tr>
<th>Study or subgroup</th>
<th>Experimental Mean</th>
<th>Control Mean</th>
<th>Std. mean difference</th>
<th>IV, random, 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Li, 2010</td>
<td>13.8</td>
<td>4.8</td>
<td>3.53 [1.19, 5.87]</td>
<td></td>
</tr>
<tr>
<td>Li, 2014</td>
<td>14</td>
<td>5.8</td>
<td>3.59 [1.22, 5.96]</td>
<td></td>
</tr>
</tbody>
</table>

Total (95% CI) 10 10 100.0% 3.56 [1.89, 5.23]

Test for overall effect: $Z = 6.49 \ (P < 0.00001)$

**Figure 6:** The forest plot: effects of salvianolic acid B that used pentobarbital as anesthetic for improving BVD compared with the control group.

<table>
<thead>
<tr>
<th>Study or subgroup</th>
<th>Experimental Mean</th>
<th>Control Mean</th>
<th>Std. mean difference</th>
<th>IV, random, 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pang, 2008</td>
<td>13.8</td>
<td>5.74</td>
<td>2.35 [1.16, 3.57]</td>
<td></td>
</tr>
<tr>
<td>Fan, 2004</td>
<td>14</td>
<td>6.96</td>
<td>2.06 [0.55, 3.57]</td>
<td></td>
</tr>
<tr>
<td>Lin, 2016</td>
<td>16</td>
<td>23</td>
<td>5.37 [1.50, 9.24]</td>
<td></td>
</tr>
</tbody>
</table>

Total (95% CI) 28 28 100.0% 2.85 [0.69, 5.00]

Test for overall effect: $Z = 4.19 \ (P < 0.00001)$

**Figure 7:** The forest plot: effects of salvianolic acid B for decreasing myocardial infarct size compared with the control group.

<table>
<thead>
<tr>
<th>Study or subgroup</th>
<th>Experimental Mean</th>
<th>Control Mean</th>
<th>Std. mean difference</th>
<th>IV, random, 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ma, 2006</td>
<td>14</td>
<td>27.6</td>
<td>6.38 [2.63, 10.13]</td>
<td></td>
</tr>
<tr>
<td>Li, 2014</td>
<td>16</td>
<td>36.6</td>
<td>8.57 [4.70, 12.44]</td>
<td></td>
</tr>
</tbody>
</table>

Total (95% CI) 39 40 100.0% -1.66 [0.82, 2.50]

Test for overall effect: $Z = 7.03 \ (P < 0.00001)$

**Figure 8:** The forest plot: effects of salvianolic acid B for decreasing myocardial infarct size compared with the control group.

<table>
<thead>
<tr>
<th>Study or subgroup</th>
<th>Experimental Mean</th>
<th>Control Mean</th>
<th>Std. mean difference</th>
<th>IV, random, 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fan, 2004</td>
<td>13.8</td>
<td>6.16</td>
<td>2.35 [1.16, 4.53]</td>
<td></td>
</tr>
<tr>
<td>Lin, 2016</td>
<td>16</td>
<td>20.7</td>
<td>6.96 [2.47, 11.45]</td>
<td></td>
</tr>
<tr>
<td>Pang, 2008</td>
<td>13.8</td>
<td>28.9</td>
<td>8.57 [4.70, 12.44]</td>
<td></td>
</tr>
</tbody>
</table>

Total (95% CI) 34 35 100.0% -2.02 [0.82, 2.50]

Test for overall effect: $Z = 6.49 \ (P < 0.00001)$
There is no doubt that study quality is an important effect factor [41]. We suggest that further design of the studies carried out should refer to the ten-item scale [38] such as random allocation, blinded induction of model, blinded assessment of outcome, and use of anesthetic without significant intrinsic vascular protection activity. In addition, we should include appropriate animals because an unsuitable animal model may affect the validity of the experiments [46]. Myocardial infarction generally occurs in elderly patients with hypertension or hyperlipidemia [42], so using appropriate models can increase the accuracy of the results.

4.4. Conclusion. The salvianolic acid including salvianolic acid A, salvianolic acid B, and a mixture of salvianolic acids can reduce myocardial infarct size and promote the expression of VEGF and BVD in animal model experiments of MI, suggesting that salvianolic acid has cardioprotective function through promoting angiogenesis in the animal model of MI. However, the positive conclusion should be treated cautiously because of the methodological flaws.

Conflicts of Interest

The authors confirm that this article’s content has no conflict of interest.

Authors’ Contributions

Long-jie Yu, Ke-Jian Zhang, and Jia-Zhen Zhu contributed equally to this work.

Acknowledgments

This project was supported by the grant of the National Natural Science Foundation of China (81473491/H2902).

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