

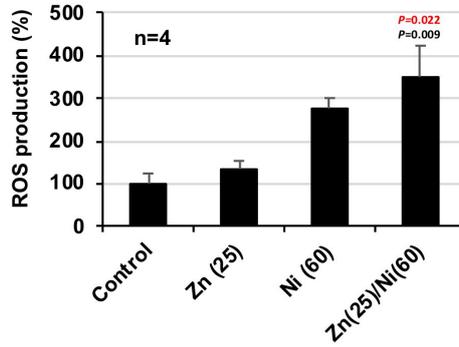
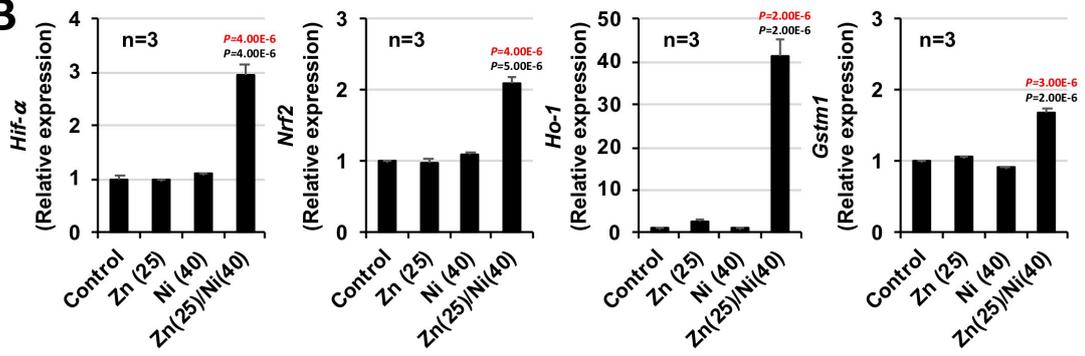
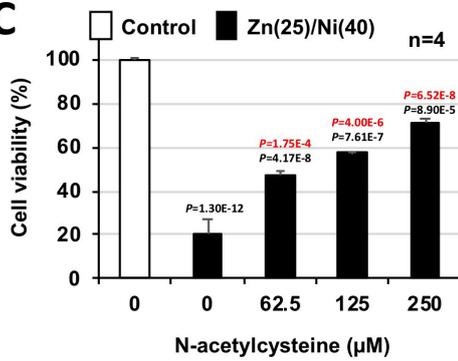
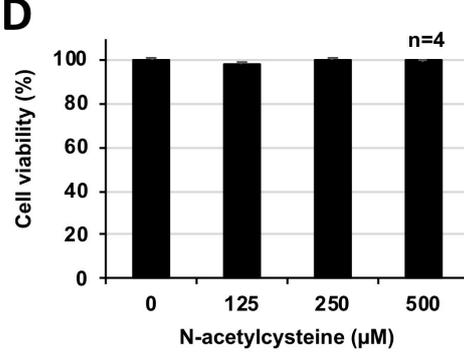
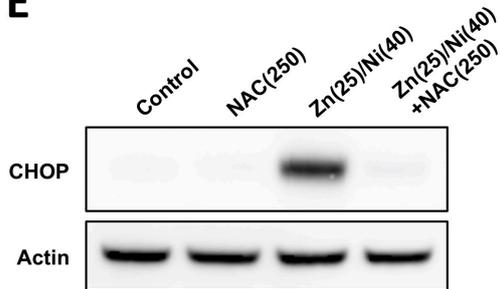
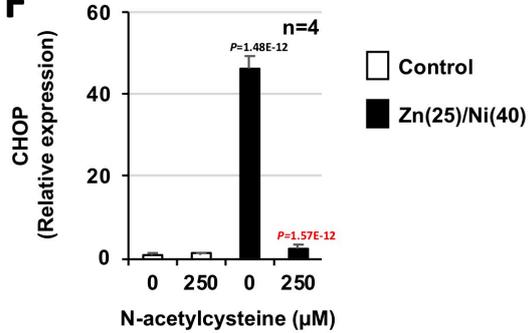
Nickel enhances zinc-induced neuronal cell death by priming the endoplasmic reticulum stress response

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A**B****C****D****E****F**

Supplementary Figure S1

GT1-7 cells (6-well culture plates at a density of 7.5×10^5 cells per well) were incubated with DCFHDA (100 μM), an indicator of ROS, in the absence (Control) or presence of NiCl_2 (Ni, 60 μM) and/or ZnCl_2 (Zn, 25 μM) for 2 h. DCFHDA fluorescence (ex 485 nm, em 530 nm) was measured using a fluorescence microplate reader (A). GT1-7 cells (6-well culture plates at a density of 7.5×10^5 cells per well) were incubated in the absence (Control) or presence of NiCl_2 (Ni, 40 μM) and ZnCl_2 (Zn, 25 μM) for 4 h. Total RNA was extracted and subjected to real-time RT-PCR using primer sets specific for each gene. Values were normalized to *Gapdh* and expressed relative to the control (B). GT1-7 cells (96-well culture plates at a density of 3×10^4 cells per well (C, D), 6-well culture plates at a density of 7.5×10^5 cells per well (E, F)) were pre-treated with the indicated concentrations (μM) of N-acetylcysteine just before $\text{Ni}^{2+}/\text{Zn}^{2+}$ treatment. Next, GT1-7 cells were incubated in the absence (Control) or presence of NiCl_2 (40 μM) and ZnCl_2 (25 μM) for 24 h (C) or 7 h (E, F). GT1-7 cells were treated with the indicated concentrations of N-acetylcysteine for 24 h (D). Cell viability was determined using CellTiter-Glo[®] 2.0 (C, D). Whole-cell extracts were analyzed by immunoblotting with an antibody against CHOP or actin (E). The band intensity of CHOP was determined using ImageJ software (F). Values represent mean \pm S.E.M. *P* values are described in the figure when *P* < 0.05. [Black: vs. Control, Red: vs. ZnCl_2 alone] (A, B) or [Black: vs. Control, Red: vs. $\text{Zn}(25)/\text{Ni}(40)$] (C, F).