

Review Article

Parkinson's Disease-Induced Zebrafish Models: Focussing on Oxidative Stress Implications and Sleep Processes

Madalina-Andreea Robea,¹ Ioana-Miruna Balmus,^{2,3} Alin Ciobica ,³ Stefan Strungaru,³ Gabriel Plavan,¹ Lucian Dragos Gorgan ,¹ Alexandra Savuca,¹ and Mircea Nicoara¹

¹Department of Biology, Faculty of Biology, "Alexandru Ioan Cuza" University of Iasi, Carol I Avenue, No. 20A, Iasi, Romania

²Department of Interdisciplinary Research in Science, "Alexandru Ioan Cuza" University of Iasi, Carol I Avenue, No. 11, Iasi, Romania

³Department of Research, Faculty of Biology, "Alexandru Ioan Cuza" University of Iasi, Carol I Avenue, No. 20A, Iasi, Romania

Correspondence should be addressed to Alin Ciobica; alin.ciobica@uaic.ro and Lucian Dragos Gorgan; lucian.gorgan@uaic.ro

Received 28 April 2020; Revised 7 July 2020; Accepted 23 July 2020; Published 18 August 2020

Academic Editor: Daniel Lopez-Malo

Copyright © 2020 Madalina-Andreea Robea et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The complex yet not fully understood pathophysiology of Parkinson's disease includes an important molecular component consisting of oxidative status changes, thus leading to oxidative stress occurrence. While no particular evidence has been reported that describes the relationship between oxidative stress and the molecular mechanisms behind Parkinson's disease development, animal model studies has shown that oxidative stress induction could modulate Parkinson's disease symptomatology. Despite the inability to perfectly replicate human disease in animals and despite that Parkinson's disease has not been reported in any animal species, animal modeling is one of the most important tools in understanding the complex mechanisms of human disorders. In this way, this study is aimed at detailing this particular relationship and describing the molecular mechanisms underlying Parkinson's disease in animal models, focusing on the potential advantages and disadvantages of zebrafish in this context. The information relevant to this topic was gathered using major scientific database research (PubMed, Google Scholar, Web of Science, and Scopus) based on related keywords and inclusion criteria. Thus, it was observed that oxidative stress possesses an important role in Parkinson's disease as shown by numerous animal model studies, many of which are based on rodent experimental models. However, an emerging impact of the zebrafish model was observed in the research of Parkinson's disease pathological mechanisms with regard to disease development factors and the cause-effect relationship between oxidative stress and comorbidities (such as depression, hyposmia, fatigue, sleep disturbances, and cognitive deficits) and also with regard to the pharmacological potential of antioxidant molecules in Parkinson's disease treatment.

1. Introduction

Oxidative stress (OS) greatly impacts the human body leading to well-known pathologies, such as diabetes, atherosclerosis, Alzheimer's disease, and Parkinson's disease (PD) [1, 2]. The main cause of OS occurrence is due to the imbalance between reactive oxygen species (ROS) production and the ability of the biological systems to transform ROS into harmless oxygen species (such as water), or to detoxify the intermediate metabolites or to repair the oxidative damage [1, 3].

The concept of OS implication in mental illness was previously described [4, 5]; however, it is gen-

erally known that, with several exceptions, the occurrence of mental illnesses consists of the interaction between genetic or developmental events and environmental factors [6, 7]. Based on the context that mitochondrial dysfunction is facilitated by several different mechanisms and also on the fact that high levels of ROS are needed in the nervous system due to autophagy and mitophagy functions [8], OS occurrence in both the normal and pathological brain functions is currently accepted as a normal yet controlled mechanism.

Considering all these aspects, PD can be defined as a complex neuropsychiatric disorder occurring mostly in elders, which is commonly characterized by dopaminergic

system malfunction leading to muscular tonus loss [9]. The exact cause of PD is unknown, but several hypotheses point at genetic inheritance, drugs, and environmental factors, such as pesticides, heavy metals, cigarette smoking, and caffeine [10, 11]. The administration of several chemical compounds can influence the entire cell metabolism leading to a cascade of events as shown in Figure 1.

The emerging use of the zebrafish model in neurological and neurodegenerative human diseases (such as PD, autism, Huntington's disease, and Alzheimer's disease) was described by Xi et al. [12] and Brennan [13]. Despite that some disease phenotypes which are caused by orthologue genes can be very different, particularly when comparing fish and humans [14], it was shown that more than 70% of all human disease genes have functional homologs in *Danio rerio* [15, 16]. In this way, the fast growing and easy-to-breed zebrafish could be a reasonable choice when contemplating to keep thousands of animals at low costs [16, 17]. While zebrafish was originally considered as a bridge connection in the experimental gap between fly/worm and mouse/human in studying embryo development, it was shown that as the new research methodologies and genetic tools were updated, the zebrafish animal model was reported to be well suited to both developmental and genetic analysis [18] as well as complex human disorders [19, 20].

In this context, experimental animal models are needed to provide additional evidence on PD etiology, mechanisms, and possible therapeutic interventions. Thus, in this study, we aimed to describe the influence of OS on the Parkinsonian nervous system, as previously shown also by our research group on rodent models [12–15]. Furthermore, considering the emerging use of zebrafish in the novel worldwide research endeavours, we aimed to compare and elaborate the zebrafish neurophysiology model in PD research with regard to disease development factors, cause-effect relationship of OS and comorbidities (focussing on sleep disturbances), and the pharmacological potential of antioxidant molecules.

2. Materials and Methods

The search strategy included the use of major scientific databases (PubMed, Google Scholar, Web of Science, and Scopus) for research of scientific articles published between 1990 and 2020. The following search keywords were used: “oxidative stress,” “Parkinson's disease,” “animal model,” “rat,” “mice,” “zebrafish,” “1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP),” “rotenone,” “paraquat,” and “neurotoxin.” The process of scientific article selection considered only reports written in English language, and the selection was conducted by four separate researchers (Robea M.-A., Balmus I.-M., Savuca A., and Ciobica A.) whose differences in opinion were resolved by common consent. After the initial scientific database research, all the scientific articles were reviewed considering some inclusion criteria, such as the reports that (1) included information/research data on the oxidative changes occurring in Parkinson's disease, or described the molecular pathways of Parkinson's disease in relation to human pathology, or presented significant results on Parkinson's disease treatments (in relation to oxidative balance pathways) and

(2) included relevant information on the Parkinson's disease animal models (in relation to oxidative balance pathways), or described the molecular pathways of Parkinson's disease symptoms' means of modulation in animals, or presented significant results and correlations on the Parkinson's disease treatments (in relation to oxidative balance pathways and antioxidant potential). Exclusion criteria were formulated to avoid duplicate studies (i.e., studies on the same antioxidant molecule), studies not related to oxidative stress and Parkinson's disease pathological pathways, and irrelevant animal model studies in Parkinson's disease research (Figure 2).

3. Parkinson's Disease and Oxidative Stress

PD is a progressive neurodegenerative disease, which predominantly occurs in the elderly population [24–26]. Characterized by loss of neurons from the substantia nigra, PD leads to inhibition of dopamine production and accumulation of Lewy bodies (LB) formed by α -synuclein aggregates, a presynaptic neuronal protein [11, 27–29]. The result of these neuromolecular changes is translated into several clinical symptoms, such as bradykinesia, resting tremor, rigidity, and postural instability [27, 29–31]. Several nonmotor symptoms, such as depression, hyposmia, fatigue, sleep disturbances, and cognitive deficits such as dementia are often considered comorbidities of PD [11, 27, 29, 32].

Dopamine (DA) synthesis starts with tyrosine and two key enzymes (tyrosine hydrolase and amino acid decarboxylase), whereas its degradation is provided by the action of three key enzymes (monoamine oxidase B, catechol-O-methyl-transferase, and dopamine β -hydroxylase) resulting in two final metabolites and norepinephrine [14, 33]. However, excess DA induces neuronal damage and cell death through ROS generation. Furthermore, ROS accumulation could lead to DNA mutations and to loss of dopaminergic neurons from the substantia nigra [33–35].

Considering that ROS can be produced by different biological structures, many ROS functions have been described mainly according to their reactivity. Mitochondria and metabolism are some of the most important sources of ROS, thus enzymes such as nitric oxide synthase, monoamine oxidase, and xanthine oxidase produce daily huge amounts of the following reactive oxygen and nitrosative species: superoxide ($O_2^{\cdot-}$), hydroxyl (OH^{\cdot}), peroxy (ROO^{\cdot}), nitric oxide (NO^{\cdot}), nitrogen dioxide (NO_2^{\cdot}), dinitrogen trioxide (N_2O_3), nitrosonium cation (NO^+), nitroxyl anion (HNO), and lipid peroxy (LOO^{\cdot}) [9, 10, 36, 37].

Olanow and Tatton [34] and Asanuma et al. [38] both reported an increase in lipid peroxidation and a decrease in the activity of antioxidant enzymes, such as catalase (CAT), glutathione (GSH), and glutathione peroxidase (GPx), in PD patients. Furthermore, dysregulated metal ion homeostasis has been often reported in PD development, particularly iron [39]. High iron levels were discovered in the substantia nigra pars compacta which were presumed to lead to hydroxyl radical (OH) generation due to iron's redox instability [23, 40, 41].

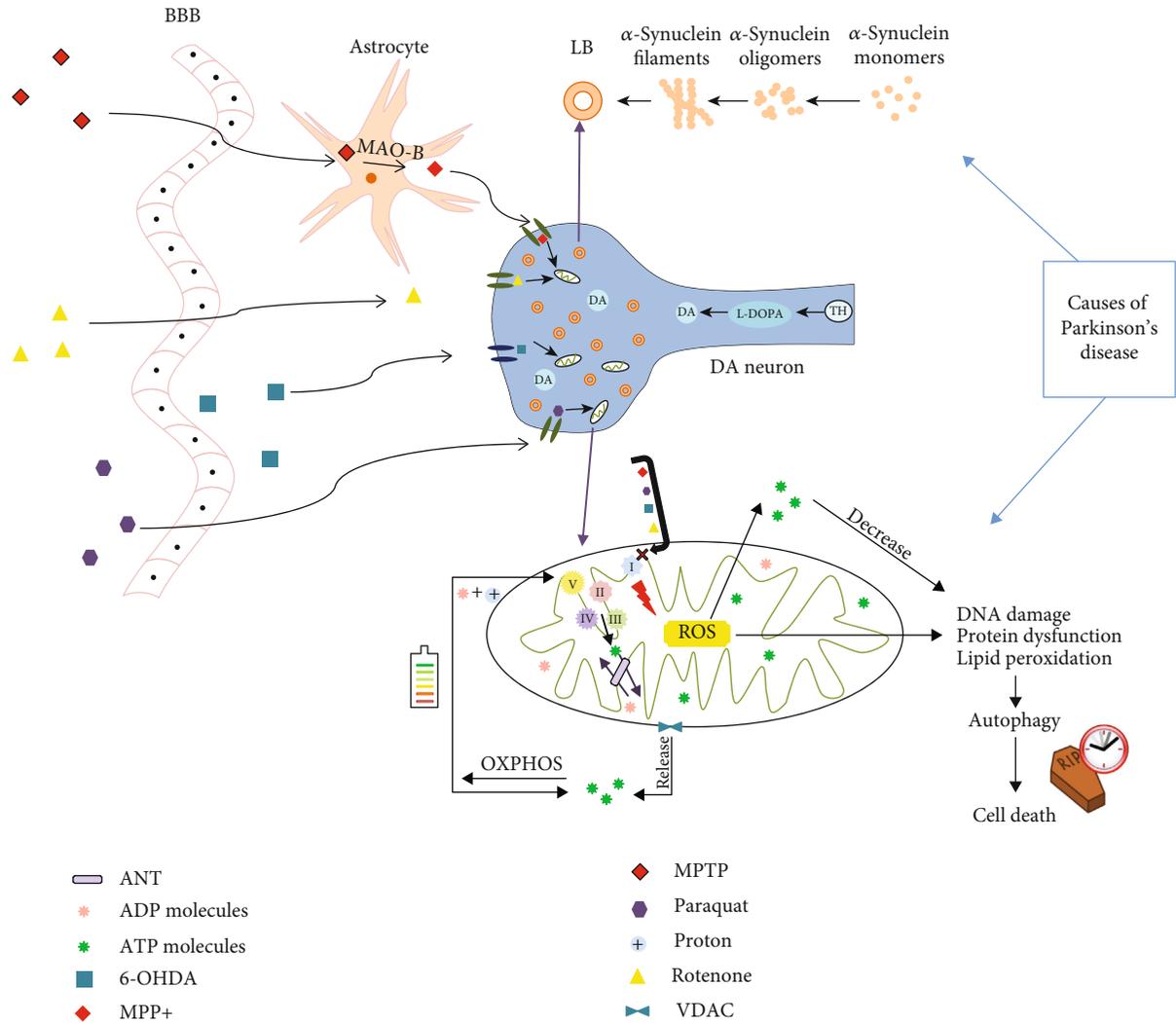


FIGURE 1: Parkinson's disease mechanism of action in the central nervous system and the pharmacokinetic effects of several agents that induce Parkinson-like symptoms. MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) crosses the blood-brain barrier to be metabolized in 1-methyl-4-phenylpyridinium (MPP+) by monoamine oxidase B in the astrocytes. Afterwards, the transportation system of the synaptic cleft assists the intraneuronal MPP+ transfer and transports it further into the mitochondria where it impairs the mitochondrial respiration chain leading to reactive oxygen species production and dopaminergic neuron loss [21]. Similar to MPTP, paraquat could increase reactive oxygen species production, but in contrast to MPTP, it could lead to Lewy body (LB) formation [22]. 6-Hydroxydopamine could also enter the dopaminergic neurons and lead to reactive oxygen species production in the absence of the Lewy body inclusions [23]. Following diffusion to intraneuronal space, rotenone inhibits mitochondrial complex I and promotes the formation of Lewy body inclusions [22, 23]. Abbreviations: 6-OHDA—6-hydroxydopamine; ADP—adenosine diphosphate; ANT—adenine nucleotide translocase; ATP—adenosine triphosphate; BBB—blood-brain barrier; DA—dopamine; LB—Lewy bodies; L-DOPA—levodopa; MAO-B—monoamine oxidase B; MPP+—1-methyl-4-phenylpyridinium; MPTP—1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; OXPHOS—oxidative phosphorylation; ROS—reactive oxygen species; TH—tyrosine; VDAC—voltage-dependent anion channel.

OS implication in PD is supported by postmortem studies and by numerous reports which suggested its active role in PD pathological processes [42]. A link between OS, mitochondrial dysfunction, and glutathione levels was suggested by Di Monte et al. [43] by the implications of DA metabolism by monoamine oxidase (MAO) during which the formation and accumulation of H₂O₂ occurs. Simultaneously, the glutathione resources are depleted while glutathione peroxidase catalyses the reduction of H₂O₂ in H₂O [37, 38]. Furthermore, as a consequence of substantia nigra glutathione depletion and mitochondrial

dysfunction, NO[•] production increases and leads to α-synuclein (α-syn) accumulation [10, 12, 29].

In physiological conditions, oxidised glutathione (GSSG) is reduced by glutathione reductase in reduced glutathione (GSH) using NADPH. However, the GSH depletion could be caused by impaired synthesis of GSH, which is a result of mitochondrial dysfunction since there is not an adequate quantity of ATP to sustain the GSH production [43, 44]. To support this hypothesis, Hauser et al. [45] proved that GSH is reduced approximately 40-50% in PD patients.

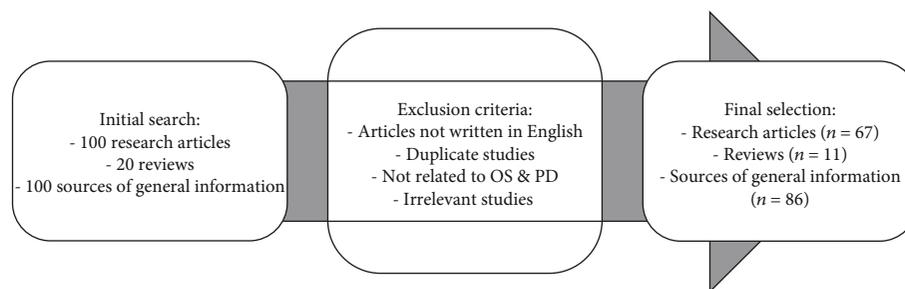


FIGURE 2: The procedure used for the selection of scientific articles.

Similarly, an important pathological mechanism underlying PD pathogenesis could be supported by nitric oxide action due to its capacity to impair synaptic activity, memory functionality, and neuronal plasticity [34, 41]. This aspect was suggested by Ravenstijn et al. [66] while showing that 7-nitroindazole could exhibit an inhibitory effect on nitric oxide synthase from the substantia nigra pars compacta leading to malonate, 3-nitropropionic acid, or MPTP-induced lesion attenuation [46].

As we previously described, mitochondrial dysfunction plays a major role in symptom persistence and disease progression [47, 48]. The decreased rate of ATP production leads to OS and further to cell death [49]. Mitochondrial complex I is often the target in neurodegenerative PD since it plays a crucial role in the mitochondrial respiratory chain [19, 48].

4. Parkinson's Disease Animal Models: Rodents versus Zebrafish

Recent studies showed that mitochondrial dysfunction is a key feature of PD pathogenesis [27, 34, 41, 47, 50]. According to numerous reports, there are several chemical compounds which could influence the activity of mitochondrial complex I. In this way, the modulation of mitochondrial activity could lead to the occurrence of PD-like symptomatology in both rodent and zebrafish models. Thus, rotenone, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), paraquat, 6-hydroxydopamine (6-OHDA), pyrethroids, and organophosphates [23, 51–53] were successfully used to increase the ROS levels and therefore to promote dopaminergic neuron degeneration [54]. This could be the reason why many PD animal models (mainly rodents) are based on the acute/chronic administration of some of the mentioned chemicals and also the evidence which ties OS to PD molecular pathways. Table 1 summarises several animal model studies on PD-like impairments, chemical inductors, and comparisons between the zebrafish and rodent models.

Accumulation of α -syn is a clear sign of PD and one of the main causes to its development [73]. Synucleins, a family of proteins naturally occurring in the nervous system, are known to contribute to vesicle synapse maintenance or DA activity [65–68]. Zebrafish possesses three genes which encode β -, γ 1-, and γ 2-synucleins [66, 67]. Milanese et al. [14] showed that if the β - and γ 1-synucleins are knocked out, zebrafish exhibits hypokinesia and low levels of DA. Zebrafish synucleins (zSynC) share a high similarity with human

synucleins, wherein zSynC is 70% identical and 82% similar to human β -synuclein [74].

Furthermore, Prabhudesai et al. [75] suggested that α -syn accumulation in the zebrafish nervous system leads to neuron apoptosis and death. Their hypothesis was confirmed since CLR01, a molecular tweezer, could increase the survival rate of embryos and suppress α -syn aggregation in a transgenic zebrafish model carrying human wild type α -syn [75].

Previous studies demonstrated that overexpression of α -syn in a hypothalamic neuronal cell line could lead to increased ROS, mitochondrial impairment, and LB [50, 73, 76]. Also, it was suggested that cholesterol metabolites resulting from ROS degradation could promote the aggregation of α -syn [76].

Due to several limitations of the classical chemical-inducing animal models, genetically engineered animal models are now generally preferred. However, the studies on the pathological mechanisms underlying PD development consider more appropriate the use of the chemical-inducing models, since the interplay between the dopaminergic neuron functions and brain redox activity remains a fine game of regulatory potentials. Thus, the study of OS was performed on animal models, such as *Drosophila*, zebrafish, mice, or rats, predominantly using common neurotoxicants (MPTP, 6-OHDA, rotenone, and paraquat) [23, 37, 77, 78]. Thus, the main mechanisms related to Parkinson's disease in the zebrafish central nervous system are schematically presented in Figure 3, together with several agents that induce Parkinson's disease and some treatment alternatives.

5. MPTP

One of the most common chemical agents which can modulate PD symptomatology in animal models is 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) [43, 83]. Following its monoamine oxidase B (MAO-B) promoted conversion to 1-methyl-4-phenylpyridinium in astrocytes [35, 38, 43, 56], MPP⁺ could easily bind DA transporters reaching mitochondria and interfering in the oxidative phosphorylation process carried out by mitochondrial complex I [35, 38, 44, 84]. Also, many reports described the MPTP neurotoxicity to be correlated with tyrosine hydrolase loss and DA transporters [35, 43]. Due to the fact that the participants to this mechanism are highly conserved, OS induction in this way was observed in zebrafish, mice, rats, cats, dogs, and nonhuman primates [35, 85].

TABLE 1: Parkinson's disease animal models based on neuromodulation agents.

PD-inducing agent	Model organism	Treatment	Route of administration	Effects	Reference
MPTP	Zebrafish	5 $\mu\text{g/ml}$ and 10 $\mu\text{g/ml}$, 3 days	Dissolved in the water	↓Locomotor activity ↓Pretecal area size ↓DA levels	[55]
		Single 20 mg/kg dose	Abdominal injection	↓DA and noradrenaline levels ↓Locomotor activity	[40]
		1 \times 50 μg and 2 \times 50 $\mu\text{g}/24\text{ h}$	Intraperitoneal injection	↓Locomotor activity Evidence of freezing bouts	[56]
	Rat	Single 20 $\mu\text{l/kg}$ dose	Intrastantia nigra injection	↑MDA ↓SOD	[57]
		Single 100 $\mu\text{g}/1\ \mu\text{l}$ dose	Bilateral infusion	↑LPO ↓GSH ↑SOD levels in the striatum	[58]
		1 $\mu\text{mol}/2\ \text{ml}$, in the 1st, 7th, and 14th day of the experiment	Intrastantia nigra injection	↑MDA ↓GSSH ↓CAT	[59]
Mice	30 $\mu\text{g/kg}$, twice at 16 h intervals	Intraperitoneal injection	↓GSH ↓SOD in substantia nigra	[60]	
6-OHDA	Zebrafish	Single dose: 25 mg/kg	Abdominal injection	↓Velocity rate and locomotor activity ↓DA neurons	[48]
		Single dose: 25 mg/kg	Intraperitoneal injection	↓DA and noradrenaline levels ↓Locomotor activity	[40]
	Rats	10 $\mu\text{g}/2\ \mu\text{l}$	Unilateral intrastriatal injection	↓GSH ↓CAT ↓SOD	[19]
		8 mg/2 ml	Intrastriatal injection	↑MDA levels ↓GSH and SOD levels in striatum	[20]
	Zebrafish	1-12 mg/kg, 7 to 36 consecutive days	Intravenous injection	↓DA neurons ↑ α -Syn aggregates	[15]
		5 $\mu\text{g/l}$, 28 consecutive days	Dissolved in water	↓Locomotor activity ↓DA neurons ↑ α -Syn aggregates	[16]
Rotenone	Rats	1.5 mg/kg and 2.5 mg/kg, 2 months	Intraperitoneal injection	↓DA neurons in posterior striatum and prefrontal cortex ↑Catalepsy	[17]
		2.2-2.5 mg/kg, 28 consecutive days	Intravenous injection	↓Locomotor activity ↑ α -Synuclein aggregates	[61]
		2.0-3.0 mg/kg, 28-56 days	Subcutaneous injection	↓DA neurons α -Syn aggregates	[62]
		Single 2.5 mg/kg dose	Intraperitoneal injection	↓Body weight ↓DA neurons in striatum	[63]
	Rats	2-12 $\mu\text{g}/\mu\text{l}$, 28-90 days	Stereotaxial infusion	↑ α -Syn aggregates ↓ROS level	[64]
		2.0 mg/kg, 28 days	Subcutaneous injection	↓Locomotor activity ↓DA neurons ↑ α -Syn aggregates	[65]
		5 μg , 21 days	Stereotaxial infusion	↓DA neurons ↑ α -Syn aggregates	[66]
		0.25-0.50 μg , 21 days	Stereotaxial infusion	↑ α -Syn aggregates	[67]
		3 mg/kg, 30 days	Intraperitoneal injection	↑MDA levels ↓GSH and SOD levels	[68]

TABLE 1: Continued.

PD-inducing agent	Model organism	Treatment	Route of administration	Effects	Reference
	Zebrafish	10 mg/kg, twice a day for 3 days	Intraperitoneal injection	↓Locomotor activity Spatial memory impairments	[69]
		1, 10, and 100 μ M, 4 days	Dissolved in the water	↓Mitochondrial respiration	[70]
Paraquat	Mice	0, 0.89, 2.67, and 8 mg/kg, 28 days	Oral administration	↑MDA in HIP ↑Mitochondrial injury	[71]
		Paraquat (10 mg/kg) + maneb (30 mg/kg), twice a week, 9 weeks	Intraperitoneal injection	↑MDA ↑NO ↓GST	[72]

↑: increase; ↓: decrease; CAT: catalase; DA: dopamine; GPx: glutathione peroxidase; GSH: glutathione; GSSG: oxidised glutathione; GST: glutathione S-transferase; LPO: lipid hydroperoxide; MDA: malondialdehyde; NO: nitric oxide; SOD: superoxide dismutase.

In zebrafish, the reports showed that MPTP could efficiently induce decreased locomotor activity, which is caused by DA activity decrease, number of DA neurons, and pretecal size reduction [45, 49, 86–88]. Also, MPTP could induce bradykinesia manifested in zebrafish as decreased velocity and abnormal swimming behaviour [49, 86, 89]. Moreover, as compared to rodent models which seemed to overcome the short-term toxin activity on locomotion or even exhibit no changes or hyperactivity [90], zebrafish showed behavioural changes even in acute MPTP administration (Table 1).

However, neuromodulation efficiency depends on several factors, such as the administration route, sex, strain, and developmental stage [78, 91]. For example, larval zebrafish is more sensitive to MPTP than adult zebrafish [92]. Also, Jackson-Lewis et al. [93] showed that MPTP administration effects could be influencing the size of central nervous system damage in a dose-dependent manner, since they demonstrated that the impairment of tyrosine hydroxylase (TH) activity following lower doses of MPTP could not lead to DA neuron loss in mice. Thus, Kirchoff et al. [94] reported that two MPTP injections (15 or 20 mg/kg) for 7 consecutive days were sufficient to cause dopaminergic neuron loss in mice. Thus, in a recent report of Mingazov et al. [95], it was demonstrated that 12 mg/kg b.w./day subcutaneous MPTP treatment for 2 weeks could not induce DA metabolism impairment or MAO-A and MAO-B enzyme activity changes even with DA neuron loss confirmation.

Similarly, the reports on DA neuron loss in zebrafish are rather controversial. In this way, it was shown that at 24 hours postfertilization, zebrafish embryos immersed in MPTP solution (800 μ M, distilled water) for 2 days exhibited posterior tuberculum DA neuron function impairment leading to behavioural deficits [96]. Also, since L-deprenyl treatment—which is a potent MAO-B inhibitor—could prevent neurodegeneration in zebrafish [55, 97], it could be suggested that a similar mammalian MPTP catabolism pathway could be present in zebrafish; however, it is a known fact that they possess a single monoamine oxidase homologous to both MAO-A and MAO-B [55].

In this way, despite the demonstrated limitations of the rodent models, mice are preferred for DA neuron loss models, since there is no previous report accounting for the

neuronal loss that was not present after MPTP administration in mice and rats.

6. 6-OHDA

Because it is incapable of passing the blood-brain barrier, the catecholamine 6-hydroxydopamine (6-OHDA) was the first neurotoxin used to model PD [23, 98] by direct/sham injection [98, 99]. The 6-OHDA mechanism of action consists of mitochondrial complex I and IV inhibition [23, 98, 100–102] and also autooxidation resulting in O_2^- and H_2O_2 . In this way, the major effect of 6-OHDA is the DA neuron impairment.

Regarding the active potential of 6-OHDA to model PD symptomatology in zebrafish, it was observed that its administration could lead to changes in both biochemical and behavioural parameters. In this way, several studies reported that the 6-OHDA administration led to a decrease in dopamine and noradrenaline levels and also suggested that OS could develop in a cause-effect relationship with regard to 6-OHDA [40, 48]. Furthermore, it seems that 6-OHDA could successfully model in zebrafish one of the main PD symptoms—motor disturbances—as it was previously suggested that zebrafish larvae exposed to a 6-OHDA solution exhibited salient motor impairments and decreases in tyrosine hydroxylase activity [40, 48, 80–82].

Also, the other PD hallmark symptom—DA neuron loss—was evaluated in zebrafish models by Parnig et al. [103] and Vijayanathan et al. [48]. Thus, they showed that irrespective of the administration route and developmental stage (e.g., Parnig et al. treated zebrafish embryos with 250 μ M 6-OHDA dissolved in the water, while, Vijayanathan et al. microinjected 25 mg/kg 6-OHDA in the ventral diencephalon of adult zebrafish), 6-OHDA treatment as short as 3 days could lead to DA neuron loss [48, 103].

Moreover, given the many differences between zebrafish and rodents, similar effects of 6-OHDA administration were reported in rodent models [101]. Also, regarding the OS implications of 6-OHDA in rodent models, the antioxidant role of GPx overexpression and thus an increase in activity for the DA neuron protection was demonstrated. In this way, Bensadoun et al. [104] suggested that one of the most

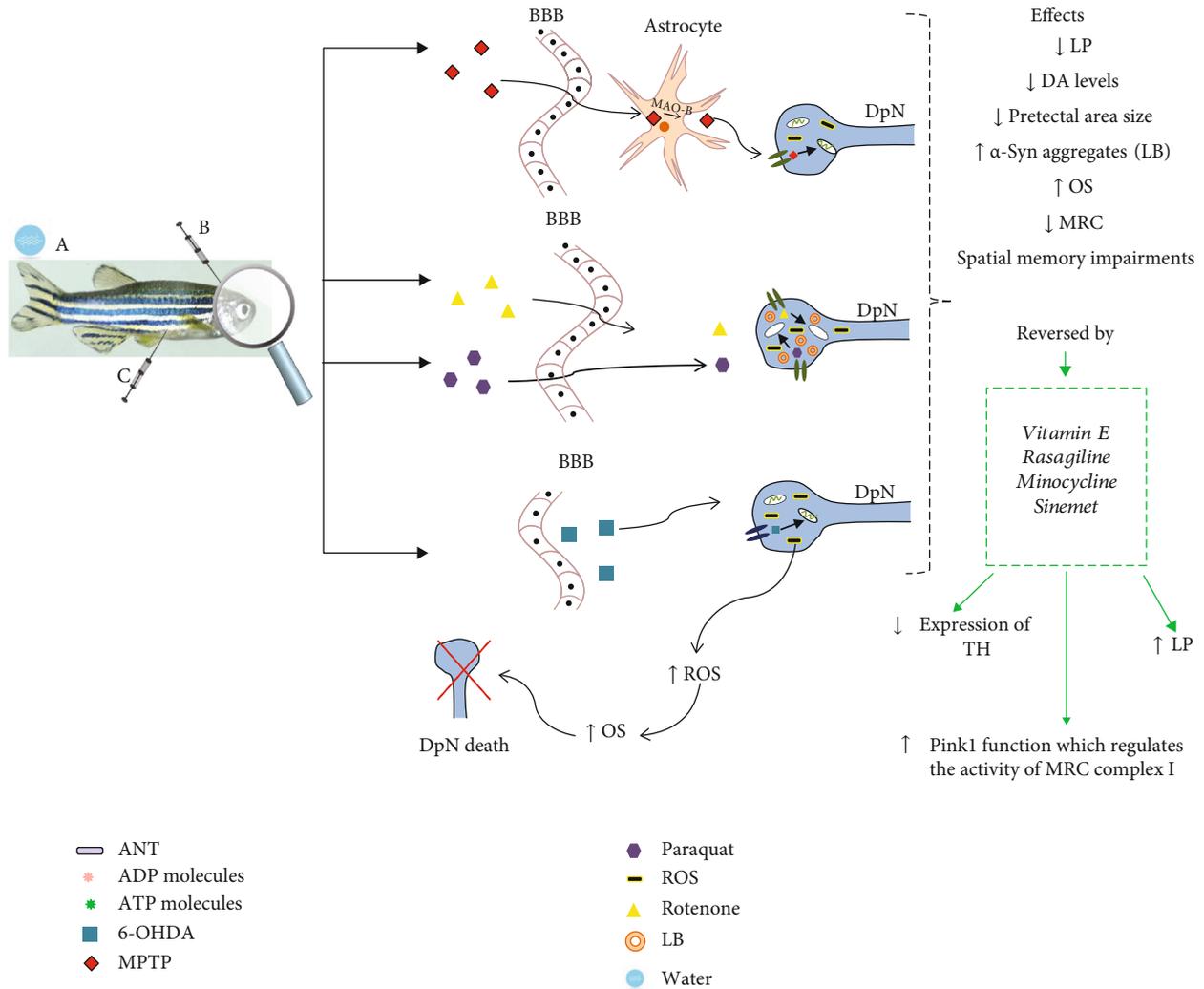


FIGURE 3: Parkinson's disease molecular mechanisms and effects in the zebrafish central nervous system induced by several Parkinson's disease agents and treatment alternatives. Administration of these chemicals (MPTP, rotenone, paraquat, and 6-OHDA) through various ways can lead to a reduction in locomotor parameter activity, a decrease of dopamine neuron number, an increase of oxidative stress, and the inhibition of mitochondrial complex I promoting the formation of Lewy body inclusions [48, 69, 70, 79]. Vitamin E, rasagiline, minocycline, and Sinemet can reverse the action of the Parkinson's disease agents mentioned above in zebrafish [80–82]. Abbreviations: ↑—increase; ↓—decrease; 6-OHDA—6-hydroxydopamine; A—dissolved in the water; ADP—adenosine diphosphate; ANT—adenine nucleotide translocase; ATP—adenosine triphosphate; B—intracerebroventricularly injection; BBB—blood-brain barrier; C—intraperitoneal injection; DA—dopamine; DpN—dopaminergic neuron; LB—Lewy bodies; L-DOPA—levodopa; LP—locomotor parameters; MPTP—1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; MAO-B—monoamine oxidase B; MRC—mitochondrial respiratory chain; OS—oxidative stress; ROS—reactive oxygen species; TH—tyrosine.

prominent components of the anti-6-OHDA neurotoxic effect is the GPx antioxidant enzyme which not only prevented the dopamine secretion inhibition but also indirectly modulated the tyrosine hydroxylase activity.

7. Rotenone

Despite that MPTP and 6-OHDA administrations are one of the most common ways to modulate PD symptomatology in animal models, more recent research revealed that several nonintended molecules possess the potential to induce PD [22, 23, 105]. For example, exposure to some extensively used herbicides and pesticides was shown to lead to mitochondrial

dysfunction and DA neuron loss. In this category stands rotenone, which is an alkaloidal pesticide used worldwide [106] and which acts on mitochondrial complex I and DA neurons [17, 63, 106–109]. According to Alam and Schmidt's report [17], rotenone could destroy DA neurons and induce PD symptomatology in rats following 2 months of intraperitoneal treatment. Also, it was shown that rotenone administration could lead to α -syn aggregate formation and increased oxidant levels [15, 61, 62, 64–67].

The cytotoxic effects of rotenone seen in fish are similar to those observed in rodents. For example, the administration of rotenone in zebrafish could lead to impaired motor ability, olfactory dysfunction, and decreased DA levels [16,

110]. Also, DA neuron reduction was reported by Martel et al.'s study [111], wherein zebrafish embryos received for 7 days a dose of 30 μM rotenone. These aspects were previously described to be a part of the natural mechanism of action of rotenone used to eradicate pests. Moreover, the OS-causing potential of rotenone was described and additional evidence was brought by Melo et al. [112] who demonstrated that 20 $\mu\text{g l}^{-1}$ rotenone can inhibit CAT, GST, and acetyl cholinesterase activity.

Another recent report showed that a 4-week administration of 2 $\mu\text{g l}^{-1}$ rotenone to adult zebrafish leads to 40–50% TH expression decrease. Also, the decrease of DA level was correlated with impaired locomotor activity as a motor symptom and anxiety behaviour specific to PD individuals [110]. As stated before, rotenone is a potent tool to induce motor and nonmotor symptoms of PD in a zebrafish model [111, 112].

8. Paraquat

Being largely used as herbicide, the paraquat mechanism of action is similar to MPTP [39]. In this way, lipid peroxidation, mitochondrial dysfunction, accumulation of α -syn, and low levels of GSH were reported in organism models such as rodents and zebrafish following paraquat administration [39, 70, 71, 79, 113]. Additionally, the loss of DA neurons is evidence of the effects of paraquat which can be observed in behavioural and biochemical deficits [70].

Moreover, it was shown that paraquat has the potential to increase the effect of ROS on other molecules, such as lipids, which are one of the main targets of oxidants [114, 115]. Thus, it was observed that 0.04 ppm paraquat administration to zebrafish could lead to more than a 15% increase in MDA levels after 96 hours, concomitantly with DA, GSH, and serotonin decrease [79]. Another important aspect on the paraquat mechanism of action in zebrafish models was that intraperitoneal administration could lead to increased CAT and GPx activity suggesting that its administration could boost some of the antioxidant enzyme activities in the effort of decreasing the cellular ROS levels [116].

9. Relevant Antioxidant Opportunities in Parkinson's Disease Treatment

Considering that healthy functioning metabolism includes a physiological anti-ROS system actively engaged in the prevention of overproduction and accumulation of ROS and that OS seems to be an important component of PD in both human and animal models, the obvious lead for a PD cure could be the antioxidant system stimulation. In this way, the antioxidant system consists of biologically active molecules responsible for ROS neutralization and cell protection against the free toxic radicals' effects [4–7]. Thus, the antioxidant enzymes, such as superoxide dismutase (SOD), glutathione peroxidase (GPx), catalase (CAT), and glutathione reductase (GRx) [10–13], and the nonenzymatic antioxidants, such as lipoic acid, coenzyme Q10, melatonin, vitamin E, vitamin C, flavonoids, and omega acids

[21], form a powerful protection system which prevents the occurrence of OS.

Regarding the implication of mitochondria in ROS metabolism, being responsible for adenosine triphosphate (ATP) production, the main chemical energy source for cellular functions [23], it was demonstrated that the reduction of the ATP synthesis and the electron transport chain impairments lead to ROS accumulation [24]. Due to the fact that mitochondria are a high-quantity ROS source, the correlation between mitochondrial dysfunction and neurodegenerative diseases was predictable. Moreover, the biochemical profile of the brain and the vital role of ROS in brain molecular signalling are further evidence that the central nervous system is continuously predisposed to OS exposure [17, 18]. Additional evidence regarding the antioxidant system and the modulatory pathways has been reported while studying antioxidant supplements, while it was observed that they could provide symptomatology relief or even to reverse oxidative changes and their effects in chronic mitochondrial diseases [24].

PD treatment is mainly based on levodopa and dopamine agonists (amantadine) [47, 114, 117, 118]. Despite the advantages of levodopa administration, dopamine agonists only partially reverse motor symptoms of PD leading to different motor oscillations. However, levodopa remains the main substance used in PD therapy [117–119].

Also, other alternatives for PD treatment are catechol-O-methyl-transferase (COMT) inhibitors and monoamine oxidase B inhibitors, such as safinamide or rasagiline [114, 120, 121]. Similarly, α -lipoic acid is used to remove the excess metals, and coenzyme Q10 is used to decrease oxidative marker activity [27, 114]. Another efficient therapeutic choice in PD treatment is selegine (L-deprenyl) [55, 122]. The carotenoid lycopene was described as a potent antioxidant by reducing the complex I inhibition in a rotenone rat model and reversing MPTP effects in a PD mice model [68, 123].

Besides the drug-centred therapies, exercise programs are an effective strategy used in PD patients to improve and delay functional decline [124]. Alongside the improvement in muscle tonus, it was demonstrated that exercise leads to OS decrease and overall metabolism improvement [125], while muscle weakness is one of the main symptoms of PD, its severity increasing with time [126]. Despite that several studies reported muscle weakness as a consequence of sedentary state and aging processes, bradykinesia seems an important and independent symptom of PD; however, its occurrence mechanism is not fully understood [124, 126]. Recently, it was demonstrated that aquatic exercise therapy could improve motor disability of PD individuals [127].

Studies made until now using animal models led to new speculations regarding PD treatment and other alternatives for it. Treatment with 100 μM vitamin E, 10 μM minocycline, and 25 $\mu\text{g/ml}$ Sinemet (a well-known drug for PD which contains carbidopa and levodopa) in the presence of 25 $\mu\text{g/ml}$ 6-OHDA of zebrafish larvae led to a reverse of locomotor disruptions and of the changes that appeared for parkin, pink1, and cd-11b mRNA expression [80]. Locomotor deficits and neuronal loss observed in

zebrafish larvae after 3 days of treatment with 250 μM 6-OHDA were prevented by 10 μM minocycline and 1 μM rasagiline coadministration [81]. Another study also reported motor and optomotor alterations and morphological changes in zebrafish larvae induced by 250 μM 6-OHDA which were ameliorated after supplementation with 1 mg l^{-1} N-acetylcysteine, which is known for its antioxidant, anti-inflammatory, and neurotrophic potential [82].

10. Is There an Oxidative Stress Correlation to Sleep Disturbances in Parkinson's Disease?

According to Porkka-Heiskanen et al. [128], sleep is the periodic physiological state characterized by temporal suppression of consciousness, partial loss of sensitivity, and decrease of several body functions, such as heart rhythm, respiratory rate, muscle relaxation, and body temperature. Due to its complex mechanism, many factors influence the quantity and quality of sleep, namely stress exposure, health conditions, or some forms of substance abuse [128].

Several recent studies reported that sleep disturbances are a common symptom of PD. In this way, both Menza et al. [129] and Selvaraj and Keshavamurthy [130] described that a direct correlation between PD severity and sleeping time could be suggested and assumed by memory deficits, depressive mood, body weakness, and involuntary sleep events during the day [128]. In this way, several sleep disturbances were described to occur in PD patients, such as insomnia, sleep-related respiratory disorders (SRD), excessive daytime sleepiness (EDS), and sleep-related motor problems often exhibited with variable intensities and durations [129–131]. The connection between these sleep impairments and PD could be certain mechanisms also occurring in OS, for an instance due to sleep apnea or poor oxygenation of the brain during sleeping time [132].

However, regarding the occurrence of sleep-related respiratory symptoms in PD patients, recent studies reported controversial results. In this way, sleep apnea episodes were not significantly present in PD patients, as compared to control groups [131], despite that other reports demonstrated that almost 50% of PD patients experienced sleep apnea incidents [129]. In a more recent study, Bohnen and Hu [132] reported a correlation between sleep apnea which leads to repeated periods of hypoxia and reoxygenation during sleeping and the occurrence of OS and inflammation through a similar mechanism with sleep apnea-induced chronic intermittent hypoxia models. On the other hand, other studies reported that the more common sleep disturbance in PD patients is insomnia [131, 133], which is present in 54–60% of cases [134]. However, Gjerstad et al. [134] discussed the results in the context of age, pathological lesions in the upper brainstem and midbrain, depression, nocturia, and medication [133, 135]. Thus, the multifactorial etiology of sleep disturbances [132] could be discussed in this context and also in the context of the possible comorbidity of sleep disturbances in PD. However, the cause-effect relationship between PD and sleep disturbances is not fully understood.

Another PD-occurring sleep disturbance is rapid eye movement sleep behaviour disorder characterized by motor

behaviours and different vocalizations [135, 136]. By comparison with the other sleep disturbance symptoms in PD, rapid eye movement sleep behaviour disorder is being considered a premotor symptom, and in some cases a disease development marker [137] due to the fact that 40 to 65% of those diagnosed with rapid eye movement sleep behaviour disorder are further later diagnosed with PD [137–140]. Excessive daytime sleepiness (EDS) and fatigue are also present in PD [129, 131, 141, 142]. Restless legs syndrome (RLS) and periodic limb movements in sleep (PLMS) are both correlated to PD [131, 133, 141, 143, 144].

RLS is a sensorimotor condition characterized by the desire to move the legs due to unpleasant sensations [133]. Often appearing in older PD patients, RLS was correlated to iron deficiency as a secondary condition for RLS onset [133, 144, 145]. However, low substantia nigra iron levels were reported in RLS while increased iron levels in PD patients were suggested to lead to OS [142, 144, 145]. Both PD and RLS are characterized by iron deficiency, which may lead to DA damage specific for PD-RLS according to a study published in 2017 [142].

The correlation between OS and PD was previously described, but so was the idea that sleep deprivation can cause OS [146–148]. Thus, it is controversial to ask if any of these correlations could be explained in the context of the presence of all three components: PD, sleep disturbance, and OS. This aspect was partly elucidated due to animal models using certain substances or genetic manipulations for PD features. Scientific literature reported various ways to study these aspects whose target were sleep disturbances [129, 130]. Disruptions in diurnal rhythms, stress, and specific alterations in sleep architecture are only three examples of methods used in animal research [149].

In this way, a recent study presented reasonable evidence which correlated sleep disorders occurring in PD and OS. Filograna et al. [150] extensively described the mechanism through which iron chelators prevent the increase of substantia nigra iron levels in PD patients. Thus, in a well-known study, an iron-chelator-treated chronic iron-loaded mice model exhibited improved OS markers and decreased iron levels. Also, the same group described the antioxidant effects of melatonin in the context of PD-occurring OS mechanisms. The authors pointed out that melatonin antioxidant activity is not described as only free radicals scavenging, but also as other indirect modulatory activities, such as expression stimulation of several antioxidant enzymes and the downregulation of prooxidant enzymes.

Another important component of this triad is the mechanism underlying intermittent hypoxemia observed in PD patients in concomitance with obstructive sleep apnea [151]. According to Kaminska et al. [151], a potential origin of OS occurrence in PD would be the exposure to intermittent episodes of hypoxemia during the sleeping periods. It was shown that intermittent hypoxemia could lead to important changes in the brain structures involved in peripheral nerve conduction, impaired learning and memory, and neuronal loss possibly through mechanisms of ischemia/reperfusion, and oxidative injury.

The recent report of Cao et al. [152] suggested that another OS-related ion could be involved in sleep disturbances. It was demonstrated that magnesium could have long-term benefits in reducing the likelihood of falling asleep in the daytime in women, but not in men, through a mechanism that is, however, unknown. Genetic predisposition would be one of the possible responses in the matter of mechanisms.

Despite that the genetic landscape of PD is currently well described, the implication of magnesium in PD has been only recently hypothesised [153]. Recent research in human and animal models showed that low magnesium levels are correlated with increased risk to develop PD [154]. Moreover, Sturgeon et al. [154] suggested that the mechanism through which this correlation is built is based on a unique genetic landscaping of magnesium homeostasis. Sustaining this hypothesis, several studies meta-analysed by Jin et al. [155] suggested that increased magnesium levels are a molecular feature of PD, therefore magnesium dishomeostasis may be considered a real risk factor in PD. Both SLC41A1 and TRPM7 are directly or indirectly modulating sleep-related behaviours [154]. Thus, SLC41A1 is being involved in rapid eye movement sleep behaviour disorder [156], while TRPM7 is suggested to be involved in sleep-wake cycle modulation through magnesium ion ligation potential [157].

Moreover, the OS and DA perturbations were also observed in gene mutations of α -syn, PINK, parkin, and DJ-1 proteins [25, 52]. The lack of PINK1 leads to a loss of DA neurons, affects the mitochondrial morphology, and is linked with OS [21, 25, 158–161]. Also, the accumulation of α -syn causes the reduction of mitochondrial activity and a high production of ROS which is completed by cell death [25, 31]. The PARK2 and LRRK2 genes represent a source of ROS production [26]. All these proteins are linked with PD pathogenesis, and their use in genetic manipulations has become a new tool in transgenic animal models [12, 161, 162].

Thus, genetic implications in PD are not new to PD research. Several recent studies suggested not only that some genetic factors give real predisposition to PD development at some point in life [163] but also that PD may be one of the disorders which run in a family [164]. Currently, more than 20 PD genetic predisposition loci are identified and extensively reviewed [163]. Among these, some target the synaptic vesicle anomalies (SNCA mutations), the protein-to-protein interaction in the cytoskeleton assembly (dardarin gene mutations), ubiquitin degradation (parkin gene mutations), and several other energetic mechanisms.

11. Conclusions

This study synthesized the current information and correlated available data on the relevance of the oxidative stress status modifications in the complex pathophysiology of Parkinson's disease with regard to the available animal models. Moreover, the importance of the zebrafish model in Parkinson's disease research was described. It was observed that OS possesses an important role in Parkinson's disease as shown by numerous animal model studies, many of which

are based on rodent experimental models. However, an emerging impact of the zebrafish model was observed in research on Parkinson's disease pathological mechanisms with regard to disease development factors, cause-effect relationship of oxidative stress and comorbidities (such as depression, hyposmia, fatigue, sleep disturbances, and cognitive deficits), and also regarding the pharmacological potential of antioxidant molecules in Parkinson's disease treatment.

Conflicts of Interest

None, except for the research grants mentioned in the Acknowledgments.

Authors' Contributions

Robea M.-A. and Balmus I.-M. equally contributed to this work as first authors.

Acknowledgments

AC and IMB are supported by a research grant for Young Teams offered by UEFISCDI, Romania, no. PN-III-P1-1.1-TE-2016-1210, contract no. 58 from 02/05/2018, called "Complex study regarding the interactions between OS, inflammation and neurological manifestations in the pathophysiology of irritable bowel syndrome (animal models and human patients)." MAR was funded by the European Social Fund, through Operational Programme Human Capital 2014-2020, project number POCU/380/6/13/123623, project title "PhD Students and Postdoctoral Researchers Prepared for the Labour Market!".

References

- [1] H. Sies, C. Berndt, and D. P. Jones, "Oxidative stress," *Annual Review of Biochemistry*, vol. 86, no. 1, pp. 715–748, 2017.
- [2] G. Cenini, A. Lloret, and R. Cascella, "Oxidative stress in neurodegenerative diseases: from a mitochondrial point of view," *Oxidative Medicine and Cellular Longevity*, vol. 2019, Article ID 2105607, 18 pages, 2019.
- [3] J. Kruk, H. Y. Aboul-Enein, A. Kladna, and J. E. Bowser, "Oxidative stress in biological systems and its relation with pathophysiological functions: the effect of physical activity on cellular redox homeostasis," *Free Radical Research*, vol. 53, no. 5, pp. 497–521, 2019.
- [4] S. Salim, "Oxidative stress and psychological disorders," *Current Neuropharmacology*, vol. 12, no. 2, pp. 140–147, 2014.
- [5] W. Hassan, H. Noreen, V. Castro-Gomes, I. Mohammadzai, J. Batista Teixeira da Rocha, and J. Landeira-Fernandez, "Association of oxidative stress with psychiatric disorders," *Current Pharmaceutical Design*, vol. 22, no. 20, pp. 2960–2974, 2016.
- [6] L. Polito, A. Greco, and D. Seripa, "Genetic profile, environmental exposure, and their interaction in Parkinson's disease," *Parkinson's Disease*, vol. 2016, article 6465793, pp. 1–9, 2016.
- [7] R. Uher and A. Zwickler, "Etiology in psychiatry: embracing the reality of poly-gene-environmental causation of mental illness," *World Psychiatry*, vol. 16, no. 2, pp. 121–129, 2017.

- [8] J. Zhang, M. L. Culp, J. G. Craver, and V. Darley-USmar, "Mitochondrial function and autophagy: integrating proteotoxic, redox, and metabolic stress in Parkinson's disease," *Journal of Neurochemistry*, vol. 144, no. 6, pp. 691–709, 2018.
- [9] M. J. Armstrong and M. S. Okun, "Diagnosis and treatment of Parkinson Disease," *JAMA*, vol. 323, no. 6, pp. 548–560, 2020.
- [10] T. Gasser, T. Wichmann, and M. R. DeLongB. D. Coyle, *Chapter 19. Parkinson disease and other synucleinopathies*, M. J. Zigmond and L. P. Rowland, Eds., Academic Press, San Diego, 2015.
- [11] W. L. Kouli, A. Torsney, and K. M. Kuan, "Parkinson's disease: etiology, neuropathology, and pathogenesis," in *Parkinson's Disease: Pathogenesis and Clinical Aspects*, J. C. Stoker and T. B. Greenland, Eds., Codon Publications, 2018.
- [12] Y. Xi, S. Noble, and M. Ekker, "Modeling neurodegeneration in zebrafish," *Current Neurology and Neuroscience Reports*, vol. 11, no. 3, pp. 274–282, 2011.
- [13] C. H. Brennan, "Zebrafish behavioural assays of translational relevance for the study of psychiatric disease," *Reviews in the Neurosciences*, vol. 22, no. 1, pp. 37–48, 2011.
- [14] C. Milanese, J. J. Sager, Q. Bai et al., "Hypokinesia and reduced dopamine levels in zebrafish lacking β - and γ 1-Synucleins," *The Journal of Biological Chemistry*, vol. 287, no. 5, pp. 2971–2983, 2012.
- [15] R. Betarbet, T. B. Sherer, G. MacKenzie, M. Garcia-Osuna, A. V. Panov, and J. T. Greenamyre, "Chronic systemic pesticide exposure reproduces features of Parkinson's disease," *Nature Neuroscience*, vol. 3, no. 12, pp. 1301–1306, 2000.
- [16] H. Khotimah, M. Ali, S. B. Sumitro, and M. A. Widodo, "Decreasing α -synuclein aggregation by methanolic extract of *Centella asiatica* in zebrafish Parkinson's model," *Asian Pacific Journal of Tropical Biomedicine*, vol. 5, no. 11, pp. 948–954, 2015.
- [17] M. Alam and W. J. Schmidt, "Rotenone destroys dopaminergic neurons and induces parkinsonian symptoms in rats," *Behavioural Brain Research*, vol. 136, no. 1, pp. 317–324, 2002.
- [18] C. Santoriello and L. I. Zon, "Hooked! Modeling human disease in zebrafish," *The Journal of Clinical Investigation*, vol. 122, no. 7, pp. 2337–2343, 2012.
- [19] M. Ahmad, et al.S. Saleem, A. S. Ahmad et al., "Ginkgo biloba affords dose-dependent protection against 6-hydroxydopamine-induced parkinsonism in rats: neurobehavioural, neurochemical and immunohistochemical evidences," *Journal of Neurochemistry*, vol. 93, no. 1, pp. 94–104, 2005.
- [20] M. Dagdelen, H. Akkaya, and E. Genc, "P.I.g.015 The role of valproic acid and levodopa on oxidative stress in a 6-hydroxydopamine lesioned rat model of Parkinson's disease," *European Neuropsychopharmacology*, vol. 23, p. S197, 2013.
- [21] V. Dias, E. Junn, and M. M. Mouradian, "The role of oxidative stress in Parkinson's disease," *Journal of Parkinson's Disease*, vol. 3, no. 4, pp. 461–491, 2013.
- [22] R. Nistico, B. Mehdawy, S. Piccirilli, and N. Mercuri, "Paraquat- and rotenone-induced models of Parkinson's disease," *International journal of immunopathology and pharmacology*, vol. 24, no. 2, pp. 313–322, 2011.
- [23] F. Blandini and M.-T. Armentero, "Animal models of Parkinson's disease," *The FEBS Journal*, vol. 279, no. 7, pp. 1156–1166, 2012.
- [24] H. Sies, "Oxidative stress: a concept in redox biology and medicine," *Redox Biology*, vol. 4, pp. 180–183, 2015.
- [25] J. Blesa, I. Trigo-Damas, A. Quiroga-Varela, and V. R. Jackson-Lewis, "Oxidative stress and Parkinson's disease," *Frontiers in Neuroanatomy*, vol. 9, 2015.
- [26] L. Puspita, S. Y. Chung, and J.-W. Shim, "Oxidative stress and cellular pathologies in Parkinson's disease," *Molecular Brain*, vol. 10, no. 1, p. 53, 2017.
- [27] G. L. Nicolson, "Mitochondrial dysfunction and chronic disease: treatment with natural supplements," *Integrative Medicine: A Clinician's Journal*, vol. 13, no. 4, pp. 35–43, 2014.
- [28] E. Niedzielska, I. Smaga, M. Gawlik et al., "Oxidative stress in neurodegenerative diseases," *Molecular Neurobiology*, vol. 53, no. 6, pp. 4094–4125, 2016.
- [29] W. Poewe, K. Seppi, C. M. Tanner et al., "Parkinson disease," *Nature Reviews Disease Primers*, vol. 3, no. 1, 2017.
- [30] H.-J. Lee, S. Y. Shin, C. Choi, Y. H. Lee, and S.-J. Lee, "Formation and removal of α -Synuclein aggregates in cells exposed to mitochondrial inhibitors," *The Journal of Biological Chemistry*, vol. 277, no. 7, pp. 5411–5417, 2002.
- [31] L. J. Martin, Y. Pan, A. C. Price et al., "Parkinson's disease alpha-synuclein transgenic mice develop neuronal mitochondrial degeneration and cell death," *The Journal of Neuroscience*, vol. 26, no. 1, pp. 41–50, 2006.
- [32] J. M. Beitz, "Parkinson's disease a review," *Frontiers in Bioscience*, vol. S6, no. 1, pp. 65–74, 2014.
- [33] J. Segura-Aguilar and I. Paris, *Mechanisms of dopamine oxidation and Parkinson's disease BT - Handbook of Neurotoxicity*, R. M. Kostrzewa, Ed., Springer New York, New York, NY, 2014.
- [34] C. W. Olanow and W. G. Tatton, "Etiology and pathogenesis of Parkinson's disease," *Annual Review of Neuroscience*, vol. 22, no. 1, pp. 123–144, 1999.
- [35] J. B. Schulz, J. Lindenau, J. Seyfried, and J. Dichgans, "Glutathione, oxidative stress and neurodegeneration," *European Journal of Biochemistry*, vol. 267, no. 16, pp. 4904–4911, 2000.
- [36] B. Halliwell, "Role of free radicals in the neurodegenerative Diseases," *Drugs & Aging*, vol. 18, no. 9, pp. 685–716, 2001.
- [37] A. Ciobica, M. Padurariu, and L. Hritcu, "The effects of short-term nicotine administration on behavioral and oxidative stress deficiencies induced in a rat model of Parkinson's disease," *Psychiatria Danubina*, vol. 24, no. 2, pp. 194–205, 2012.
- [38] M. Asanuma, I. Miyazaki, and N. Ogawa, "Dopamine- or L-DOPA-induced neurotoxicity: the role of dopamine quinone formation and tyrosinase in a model of Parkinson's disease," *Neurotoxicity Research*, vol. 5, no. 3, pp. 165–176, 2003.
- [39] S. M. Goldman, "Environmental toxins and Parkinson's disease," *Annual Review of Pharmacology and Toxicology*, vol. 54, no. 1, pp. 141–164, 2014.
- [40] O. V. Anichtchik, J. Kaslin, N. Peitsaro, M. Scheinin, and P. Panula, "Neurochemical and behavioural changes in zebrafish Danio rerio after systemic administration of 6-hydroxydopamine and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine," *Journal of Neurochemistry*, vol. 88, no. 2, pp. 443–453, 2004.
- [41] T. Kitada, A. Pisani, D. R. Porter et al., "Impaired dopamine release and synaptic plasticity in the striatum of PINK1-deficient mice," in *Proceedings of the National Academy of Sciences of the United States of America*, vol. 104no. 27, pp. 11441–11446, 2007.

- [42] M. Valko, D. Leibfritz, J. Moncol, M. T. D. Cronin, M. Mazur, and J. Telser, "Free radicals and antioxidants in normal physiological functions and human disease," *The International Journal of Biochemistry & Cell Biology*, vol. 39, no. 1, pp. 44–84, 2007.
- [43] D. A. Di Monte, P. Chan, and M. S. Sandy, "Glutathione in Parkinson's disease: a link between oxidative stress and mitochondrial damage?," *Annals of Neurology*, vol. 32, no. S1, pp. S111–S115, 1992.
- [44] S. Bharath, M. Hsu, D. Kaur, S. Rajagopalan, and J. K. Andersen, "Glutathione, iron and Parkinson's disease," *Biochemical Pharmacology*, vol. 64, no. 5–6, pp. 1037–1048, 2002.
- [45] R. A. Hauser, K. E. Lyons, T. McClain, S. Carter, and D. Perlmutter, "Randomized, double-blind, pilot evaluation of intravenous glutathione in Parkinson's disease," *Movement Disorders*, vol. 24, no. 7, pp. 979–983, 2009.
- [46] J. B. Schulz, R. T. Matthews, T. Klockgether, J. Dichgans, and M. F. Beal, "The role of mitochondrial dysfunction and neuronal nitric oxide in animal models of neurodegenerative diseases," *Detection of Mitochondrial Diseases*, pp. 193–197, 1997.
- [47] B. S. Connolly and A. E. Lang, "Pharmacological treatment of Parkinson disease: a review," *JAMA*, vol. 311, no. 16, pp. 1670–1683, 2014.
- [48] Y. Vijayanathan, F. T. Lim, S. M. Lim et al., "6-OHDA-lesioned adult zebrafish as a useful Parkinson's disease model for dopaminergic neuroregeneration," *Neurotoxicity Research*, vol. 32, no. 3, pp. 496–508, 2017.
- [49] K. Dooley and L. I. Zon, "Zebrafish: a model system for the study of human disease," *Current Opinion in Genetics & Development*, vol. 10, no. 3, pp. 252–256, 2000.
- [50] K. F. Winklhofer and C. Haass, "Mitochondrial dysfunction in Parkinson's disease," *Biochimica et Biophysica Acta*, vol. 1802, no. 1, pp. 29–44, 2010.
- [51] C. M. Feitosa, G. L. da Silva Oliveira, A. do Nascimento Cavalcante, S. K. Morais Chaves, and M. Rai, "Determination of parameters of oxidative stress in vitro models of neurodegenerative diseases—a review," *Current Clinical Pharmacology*, vol. 13, no. 2, pp. 100–109, 2018.
- [52] O. Hwang, "Role of oxidative stress in Parkinson's disease," *Experimental Neurobiology*, vol. 22, no. 1, pp. 11–17, 2013.
- [53] M. Hemmati-Dinarvand, S. saedi, M. Valilo et al., "Oxidative stress and Parkinson's disease: conflict of oxidant-antioxidant systems," *Neuroscience Letters*, vol. 709, p. 134296, 2019.
- [54] R. M. Giráldez-Pérez, M. Antolín-Vallespín, M. D. Muñoz, and A. Sánchez-Capelo, "Models of α -synuclein aggregation in Parkinson's disease," *Acta Neuropathologica Communications*, vol. 2, no. 1, p. 176, 2014.
- [55] E. T. McKinley, T. C. Baranowski, D. O. Blavo, C. Cato, T. N. Doan, and A. L. Rubinstein, "Neuroprotection of MPTP-induced toxicity in zebrafish dopaminergic neurons," *Brain Research. Molecular Brain Research*, vol. 141, no. 2, pp. 128–137, 2005.
- [56] N. S. Babu, C. L. N. Murthy, S. Kakara, R. Sharma, C. V. Brahmendra Swamy, and M. M. Idris, "1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine induced Parkinson's disease in zebrafish," *PROTEOMICS*, vol. 16, no. 9, pp. 1407–1420, 2016.
- [57] S. F. Yang et al., "Protective effect and mechanism of Ginkgo biloba leaf extracts for Parkinson disease induced by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine," *Acta Pharmacologica Sinica*, vol. 22, no. 12, pp. 1089–1093, 2016.
- [58] J. K. Barbiero, R. Santiago, F. S. Tonin et al., "PPAR-alpha agonist fenofibrate protects against the damaging effects of MPTP in a rat model of Parkinson's disease," *Progress in Neuro-Psychopharmacology & Biological Psychiatry*, vol. 53, pp. 35–44, 2014.
- [59] R. Bisht, B. Kaur, H. Gupta, and A. Prakash, "Ceftriaxone mediated rescue of nigral oxidative damage and motor deficits in MPTP model of Parkinson's disease in rats," *Neurotoxicology*, vol. 44, pp. 71–79, 2014.
- [60] B. Thomas and K. P. Mohanakumar, "Melatonin protects against oxidative stress caused by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine in the mouse nigrostriatum," *Journal of Pineal Research*, vol. 36, no. 1, pp. 25–32, 2004.
- [61] G. U. Höglinger, J. Féger, A. Prigent et al., "Chronic systemic complex I inhibition induces a hypokinetic multisystem degeneration in rats," *Journal of Neurochemistry*, vol. 84, no. 3, pp. 491–502, 2003.
- [62] T. B. Sherer, J. H. Kim, R. Betarbet, and J. T. Greenamyre, "Subcutaneous Rotenone Exposure Causes Highly Selective Dopaminergic Degeneration and α -Synuclein Aggregation," *Experimental Neurology*, vol. 179, no. 1, pp. 9–16, 2003.
- [63] N. Lapointe, M. St-Hilaire, M. G. Martinoli et al., "Rotenone induces non-specific central nervous system and systemic toxicity," *The FASEB Journal*, vol. 18, no. 6, pp. 717–719, 2004.
- [64] K. S. Saravanan, K. M. Sindhu, and K. P. Mohanakumar, "Acute intranigral infusion of rotenone in rats causes progressive biochemical lesions in the striatum similar to Parkinson's disease," *Brain Research*, vol. 1049, no. 2, pp. 147–155, 2005.
- [65] Y. Feng, Z. H. Liang, T. Wang, X. Qiao, H. J. Liu, and S. G. Sun, "alpha-Synuclein redistributed and aggregated in rotenone-induced Parkinson's disease rats," *Neuroscience Bulletin*, vol. 22, no. 5, pp. 288–293, 2006.
- [66] P. G. M. Ravenstijn, M. Merlini, M. Hameetman et al., "The exploration of rotenone as a toxin for inducing Parkinson's disease in rats, for application in BBB transport and PK-PD experiments," *Journal of Pharmacological and Toxicological Methods*, vol. 57, no. 2, pp. 114–130, 2008.
- [67] A. Norazit, A. C. B. Meedeniya, M. N. Nguyen, and A. Mackay-Sim, "Progressive loss of dopaminergic neurons induced by unilateral rotenone infusion into the medial forebrain bundle," *Brain Research*, vol. 1360, pp. 119–129, 2010.
- [68] H. Kaur, S. Chauhan, and R. Sandhir, "Protective effect of lycopene on oxidative stress and cognitive decline in rotenone induced model of Parkinson's disease," *Neurochemical Research*, vol. 36, no. 8, pp. 1435–1443, 2011.
- [69] J. W. Bortolotto, G. P. Cognato, R. R. Christoff et al., "Long-term exposure to paraquat alters behavioral parameters and dopamine levels in adult zebrafish (*Danio rerio*)," *Zebrafish*, vol. 11, no. 2, pp. 142–153, 2014.
- [70] X. H. Wang, C. L. Souders II, Y. H. Zhao, and C. J. Martyniuk, "Paraquat affects mitochondrial bioenergetics, dopamine system expression, and locomotor activity in zebrafish (*Danio rerio*)," *Chemosphere*, vol. 191, pp. 106–117, 2018.
- [71] Q. Chen, Y. Niu, R. Zhang et al., "The toxic influence of paraquat on hippocampus of mice: Involvement of oxidative stress," *Neurotoxicology*, vol. 31, no. 3, pp. 310–316, 2010.

- [72] N. K. Singhal, G. Srivastava, D. K. Patel, S. K. Jain, and M. P. Singh, "Melatonin or silymarin reduces maneb- and paraquat-induced Parkinson's disease phenotype in the mouse," *Journal of Pineal Research*, 2010.
- [73] L. J. Hsu, Y. Sagara, A. Arroyo et al., " α -Synuclein Promotes Mitochondrial Deficit and Oxidative Stress," *The American Journal of Pathology*, vol. 157, no. 2, pp. 401–410, 2000.
- [74] Z. Sun and A. D. Gitler, "Discovery and characterization of three novel synuclein genes in zebrafish," *Developmental Dynamics*, vol. 237, no. 9, pp. 2490–2495, 2008.
- [75] S. Prabhudesai, S. Sinha, A. Attar et al., "A novel "molecular tweezer" inhibitor of α -Synuclein neurotoxicity in vitro and in vivo," *Neurotherapeutics*, vol. 9, no. 2, pp. 464–476, 2012.
- [76] D. A. Bosco, D. M. Fowler, Q. Zhang et al., "Elevated levels of oxidized cholesterol metabolites in Lewy body disease brains accelerate α -synuclein fibrilization," *Nature Chemical Biology*, vol. 2, no. 5, pp. 249–253, 2006.
- [77] J. Sian, M. B. H. Youdim, P. Riederer, and M. Gerlach, "MPTP-induced Parkinsonian syndrome," in *Basic Neurochemistry: Molecular, Cellular and Medical Aspects*, 6th ed., Lippincott-Raven, Philadelphia, 1999.
- [78] R. Betarbet, T. B. Sherer, and J. T. Greenamyre, "Animal models of Parkinson's disease," *BioEssays*, vol. 24, no. 4, pp. 308–318, 2002.
- [79] J. Nellore and N. P., "Paraquat exposure induces behavioral deficits in larval zebrafish during the window of dopamine neurogenesis," *Toxicology Reports*, vol. 2, pp. 950–956, 2015.
- [80] C.-W. Feng, et al. Z.-H. Wen, S.-Y. Huang et al., "Effects of 6-hydroxydopamine exposure on motor activity and biochemical expression in zebrafish (*Danio rerio*) larvae," *Zebrafish*, vol. 11, no. 3, pp. 227–239, 2014.
- [81] A. Cronin and M. Grealy, "Neuroprotective and Neurorestorative Effects of Minocycline and Rasagiline in a Zebrafish 6-Hydroxydopamine Model of Parkinson's Disease," *Neuroscience*, vol. 367, pp. 34–46, 2017.
- [82] R. Benvenuti, M. Marcon, C. G. Reis et al., "N-acetylcysteine protects against motor, optomotor and morphological deficits induced by 6-OHDA in zebrafish larvae," *PeerJ*, vol. 6, p. e4957, 2018.
- [83] J. W. Langston, "The MPTP story," *Journal of Parkinson's Disease*, vol. 7, no. s1, pp. S11–S19, 2017.
- [84] A. Masato, N. Plotegher, D. Boassa, and L. Bubacco, "Impaired dopamine metabolism in Parkinson's disease pathogenesis," *Molecular Neurodegeneration*, vol. 14, no. 1, p. 35, 2019.
- [85] T. Farooqui and A. A. Farooqui, "Lipid-Mediated Oxidative Stress and Inflammation in the Pathogenesis of Parkinson's Disease," *Parkinson's Disease*, vol. 2011, pp. 1–9, 2011.
- [86] G. Cohen, "Oxidative stress, mitochondrial respiration, and Parkinson's disease," *Annals of the New York Academy of Sciences*, vol. 899, no. 1, pp. 112–120, 2000.
- [87] V. Calabrese, C. Mancuso, M. Calvani, E. Rizzarelli, D. A. Butterfield, and A. M. Giuffrida Stella, "Nitric oxide in the central nervous system: neuroprotection versus neurotoxicity," *Nature Reviews. Neuroscience*, vol. 8, no. 10, pp. 766–775, 2007.
- [88] J.-S. Park, R. L. Davis, and C. M. Sue, "Mitochondrial Dysfunction in Parkinson's Disease: New Mechanistic Insights and Therapeutic Perspectives," *Current Neurology and Neuroscience Reports*, vol. 18, no. 5, p. 21, 2018.
- [89] S. Shams, J. Rihel, J. G. Ortiz, and R. Gerlai, "The zebrafish as a promising tool for modeling human brain disorders: a review based upon an IBNS symposium," *Neuroscience and Biobehavioral Reviews*, vol. 85, pp. 176–190, 2018.
- [90] G. E. Meredith and D. J. Rademacher, "MPTP mouse models of Parkinson's disease: an update," *Journal of Parkinson's Disease*, vol. 1, no. 1, pp. 19–33, 2011.
- [91] M. Sedelis, R. K. W. Schwarting, and J. P. Huston, "Behavioral phenotyping of the MPTP mouse model of Parkinson's disease," *Behavioural Brain Research*, vol. 125, no. 1–2, pp. 109–125, 2001.
- [92] R. L. Vaz, T. F. Outeiro, and J. J. Ferreira, "Zebrafish as an animal model for drug discovery in Parkinson's disease and other movement disorders: a systematic review," *Frontiers in Neurology*, vol. 9, 2018.
- [93] V. Jackson-Lewis, J. Blesa, and S. Przedborski, "Animal models of Parkinson's disease," *Parkinsonism & Related Disorders*, vol. 18, pp. S183–S185, 2012.
- [94] J. Kirchoff, A. Mork, L. T. Brennum, and T. N. Sager, "Striatal extracellular dopamine levels and behavioural reversal in MPTP-lesioned mice," *NeuroReport*, vol. 20, no. 5, pp. 482–486, 2009.
- [95] E. R. Mingazov, G. R. Khakimova, E. A. Kozina et al., "MPTP mouse model of preclinical and clinical Parkinson's disease as an instrument for translational medicine," *Molecular Neurobiology*, vol. 55, no. 4, pp. 2991–3006, 2018.
- [96] C. S. Lam, V. Korzh, and U. Strahle, "Zebrafish embryos are susceptible to the dopaminergic neurotoxin MPTP," *The European Journal of Neuroscience*, vol. 21, no. 6, pp. 1758–1762, 2005.
- [97] E. Kabashi, E. Brustein, N. Champagne, and P. Drapeau, "Zebrafish models for the functional genomics of neurogenetic disorders," *Biochimica et Biophysica Acta*, vol. 1812, no. 3, pp. 335–345, 2011.
- [98] N. Simola, M. Morelli, and A. R. Carta, "The 6-hydroxydopamine model of Parkinson's disease," *Neurotoxicity Research*, vol. 11, no. 3–4, pp. 151–167, 2007.
- [99] A. Schober, "Classic toxin-induced animal models of Parkinson's disease: 6-OHDA and MPTP," *Cell and Tissue Research*, vol. 318, no. 1, pp. 215–224, 2004.
- [100] Y. Y. Glinka and M. B. H. Youdim, "Inhibition of mitochondrial complexes I and IV by 6-hydroxydopamine," *European Journal of Pharmacology*, vol. 292, no. 3–4, pp. 329–332, 1995.
- [101] R. Deumens, A. Blokland, and J. Prickaerts, "Modeling Parkinson's Disease in Rats: An Evaluation of 6-OHDA Lesions of the Nigrostriatal Pathway," *Experimental Neurology*, vol. 175, no. 2, pp. 303–317, 2002.
- [102] J. Jordán, M. E. Solesio, and M. F. Galindo, "Mitochondrial alterations and mitophagy in response to 6-hydroxydopamine," in *Autophagy: Cancer, Other Pathologies, Inflammation, Immunity, Infection, and Aging*, M. A. B. T.-A. C. Hayat, Ed., pp. 201–209, Academic Press, Amsterdam, 2014.
- [103] C. Parnig, N. M. Roy, C. Ton, Y. Lin, and P. McGrath, "Neurotoxicity assessment using zebrafish," *Journal of Pharmacological and Toxicological Methods*, vol. 55, no. 1, pp. 103–112, 2007.
- [104] J. C. Bensadoun, O. Mirochnitchenko, M. Inouye, P. Aebischer, and A. D. Zurn, "Attenuation of 6-OHDA-induced neurotoxicity in glutathione peroxidase transgenic

- mice," *The European Journal of Neuroscience*, vol. 10, no. 10, pp. 3231–3236, 1998.
- [105] M. A. Robea, S.-A. Strungaru, C. Lenzi, M. Nicoara, and Ciobica, "The importance of rotenone in generating neurological and psychiatric features in zebrafish-relevance for a Parkinson's disease model," *Academy of Romanian Scientists*, vol. 7, no. 1, pp. 59–67, 2018.
- [106] N. Xiong, X. Long, J. Xiong et al., "Mitochondrial complex I inhibitor rotenone-induced toxicity and its potential mechanisms in Parkinson's disease models," *Critical Reviews in Toxicology*, vol. 42, no. 7, pp. 613–632, 2012.
- [107] R. E. Heikkila, W. J. Nicklas, I. Vyas, and R. C. Duvoisin, "Dopaminergic toxicity of rotenone and the 1-methyl-4-phenylpyridinium ion after their stereotaxic administration to rats: implication for the mechanism of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine toxicity," *Neuroscience Letters*, vol. 62, no. 3, pp. 389–394, 1985.
- [108] N. Li, K. Ragheb, G. Lawler et al., "Mitochondrial complex I inhibitor rotenone induces apoptosis through enhancing mitochondrial reactive oxygen species production," *The Journal of Biological Chemistry*, vol. 278, no. 10, pp. 8516–8525, 2003.
- [109] M. ALAM and W. SCHMIDT, "Mitochondrial complex I inhibition depletes plasma testosterone in the rotenone model of Parkinson's disease," *Physiology & Behavior*, vol. 83, no. 3, pp. 395–400, 2004.
- [110] Y. Wang, W. Liu, J. Yang et al., "Parkinson's disease-like motor and non-motor symptoms in rotenone-treated zebrafish," *Neurotoxicology*, vol. 58, pp. 103–109, 2017.
- [111] S. Martel, J. Y. Keow, and M. Ekker, "Rotenone Neurotoxicity Causes Dopamine Neuron Loss in Zebrafish," *University of Ottawa Journal of Medicine*, vol. 5, no. 2, pp. 16–21, 2015.
- [112] K. M. Melo, R. Oliveira, C. K. Grisolia et al., "Short-term exposure to low doses of rotenone induces developmental, biochemical, behavioral, and histological changes in fish," *Environmental Science and Pollution Research International*, vol. 22, no. 18, pp. 13926–13938, 2015.
- [113] H. M. Cocheme and M. P. Murphy, "Complex I is the major site of mitochondrial superoxide production by paraquat," *The Journal of Biological Chemistry*, vol. 283, no. 4, pp. 1786–1798, 2008.
- [114] R. M. Adibhatla and J. F. Hatcher, "Lipid oxidation and peroxidation in CNS health and disease: from molecular mechanisms to therapeutic opportunities," *Antioxidants & Redox Signaling*, vol. 12, no. 1, pp. 125–169, 2010.
- [115] M. Shichiri, "The role of lipid peroxidation in neurological disorders," *Journal of Clinical Biochemistry and Nutrition*, vol. 54, no. 3, pp. 151–160, 2014.
- [116] M. E. Nunes, T. E. Müller, M. M. Braga et al., "Chronic treatment with Paraquat induces brain injury, changes in antioxidant defenses system, and modulates behavioral functions in zebrafish," *Molecular Neurobiology*, vol. 54, no. 6, pp. 3925–3934, 2017.
- [117] W. Poewe, A. Antonini, J. C. Zijlmans, P. R. Burkhard, and F. Vingerhoets, "Levodopa in the treatment of Parkinson's disease: an old drug still going strong," *Clinical Interventions in Aging*, vol. 5, pp. 229–238, 2010.
- [118] R. Dhall and D. L. Kreitzman, "Advances in levodopa therapy for Parkinson disease," *Neurology*, vol. 86, 14 Supplement 1, pp. S13–S24, 2016.
- [119] N. Ogawa, "Levodopa and Dopamine Agonists in the Treatment of Parkinson's Disease: Advantages and Disadvantages," *European Neurology*, vol. 34, no. 3, pp. 20–28, 2004.
- [120] P. Riederer and G. Laux, "MAO-inhibitors in Parkinson's disease," *Experimental Neurobiology*, vol. 20, no. 1, pp. 1–17, 2011.
- [121] L. Dezsi and L. Vecsei, "Monoamine oxidase B inhibitors in Parkinson's disease," *CNS & Neurological Disorders Drug Targets*, vol. 16, no. 4, pp. 425–439, 2017.
- [122] S. G. Kim, C. H. Lee, and J. W. Park, "Deprenyl, a therapeutic agent for Parkinson's disease, inhibits arsenic toxicity potentiated by GSH depletion via inhibition of JNK activation," *Journal of Toxicology and Environmental Health. Part A*, vol. 67, no. 23–24, pp. 2013–2024, 2004.
- [123] A. Prema, U. Janakiraman, T. Manivasagam, and A. Justin Thenmozhi, "Neuroprotective effect of lycopene against MPTP induced experimental Parkinson's disease in mice," *Neuroscience Letters*, vol. 599, pp. 12–19, 2015.
- [124] V. A. Goodwin, S. H. Richards, R. S. Taylor, A. H. Taylor, and J. L. Campbell, "The effectiveness of exercise interventions for people with Parkinson's disease: a systematic review and meta-analysis," *Movement Disorders*, vol. 23, no. 5, pp. 631–640, 2008.
- [125] D. Timofte, A. Ciobica, C. Honceriu, F. Trofin, A. Ciobica, and Cojocar, "A mini-review on the effects of exercising and obesity in Parkinson's disease," *Ann. Ser. Biol. Sci.*, vol. 6, no. 1, pp. 100–113, 2017.
- [126] R. Cano-de-la-Cuerda, M. Pérez-de-Heredia, J. C. Miangolarra-Page, E. Muñoz-Hellín, and C. Fernández-de-las-Peñas, "Is there muscular weakness in Parkinson's disease?," *American Journal of Physical Medicine & Rehabilitation*, vol. 89, no. 1, pp. 70–76, 2010.
- [127] L. M. Carroll, D. Volpe, M. E. Morris, J. Saunders, and A. M. Clifford, "Aquatic exercise therapy for people with Parkinson disease: a randomized controlled trial," *Archives of Physical Medicine and Rehabilitation*, vol. 98, no. 4, pp. 631–638, 2017.
- [128] T. Porkka-Heiskanen, K.-M. Zitting, and H.-K. Wigren, "Sleep, its regulation and possible mechanisms of sleep disturbances," *Acta Physiologica (Oxford, England)*, vol. 208, no. 4, pp. 311–328, 2013.
- [129] M. Menza, R. D. Dobkin, H. Marin, and K. Bienfait, "Sleep disturbances in Parkinson's disease," *Movement Disorders*, vol. 25, no. S1, Suppl 1, pp. S117–S122, 2010.
- [130] V. K. Selvaraj, "Sleep Dysfunction in Parkinson's Disease," *JOURNAL OF CLINICAL AND DIAGNOSTIC RESEARCH*, 2016.
- [131] J.-F. Gagnon, D. Petit, V. Latreille, and J. Montplaisir, "Neurobiology of sleep disturbances in neurodegenerative disorders," *Current Pharmaceutical Design*, vol. 14, no. 32, pp. 3430–3445, 2008.
- [132] N. I. Bohnen and M. T. M. Hu, "Sleep disturbance as potential risk and progression factor for Parkinson's disease," *Journal of Parkinson's Disease*, vol. 9, no. 3, pp. 603–614, 2019.
- [133] G. Loddo, G. Calandra-Buonaura, L. Sambati et al., "The treatment of sleep disorders in Parkinson's disease: from research to clinical practice," *Frontiers in Neurology*, vol. 8, 2017.
- [134] M. D. Gjerstad, T. Wentzel-Larsen, D. Aarsland, and J. P. Larsen, "Insomnia in Parkinson's disease: frequency and

- progression over time,” *Journal of Neurology, Neurosurgery, and Psychiatry*, vol. 78, no. 5, pp. 476–479, 2006.
- [135] R.-l. Zhu, C.-j. Xie, P.-p. Hu, and K. Wang, “Clinical variations in Parkinson’s disease patients with or without REM sleep behaviour disorder: a meta-analysis,” *Scientific Reports*, vol. 7, no. 1, 2017.
- [136] C. L. Rodriguez, N. Jaimcharyatam, and K. Budur, “Rapid eye movement sleep behavior disorder: a review of the literature and update on current concepts,” *Chest*, vol. 152, no. 3, pp. 650–662, 2017.
- [137] C. H. Adler, J. G. Hentz, H. A. Shill et al., “Probable RBD is increased in Parkinson’s disease but not in essential tremor or restless legs syndrome,” *Parkinsonism & Related Disorders*, vol. 17, no. 6, pp. 456–458, 2011.
- [138] Y. E. Kim and B. S. Jeon, “Clinical implication of REM sleep behavior disorder in Parkinson’s disease,” *Journal of Parkinson’s Disease*, vol. 4, no. 2, pp. 237–244, 2014.
- [139] J. Zhang, C.-Y. Xu, and J. Liu, “Meta-analysis on the prevalence of REM sleep behavior disorder symptoms in Parkinson’s disease,” *BMC Neurology*, vol. 17, no. 1, p. 23, 2017.
- [140] F. Ghazi Sherbaf, Y. Rostam Abadi, M. Mojtabeh Zadeh, A. Ashraf-Ganjouei, H. Sanjari Moghaddam, and M. H. Aarabi, “Microstructural changes in patients with Parkinson’s disease comorbid with REM sleep behaviour disorder and depressive symptoms,” *Frontiers in Neurology*, vol. 9, 2018.
- [141] S. Happe and C. Trenkwalder, “Movement disorders in sleep: Parkinson’s disease and restless legs syndrome,” *Biomedizinische Technik. Biomedical Engineering*, vol. 48, no. 3, pp. 62–67, 2003.
- [142] Y.-S. Piao, T. H. Lian, Y. Hu et al., “Restless legs syndrome in Parkinson disease: clinical characteristics, abnormal iron metabolism and altered neurotransmitters,” *Scientific Reports*, vol. 7, no. 1, p. 10547, 2017.
- [143] T. C. Wetter, V. Collado-Seidel, T. Pollmacher, A. Yassouridis, and C. Trenkwalder, “Sleep and periodic leg movement patterns in drug-free patients with Parkinson’s disease and multiple system atrophy,” *Sleep*, vol. 23, no. 3, pp. 361–367, 2000.
- [144] W. G. Ondo, K. D. Vuong, and J. Jankovic, “Exploring the relationship between Parkinson disease and restless legs syndrome,” *Archives of Neurology*, vol. 59, no. 3, pp. 421–424, 2002.
- [145] T. Peeraully and E.-K. Tan, “Linking restless legs syndrome with Parkinson’s disease: clinical, imaging and genetic evidence,” *Translational Neurodegeneration*, vol. 1, no. 1, 2012.
- [146] L. Lungato, M. S. Marques, V. G. Pereira et al., “Sleep deprivation alters gene expression and antioxidant enzyme activity in mice splenocytes,” *Scandinavian Journal of Immunology*, vol. 77, no. 3, pp. 195–199, 2013.
- [147] G. Villafuerte, A. Miguel-Puga, E. Murillo Rodríguez, S. Machado, E. Manjarrez, and O. Arias-Carrión, “Sleep deprivation and oxidative stress in animal models: a systematic review,” *Oxidative Medicine and Cellular Longevity*, vol. 2015, 15 pages, 2015.
- [148] M. S. Trivedi, D. Holger, A. T. Bui, T. J. A. Craddock, and J. L. Tartar, “Short-term sleep deprivation leads to decreased systemic redox metabolites and altered epigenetic status,” *PLOS ONE*, vol. 12, no. 7, p. e0181978, 2017.
- [149] B. N. Greenwood, R. S. Thompson, M. R. Opp, and M. Fleshner, “Repeated exposure to conditioned fear stress increases anxiety and delays sleep recovery following exposure to an acute traumatic stressor,” *Frontiers in Psychiatry*, vol. 5, 2014.
- [150] R. Filograna, M. Beltramini, L. Bubacco, and M. Bisaglia, “Anti-oxidants in Parkinson’s disease therapy: a critical point of view,” *Current Neuropharmacology*, vol. 14, no. 3, pp. 260–271, 2016.
- [151] M. Kaminska, A.-L. Lafontaine, and R. J. Kimoff, “The interaction between obstructive sleep apnea and Parkinson’s disease: possible mechanisms and implications for cognitive function,” *Parkinson’s Disease*, vol. 2015, pp. 1–11, 2015.
- [152] Y. Cao, S. Zhen, A. Taylor, S. Appleton, E. Atlantis, and Z. Shi, “Magnesium intake and sleep disorder symptoms: findings from the Jiangsu Nutrition Study of Chinese adults at five-year follow-up,” *Nutrients*, vol. 10, no. 10, p. 1354, 2018.
- [153] T. Oyanagi and K. Hashimoto, “Magnesium in Parkinson’s disease: an update in clinical and basic aspects,” in *Magnesium in the Central Nervous System*, University of Adelaide Press, 2011.
- [154] M. Sturgeon, P. Wu, and R. Cornell, “SLC41A1 and TRPM7 in magnesium homeostasis and genetic risk for Parkinson’s disease,” *J. Neurol. neuromedicine*, vol. 1, no. 9, pp. 23–28, 2016.
- [155] X. Jin, M.-Y. Liu, D.-F. Zhang, H. Gao, and M.-J. Wei, “Elevated circulating magnesium levels in patients with Parkinson’s disease: a meta-analysis,” *Neuropsychiatric Disease and Treatment*, vol. Volume 14, pp. 3159–3168, 2018.
- [156] Z. Gan-Or, S. L. Girard, A. Noreau et al., “Parkinson’s disease genetic loci in rapid eye movement sleep behavior disorder,” *Journal of Molecular Neuroscience*, vol. 56, no. 3, pp. 617–622, 2015.
- [157] F. Ding, J. O’Donnell, Q. Xu, N. Kang, N. Goldman, and M. Nedergaard, “Changes in the composition of brain interstitial ions control the sleep-wake cycle,” *Science*, vol. 352, no. 6285, pp. 550–555, 2016.
- [158] O. Anichtchik, H. Diekmann, A. Fleming, A. Roach, P. Goldsmith, and D. C. Rubinsztein, “Loss of PINK1 function affects development and results in neurodegeneration in zebrafish,” *The Journal of Neuroscience*, vol. 28, no. 33, pp. 8199–8207, 2008.
- [159] C. A. Gautier, T. Kitada, and J. Shen, “Loss of PINK1 causes mitochondrial functional defects and increased sensitivity to oxidative stress,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 105, no. 32, pp. 11364–11369, 2008.
- [160] M. Priyadarshini, L. A. Orosco, and P. J. Panula, “Oxidative Stress and Regulation of Pink1 in Zebrafish (*Danio rerio*),” *PLoS ONE*, vol. 8, no. 11, p. e81851, 2013.
- [161] L. Zuo and M. S. Motherwell, “The impact of reactive oxygen species and genetic mitochondrial mutations in Parkinson’s disease,” *Gene*, vol. 532, no. 1, pp. 18–23, 2013.
- [162] M. M. M. Wilhelmus, P. G. Nijland, B. Drukarch, H. E. de Vries, and J. van Horssen, “Involvement and interplay of Parkin, PINK1, and DJ1 in neurodegenerative and neuroinflammatory disorders,” *Free Radical Biology & Medicine*, vol. 53, no. 4, pp. 983–992, 2012.
- [163] C. Klein and A. Westenberger, “Genetics of Parkinson’s Disease,” *Cold Spring Harbor Perspectives in Medicine*, vol. 2, no. 1, pp. a008888–a008888, 2012.
- [164] H.-X. Deng, Y. Shi, Y. Yang et al., “Identification of *_TMEM230_* mutations in familial Parkinson’s disease,” *Nature Genetics*, vol. 48, no. 7, pp. 733–739, 2016.