Thymoquinone (TQ) is a major bioactive compound mainly found in the Nigella sativa L. [1–3]. N. sativa is an emerging miracle herb that belongs to the Ranunculaceae family and is known as an annual herbaceous plant that is frequently grown in Western Asia, Middle East, Eastern Europe, and Mediterranean countries [4, 5]. Traditionally, the seeds of N. sativa are used in the Middle East, India, and Northern Africa for the treatment of cough, asthma, bronchitis, influenza, fever, headache and rheumatism [6–8]. Furthermore, it is also employed as an appetite stimulant, digestive, liver tonic, anti diarrheal, and emmenagogue to increase milk production in a lactating mother. Also, it has an immune system protecting activity against parasitic infections [5].

Growing evidence concerning the versatile TQ pharmacological activities has been established. In a number of preclinical studies, it elicits antitussive, gastroprotective, anti-inflammatory, antinociceptive, antihistaminic, antibacterial, anthelmintic, antioxidant, immunomodulatory, anticancer, hepatoprotective, cardioprotective, antidiabetic, otoxicity protective, and nephroprotective effects [9–31]. N. sativa and TQ have numerous advantageous properties with relevance to various neurological illnesses. For instance, they have anticonvulsant, anxiety, antidepressant, and antipsychotic potential. They also counter memory impairments and enhance cognitive functioning as well as attenuate drug tolerance and dependence [32–47]. Moreover, many recent studies have described the neuropharmacological properties of TQ such as anti-inflammatory and antioxidative roles in designed neurological models [48–50]. Besides, it alleviates neuropathy in an experimental peripheral diabetic model [51]. Yet, the pharmacological potential and delivery prospects of TQ in neurological illnesses have not...
been documented to date. Therefore, in this article, considering the neurological disorders, we deliberate the neuropharmacological potential, challenges, and delivery concepts of TQ along with its chemistry, pharmacokinetics, and toxicity.

2. Chemistry of Thymoquinone

Thymoquinone is a monoterpenic diketone and an important bioactive compound that forms 18.4%–24% of these essential oils with a boiling point and melting point of 230–232°C and 44–45°C, respectively. Its molecular weight is 164.204 g/mol, and value of Log P is 2.20 denoting lipophilicity of TQ. Moreover, it has the ability to penetrate the blood-brain barrier (BBB) owing to its molecular weight (less than 500 g/mol) and Log P (less than five) value. Thus, it might be suitable for clinical trials [52–56]. Structurally, it is homologous with coenzyme Q, an important antioxidant of the electron transport chain [57]. By oxidation of thymol with hydrogen peroxide, TQ can be readily synthesized in gram quantities [58]. Moreover, according to one hypothesis based on chemical composition, oxidation of quinone compounds caused by thermal processing leads to modification and conversion between compounds and accumulation of more powerful compounds. The controlled heating causes oxidation of thymol and converts it into thymohydroquinone. The heating process persistence leads to a further oxidative process that converts thymohydroquinone into thymoquinone, resulting in the accumulation of larger amounts of TQ. Based on the presence of light, the photoisomerization of TQ may cause an accumulation of its dimer, dithymoquinone [57]. Besides this, during quinone separation and extraction from seed, thymoquinone photodimerization as a consequence of exposure to sunlight produces dithymoquinone [11]. Conversions of different quinones by the chemical reaction are shown in Figure 1.

3. Pharmacokinetics and Toxicity of Thymoquinone

Thymoquinone’s pharmacokinetic profiles have been reported in several studies [59, 60]. Its solubility in water has reported as <1.0 mg/ml at room temperature [61]. TQ has poor oral bioavailability based on its low aqueous solubility and dissolution rate [62]. Moreover, TQ represents a compound with rapid plasma concentration-time curves demonstrating a rapid polyexponential decline following intravenous dosing [63]. The plasma concentration of glibenclamide increases through the coadministration of TQ as single and multiple doses, respectively [64]. In addition, TQ binds with major plasma proteins including bovine serum albumin (BSA) and alpha-1 acid glycoprotein (AGP) [65–68]. Molecular modeling and fluorescence quenching studies have confirmed the binding interaction of TQ with BSA and AGP [65, 66]. Through covalent bonding, TQ binds to both BSA and AGP. Covalent binding of TQ to BSA leads to eliminating the TQ anticancer activity against various tested cancer cell lines, while on the other hand, TQ anticancer activity was not affected when TQ is bound to AGP [67, 68]. There could be biotransformation owing to the metabolizing activity of liver enzymes, such as DT-diaphorase (a quinone reductase), which catalyzes the reduction of TQ into a dihydrothymoquinone [69]. This metabolite exhibits antioxidant properties that are more potent than those of TQ and similar to those of Trolox, which considers a standard antioxidant compound [70]. With this, TQ-treated diabetic rats show a marked decrease in hepatic protein expressions of cytochrome P450 3A2 and cytochrome P450 2C11 enzymes that are responsible for the metabolism of glibenclamide. At the glucose level, TQ has a synergistic effect with glibenclamide, which could be explained by reduction cytochrome P450 activity at the protein level. In addition, the area under the plasma drug concentration-time curve and plasma half-life of glibenclamide were also elevated, respectively, by a single dose of TQ as well as after chronic treatment [64]. A variety of in vitro and in vivo studies has evaluated thymoquinone toxicity profile [71, 72]. The oral and intraperitoneal (i.p.) median lethal doses (LD50) for TQ in rats as well as in mice were successfully determined by several research groups [73, 74]. At higher doses, it is well tolerated by male and female rats. TQ at 10 mg/kg body weight was found to have no observed adverse effect level [75]. Moreover, the administration of 20 ml develops a TQ-rich fraction nanoemulsion per kg that was not toxic following acute exposure in Sprague-Dawley rats [76]. In terms of acute toxicity, TQ-loaded nanostructured lipid carriers (TQ-NLC) have less toxicity than pure TQ. In a subacute toxicity study, oral administration of 100 mg/kg of TQ-NLC caused mortality in neither male nor female mice but resulted in liver toxicity. For long-term oral consumption, TQ and TQ-NLC at doses of 10 mg/kg are safe in mice. These results provided safety information pertaining to TQ-NLC, which further aids researchers in clinical use [77]. Furthermore, another study determined LD50 = 104.7 mg/kg in mice i.p. and 870.9 mg/kg after oral ingestion of TQ. In rats, the value of LD50 was detected to be 57.5 mg/kg while it was 794.3 mg/kg after i.p. and oral administration, respectively [73]. Interestingly, using TQ in combination with ionizing radiation such as γ-radiation was found to exert a synergistic cytotoxic effect on breast cancer cells in vitro [78]. Cisplatin administration and a high dose of TQ act synergistically to produce nephotoxicity where the involvement of the apoptotic pathway and proximal tubule damage might be a primary determinant of this effect [79].

4. Neuropharmacological Potential of Thymoquinone

4.1. Activities in Neuroinflammation Model. Neuroinflammation is the term used to describe the inflammatory response originating in the central nervous system (CNS) after suffering an injury, where there is an accumulation of glial cells (microglia and astrocytes) [80]. TQ possesses antineuroinflammatory activity, specifically via lipopolysaccharide- (LPS-) induced BV-2 microglial cells [49, 81, 82]. It is effective in reducing nitrite (NO2−) with a half maximal inhibitory concentration (IC50) of 5.04 μM, relative to selective inducible nitric oxide synthase (iNOS) inhibitor
LNIL-L-N6-(1-iminoethyl) lysine (IC$_{50}$ = 4.09 μM). TQ reduces the level of NO$^{2−}$ found to parallel the decline of iNOS protein expression. In activated microglia, TQ (10 μM) attenuates the increased levels of proinflammatory cytokines/chemokines including interleukin- (IL-) 6, IL-12p40/70, chemokine (C-C motif) ligand 12/monocyte chemotactic protein 5, chemokine (C-C motif) ligand 2/monocyte chemoattractant protein 1, and granulocyte colony-stimulating factor. TQ has also been observed to mediate attenuation of monocyte chemotactic protein 5, monocyte chemoattractant protein 1, and IL-6 protein in supernatants from activated BV-2 cells. In addition, TQ reduces LPS-mediated elevation in gene expression of C-X-C motif chemokine 10 and a number of other cytokines in the panel [81]. Considering the recent study on nuclear factor E2-related factor 2 (Nrf2)/antioxidant responsive element signaling pathway, TQ activates this signal by elevating the nuclear localization, DNA binding, and Nrf2 transcriotional activity, along with increasing protein levels of heme oxygenase-1 (HO-1) and NAD(P)H:quinone oxidoreductase. In this study, it also inhibits nuclear factor kappa-light-chain-enhancer of activated B cell- (NF-κB-) dependent neuroinflammation in BV-2 microglia [82].

Regarding recent mechanistic studies, TQ produces antineuroinflammatory activity through the AMP-activated protein kinase (AMPK) and nicotinamide adenine dinucleotide (oxidized)/sirtuin 1 (SIRT1) pathways. Further, TQ diminishes cellular ROS generation, possibly through inhibition of p40$^{phox}$ and gp91$^{phox}$ protein. Treatment of BV-2 microglia with TQ also results in an elevation in the levels of liver kinase B1 and phospho-AMPK proteins. Furthermore, TQ reduces cytoplasmic levels and increases nuclear accumulation of SIRT1 protein and enhances nicotinamide adenine dinucleotide (oxidized) concentration. Besides this, the effect of RNAi or pharmacological antagonists suppressing the anti-inflammatory activity of TQ was abolished with expressions of AMPK and SIRT1. Pharmacological antagonism of AMPK reverse TQ-induced increases in SIRT1 [49]. Another recent investigation has offered evidence for TQ’s anti-inflammatory activity. At different concentrations, TQ strongly inhibits the nitric oxide (NO) production and represses iNOS, tumor necrosis factor- (TNF-) α, cyclooxygenase-2, IL-6, and IL-1β expression in LPS-activated RAW264.7 cells [83]. Moreover, the protective effects of TQ on LPS-induced brain tissues oxidative stress status induced in rats were reported. Compared with a control group, LPS
elevates malondialdehyde (MDA) and NO metabolites while decreasing thiol content, superoxide dismutase (SOD), and catalase (CAT) in both the hippocampus and cortex. Treatments with TQ (2, 5, or 10 mg/kg) decrease IL-6, TNF-α, MDA, and NO metabolites and increase thiol content along with SOD and CAT compared to the LPS group [84].

4.2. Activities in Induced Disease Models

4.2.1. Model of Alzheimer’s Disease. Alzheimer’s disease (AD) is a chronic neurodegenerative disorder where the major risk factor is age with data showing a considerable increase in incidence after the age of 60 years [85, 86]. Its impact on public health and society as a whole is devastating [86]. It is also known as the most common type of age-related dementia and is characterized by memory loss, cognitive decline, and debilitating neurodegeneration [86–88].

Regarding research studies, TQ might be a potential agent against AD. In a model of amyloid beta (Aβ) induced neurotoxicity, cultured hippocampal and cortical neurons were treated with Aβ1–42 and TQ simultaneously for 72 hours. Treatment with TQ (100 nM) efficiently attenuates Aβ1–42-induced neurotoxicity, as evidenced by improving cell viability. Aβ1–42 causes mitochondrial membrane potential depolarization and reactive oxygen species (ROS) generation while TQ inhibited these changes. In addition, TQ restores synaptic vesicle recycling inhibition, partially reversing activity while TQ inhibited these changes. In addition, TQ restores synaptic vesicle recycling inhibition, partially upturns the loss of spontaneous firing activity, and also inhibits Aβ1–42 aggregation in vitro [89]. Another study described the inhibitory effects of TQ in comparison to tannic acid (TA) on Aβ fibril formation. The effects of TQ on the formation of Aβ1–40 were studied spectrophotometrically, over a duration of 16 days at a pH of 7.2 and 37°C. TQ and TA, with an IC50 of 0.1 and 29 μM at day 0 (within 5 h), respectively, inhibit Aβ aggregation in a dose-dependent way at different concentrations (1, 10, and 50 μM). However, at the day 16, IC50 of TQ and TA was found to be 0.2 and 0.01 μM, respectively. In an electron microscopic study, coincubation of TQ with Aβ reduces the numbers of fibrils to a certain degree with shorter fibrils and small amorphous aggregates. TQ pretreatment protects Aβ1–40 cytotoxic effects in primary cultured cerebellar granule neurons [90]. Furthermore, in combination with α7 nicotinic acetylcholine receptor’s positive allosteric modulators (PAM) with a known agonist (PNU-282987) or with TQ as a possible treatment for AD in a rat model has evaluated. LPS causes acidophilic masses, deformed neurons, Congo red (+ve) masses, and reduced CAMP response element-binding protein (CREB) phosphorylation (phospho-CREB) immunoreactivity. Concerning histological, histochemical, and immunohistochemical studies, TQ or α7 nicotinic acetylcholine receptor agonist combined with PAM can have an important role in treatment of AD [91]. In a recent study, TQ improves LPS-induced learning and memory impairments in rats by attenuating hippocampal cytokine levels and brain tissue oxidative damage [84].

4.2.2. Model of Parkinson’s Disease. Parkinson’s disease (PD) is considered a progressive neurodegenerative disorder characterized by the dopaminergic neuron loss in the substantia nigra pars compacta (SNpc) and neuroinflammation through microglial malfunction [92, 93]. TQ exhibits protective activities in PD models. The neuroprotective effects of TQ on primary dopaminergic cultures from mouse mesencephalon using 1-methyl-4-phenylpyridinium (MPP+) and rotenone toxicities were studied previously [94, 95]. MPP+ (10 μM on day 10 in vitro for 48 h) significantly decreases the number of tyrosine hydroxylase immunoreactive (THir) by 40% compared with untreated control cultures. Rotenone with both short- (20 nM on day 10 in vitro for 48 h) and long-term (1 μM on day six in vitro for six consecutive days) toxicities reduces the number of THir neurons by 33% and 24%, respectively. Treatment of cultures with TQ (0.01, 0.1, 1, and 10 μM on day eight for four days) rescues approximately 25% of THir neurons at concentrations of 0.1 μM and 1 μM against MPP+-induced cell death. Against rotenone, TQ afforded significant protection in both short- and long-term models. In case of short-term rotenone toxicity, TQ (from days 8 to 12) saves about 65%, 74%, and 79% of THir neurons at concentrations of 0.01, 0.1, and 1 μM, respectively, compared with cell loss induced by rotenone. In long-term rotenone toxicity scenarios, concomitant treatment of cultures with TQ significantly rescues about 83 to 100% of THir neurons compared with rotenone-treated cultures [94]. Additionally, MPP+ decreases the number of dopaminergic neurons by 40% and enhances the release of lactate dehydrogenase (LDH) into the culture medium. TQ significantly rescues dopaminergic neurons and decreases the release of LDH at the concentrations of 0.1 and 1 μM. Moreover, TQ treatment significantly shifts the red fluorescent intensity of the LysoTracker® Deep Red, increasing the mitochondrial membrane potential as it increased the red: green fluorescent ratio of JC-1 and decreases MPP+-induced apoptotic cell death. TQ is thought to protect dopaminergic neurons in primary mesencephalic cultures by enhancing lysosomal degradation that clears damaged mitochondria and inhibits mitochondria-mediated apoptotic cell death [95].

Furthermore, TQ produces neuroprotective activity in a 6-hydroxydopamine- (6-OHDA-) induced PD model. Unilateral intrastriatal 6-OHDA-lesioned rats were daily pretreated per oral (p.o.) with TQ at doses of 5 and/or 10 mg/kg three times at an interval of 24 h. TQ pretreatment significantly improves turning behavior, prevents loss of SNpc neurons, and lowers level of MDA. Hence, TQ offers neuroprotection against 6-OHDA-induced neurotoxicity that is partly based on the attenuation of lipid peroxidation [96]. Regarding the recent study, TQ (7.5 and 15 mg/kg/day, p.o.) pretreatment was administered one hour prior to rotenone injection, affecting motor test results (rotarod, rearing, and bar tests). TQ significantly prevents rotenone-induced motor defects and modifications in the Parkin, dynamin-related protein 1, dopamine, and tyrosine hydroxylase levels in the studied areas [97]. A summary of the effects of TQ in PD models is located in Table 1.

4.2.3. Model of Autoimmune Encephalomyelitis. Experimental autoimmune encephalomyelitis (EAE) is a CD4+ T cell-
mediated inflammatory demyelinating condition of the CNS. It is frequently employed as a model of multiple sclerosis, because this model has great importance for understanding multiple sclerosis pathogenesis and its therapy design [98, 99]. In the EAE experimental model, oxidative stress plays a significant role in EAE onset and progression. Therefore, reducing oxidative stress might ameliorate EAE signs and symptoms. Through utilizing myelin basic protein emulsified with complete Freund’s adjuvant, EAE was induced in female Lewis rats and TQ (1 mg/kg) was injected into the tail vein. Oxidative stress inhibition is observed, and TQ might improve the symptoms in EAE animals [100]. TQ concomitant with myelin basic protein after the appearance of clinical signs resulted in preventing and ameliorating EAE. TQ (1 mg/kg per day) was able to counter perivascular cuffing and infiltration of mononuclear cells in the brain and spinal cord, increase the red blood cell glutathione (GSH), and inhibit the activation of NF-κB in the brain and spinal cord [101]. In addition, in a myelin oligodendrocyte glycoprotein-induced EAE model, the antioxidant effect of TQ has nearly 90% preventive and 50% therapeutic effects in chronic relapsing EAE [102].

4.2.4. Model of Stroke. Ischemia/reperfusion (I/R) injury is the cascade of events leading to neuronal injury and death. It involves NO, excitatory amino acids, cytokine and free radical release, mitochondrial respiratory enzyme damage, induction of programmed cell death, and activation of microglia [103]. It is known that redox imbalance, inflammation, and apoptosis are the major mechanisms of I/R injury [104]. TQ shows potential activity against I/R injury and neuronal damage. Five days before ischemia, TQ (5 mg/kg) administered orally and sustained during reperfusion attenuated forebrain ischemia-induced neuronal damage. TQ treatment significantly diminishes the dead hippocampal neuronal cell number (24% in TQ-treated versus 77% for ischemia), thereby supporting TQ’s protective role in I/R injury. Accordingly, TQ also shows an effect against oxidative stress [105]. Moreover, treatment with TQ (10 mg/kg i.p.) for seven days before induction of spinal cord I/R injury improves neurological outcomes. Specifically, TQ produces effect against oxidative stress and neuroinflammation and also reduces the motor neuron apoptosis. TQ treatment also shows preservation of tissue structure [50]. With this, after TQ administration via the noninvasive nasal route of TQ mucoadhesive nanoemulsion improves neurobehavioral activity (locomotor and grip strength) was observed in middle cerebral artery occlusion-induced cerebral ischemia [106]. Additionally, in a rodent cerebral I/R model, TQ-loaded PLGA-chitosan nanoparticles optimized delivery through the nose-to-brain route exhibit the neuroprotective efficacy against cerebral ischemia [107].

4.2.5. Schizophrenia Model. The mental disorder, schizophrenia, is characterized by severe cognitive impairment, as well as positive and negative symptoms such as hallucination, delusion, thought disorders, apathy, and social withdrawal [108, 109]. Moreover, regarding the adverse effects of available antipsychotic medications, recent investigations have focused on the search for well-tolerated, safe molecules from natural sources to control the severity and progression of the disorder [110]. A potential role of TQ in combating schizophrenia has been described. With this, TQ demonstrates potential activity against schizophrenia in mouse models. In one study for 28 days, i.p. doses of TQ (20 mg/kg) were administered. The pretreatment of TQ alone and in combination with haloperidol led to observed cataleptic behavior. Moreover, in an apomorphine-induced climbing behavior model, the administration of TQ significantly reduces the ultimate time taken for single climb and climbing index. TQ reduces scopolamine-induced prolongation of transfer latency. However, a significant rise in possible alternation was observed, suggesting its antiannestic effects. TQ possesses antiannestic effect with a decrease in acetylcholinesterase (AChE) enzymatic activity in mouse brain. Decreased concentration of thiobarbituric acid reactive substance (TBARS) and the rise in glutathione and CAT levels were observed in all models [40]. Additionally, TQ administration also reduces dopamine levels, indicating the involvement of dopamine receptors in all three models, hence, demonstrating its antipsychotic-like activities [40].

4.2.6. Model of Epilepsy. Epilepsy is a brain disorder characterized by convulsive seizures or loss of consciousness or both and induced by a complex of neurotransmitter systems

<table>
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<tr>
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<th>Doses</th>
<th>Mechanistic actions</th>
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</tr>
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<tbody>
<tr>
<td>MPP+ and rotenone toxicities in dopaminergic neurons</td>
<td>0.01, 0.1, 1, and 10 μM</td>
<td>Increases the number of THir compared with untreated control cultures</td>
<td>[94]</td>
</tr>
<tr>
<td>MPP+ toxicity in dopaminergic neurons</td>
<td>0.1 and 1 μM</td>
<td>Decreases the number of dopaminergic neurons and increases the release of LDH primary mesencephalic culture by enhancing lysosomal degradation that clears damaged mitochondria and inhibits mitochondria-mediated apoptotic cell death</td>
<td>[95]</td>
</tr>
<tr>
<td>6-OHDA-induced PD model</td>
<td>5 and/or 10 mg/kg</td>
<td>Significantly improves turning behavior, prevents loss of SNpc neurons, and lowers the level of MDA</td>
<td>[96]</td>
</tr>
<tr>
<td>Rotenone-induced PD model</td>
<td>7.5 and 15 mg/kg</td>
<td>Significantly prevents rotenone-induced motor defects and modifications in the Parkin, Drp1, dopamine, and TH concentrations</td>
<td>[97]</td>
</tr>
</tbody>
</table>

MPP+: 1-methyl-4-phenylpyridinium; 6-OHDA: 6-hydroxydopamine; THir: tyrosine hydroxylase immunoreactive; LDH: lactate dehydrogenase; MDA: malondialdehyde; SNpc: substantia nigra pars compacta; Drp1: dynamin-related protein 1; TH: tyrosine hydroxylase.
However, at both 50 and 100 mg/kg doses, TQ significantly potentiates SVP antiepileptic response in PTZ model [114]. In another epileptic model, naltrexone (0.5 μg/ventricle i.c.v.) antagonizes the tonic-clonic seizure latency prolongation and the reduction in duration of seizure induced by TQ at dose of 200 μM. The anticonvulsant activity of TQ is probably through an opioid receptor-mediated increase in GABAergic tone [113]. Moreover, another study reported the antiepileptic potential of TQ in mice using PTZ and maximal electroshock seizure- (MES-) induced convulsion models. TQ in both PTZ and MES models increased sodium valproate potency (SVP). In addition, TQ reduces the ED50 of the SVP in both the models. However, at both 50 and 100 mg/kg doses, TQ significantly potentiates SVP antiepileptic response in PTZ model [114]. Furthermore, TQ along with phenobarbital combination therapy produces additive anticonvulsant effect compared to monotherapy with phenobarbital in PTZ-induced rat model [115]. According to the another study, TQ and vitamin C exhibit anticonvulsant effects via activation of the gamma-aminobutyric acid B1 receptor (GABAB1R)/calcium/calmodulin-dependent protein kinase II (CaMKII)/CREB pathway, suggesting a potential therapeutic role in epilepsy. Animals pretreated with either p.o. administration of TQ (40 mg/kg administered for seven days) or vitamin C (vitamin C 250 mg/kg i.p. 2 h before PTZ i.p. injection) or in combination (vitamin C 250 mg/kg i.p. and TQ 40 mg/kg orally 2 h before PTZ). Compared to PTZ, TQ and vitamin C significantly prolong the onset of seizures, as well decrease the incidence of high-grade seizures. Electroencephalogram (EEG) analysis indicates that TQ or vitamin C supplementation significantly reduces polyspike and epileptiform discharges. In comparison with the control group, SE cause a decline in GABAB1R expression but no change expression of protein kinase A. Additionally, seizures also decrease CaMKII and prevent phospho-CREB in both the cortex and hippocampus areas, respectively. Meanwhile, TQ and vitamin C supplementation reversed the PTZ-induced changes in expression of GABAB1R, CaMKII, and CREB. Moreover, PTZ significantly increases Bcl-2-associated X protein (Bax) concentrations, decreases a crucial antiapoptotic protein and B-cell lymphoma protein-2 (Bcl-2) expression, and diminishes the activation of caspase-3 [116]. In a penicillin-induced epilepsy model, TQ also has antiepileptic activity in rats. Various doses of TQ (10, 50, and 100 mg/kg) significantly increase the latency time to the onset of the first spike wave and decrease the frequency and amplitude of epileptiform activity over the first 20 minutes compared with the control group [33]. In another epileptic model, kainic acid (0.5 μg/ventricle i.c.v.) bilaterally induces SE in male Wistar rats. The lesioned rats were treated with TQ (10 mg/kg i.p.) for four days. TQ significantly decreases suppression of spontaneous recurrent motor seizures, reduces neuron degeneration (25%) in CA1, CA3, and the dentate hilus, and suppresses mossy fiber sprouting (30–40%) both in four-month and 12-month age groups. TQ also increases neurogenesis in both groups; however, in middle-aged rats, it is significantly more than in young rats [117].

Recently, in a lithium-pilocarpine rat model, the protective impact of TQ is observed in brain injury of SE. TQ pretreatment (10 mg/kg twice) latency to SE increases compared with rats in the model group, whereas the total power was suggestively lesser. According to the behavioral experiments, TQ might also have a protective outcome on the function of learning and memory. Additionally, pretreatment with TQ significantly improves the Nrf2 and HO-1 protein expression as well as SOD in the hippocampus [48]. TQ activities in epilepsy models are described in Table 2.

4.2.7. Models of Depression and Anxiety. Regarding the reported studies, TQ possesses both antidepressant and anxiolytic activities according to mouse models. In one mouse model of depression, TQ modulates the levels of neurotransmitters and reduces oxidative stress. GSH level reduces and TBARS level increases in a modified forced swim test and tail suspension test in mice. Pretreatment of TQ (20 mg/kg) restores GSH and decreases TBARS levels. TQ combination with fluoxetine (10 mg/kg) also led to a reduction in TBARS and elevates GSH levels [39]. Moreover, TQ yields protective effects against oxidative stress via restraint stress-induced biochemical alterations in Wistar albino rats [118]. Both the doses of TQ (5 and 10 mg/kg) treatment significantly ameliorate restraint stress-induced increased IL level and enzymatic activities in serum. In the brain, restraint stress significantly increases TBARS, NO2−, and nitrate levels and TQ treatment brought back the concentration back to the normal levels. Besides, restraint stress significantly decreases oxidative enzyme activities, such as those of the SOD, CAT, glutathione-s-transferase, glutathione peroxidase (GSH-Px), and glutathione reductase (GR). With this, TQ treatments significantly enhance the activities compared to untreated stressed rats [118]. Furthermore, in the unstressed mice, TQ (10 and 20 mg/kg) elicits the anxiolytic effects without changing NO2− levels, but only at the higher dose (20 mg/kg) improving the GABA content. On the other hand, in the stressed mice, TQ (20 mg/kg) produces anxiolytic effects with a major reduction in plasma NO2− and reversal of the reduced brain GABA content. Nitric oxide-cyclic guanosine monophosphate and GABAergic pathways are considered crucial in the anxiolytic activities of TQ [36].

4.2.8. Model of Neuropathic Pain. At present, pain is one of the main health problems among the world’s population. It is also one of the most pervasive issues of high social and economic impacts in society [119–121]. The generation of neuropathic pain is through lesions to the somatosensory nervous system that changes its structure and function so that pain happens spontaneously and via responses to noxious and innocuous stimuli that pathologically amplified...
Pain. Repeated treatment with TQ (2.5 and 5 mg/kg) significantly alleviates behavioral signs of neuropathic pain. In the sham group following a mechanical and heat-cold test, the withdrawal threshold and withdrawal latency values were lower than those of the spinal cord injury group [130]. Moreover, administration of 30 mg/kg of TQ significantly decreases Bax and ionized calcium-binding adapter molecule in terms of concentrations on day three, while stimulating the production of Bcl-2. In addition, ionized calcium-binding adapter molecule and Bax/Bcl-2 ratio were lower than those of the control group at all time points. All of the TQ treatment groups increased the paraoxonase activities of TQ in epilepsy models. Table 2: Beneficial effects of TQ in epilepsy models.

<table>
<thead>
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<tbody>
<tr>
<td>PTZ-induced epilepsy model</td>
<td>200 and 400 μM</td>
<td>Extends the onset and reduces the tonic-clonic seizure duration</td>
<td>[113]</td>
</tr>
<tr>
<td>PTZ- and MES-induced epilepsy model</td>
<td>50 and 100 mg/kg</td>
<td>Potentiates SVP antiepileptic response</td>
<td>[114]</td>
</tr>
<tr>
<td>PTZ-induced epilepsy model</td>
<td>20 and 30 mg/kg</td>
<td>TQ and PB combination therapy produces additive anticonvulsant effect</td>
<td>[115]</td>
</tr>
<tr>
<td>PTZ-induced epilepsy model</td>
<td>TQ: 40 mg/kg; vitamin C: 250 mg/kg</td>
<td>Activates the GABAB1R/CaMKII/CREB pathway, significantly decreases Bax concentrations, increases Bcl-2 expression, and activates caspase-3</td>
<td>[116]</td>
</tr>
<tr>
<td>Penicillin-induced epilepsy model</td>
<td>10, 50, and 100 mg/kg</td>
<td>Prolongs latency time and reduces the spike wave frequency and amplitude of epileptiform activity</td>
<td>[33]</td>
</tr>
<tr>
<td>Kainic acid-induced epileptic model</td>
<td>10 mg/kg</td>
<td>Reduces neuronal degeneration (25%) in CA1, CA3, and the dentate hilus; suppresses mossy fiber sprouting (30–40%); treatment also enhances the neurogenesis</td>
<td>[117]</td>
</tr>
<tr>
<td>Lithium-pilocarpine rat model</td>
<td>10 mg/kg</td>
<td>Significantly lowered the severity of seizures and significantly elevated Nrf2, HO-1, and SOD expressions</td>
<td>[48]</td>
</tr>
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</table>

PTZ: pentyletetrazole; PB: phenobarbital; SVP: sodium valproate; GABAB1R: gamma-aminobutyric acid B1 receptor; CaMKII: calmodulin-dependent protein kinase II; CREB: cAMP response element-binding protein; MES: maximal electric shock; Bax: Bcl-2-associated X protein; Bcl-2: B-cell lymphoma protein-2; Nrf2: nuclear factor E2-related factor 2; HO-1: heme oxygenase-1; SOD: superoxide dismutase.

TQ has potential activity against neuropathic pain. Moreover, the role of oxidative stress, spinal glial activation, and cell death is involved in the pathogenesis of neuropathic pain. Repeated treatment with TQ (2.5 and 5 mg/kg) significantly alleviates behavioral signs of neuropathic pain. In the lumbar spinal cord of neuropathic rats, repeated TQ administration reduces elevated Bax and ionized calcium-binding adapter molecule in terms of concentrations on day three, while stimulating the production of Bcl-2. In addition, ionized calcium-binding adapter molecule and Bax/Bcl-2 ratio have declined by days 7 and 14 as a consequence. Furthermore, TQ treatment (2.5 and 5 mg/kg) restores the levels of MDA, and a high dose of TQ (5 mg/kg) also reverses the decreased GSH in injured animals. Microglia, apoptotic factors, and oxidative stress contribute to the pathogenesis of chronic constrictive injury, and TQ plays an antinociceptive role, possibly through antioxidant and antineuroinflammatory effects [123].

5. Miscellaneous Activities

5.1. Against Traumatic Brain and Spinal Cord Injuries. As a highly complex disorder, traumatic brain injury is the leading cause of mortality and disability in people under the age of 45 years [126, 127]. Spinal cord injury, on the other hand, is quite traumatic and results in disturbances to normal sensory, motor, or autonomic function and ultimately impacts a patient’s physical, psychological, and social well-being [128]. We already discussed that TQ has activity in spinal cord injury-induced neuropathic pain. Neuroprotective activity of TQ against damages due to trauma-induced both traumatic brain and spinal cord injuries has also reported. In a rat model, after TQ treatment (5 mg/kg, for seven days), neuronal density in contralateral hippocampal regions (CA1, CA2–3, and CA4) significantly increased in comparison to the control group. TQ diminishes the level of MDA in the nuclei and mitochondrial membranes of neurons [129]. Moreover, administration of 30 mg/kg of TQ significantly decreases the histological features of spinal cord damage versus the spinal cord injury group [130].

5.2. Protective Activities in Chemical-Induced Neurotoxicity Models. Lead (Pb) is known as a highly neurotoxic agent that causes functional and structural abnormalities in the brains [131]. In addition, Pb induces neurotoxicity in the cerebrum, cerebellum, and medulla oblongata of rat brain. DNA fragmentation and pathological alteration have also evidenced...
in Pb-intoxicated rat brain [132]. However, TQ protects Pb-induced brain damage. In Sprague-Dawley rats, at a concentration of 0.5 g/l (500 ppm), Pb acetate was given in drinking water. TQ was administered daily at a dose of 20 mg/kg. Regarding brain histology, the control and TQ-treated rats appear normal. Pb acetate treatment results in compromised endothelial lining of the brain blood vessel degeneration with perivascular cuffing of mononuclear cells consistent with lymphocytes, ischemic brain infarction, choroid plexus blood vessel congestion, chromatolysis, and neuronal degeneration and also causes microglial reaction and neuronophagia, hippocampal and cerebellar neuron degeneration, and demyelination of the axon. In contrast, TQ coadministration (20 mg/kg) with Pb acetate markedly reduces the Pb acetate-induced incidence of pathological lesions [133].

Arsenic (As) is capable of crossing the blood-brain barrier and accumulates in different regions of the brain. As exposure is associated with ROS generation, which is supposed to be one of the mechanisms of As-induced oxidative stress [134]. TQ has notable potential effects against As-induced neurotoxicity. Significant and dose-dependent alterations in the level of enzymatic and biochemical biomarkers of oxidative stress were observed in As-treated system. However, pretreatment with TQ (10 μM) significantly reduces As-induced neurotoxicity. Further, a significant decline in As-induced DNA damage was documented with pretreatment of TQ in a comet assay [135]. And also, a recent study reported the ameliorative effect of TQ in As-induced neurotoxicity through antioxidative and antineuroinflammatory mechanisms in female mice. Treatment with arsenate (20 mg/kg) diminishes the levels of norepinephrine, dopamine, AChE, and Na⁺-K⁺-ATPase activities in the cerebral cortex, cerebellum, and brain stem of rats and, likewise, decreases GSH, GSH-Px, CAT, SOD, and GR concentrations. In contrast, As treatment subsequently elevates the levels of 5-hydroxytryptamine, MDA, NO²/nitrate, and TNF-α. The presence of degenerated Purkinje cells in the cerebellum was also noticed. Posttreatment with TQ (10 mg/kg) suppresses arsenate-induced neurotoxic effects as decreasing the levels of 5-hydroxytryptamine, MDA, NO, and TNF-α and raising the concentrations of norepinephrine, dopamine, GSH, GSH-Px, GR, SOD, and CAT in the cerebral cortex, cerebellum, and brain stem. Likewise, TQ posttreatment increases AChE and Na⁺-K⁺-ATPase activities [136].

A variety of mechanisms has proposed to contribute to ethanol-induced neurotoxicity including excitotoxicity, disruption of cell-cell interactions, apoptosis induction, oxidative stress, and interference with growth factor activity [137]. TQ exhibits protective effects against ethanol-induced neurotoxicity, specifically against apoptosis and cell death. After ethanol (100 mM) exposure for 12 hours, TQ (25 μM) separately and synergistically with metformin (10 mM) improves cell viability and attenuates cytotoxic-free calcium [Ca²⁺]c elevation. Additionally, ethanol exposures typically lower the physiological mitochondrial transmembrane potential (ΔΨm) but metformin and TQ maintain these conditions. In addition, ethanol reduces the expression of Bcl-2 and stimulates the release of cytochrome c from the mitochondria. Through the rise of Bcl-2 expression, metformin and TQ treatment prevents the apoptotic cascade. TQ also suppresses caspase-9 and caspase-3 activation and reduces poly [ADP-ribose] polymerase 1 cleavage. With respect to the relevant staining methods, ethanol treatment confirms more cell death, while metformin and TQ separately or metformin with TQ combined prevents ethanol-induced apoptotic cell death [138].

As a volatile organic compound, toluene is extensively used as an industrial solvent. It is capable of impacting the CNS like depressants, through excitation and psychomotor impairment, followed by inhibition of locomotor activity and sedation. It also brings about oxidative stress, memory loss, and progressive nerve and brain damage [139]. TQ has potential in mediating the effects of toluene-induced neurodegeneration in rat hippocampus and frontal cortex. Chronic toluene exposure causes severe degenerative changes, shrunken cytoplasm, slightly dilated cisternae of the endoplasmic reticulum, markedly swollen mitochondria with degenerated cristae, and nuclear membrane breakdown with chromatin disorganization in neurons of the hippocampus and frontal cortex. TQ (50 mg/kg) treatment protects against such effects [139, 140].

Acrylamide (ACR) is considered a well-documented neurotoxic agent in both humans and laboratory animals, and its existence is a worldwide concern. In addition, sub-chronic, low-level of occupational human exposure to ACR can lead to neurotoxicity defined by ataxia, skeletal muscle weakness, and numbness of the hands and feet [141]. Oxidative damage has suggested as a mechanism of neurotoxicity induced by ACR [142]. With this, TQ features protective effects in ACR-induced neurotoxicity through preventing oxidative stress. In one experiment, male Wistar rats were treated with ACR (50 mg/kg i.p.) alone or with TQ (2.5, 5, and 10 mg/kg i.p.) for 11 days. Exposure to ACR resulted in severe gait abnormalities while treatment with TQ significantly decreases them. ACR exposures elevate the concentrations of MDA in the cerebral cortex while TQ treatment significantly and dose dependently reduces lipid peroxidation [143].

5.3. Radiation-Induced Brain Damage. Through interacting with living cells, ionizing radiation (IR) results in several cytotoxic properties that are mediated through ROS/reactive nitrogen species production. In the brain, TQ modulates IR-induced reactive nitrogen species. One study described a single dose (5 Gy) of total cranial gamma irradiation given to rats and in therapy groups, the dosing of TQ (30 mg/kg/day i.p.) starting 30 min before the radiation dose and after the IR continuing daily for 10 days. In treated rats, N. sativa oil or TQ lowers the NO and peroxynitrite levels as well as nitric oxide synthases enzyme activity in brain tissue than in those animals that received ionizing radiation alone [144].

5.4. Morphine-Induced Tolerance and Dependence. Repeated administration of morphine and related opiates causes tolerance and dependence [145]. TQ combats morphine-induced
tolerance and dependence in mice [44, 146]. In treatment
groups, the i.p. doses of TQ (10 mg/kg) administered
30 min before morphine injections continued for up to seven
days. TQ inhibits the increase in brain MDA concentrations
and NO production, as well as the declines in brain GSH level
and GSH-Px activity, induced by repetitive morphine and
associated naloxone-induced withdrawal [146]. Furthermore,
administration of single or repeated doses of TQ (20
and 40 mg/kg i.p.) significantly decreases the number of
jumps in morphine-dependent animals. In addition, TQ
attenuates tolerance to the analgesic effect of morphine while
high dose of TQ (40 mg/kg) impairs the motor coordination
of animals [44].

A summary of TQ’s protective effects against chemical-
induced neurotoxicity, radiation-induced brain damage,
and morphine-induced drug tolerance and dependence is
found in Table 3.

### 5.5. Protective Activity in Streptozotocin-Induced Model

In a streptozotocin-induced diabetes model, in the rat’s brain,
pretreatment with TQ (10 mg/kg p.o.) significantly attenuates
increased NO and MDA concentration compared to the
control group. In addition, streptozotocin-induced diabetes
also significantly decreases GSH and glutathione
s-transferase and CAT; TQ reverses these changes. The
increase in GSH and CAT activity with TQ administration
may reduce oxidative stress in diabetes mellitus. Moreover,
because of chemical nature, TQ reacts with GSH, nicotin-
amide adenine dinucleotide, and nicotinamide adenine
dinucleotide phosphate. Henceforth, a probable intracellu-
lar nonenzymatic activation of TQ was presumed depend-
ent upon GSH, nicotinamide adenine dinucleotide, and
nicotinamide adenine dinucleotide phosphate representing
the “cellular switch” for moderating cellular antioxidant
defenses [147].

### 6. Challenges and Delivery Prospects of Thymoquinone in Neurological Disorders

Several clinical trials of TQ have been conducted, but owing
to its hydrophobic properties, many phase studies have not
been carried out [77]. Thus, TQ clinical studies are a major
challenge for researchers. In a phase I trial, it was found safe
and well tolerated in patients up to 10 mg/kg/day [148]. In
2010, a study was performed that TQ was administered to a
group of children with epilepsy of a sample size of 22 [149].
Regarding this study, TQ had antiepileptic effects in children
with refractory seizures [149]. Recently, a study was con-
ducted on sialidosis type 1, an autosomal recessive disorder
causing neuraminidase deficiency that causes sialic acid accu-
mulation in tissues. At present, there is no disease-modifying
treatment. As a weak histone deacetylase inhibitor, TQ is a
Neu 4 sialidase activity agonist and vorinostat is a more
potent histone deacetylase inhibitor. However, a clear physi-
ological effect was evidence with TQ on a patient with
sialidosis type 1, but not with vorinostat [150].

With respect to the neuropharmacological potential of
TQ, it might be a valuable agent against neurological disor-
ders, hence, its formulations and delivery merit investigation.
Delivery is also relevant to important to clinical trial studies
of TQ against neurological disorders. As per our previous
discussion, the chemical nature of TQ is hydrophobic, which
causes poor water solubility specifically leading poor formul-
aton characteristics. In human, because of its chemical
nature, it has weak membrane penetration [151]. Moreover,
TQ has less bioavailability owing to its slow absorption and
rapid elimination behavior [60]. Therefore, increasing bio-
availability is essential for better pharmacological activities.

In a liver cirrhosis model, encapsulated solid lipid nano-
particles (SLN) of TQ (TQ-SLN) improve oral delivery.
With this formulation, an enhanced bioavailability and
pharmacodynamics profile could be achieved [152]. In a pharmacokinetic study model, after oral administration to animals, TQ-SLN bioavailability was increased nearly a five-fold. According to the drug distribution analysis, TQ-SLN were found to accumulate more so in the brain than other organs, henceforth, its suitability for brain-targeted drug delivery [153]. Several self-nanoemulsifying drug delivery systems (SNEDDS) of TQ have also been developed and evaluated with regard to oral bioavailability. The oral administration of optimized SNEDDS shows significant improvement in in vivo absorption, as well as bioavailability (3.87-fold) of TQ versus TQ suspension [62]. TQ-loaded liposomes and TQ loaded in liposomes modified with Triton X-100 with diameters of roughly 100 nm were found to maintain stability and improve bioavailability [151]. TQ-encapsulated nanoparticles with 97.5% effectiveness in a biodegradable nanoparticulate formulation overcome the poor solubility, thermal and light sensitivity, and minimal systemic bioavailability [154–156]. Additionally, TQ-NLC have been developed to improve bioavailability (elimination half-life~five hours) [157, 158].

A modified form of TQ through chemical conjugation of the quinones with penetrating cations produces neuroprotective activity in a model of middle cerebral artery occlusion. Concerning the study assumption, the mitochondria-targeted delivery of modified TQ significantly increases the drugs’ antioxidant efficiency leading to neuroprotective activity against I/R injury [159]. In one investigation of optimized TQ-loaded PLGA-chitosan nanoparticles delivered via nose-to-brain route, TQ elicits neuroprotective activity in a rodent cerebral I/R model. Intranasal delivery of optimized TQ-loaded PLGA-chitosan nanoparticles to the brain significantly reduces the ischemic infarct volume and enhances the locomotor activity and grip strength in the middle cerebral artery-occluded rats. With respect to the biochemical studies, the intranasal delivery of TQ-loaded PLGA-chitosan nanoparticles significantly diminishes lipid peroxidation but elevates antioxidant enzyme activity in the brain. Pharmacokinetic and localization studies showed that TQ-loaded PLGA-chitosan nanoparticles facilitate the delivery of TQ to the brain by an intranasal nose-to-brain transport pathway and enhanced their pharmacokinetic profile in brain tissues [107].

Many novel formulations of TQ have not yet been assessed for neurological disease models. Accordingly, we expect that the aforementioned TQ formulations might be probable candidates for the different neuropharmacological models. As a probable candidate, different formulations of thymoquinone for neurological disorders are portrayed in Figure 2.
7. Concluding Remarks

In recent times, phytochemicals have been reported as potential therapeutic alternative candidates for neurological disorders. Regarding the side-effects of modern medicine versus its benefits, the discovery of lesser and/or nontoxic drugs is a principle objective for researchers. Extracts from the various plant parts have potential in various neurological disorders. At the present time, the identification of compounds in the extract responsible for potential activity is also challenging. As a major constituent in *N. sativa*, last few years of research have emphasized research on TQ in various disorders.

In the discussion, by demarcating neurological disorders, we have presented the data concerning the updated pharmacological potential and delivery prospects of TQ. As per our overview, based on neuropharmacological potential, TQ is an indeed an important therapeutic candidate in the treatment of neurological disorders. It features emerging activities in various disease models as well as trauma- and chemical-induced CNS impairments. Further, the effects of TQ on neuroinflammation, neurotoxicity, and neurological disorders, where oxidative stress reduction is a significant effect. Apart from its neuropharmacological activities, the formulation and delivery concepts of TQ as related to neurological disorders are also vital. Henceforth, this study discussed the formulation and delivery prospects of TQ in neurological disorders. This study might be helpful (i) to foster deeper study of TQ in neurological disorders, (ii) to identify the pharmacodynamic profile of TQ, (iii) to discover potential analogs of TQ through various synthetic pathways, and (iv) to design clinical trials to assess TQ in treating neurological disorders. However, many studies have been conducted but further investigations including laboratory-based and bioinformatics approaches are recommended. These investigations might be helpful to establish the precise pharmacological safety, efficacy, and potency profiles of TQ in the therapy for neurological disorders. Overall, in the near future, TQ and its modified forms might contribute in the field of neuromedicine to cure of various neurological diseases and disorders.

**Abbreviations**

6-OHDA: 6-Hydroxydopamine  
Aβ: Amyloid beta  
AD: Alzheimer’s disease  
AMPK: AMP-activated protein kinase  
AMP: Adenosine monophosphate  
ACHE: Acetylcholine esterase  
CAT: Catalase  
EAE: Experimental autoimmune encephalomyelitis  
GSH: Glutathione  
Gr: Glutathione reductase  
GSH-Px: Glutathione peroxidase  
GABA: γ-Aminobutyric acid  
i.p.: Per intraperitoneal administration  
i.c.v.: Per intracerebroventricular administration  
IC$_{50}$: Half maximal inhibitory concentration  
iNOS: Inducible nitric oxide synthase  
I/R: Ischemia/reperfusion  
IL: Interleukin  
LD$_{50}$: Median lethal dose  
LPS: Lipopolysaccharide  
LDH: Lactate dehydrogenase  
MDA: Malondialdehyde  
MPP$: 1-Methyl-4-phenylpyridinium ion  
NF-κB: Nuclear factor kappa-light-chain-enhancer of activated B cells  
NO: Nitric oxide  
NO$_2$–: Nitrite  
ONOO (−): Peroxynitrite  
PD: Parkinson’s disease  
PTZ: Pentylentetrazole  
p.o.: Per oral administration  
ROS: Reactive oxygen species  
SOD: Superoxide dismutase  
SE: Status epilepticus  
SIRT1: Sirtuin 1  
TQ: Thymoquinone  
TNF-α: Tumor necrosis factor-α  
TBARS: Thiobarbituric acid reactive substance  
THir: Tyrosine hydroxylase immunoreactive.

**Conflicts of Interest**

The authors declare that there is no conflict of interests regarding the publication of this paper.

**Authors’ Contributions**

Md. Jakaria and Dong-Kug Choi conceived and designed the study. Md. Jakaria also wrote the manuscript including reviewing the literature, drawing the figures, and making the tables. Duk-Yeon Cho and Md. Ezazul Haque contributed by arranging the data. Govindarajan Karthivashan and In-Su Kim contributed by drawing the figures. Palanivel Ganesan contributed by arranging and editing the data. Dong-Kug Choi also performed supervision and correspondence. All authors read and approved the final manuscript.

**Acknowledgments**

This research was supported by The Leading Human Resource Training Program of Regional Neo Industry through the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT and Future Planning (Grant no. NRF-2016H1D5A1909610). This research was also supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Science and ICT (Grant no. 2017R1A2A2A07001035).

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