

Review Article

Role of p38 MAPK Signalling in Testis Development and Male Fertility

Dandan Luo,^{1,2,3} Zhao He ,^{1,3} Chunxiao Yu ,^{1,2,3} and Qingbo Guan ,^{1,2,3}

¹Department of Endocrinology, Shandong Provincial Hospital Affiliated to Shandong First Medical University, Jinan, Shandong 250021, China

²Department of Endocrinology, Shandong Provincial Hospital, Shandong University, Jinan, Shandong 250021, China

³Shandong Provincial Key Laboratory of Endocrinology and Lipid Metabolism, Institute of Endocrinology and Metabolism, Shandong Academy of Clinical Medicine, Jinan, Shandong 250021, China

Correspondence should be addressed to Chunxiao Yu; yuchx08@163.com and Qingbo Guan; doctorguanqingbo@163.com

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The testis is an important male reproductive organ, which ensures reproductive function via the secretion of testosterone and the generation of spermatozoa. Testis development begins in the embryonic period, continues after birth, and generally reaches functional maturation at puberty. The stress-activated kinase, p38 mitogen-activated protein kinase (MAPK), regulates multiple cell processes including proliferation, differentiation, apoptosis, and cellular stress responses. p38 MAPK signalling plays a crucial role in testis development by regulating spermatogenesis, the fate determination of pre-Sertoli, and primordial germ cells during embryogenesis, the proliferation of testicular cells in the postnatal period, and the functions of mature Sertoli and Leydig cells. In addition, p38 MAPK signalling is involved in decreased male fertility when exposed to various harmful stimuli. This review will describe in detail the biological functions of p38 MAPK signalling in testis development and male reproduction, together with its pathological role in male infertility.

1. Introduction

The p38 mitogen-activated protein kinase (p38 MAPK) belongs to the family of MAPKs, which are involved in a variety of cellular processes including cell proliferation, differentiation, apoptosis, and cellular stress responses [1]. There are four isoforms of the p38 MAPK family, which are encoded by distinct genes: *p38 α* (*Mapk14*), *p38 β* (*Mapk11*), *p38 γ* (*Mapk12*), and *p38 δ* (*Mapk13*) [2–4]. Each of the p38 MAPK isoforms have been identified in the testis [5]. Recent studies have indicated that p38 MAPK signalling has broad physiological and pathological effects on male reproduction.

The testis is the male reproductive organ responsible for the generation of testosterone and sperm [6]. Testis organogenesis begins in the embryo with the development of the bipotential gonad into a testis, as opposed to an ovary, in a process called sex determination. In the XY gonad, male sex determination and embryonic testis development are

regulated by an assortment of complex and interconnected molecular regulatory networks [6]. A defect in any XY gonad cells during testis development causes embryonic gonadal reversal and results in an improper testis [7]. In previous decades, studies have indicated that p38 MAPK signalling plays a vital role in male sex determination and embryonic testis development.

Postnatal testis development is a complex process involving the proliferation and differentiation of testicular somatic cells (Leydig cells and Sertoli cells), which culminates in testis maturation at puberty [8]. In adults, Leydig cells located at the testicular interstitium are the primary source of testosterone. Testosterone is essential for the development of the male phenotype and for spermatogenesis, alongside having various systemic endocrine effects outside of the testis [9–11]. Sertoli cells, the only somatic constituent of the testicular seminiferous epithelium, provide physical and nutritional support, as well as an immune-protective environment for the germ cells [12, 13]. Recent studies have

indicated that p38 MAPK signalling has broad physiological effects on the function of both Sertoli and Leydig cells.

Over several decades, there has been a significant decreasing trend in male fertility, including a decline in the concentration of testosterone and the quality of sperm. The etiology of male infertility is very diverse, including environmental factors, sedentary lifestyle, aging, and systemic or testicular diseases. Testicular oxidative stress is considered to be the central mechanism leading to male infertility caused by these factors. As a stress-activated protein, p38 MAPK has a substantial role in the pathogenesis of many diseases. Growing evidence indicates that p38 MAPK signalling is involved in the reduction of male fertility in hazardous situations.

In this review, we described the key biological effect of p38 MAPK signalling on embryonic testis development and adult male fertility, in addition to discussing its potential role in male infertility during oxidative stress.

2. p38 MAPK Signalling Is Required for Male Sex Determination and Testis Development

In the majority of mammals, testes and ovaries are derived from the common bipotential gonads. Male fate is determined by SRY (sex-determining region, Y gene), which triggers the testis developmental pathway [14, 15]. SRY activates SRY-box transcription factor 9 (SOX9), which triggers supporting cells to differentiate into male Sertoli cells rather than female pregranulosa cells [16]. Sertoli cells subsequently signal embryonic primordial germ cells (PGCs) to differentiate into male germ cells. It is currently accepted that p38 MAPK signalling has a crucial effect on the fate determination of both somatic cells and PGCs in the XY gonad.

2.1. p38 MAPK Signalling Is Indispensable for the Fate of Sertoli Cell during Male Sex Determination. In 2009, a gene mutation was identified in mice that caused gonadal sex reversal, resulting in ovarian development in an XY embryo. This genetic mutation for boygirl is an early stop codon which disrupts the autosomal gene encoding mitogen-activated protein kinase kinase kinase 4 (MAP3K4) [17]. This team subsequently revealed that haploinsufficiency of MAP3K4 gives rise to the sex reversal phenotype in mice [18]. p38 MAPK is downstream of MAP3K4 in several cellular processes and is therefore inhibited in the testes of mice with a *Map3k4* mutation [19]. Mice with genetic ablation of both *p38 α* and *p38 β* MAPK cause XY gonadal sex reversal, which confirms that MAP3K4 regulates male sex determination via p38 MAPK signalling [20, 21].

Recent data has determined that growth arrest and DNA damage inducible gamma (GADD45 γ) is the upstream kinase of MAP3K4 in male sex determination, since mice with a GADD45 γ defect display XY gonadal sex reversal via the MAP3K4/p38 MAPK signalling pathway [20, 21]. Additionally, mitogen-activated protein kinase kinase 6 (MAP2K6) is the upstream kinase of the p38 MAPK-mediated male sex determination [19, 22]. Therefore, GADD45 γ /MAP3K4/MAP2K6/p38 MAPK signalling determines the male sex determination (Figure 1(a)).

p38 MAPK signalling triggers the expression of SRY, which controls male sex determination. In XY gonads, a transient burst of SRY expression between 10.5 and 12.5 days postcoitum (dpc) in supporting cell progenitors initiates their commitment to a testicular fate (Sertoli cells) as opposed to ovarian development (pregranulosa cells) [23, 24]. In the absence of SRY, the gonadal primordium follows the alleged “default” pathway and develops into an ovary [15, 25]. p38 MAPK signalling ensures punctual expression of SRY via the transcription factor GATA-binding protein 4 (GATA4) in supporting cell progenitors, thereby prompting their commitment to a Sertoli cell fate [26]. Genetic ablation of p38 MAPK in an XY embryo causes XY gonadal sex reversal due to the delayed onset and reduced expression of SRY [20, 21].

Surprisingly, men with an overactivation of the p38 MAPK pathway caused by mutation in mitogen-activated protein kinase kinase kinase 1 (MAP3K1) display the male-to-female reversal phenotype [27]. MAP3K1 is one of upstream kinases of p38 MAPK. The mutation in MAP3K1 results in a gain of function and activates p38 MAPK. The overactivated p38 MAPK downregulates several male-specific genes, including SRY, SOX9, and anti-Müllerian hormone (AMH), and upregulates female-specific genes, including Wnt family member 4 (WNT4)/ β -catenin and stimulated by retinoic acid 8 (STRA8), causing XY gonadal dysgenesis (Figure 1(b)) [28, 29].

Overall, both inhibition and overactivation of p38 MAPK result in male-to-female sex reversal. This reveals that the activity of p38 MAPK is vital for male sex determination. Minor changes in p38 MAPK activity can tilt the balance from testis-determining to ovary-promoting signalling. Additional experiments are therefore required to clarify the underlying mechanisms.

2.2. p38 MAPK Signalling Is Necessary for XY Germ Cell Fate Determination in Foetal Testis Development. Mammalian spermatozoa and oocytes are derived from the common embryonic PGCs, which colonise the nascent gonad, and later undergo sex-specific fate determination [30]. In foetal ovaries, the presence of retinoic acid (RA) and STRA8 signals PGCs to enter meiosis and induces them to differentiate into female germ cells at 12.5 dpc [31–34]. In foetal testes, fibroblast growth factors 9 (FGF9) in Sertoli cells blocks the expression of RA/STRA8 signals by inducing the expression of cytochrome p450 family 26 subfamily b member 1 (CYP26B1), a gene encoding an enzyme which degrades RA [35, 36].

p38 MAPK signalling is required for the fate determination of XY PGCs in embryonic testis development by inhibiting RA/STRA8 signalling. p38 MAPK signalling is activated in PGCs of XY gonads from around 11.5 dpc and induces the expression of Nanos2 (Nanos C2HC-Type Zinc Finger 2) in mice [35, 36]. Nanos2 inhibits the upregulation of STRA8 to block PGCs from entering meiosis and promotes male differentiation in XY PGCs [37]. Thus, inhibition of p38 MAPK permits the expression of STRA8 in PGCs [38]. However, it remains unclear whether the

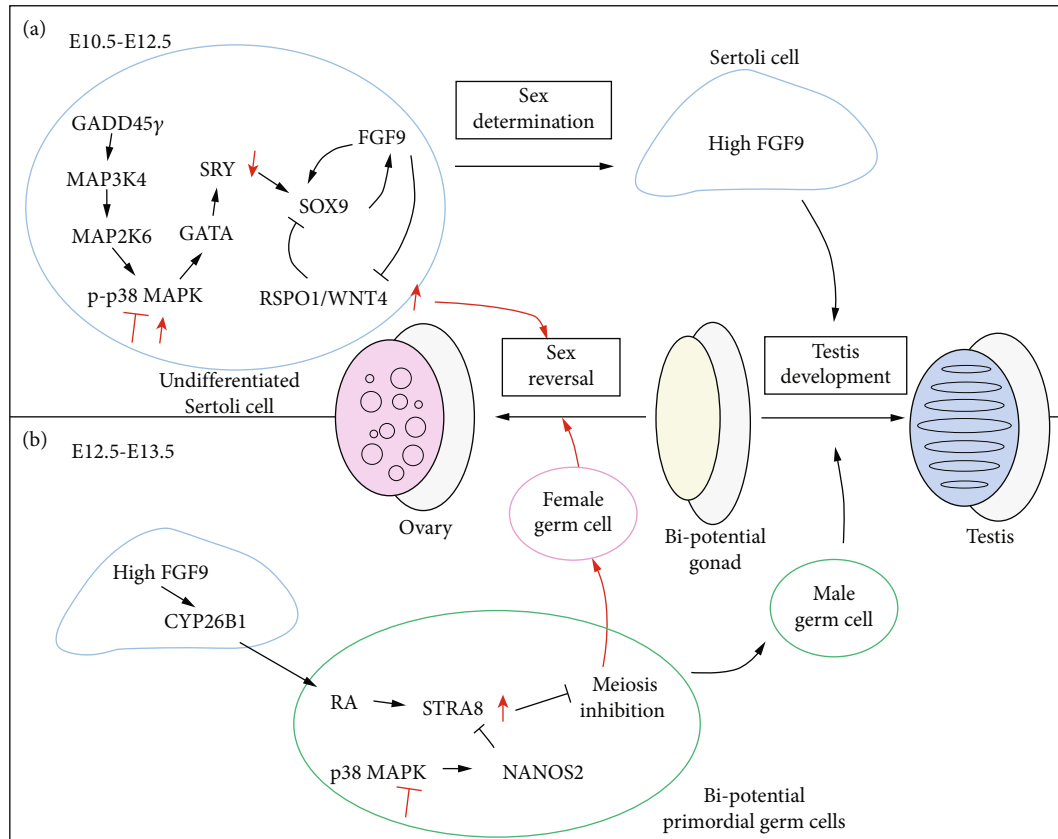


FIGURE 1: A summary of p38 MAPK signalling in sex determination in mice. (a) A diagrammatic representation of the sex fate decision of Sertoli cells. At around 10.5 dpc, p38 MAPK is activated by the GADD45 γ /MAP3K4/MAP2K6 pathway. Then, p-p38 MAPK activates GATA4 by phosphorylation and subsequent expression of SRY. SRY upregulates SOX9 and FGF9 expression to induce differentiation into Sertoli cells (black arrows). At 12.5 dpc, testis cords have formed and morphological differences between the testis (blue) and the ovary (pink) are evident. Both inhibition and overactivation of p38 MAPK cause delayed onset and reduced expression of SRY, which permits WNT4 and RSPO1 to express in a female-specific manner, inducing ovarian development (red arrows). (b) A schema for the sex fate decision of XY PGCs. In XY gonads, the SRY-SOX9-FGF9 pathway prevents XY PGCs from differentiating into female germ cells by blocking the network of RA/STRA8 through CYP26B1. p38 MAPK signalling is activated in PGCs of XY gonads around E11.5 dpc and induces the expression of Nanos2. Nanos2 inhibits the expression of STRA8 to block PGCs from entering meiosis and promotes a male differentiation in XY PGCs (black arrows). The inhibition of p38 MAPK permits the expression of STRA8 in PGCs (red arrows).

upstream kinase enzymes of p38 MAPK are involved in XY PGC fate determination.

As summarised in Figure 1(a), p38 MAPK is clearly necessary for determining the fate of pre-Sertoli cells and PGCs in XY gonads. Further research is needed in this area to clarify the underlying mechanisms at work.

3. The Role of p38 MAPK Signalling in Postnatal Testis Development

Testis development continues after birth. During this period, testicular somatic cells undergo proliferation and maturation, and spermatogenesis begins. p38 MAPK has a key role in proliferation, but not differentiation, of testicular cells during postnatal testis development.

Differential transcriptional profile analysis revealed that the signalling levels of the majority of MAPK genes are downregulated in mature and maturing Sertoli cells compared to immature Sertoli cells [39]. Perhaps p38 MAPK promotes Sertoli cell proliferation and blocks their maturation.

Cecilia et al. discovered that inhibiting p38 MAPK attenuates the proliferation of cultured immature Sertoli cells [40]. The downregulation of p38 MAPK is required for Sertoli cell maturation. The overactivation of p38 MAPK caused by uninterrupted expression of tetraspanin-8 inhibits the maturation of Sertoli cells. The natural downregulation of tetraspanin-8 during puberty is considered a prerequisite for Sertoli cell maturation [41].

Research investigating the role of p38 MAPK in the proliferation and maturation of Leydig cells and the initiation of spermatogenesis is limited. However, the effect of another member of the MAPK family, extracellular-signal-regulated kinase (ERK), on postnatal testicular development has been well studied. ERK promotes immature Sertoli cell proliferation, suggesting p38 MAPK may have a similar role in postnatal testis development [42].

Regarding Leydig cells, ERK signalling is critical for maintaining its population in the adult testis. The Leydig cell-specific deletion of ERK1/2 results in Leydig cell hypoplasia, hypergonadotropic hypogonadism, and loss of

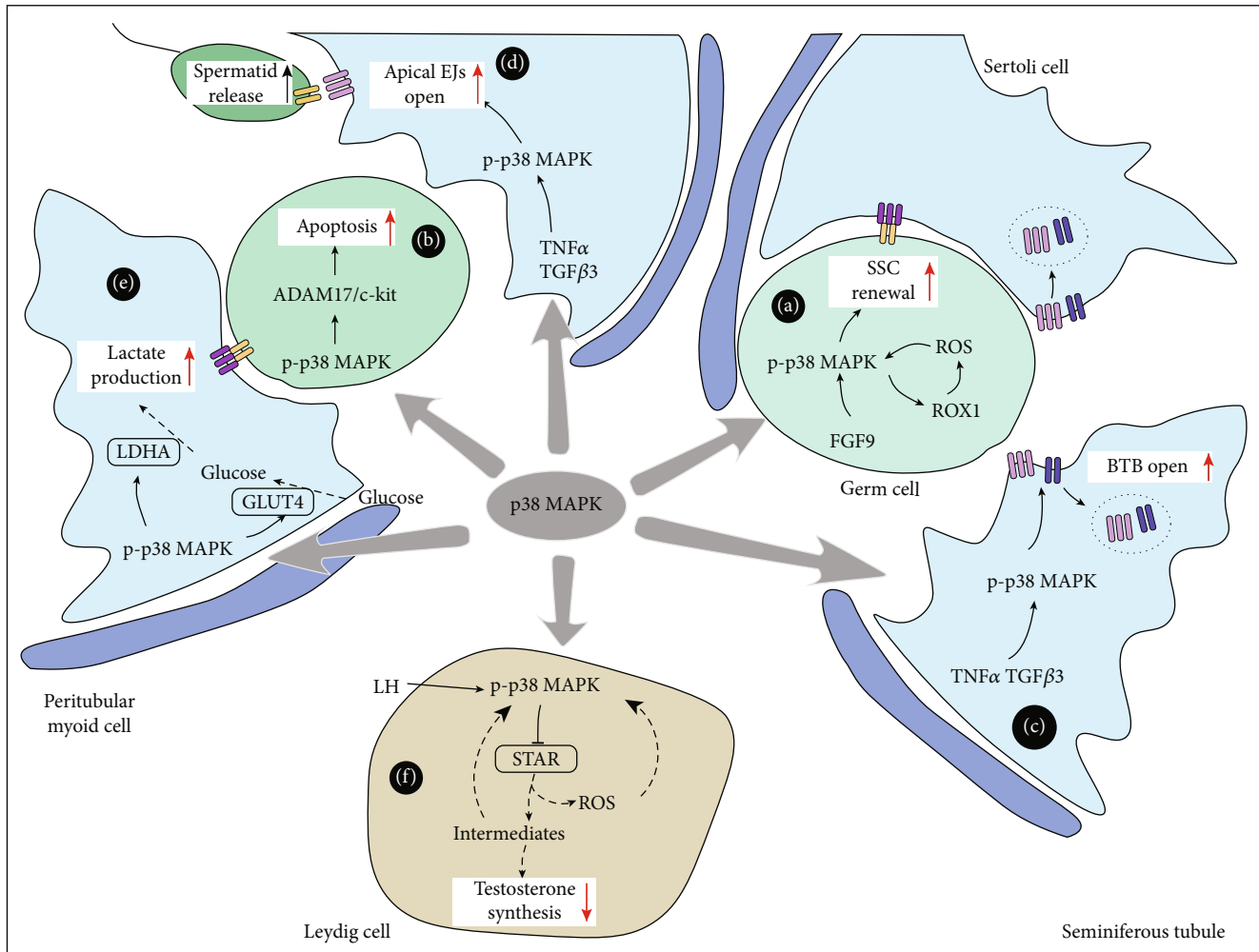


FIGURE 2: The molecular mechanisms of how p38 MAPK signalling regulates male fertility. p38 MAPK signalling is involved in (a) ROS-mediated SSC proliferation, (b) germ cell apoptosis, (c, d) dynamics of BTB and apical ESs, (e) glucose metabolism in Sertoli cells, and (f) negatively regulating testosterone synthesis in Leydig cells.

fertility in adult mice [43]. Regarding germ cells, single-cell RNA sequencing data reveals that ERK1/2 signalling is activated in undifferentiated spermatogonia and begins to decrease during the spermatogonial stem cell- (SSC-) to-progenitor transition [44]. In addition, the results from mice with germ cell-specific deletion of ERK1/2 confirm that ERK1/2 signalling is predominantly activated in SSCs to maintain their undifferentiated state [45]. Therefore, ERK1/2 is necessary for SSC self-renewal and proliferation, but not for the initiation of spermatogenesis.

As a kinase essential for regulating cell proliferation, p38 MAPK appears to promote the proliferation but not the differentiation of Sertoli, Leydig, and germ cells in postnatal testis development. A testicular cell conditional p38 MAPK knockout mouse is required to prove this inference.

4. p38 MAPK Signalling Regulates Male Fertility

Male fertility depends upon spermatogenesis, which is the generation of mature spermatozoa. Spermatogenesis is

tightly regulated by mature Sertoli and Leydig cells. Recent studies have indicated that p38 MAPK has a broad range of physiological roles in male fertility, including self-renewal and differentiation of SSCs and regulation of the functions of testicular somatic cells.

4.1. p38 MAPK Signalling Plays Multiple Roles in Various Biological Processes of Germ Cells. In adult testes, some SSCs maintain the stem cell pool through self-renewal, while others differentiate into spermatogonia to generate spermatozoa [46]. Spermatogonia proliferate and differentiate into primary spermatocytes, which undergo meiosis to produce haploid round spermatids [47]. These spermatids undergo metamorphosis into spermatozoa. Spermatozoa are subsequently released into the epididymis, where they undergo maturation to gain ability of motility [48]. p38 MAPK signalling has multiple roles in the biological processes of germ cells.

4.1.1. Self-Renewal and Differentiation of SSCs. As displayed in Figure 2(a), p38 MAPK is involved in SSC self-renewal

under the regulation of FGF9, which is vital for SSC self-renewal [49]. FGF9 activates p38 MAPK to promote the expression of ETS variant transcription factor 5 (ETV5) and B cell lymphoma 6 (BCL6), which are genes required for SSC self-renewal [50]. Inhibition of p38 MAPK inhibits FGF9-mediated SSC growth [51]. Similarly, an inhibitor of p38 α/β MAPK (SB202190) prevents SSC self-renewal in mice *in vitro* [52].

A moderate concentration of reactive oxygen species (ROS) is required for SSC proliferation. ROS deprivation inhibits SSC proliferation by using ROS scavengers or ablating the gene NADPH oxidase 1 (NOX1), which is required for ROS generation. In SSC proliferation, p38 MAPK is activated when exposed to moderate ROS concentrations, while the inhibition of p38 MAPK prevents the expression of NOX1. These results suggest that NOX1/ROS/p38 MAPK is involved in ROS-mediated SSC proliferation (Figure 2(a)) [53].

4.1.2. Spermatocyte Meiosis. DAZL (deleted in azoospermia-like) is a germ cell-specific RNA-binding protein, which has a vital role in spermatocyte meiosis [54, 55]. Recent studies suggest that p38 MAPK negatively regulates meiosis through DAZL. p38 MAPK activates MAPKAP kinase 2 (MK2), which phosphorylates DAZL and reduces the translation of DAZL-regulated target RNAs, resulting in a disorder in meiosis [56, 57].

4.1.3. Apoptosis of Germ Cells. During spermatogenesis, germ cell apoptosis is a key event which controls sperm output by eliminating damaged or unwanted sperm. p38 MAPK induces germ cell apoptosis in a process mediated by several proteins (Figure 2(b)). p38 MAPK activates ADAM17 (A disintegrin and metalloprotease-17), a widely distributed extracellular metalloprotease. ADAM17 induces shedding of the extracellular domains of c-KIT, a glycosylated transmembrane protein before inhibiting cell survival signals [57–59]. p38 MAPK signalling is activated by downregulation of cold-inducible RNA-binding protein (CIRP), an RNA-binding protein expressed in normal testes and downregulated following heat stress, promoting germ cell apoptosis [53].

4.1.4. Spermatozoan Maturation in the Epididymis. Spermatozoa in most mammalian species are kept completely motionless and viable for up to a few weeks in the cauda epididymis before ejaculation. Vigorous motility is initiated almost instantly upon sperm release from cauda during ejaculation. To gain fertilizing competence, they must go through a process called sperm capacitation [60]. In this process, sperm switch from progressive to hyperactivated motility and undergo a regulated release of acrosomal content in a process called the acrosome reaction (AR).

In both caudal and ejaculated spermatozoa, p-p38 MAPK is primarily localised to the upper midpiece of the spermatozoan tail where numerous mitochondria reside [61, 62], which suggests that p38 MAPK is closely related to sperm viability and motility. In caudal spermatozoa, p-p38 MAPK suppresses spermatozoan motility to ensure quiescence and survival via inhibiting mitochondrial respiratory capacity [63].

In ejaculated spermatozoa, the concentration of p-p38 MAPK is negatively associated with spermatozoan motility [64]. Consistent with this, p38 MAPK signalling inhibits both total and progressive spermatozoan motility [62]. Activated-p-p38 MAPK is involved in heat stress, and arachidonic acid (AA) caused the decline in sperm motility [65, 66]. In addition, p-p38 MAPK inhibits spermatozoan hyperactivated motility, which is a type of motility unique to capacitated spermatozoa [67]. Although p-p38 MAPK inhibits spermatozoan capacitation, it promotes the spermatozoan acrosome reaction [62].

4.2. p38 MAPK Signalling Regulates the Dynamics of the Blood-Testis Barrier (BTB) and Lactate Production in Sertoli Cells. The BTB consists of various types of junction connecting to adjacent Sertoli cells in proximity to the basement membrane, including tight junctions (TJs), adherens junctions (AJs), and gap junctions [68, 69]. The BTB separates the seminiferous epithelium into two distinct sections, the adluminal and basal compartments. Current researches indicate that p38 MAPK may regulate the dynamics of the BTB in a process mediated by transforming growth factor β (TGF- β) and tumor necrosis factor α (TNF- α) [70, 71]. TGF- β and TNF- α are crucial regulators of BTB dynamics [72].

As exhibited in Figure 2(c), p38 MAPK signalling is involved in the regulation of the BTB, mediated by TGF- β 3. When TGF- β 3 and its receptor T β R1 simultaneously bind to TAB1 and CD2AP, this complex activates p38 MAPK increases and subsequently disrupts the TJs by increasing the loss of TJ-associated proteins, such as occludin and zonula occludens-1, in cultured Sertoli cells [73]. This disruption is partially rescued by p38 MAPK inhibitor SB202190 [74]. Additionally, TGF- β 3/p38 MAPK disturbs the dynamics of apical ectoplasmic specialisations (ESs), mediated by downregulation of the apical ES-associated proteins including cadherins and catenins (Figure 2(d)) [73]. The disassembly of apical ESs, the junctions restricted to Sertoli cells and spermatids, facilitates the release of spermatozoa [75, 76].

p38 MAPK is also involved in the dynamics of TJs and apical ESs, mediated by TNF- α [77, 78]. TNF- α transiently inhibits the steady-state protein concentrations of occludin, zonula occludens-1, and N-cadherin via activating p38 MAPK [79]. In addition, p38 MAPK promotes the transcription of a BTB protein, junctional adhesion molecule-B, in Sertoli cells under the regulation of IL-1 α [80].

In addition, p38 MAPK in Sertoli cells serves as a significant regulator for glucose metabolism, making sure sufficient lactate supply for germ cells' energy substrate [81, 82]. Glucose deprivation activates p38 MAPK signalling in Sertoli cells, which subsequently increases the expression of glucose transporter type 1 (GLUT1), ensuring the uptake of glucose (Figure 2(e)) [83]. Activated p38 MAPK also promotes the expression and activity of lactate dehydrogenase (LDH) in Sertoli cells (Figure 2(e)) [84].

Generally, p38 MAPK is considered to be involved in the dynamics of the BTB and ESs to facilitate the migration of spermatocytes across the BTB and the release of spermatid. However, considerable *in vivo* research remains to be completed in this area.

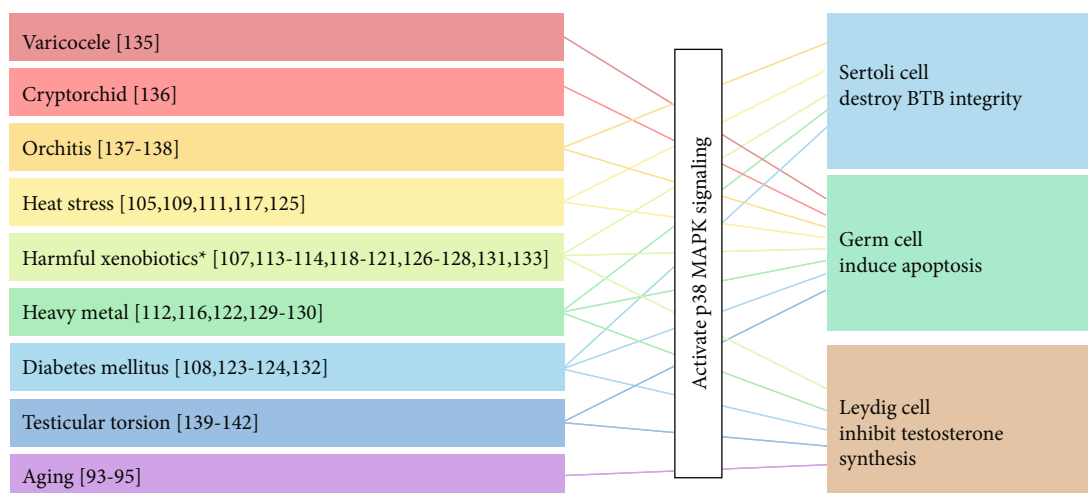


FIGURE 3: A summary of the stimuli which induce oxidative stress and activate the p38 MAPK pathway to injure testicular cells. *Harmful xenobiotics include PM2.5, organic compounds, pesticides, and plasticisers.

4.3. *p38 MAPK Is a Negative Regulator of Testosterone Synthesis in Leydig Cells.* In Leydig cells, a series of steroidogenic enzymes are responsible for testosterone biosynthesis. The steroidogenic acute regulatory (StAR) protein regulates the rate-limiting step in steroidogenesis. Both LH/human chorionic gonadotropin (hCG) and cAMP enhance StAR expression [85]. Currently, p38 MAPK is considered a negative regulator of testosterone synthesis in Leydig cells (Figure 2(f)).

In Leydig cells and adrenal cell lines, which also produce testosterone, p38 MAPK inhibits the expression of StAR [86]. Overexpression of wild type, as opposed to the dominant negative form of p38 MAPK, significantly reduced the basal and cAMP-sensitive activity of the StAR promoter [87]. During steroidogenesis, mitochondria generate vast quantities of ROS. ROS and steroid precursors/metabolites lead to the activation of p38 MAPK. Activated-p38 MAPK inhibits the transcriptional activity of the cAMP-response element-binding protein (CREB) by phosphorylating CREB (p-CREB), which results in reduced expression of StAR via a feedback loop [88].

In males, serum testosterone levels decline with advancing age. This is referred to as age-related testosterone decline, and p38 MAPK is reportedly involved in this process [89, 90]. Elevated oxidative stress in aging Leydig cells suppresses steroidogenesis through the activation of the p38 MAPK/CREB/StAR pathway, a key mechanism behind age-related testosterone decline [91, 92]. Additionally, p38 MAPK upregulates the expression of cyclooxygenase-2 (COX2) in aging cells [93], which has an inhibitory effect on steroidogenesis in both young and old Leydig cells [92–95].

5. p38 MAPK Signalling Is Involved in Male Infertility Caused by Testicular Oxidative Stress Insult

In recent decades, there has been a decline in the spermatozoan count and testosterone concentration worldwide

[96–98]. These declines are linked to the effects of environmental contaminants, lifestyle factors, aging, and systemic and testicular diseases. Virtually, all of these factors cause overproduction of ROS, leading to oxidative stress in the testicular cells, which is the leading cause of male infertility [99–101]. As summarised in Figure 3, compelling evidence suggests that p38 MAPK, as a major regulator under oxidative stress, exacerbates the decline in male fertility.

5.1. *Induce Germ Cell Apoptosis.* Testicular cells, particularly germ cells, are highly sensitive to oxidative stress and undergo apoptosis in response to ROS. p38 MAPK participates in germ cell apoptosis via various pathways. Activated intrinsic apoptotic pathways, p38 MAPK/ROS/BAX/caspases, induce apoptosis of germ cells or ejaculated spermatozoan as a result of heat stress, hormonal stimulation, PM2.5, diabetes mellitus, and other stimuli [53, 78, 102–108]. Activation of BAX leads to the subsequent initiation of mitochondria-dependent death processes. Conversely, activated extrinsic apoptotic pathways triggered by p38 MAPK mediate germ cell apoptosis when exposed to selenium and lead [109, 110]. Additionally, germ cell apoptosis in response to bisphenol-A and nonylphenol is regulated by the activation of p38 MAPK/ADAM17/c-kit signalling [111].

However, contradictory results also exist. Activated-p38 MAPK alleviates heat stress-induced germ cell damage. p38 MAPK and its downstream substrate MAPKAP kinase 2 (MK2) phosphorylate HSPA1 of the HSP70 family, which renders germ cells more resistant to heat stress-induced apoptosis [112].

5.2. *Disrupted BTB of Sertoli Cells.* The BTB is the most significantly affected structure when Sertoli cells are exposed to oxidative stress from various harmful stimuli. Activated-p38 MAPK is involved in the disruption of the BTB integrity in multiple pathways. TGF- β 3/p38 MAPK mediates the disruption of the BTB caused by physical/chemical factors, including PM 2.5, CdCl₂, and heat stress [113–115]. p38 MAPK/Nrf2 is involved in BTB damage caused by PM 2.5

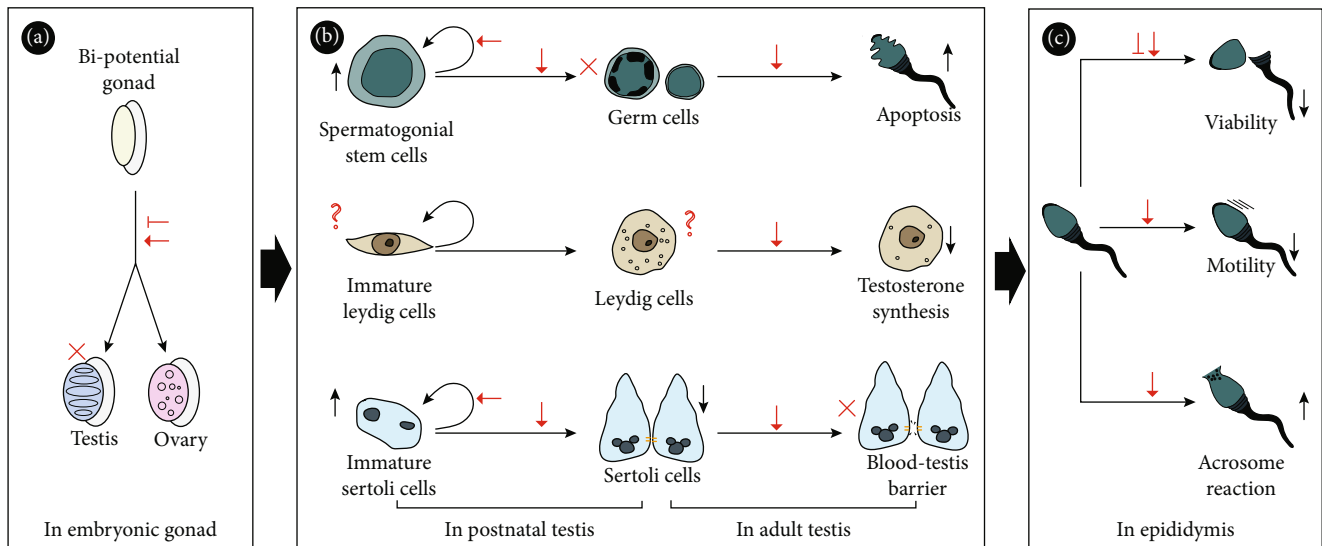


FIGURE 4: The roles of p38 MAPK in testis development and testicular function. (a) Both inhibition and overactivation of p38 MAPK disturb testis formation in embryonic gonad. (b) The effects of p38 MAPK on postnatal and adult testis: it maintains spermatogonial stem cell renewal and immature Sertoli cell proliferation; it inhibits spermatogenesis and Sertoli cell maturation; it reduces male fertility by promoting germ cell apoptosis, inhibiting testosterone synthesis, and disrupting integrity of the blood-testis barrier. (c) p38 MAPK regulates spermatozoan maturation, including maintaining spermatozoan viability, inhibiting spermatozoan motility, and improving spermatozoan capacitation in the epididymis. Red arrow indicates overactivation of p38 MAPK. Red line with end bar: inhibition of p38 MAPK. Shorter black arrow indicates upregulation or downregulation. Long black arrow: direction or flow. Red cross indicates a block. Red question mark indicates unknown.

and polystyrene microplastics [116, 117]. Aroclor1254 disturbs the BTB by promoting endocytosis and degradation of junction proteins, including JAM-A, N-cadherin, and β -catenin, via the p38 MAPK pathway [118]. Glyphosate initiates calcium-mediated cell death in Sertoli cells by activating p38 MAPK [119]. ROS/p38 MAPK in the testes destroys the integrity of the BTB in diabetes mellitus patients, though the exact mechanism is unclear [120, 121].

5.3. Inhibited Testosterone Synthesis in Leydig Cells. p38 MAPK contributes to reduced testosterone concentrations in Leydig cells through the inhibition of expression or activity of enzymes related to testosterone synthesis or by inducing Leydig cell apoptosis [122]. Benzo (a) pyrene and beta-cypermethrin exposure prompted a ROS imbalance and activated p38 MAPK, which suppressed testosterone synthesis by preventing the expression of steroidogenic enzymes, such as cytochrome p450 family 11 subfamily a member 1 (CYP11A1), 3β -hydroxysteroid dehydrogenase (3β -HSD), and 17β -HSD [123, 124]. In Leydig cells, perfluorooctane sulfonate disrupts testosterone biosynthesis via the p38 MAPK/CREB/CRTC2/StAR signalling pathway [125]. Cadmium induces testosterone synthesis disorder via the TLR4/MAPK/NF- κ B signalling pathway [126]. p38 MAPK activation contributes to the Ni-induced testosterone synthesis disruption in rat Leydig cells [127]. Microcystin-LR (MC-LR) and elevated concentrations of glucose activate testicular macrophages, promoting their release of TNF- α [128, 129]. TNF- α binds to the TNF receptor 1 on the Leydig cells, thus activating the ROS/p38 MAPK signalling pathway, resulting in reduced serum testosterone levels. Both cordycepin and acrylamide induced

apoptosis of Leydig cells by activating p38 MAPK signalling [130, 131].

In addition, ROS/p38 MAPK signalling has a role in male infertility in various testicular diseases, including varicocele [132], cryptorchidism [133], autoimmune orchitis [134, 135], and testicular torsion [136–139].

6. Concluding Remarks and Prospects for the Future

The p38 MAPK signalling pathway plays important roles in testis formation and male fertility at the testis and epididymis levels (Figure 4). It is known that testis development and male fertility are regulated by the hypothalamus-hypophysis axis. Few studies revealed that p38 MAPK is involved in developmental migration and maturation of gonadotropin-releasing hormone (GnRH) neurons [140, 141]. In addition, p38 MAPK is expressed in kiss neuron and participants in the initiation of puberty [142, 143]. Further studies are needed to clarify the exact effect of p38 MAPK on the hypothalamus-hypophysis axis.

Furthermore, more rigorous studies based on gene-editing experiments are necessary to validate the present conclusion. Because the use of p38 MAPK inhibitors is a prerequisite for the generation of meaningful results, it may not be a true reflection of the testis *in vivo*, since the testis is a heterogeneous organ comprised of numerous cell types.

In addition, there is no doubt that p38 MAPK contributes to the reduced male fertility caused by various harmful stimuli such as environmental contaminants, systemic

diseases, and aging. Several articles suggested that p38 MAPK signalling might be a potential effective therapeutical target. However, we do not think p38 MAPK per se is an ideal candidate, since p38 MAPK has both protective and detrimental effects in different testicular cells. The treatment based on the etiologies and ameliorating oxidative stress are more accessible, safe, and efficacious.

Data Availability

The data supporting this systematic review are from previously reported studies and datasets, which have been cited.

Conflicts of Interest

The authors declare no conflict of interest.

Authors' Contributions

Zhao He, Chunxiao Yu, and Qingbo Guan conceived of and designed the review. Dandan Luo wrote the paper.

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References

- [1] M. Cargnello and P. P. Roux, "Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases," *Microbiology and Molecular Biology Reviews*, vol. 75, pp. 50–83, 2011.
- [2] J. M. Kyriakis and J. Avruch, "Mammalian MAPK signal transduction pathways activated by stress and inflammation: a 10-year update," *Physiological Reviews*, vol. 92, pp. 689–737, 2012.
- [3] A. Cuadrado and A. R. Nebreda, "Mechanisms and functions of p38 MAPK signalling," *The Biochemical Journal*, vol. 429, pp. 403–417, 2010.
- [4] A. Cuenda and S. Rousseau, "p38 MAP-kinases pathway regulation, function and role in human diseases," *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, vol. 1773, no. 8, pp. 1358–1375, 2007.
- [5] C. H. Wong and C. Y. Cheng, "Mitogen-activated protein kinases, adherens junction dynamics, and spermatogenesis: a review of recent data," *Developmental Biology*, vol. 286, pp. 1–15, 2005.
- [6] A. K. Lucas-Herald and A. Bashamboo, "Gonadal development," *Endocrine Development*, vol. 27, pp. 1–16, 2014.
- [7] N. Warr and A. Greenfield, "The molecular and cellular basis of gonadal sex reversal in mice and humans," *Wiley Interdisciplinary Reviews: Developmental Biology*, vol. 1, no. 4, pp. 559–577, 2012.
- [8] F. E. Franke, K. Pauls, R. Rey, A. Marks, M. Bergmann, and K. Steger, "Differentiation markers of Sertoli cells and germ cells in fetal and early postnatal human testis," *Anatomy and embryology*, vol. 209, no. 2, pp. 169–177, 2004.
- [9] W. H. Walker, "Testosterone signaling and the regulation of spermatogenesis," *Spermatogenesis*, vol. 1, pp. 116–120, 2011.
- [10] A. Tsujimura, "The relationship between testosterone deficiency and men's health," *The world journal of men's health*, vol. 31, no. 2, pp. 126–135, 2013.
- [11] L. B. Smith and W. H. Walker, "The regulation of spermatogenesis by androgens," *Seminars in Cell & Developmental Biology*, vol. 30, pp. 2–13, 2014.
- [12] N. F. Da, S. L. Hao, and W. X. Yang, "Multiple signaling pathways in Sertoli cells: recent findings in spermatogenesis," *Cell Death & Disease*, vol. 10, no. 8, p. 541, 2019.
- [13] M. D. Griswold, "50 years of spermatogenesis: Sertoli cells and their interactions with germ cells," *Biology of Reproduction*, vol. 99, no. 1, pp. 87–100, 2018.
- [14] J. Bowles and P. Koopman, "Sex determination in mammalian germ cells: extrinsic versus intrinsic factors," *Reproduction*, vol. 139, no. 6, pp. 943–958, 2010.
- [15] K. Kashimada and P. Koopman, "Sry: the master switch in mammalian sex determination," *Development*, vol. 137, pp. 3921–3930, 2010.
- [16] N. Gonen and R. Lovell-Badge, *The regulation of Sox9 expression in the gonad*, Aufl. Elsevier Inc., 2019.
- [17] D. Bogani, P. Siggers, R. Brixey et al., "Loss of mitogen-activated protein kinase kinase 4 (MAP3K4) reveals a requirement for MAPK signalling in mouse sex determination," *PLoS Biology*, vol. 7, no. 9, article e1000196, 2009.
- [18] N. Warr, P. Siggers, G. A. Carré et al., "Transgenic expression of *Map3k4* rescues T-associated sex reversal (Tas) in mice," *Human Molecular Genetics*, vol. 23, pp. 3035–3044, 2014.
- [19] N. A. Shendy, A. L. Broadhurst, K. Shoemaker, R. Read, and A. N. Abell, "MAP3K4 kinase activity dependent control of mouse gonadal sex determination," *Biology of Reproduction*, vol. 105, pp. 491–502, 2021.
- [20] N. Warr, G. A. Carre, P. Siggers et al., "*Gadd45* γ and *Map3k4* interactions regulate mouse testis determination via p38 MAPK-mediated control of *Sry* expression," *Developmental Cell*, vol. 23, no. 5, pp. 1020–1031, 2012.
- [21] M. S. Gierl, W. H. Gruhn, A. von Seggern, N. Maltry, and C. Niehrs, "GADD45G functions in male sex determination by promoting p38 signaling and *Sry* expression," *Developmental Cell*, vol. 23, no. 5, pp. 1032–1042, 2012.
- [22] N. Warr, P. Siggers, G. A. Carré, S. Wells, and A. Greenfield, "Genetic analyses reveal functions for MAP2K3 and MAP2K6 in mouse testis determination1," *Biology of Reproduction*, vol. 94, no. 5, pp. 1–7, 2016.
- [23] J. Schmahl, E. M. Eicher, L. L. Washburn, and B. Capel, "Sry induces cell proliferation in the mouse gonad," *Development*, vol. 127, no. 1, pp. 65–73, 2000.
- [24] E. Yildirim, S. Aksoy, T. Onel, and A. Yaba, "Gonadal development and sex determination in mouse," *Reproductive Biology*, vol. 20, pp. 115–126, 2020.
- [25] R. Sekido and R. Lovell-Badge, "Sex determination and SRY: down to a wink and a nudge?," *Trends in Genetics*, vol. 25, no. 1, pp. 19–29, 2009.
- [26] S. G. Tevosian, K. H. Albrecht, J. D. Crispino, Y. Fujiwara, E. M. Eicher, and S. H. Orkin, "Gonadal differentiation, sex determination and normal *Sry* expression in mice require direct interaction between transcription partners GATA4 and FOG2," *Development*, vol. 129, pp. 4627–4634, 2002.
- [27] H. Ostrer, "Pathogenic variants in MAP3K1 cause 46, XY gonadal dysgenesis: a review," *Sexual Development*, vol. mar, pp. 1–6, 2022.

- [28] J. Loke, A. Pearlman, O. Radi et al., "Mutations in MAP3K1 tilt the balance from SOX9/FGF9 to WNT/ β -catenin signaling," *Human Molecular Genetics*, vol. 23, no. 4, pp. 1073–1083, 2014.
- [29] X. Y. Dsd, "MAP3K1 variant causes hyperactivation of Wnt4/ β -catenin/FOXL2 signaling contributing to 46,XY disorders/differences of sex development," *Frontiers in Genetics*, vol. 13, pp. 1–15, 2022.
- [30] P. K. Nicholls, H. Schorle, S. Naqvi et al., "Mammalian germ cells are determined after PGC colonization of the nascent gonad," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 116, pp. 25677–25687, 2019.
- [31] T. Endo, M. M. Mikedis, P. K. Nicholls, D. C. Page, and D. G. de Rooij, "Retinoic acid and germ cell development in the ovary and testis," *Biomolecules*, vol. 9, no. 12, pp. 1–20, 2019.
- [32] J. Koubova, D. B. Menke, Q. Zhou, B. Capel, M. D. Griswold, and D. C. Page, "Retinoic acid regulates sex-specific timing of meiotic initiation in mice," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 8, pp. 2474–2479, 2006.
- [33] J. Bowles, D. Knight, C. Smith et al., "Retinoid signaling determines germ cell fate in mice," *Science (80-)*, vol. 312, no. 5773, pp. 596–600, 2006.
- [34] M. Mark, H. Jacobs, M. Oulad-Abdelghani et al., "STRA8-deficient spermatocytes initiate, but fail to complete, meiosis and undergo premature chromosome condensation," *Journal of Cell Science*, vol. 121, pp. 3233–3242, 2008.
- [35] G. MacLean, H. Li, D. Metzger, P. Chambon, and M. Petkovich, "Apoptotic extinction of germ cells in testes of *Cyp26b1* knockout mice," *Endocrinology*, vol. 148, pp. 4560–4567, 2007.
- [36] J. Bowles, C. W. Feng, C. Spiller, T. L. Davidson, A. Jackson, and P. Koopman, "FGF9 suppresses meiosis and promotes male germ cell fate in mice," *Developmental Cell*, vol. 19, no. 3, pp. 440–449, 2010.
- [37] A. Suzuki and Y. Saga, "Nanos 2 suppresses meiosis and promotes male germ cell differentiation," *Genes & Development*, vol. 22, pp. 430–435, 2008.
- [38] K. Ewen, A. Jackson, D. Wilhelm, and P. Koopman, "A male-specific role for p38 mitogen-activated protein kinase in germ cell sex differentiation in mice," *Biology of Reproduction*, vol. 83, no. 6, pp. 1005–1014, 2010.
- [39] M. Gautam, I. Bhattacharya, U. Rai, and S. S. Majumdar, "Hormone induced differential transcriptome analysis of Sertoli cells during postnatal maturation of rat testes," *PLoS One*, vol. 13, 2018.
- [40] C. Petersen, K. Svechnikov, B. Fröysa, and O. Söder, "The p38 MAPK pathway mediates interleukin-1-induced Sertoli cell proliferation," *Cytokine*, vol. 32, pp. 51–59, 2005.
- [41] B. S. Pradhan, I. Bhattacharya, R. Sarkar, and S. S. Majumdar, "Pubertal down-regulation of Tetrastin 8 in testicular Sertoli cells is crucial for male fertility," *Molecular Human Reproduction*, vol. 26, no. 10, pp. 760–772, 2020.
- [42] P. Crépieux, S. Marion, N. Martinat et al., "The ERK-dependent signalling is stage-specifically modulated by FSH, during primary Sertoli cell maturation," *Oncogene*, vol. 20, no. 34, pp. 4696–4709, 2001.
- [43] S. Yamashita, P. Tai, J. Charron, C. M. Ko, and M. Ascoli, "The Leydig cell MEK/ERK pathway is critical for maintaining a functional population of adult Leydig cells and for fertility," *Molecular Endocrinology*, vol. 25, no. 7, pp. 1211–1222, 2011.
- [44] J. Liao, S. H. Ng, A. C. Luk et al., "Revealing cellular and molecular transitions in neonatal germ cell differentiation using single cell RNA sequencing," *Development*, vol. 146, no. 6, 2019.
- [45] K. Hasegawa, S. H. Namekawa, and Y. Saga, "MEK/ERK signaling directly and indirectly contributes to the cyclical self-renewal of spermatogonial stem cells," *Stem Cells*, vol. 31, pp. 2517–2527, 2013.
- [46] D. G. De Rooij, "The nature and dynamics of spermatogonial stem cells," *Development*, vol. 144, pp. 3022–3030, 2017.
- [47] M. D. Griswold and M. D. Griswold, "Spermatogenesis: the commitment to meiosis," *Physiological Reviews*, vol. 96, pp. 1–17, 2016.
- [48] R. Sullivan and R. Miesusset, "The human epididymis: its function in sperm maturation," *Human Reproduction Update*, vol. 22, no. 5, pp. 574–587, 2016.
- [49] J. M. Oatley and R. L. Brinster, "The germline stem cell niche unit in mammalian testes," *Physiological Reviews*, vol. 92, pp. 577–595, 2012.
- [50] J. M. Oatley, M. R. Avarbock, A. I. Telaranta, D. T. Fearon, and R. L. Brinster, "Identifying genes important for spermatogonial stem cell self-renewal and survival," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, pp. 9524–9529, 2006.
- [51] F. Yang, E. C. Whelan, X. Guan et al., "FGF9 promotes mouse spermatogonial stem cell proliferation mediated by p38 MAPK signalling," *Cell Proliferation*, vol. 54, no. 1, article e12933, 2021.
- [52] Z. Niu, H. Mu, H. Zhu, J. Wu, and J. Hua, "p38 MAPK pathway is essential for self-renewal of mouse male germline stem cells (mGSCs)," *Cell Proliferation*, vol. 50, no. 1, pp. e12314–e12315, 2017.
- [53] Z. P. Xia, X. M. Zheng, H. Zheng, X. J. Liu, G. Y. Liu, and X. H. Wang, "Downregulation of cold-inducible RNA-binding protein activates mitogen-activated protein kinases and impairs spermatogenic function in mouse testes," *Asian Journal of Andrology*, vol. 14, no. 6, pp. 884–889, 2012.
- [54] H. Li, Z. Liang, J. Yang et al., "DAZL is a master translational regulator of murine spermatogenesis," *National Science Review*, vol. 344, pp. 455–468, 2019.
- [55] M. M. Mikedis, Y. Fan, P. K. Nicholls et al., "Dazl mediates a broad translational program regulating expansion and differentiation of spermatogonial progenitors," *eLife*, vol. 9, pp. 1–96, 2020.
- [56] P. A. Williams, M. S. Krug, E. A. McMillan et al., "Phosphorylation of the RNA-binding protein dazl by MAPKAP kinase 2 regulates spermatogenesis," *Molecular Biology of the Cell*, vol. 27, no. 15, pp. 2341–2350, 2016.
- [57] J. Matsui, T. Wakabayashi, M. Asada, K. Yoshimatsu, and M. Okada, "Stem cell factor/c-kit signaling promotes the survival, migration, and capillary tube formation of human umbilical vein endothelial cells," *The Journal of Biological Chemistry*, vol. 279, pp. 18600–18607, 2004.
- [58] M. A. Bedell and A. M. Zama, "Genetic analysis of kit ligand functions during mouse spermatogenesis," *Journal of Andrology*, vol. 25, no. 2, pp. 188–199, 2004.
- [59] L. Rönstrand, "Signal transduction via the stem cell factor receptor/c-kit," *Cellular and Molecular Life Sciences*, vol. 61, no. 19–20, pp. 2535–2548, 2004.

- [60] P. Tienthai, A. Johannisson, and H. Rodriguez-Martinez, "Sperm capacitation in the porcine oviduct," *Animal Reproduction Science*, vol. 80, no. 1-2, pp. 131-146, 2004.
- [61] Y. Shima and K. Morohashi, "Leydig progenitor cells in fetal testis," *Molecular and Cellular Endocrinology*, vol. 445, pp. 55-64, 2017.
- [62] T. Almog, S. Lazar, N. Reiss et al., "Identification of extracellular signal-regulated kinase 1/2 and p38 MAPK as regulators of human sperm motility and acrosome reaction and as predictors of poor spermatozoan quality," *The Journal of Biological Chemistry*, vol. 283, no. 21, pp. 14479-14489, 2008.
- [63] L. Kumar, S. K. Yadav, B. Kushwaha et al., "Energy utilization for survival and fertilization-parsimonious quiescent sperm turn extravagant on motility activation in rat," *Biology of Reproduction*, vol. 94, pp. 1-9, 2016.
- [64] J. V. Silva, M. J. Freitas, B. R. Correia et al., "Profiling signaling proteins in human spermatozoa: biomarker identification for sperm quality evaluation," *Fertility and Sterility*, vol. 104, pp. 845-856.e8, 2015.
- [65] M. B. Rahman, L. Vandaele, T. Rijsselaere et al., "Bovine spermatozoa react to in vitro heat stress by activating the mitogen-activated protein kinase 14 signalling pathway," *Reproduction, Fertility, and Development*, vol. 26, pp. 245-257, 2014.
- [66] L. Yu, X. Yang, B. Ma et al., "Abnormal arachidonic acid metabolic network may reduce sperm motility via P38 MAPK," *Open Biology*, vol. 9, no. 4, p. 9, 2019.
- [67] P. Sun, Y. Wang, T. Gao et al., "Hsp90 modulates human sperm capacitation via the Erk1/2 and p38 MAPK signaling pathways," *Reproductive Biology and Endocrinology*, vol. 19, pp. 1-11, 2021.
- [68] P. P. Y. Lie, C. Y. Cheng, and D. D. Mruk, "Signalling pathways regulating the blood-testis barrier," *The International Journal of Biochemistry & Cell Biology*, vol. 45, pp. 621-625, 2013.
- [69] C. Yan Cheng and D. D. Mruk, "The blood-testis barrier and its implications for male contraception," *Pharmacological Reviews*, vol. 64, pp. 16-64, 2012.
- [70] C. Cheng, M. Li, and D. Mruk, "Unlocking" the blood-testis barrier and the ectoplasmic specialization by cytokines during spermatogenesis: emerging targets for male contraception," *Immunology, Endocrine & Metabolic Agents in Medicinal Chemistry (Formerly Current Medicinal Chemistry-Immunology, Endocrine and Metabolic Agents)*, vol. 8, pp. 20-27, 2008.
- [71] W. Y. Lui and C. Y. Cheng, "Regulation of cell junction dynamics by cytokines in the testis—a molecular and biochemical perspective," *Cytokine & Growth Factor Reviews*, vol. 18, pp. 299-311, 2007.
- [72] M. W. M. Li, D. D. Mruk, W. M. Lee, and C. Y. Cheng, "Cytokines and junction restructuring events during spermatogenesis in the testis: an emerging concept of regulation," *Cytokine & Growth Factor Reviews*, vol. 20, no. 4, pp. 329-338, 2009.
- [73] W. Xia, D. D. Mruk, W. M. Lee, and C. Y. Cheng, "Differential interactions between transforming growth factor- β 3/T β R1, TAB1, and CD2AP disrupt blood-testis barrier and Sertoli-germ cell adhesion," *The Journal of Biological Chemistry*, vol. 281, pp. 16799-16813, 2006.
- [74] W. Lui, W. M. Lee, and C. Y. Cheng, "Transforming growth factor β 3 regulates the dynamics of Sertoli cell tight junctions via the p38 mitogen-activated protein kinase pathway," *Biology of Reproduction*, vol. 68, pp. 1597-1612, 2003.
- [75] B. Zheng, J. Yu, Y. Guo et al., "Cellular nucleic acid-binding protein is vital to testis development and spermatogenesis in mice," *Reproduction*, vol. 156, no. 1, pp. 59-69, 2018.
- [76] C. H. Wong, W. Xia, N. P. Y. Lee, D. D. Mruk, W. M. Lee, and C. Y. Cheng, "Regulation of ectoplasmic specialization dynamics in the seminiferous epithelium by focal adhesion-associated proteins in testosterone-suppressed rat testes," *Endocrinology*, vol. 146, no. 3, pp. 1192-1204, 2005.
- [77] A. Hellani, J. Ji, C. Mauduit, C. Deschildre, E. Tabone, and M. Benahmed, "Developmental and hormonal regulation of the expression of oligodendrocyte-specific protein/claudin 11 in mouse testis," *Endocrinology*, vol. 141, pp. 3012-3019, 2000.
- [78] M. K. Y. Siu, W. M. Lee, and C. Y. Cheng, "The interplay of collagen IV, tumor necrosis factor- α , gelatinase B (matrix metalloprotease-9), and tissue inhibitor of metalloproteases-1 in the basal lamina regulates sertoli cell-tight junction dynamics in the rat testis," *Endocrinology*, vol. 144, pp. 371-387, 2003.
- [79] M. W. Li, W. Xia, D. D. Mruk et al., "Tumor necrosis factor α reversibly disrupts the blood-testis barrier and impairs Sertoli-germ cell adhesion in the seminiferous epithelium of adult rat testes," *The Journal of Endocrinology*, vol. 190, pp. 313-329, 2006.
- [80] Y. Wang and W. Y. Lui, "Opposite effects of interleukin-1 α and transforming growth factor- β 2 induce stage-specific regulation of junctional adhesion molecule-B gene in Sertoli cells," *Endocrinology*, vol. 150, no. 5, pp. 2404-2412, 2009.
- [81] A. Kishimoto, T. Ishiguro-Oonuma, R. Takahashi et al., "Immunohistochemical localization of GLUT3, MCT1, and MCT2 in the testes of mice and rats: the use of different energy sources in spermatogenesis," *Biomedical Research*, vol. 36, pp. 225-234, 2015.
- [82] S. Brauchi, M. C. Rauch, I. E. Alfaro et al., "Kinetics, molecular basis, and differentiation of L-lactate transport in spermatogenic cells," *American journal of physiology-Cell physiology*, vol. 288, no. 3, pp. C523-C534, 2005.
- [83] M. F. Riera, M. N. Galardo, E. H. Pellizzari, S. B. Meroni, and S. B. Cigorruga, "Molecular mechanisms involved in Sertoli cell adaptation to glucose deprivation," *American journal of physiology-endocrinology and metabolism*, vol. 297, no. 4, pp. E907-E914, 2009.
- [84] M. N. Galardo, M. F. Riera, M. Regueira, E. H. Pellizzari, S. B. Cigorruga, and S. B. Meroni, "Different signal transduction pathways elicited by basic fibroblast growth factor and interleukin 1 β regulate CREB phosphorylation in Sertoli cells," *Journal of Endocrinological Investigation*, vol. 36, pp. 331-338, 2013.
- [85] A. J. Reinhart, S. C. Williams, and D. M. Stocco, "Transcriptional regulation of the STAR gene," *Molecular and Cellular Endocrinology*, vol. 151, pp. 161-169, 1999.
- [86] A. Turcu, J. M. Smith, R. Auchus, and W. E. Rainey, "Adrenal androgens and androgen precursors—definition, synthesis, regulation and physiologic actions," *Comprehensive Physiology*, vol. 4, no. 4, pp. 1369-1381, 2014.
- [87] S. K. Zaidi, W. J. Shen, S. Bittner et al., "p38 MAPK regulates steroidogenesis through transcriptional repression of STAR gene," *Journal of Molecular Endocrinology*, vol. 53, pp. 1-16, 2014.

- [88] J. Li, Q. Zhou, Z. Ma et al., "Feedback inhibition of CREB signaling by p38 MAPK contributes to the negative regulation of steroidogenesis," *Reproductive Biology and Endocrinology*, vol. 15, pp. 1–13, 2017.
- [89] S. I. Liochev, "Reactive oxygen species and the free radical theory of aging," *Free Radical Biology & Medicine*, vol. 60, pp. 1–4, 2013.
- [90] D. P. Jones, "Radical-free biology of oxidative stress," *American Journal of Physiology-Cell Physiology*, vol. 295, no. 4, pp. C849–C868, 2008.
- [91] P. Abidi, S. Leers-Sucheta, Y. Cortez, J. Han, and S. Azhar, "Evidence that age-related changes in p38 MAP kinase contribute to the decreased steroid production by the adrenocortical cells from old rats," *Aging Cell*, vol. 7, no. 2, pp. 168–178, 2008.
- [92] P. Abidi, H. Zhang, S. M. Zaidi et al., "Oxidative stress-induced inhibition of adrenal steroidogenesis requires participation of p38 mitogen-activated protein kinase signaling pathway," *The Journal of Endocrinology*, vol. 198, no. 1, pp. 193–207, 2008.
- [93] Y. Zhao, X. Liu, Y. Qu et al., "The roles of p38 MAPK → COX2 and NF-κB → COX2 signal pathways in age-related testosterone reduction," *Scientific Reports*, vol. 9, no. 1, pp. 1–11, 2019.
- [94] B. R. Zirkin, "Cyclooxygenase-2 and reduced steroidogenesis in the aging male," *Endocrinology*, vol. 146, pp. 4200–4201, 2005.
- [95] H. Chen, L. Luo, J. Liu, and B. R. Zirkin, "Cyclooxygenases in rat Leydig cells: effects of luteinizing hormone and aging," *Endocrinology*, vol. 148, no. 2, pp. 735–742, 2007.
- [96] P. Sengupta, E. Borges Jr., S. Dutta, and E. Krajewska-Kulak, "Decline in sperm count in European men during the past 50 years," *Human & Experimental Toxicology*, vol. 37, pp. 247–255, 2018.
- [97] P. Sengupta, U. Nwagha, S. Dutta, E. Krajewska-Kulak, and E. Izuka, "Evidence for decreasing sperm count in African population from 1965 to 2015," *African Health Sciences*, vol. 17, pp. 418–427, 2017.
- [98] P. Sengupta, S. Dutta, and E. Krajewska-Kulak, "The disappearing sperms: analysis of reports published between 1980 and 2015," *American Journal of Men's Health*, vol. 11, pp. 1279–1304, 2017.
- [99] S. Kumar, S. Murarka, V. V. Mishra, and A. K. Gautam, "Environmental & lifestyle factors in deterioration of male reproductive health," *The Indian Journal of Medical Research*, vol. 140, pp. 29–35, 2014.
- [100] A. Salas-Huetos, E. R. James, K. I. Aston, T. G. Jenkins, and D. T. Carrell, "Diet and sperm quality: nutrients, foods and dietary patterns," *Reproductive Biology*, vol. 19, pp. 219–224, 2019.
- [101] L. Nordkap, U. N. Joensen, M. B. Jensen, and N. Jørgensen, "Regional differences and temporal trends in male reproductive health disorders: semen quality may be a sensitive marker of environmental exposures," *Molecular and Cellular Endocrinology*, vol. 355, pp. 221–230, 2012.
- [102] P. J. Hansen, "Effects of heat stress on mammalian reproduction," *Philosophical Transactions of the Royal Society B: Biological Sciences*, vol. 364, no. 1534, pp. 3341–3350, 2009.
- [103] Y. Vera, K. Erkkilä, C. Wang et al., "Involvement of p38 mitogen-activated protein kinase and inducible nitric oxide synthase in apoptotic signaling of murine and human male germ cells after hormone deprivation," *Molecular Endocrinology*, vol. 20, pp. 1597–1609, 2006.
- [104] B. Liu, S. D. Wu, L. J. Shen et al., "Spermatogenesis dysfunction induced by PM_{2.5} from automobile exhaust via the ROS-mediated MAPK signaling pathway," *Ecotoxicology and Environmental Safety*, vol. 167, pp. 161–168, 2019.
- [105] Y. Zhao, Y. Tan, J. Dai et al., "Exacerbation of diabetes-induced testicular apoptosis by zinc deficiency is most likely associated with oxidative stress, p38 MAPK activation, and p53 activation in mice," *Toxicology Letters*, vol. 200, pp. 100–106, 2011.
- [106] C. Lizama, C. F. Lagos, R. Lagos-Cabré et al., "Calpain inhibitors prevent p38 MAPK activation and germ cell apoptosis after heat stress in pubertal rat testes," *Journal of Cellular Physiology*, vol. 221, pp. 296–305, 2009.
- [107] Y. Jia, J. Castellanos, C. Wang et al., "Mitogen-activated protein kinase signaling in male germ cell apoptosis in the rat," *Biology of Reproduction*, vol. 80, pp. 771–780, 2009.
- [108] L. Zou, G. Cheng, C. Xu et al., "The role of miR-128-3p through MAPK14 activation in the apoptosis of GC2 spermatocyte cell line following heat stress," *Andrology*, vol. 9, no. 2, pp. 665–672, 2021.
- [109] S. Dong, D. Liang, N. An et al., "The role of MAPK and FAS death receptor pathways in testicular germ cell apoptosis induced by lead," *Acta Biochim Biophys Sin (Shanghai)*, vol. 41, pp. 800–807, 2009.
- [110] P. Ranawat and M. P. Bansal, "Apoptosis induced by modulation in selenium status involves p38 MAPK and ROS: implications in spermatogenesis," *Molecular and Cellular Biochemistry*, vol. 330, no. 1–2, pp. 83–95, 2009.
- [111] P. Urriola-Muñoz, R. Lagos-Cabré, and R. D. Moreno, "A mechanism of male germ cell apoptosis induced by bisphenol-a and nonylphenol involving ADAM17 and p38 MAPK activation," *PLoS One*, vol. 9, no. 12, pp. 1–27, 2014.
- [112] P. A. Williams, H. E. Kobilnyk, E. A. McMillan, and T. I. Strohlic, "MAPKAP kinase 2-mediated phosphorylation of HspA1L protects male germ cells from heat stress-induced apoptosis," *Cell Stress & Chaperones*, vol. 24, no. 6, pp. 1127–1136, 2019.
- [113] C. H. Wong, D. D. Mruk, W. Y. Lui, and C. Y. Cheng, "Regulation of blood-testis barrier dynamics: an in vivo study," *Journal of Cell Science*, vol. 117, pp. 783–798, 2004.
- [114] H. Cai, Y. Ren, X. X. Li et al., "Scrotal heat stress causes a transient alteration in tight junctions and induction of tgf-β expression," *International Journal of Andrology*, vol. 34, 4 part 1, pp. 352–362, 2011.
- [115] J. Liu, L. Ren, J. Wei et al., "Fine particle matter disrupts the blood-testis barrier by activating TGF-β3/p38 MAPK pathway and decreasing testosterone secretion in rat," *Environmental Toxicology*, vol. 33, no. 7, pp. 711–719, 2018.
- [116] B. Liu, L. J. Shen, T. X. Zhao et al., "Automobile exhaust-derived PM_{2.5} induces blood-testis barrier damage through ROS-MAPK-Nrf2 pathway in Sertoli cells of rats," *Ecotoxicology and Environmental Safety*, vol. 189, article 110053, 2020.
- [117] S. Li, Q. Wang, H. Yu et al., "Polystyrene microplastics induce blood-testis barrier disruption regulated by the MAPK-Nrf2 signaling pathway in rats," *Environmental Science and Pollution Research*, vol. 28, pp. 47921–47931, 2021.
- [118] X. Jia, Y. Xu, W. Wu et al., "Aroclor1254 disrupts the blood-testis barrier by promoting endocytosis and degradation of junction proteins via p38 MAPK pathway," *Cell Death & Disease*, vol. 8, no. 5, article e2823, 2017.

- [119] V. L. de Liz Oliveira Cavalli, V. Lucia, D. Cattani et al., "Roundup disrupts male reproductive functions by triggering calcium-mediated cell death in rat testis and Sertoli cells," *Free Radical Biology & Medicine*, vol. 65, pp. 335–346, 2013.
- [120] Y. P. Jiang, R. J. Ye, J. M. Yang et al., "Protective effects of Saldroside on spermatogenesis in streptozotocin induced type-1 diabetic male mice by inhibiting oxidative stress mediated blood-testis barrier damage," *Chemico-Biological Interactions*, vol. 315, article 108869, 2020.
- [121] Y. P. Jiang, J. M. Yang, R. J. Ye et al., "Protective effects of betaine on diabetic induced disruption of the male mice blood-testis barrier by regulating oxidative stress-mediated p38 MAPK pathways," *Biomedicine & Pharmacotherapy*, vol. 120, p. 109474, 2019.
- [122] Y. Wang, F. Chen, L. Ye, B. Zirkin, and H. Chen, "Steroidogenesis in Leydig cells: effects of aging and environmental factors," *Reproduction*, vol. 154, pp. R111–R122, 2017.
- [123] P. Duan, M. Ha, X. Huang, P. Zhang, and C. Liu, "Intronic miR-140-5p contributes to beta-cypermethrin-mediated testosterone decline," *Science of The Total Environment*, vol. 806, Part 1, article 150517, 2022.
- [124] B. Banerjee, S. Chakraborty, P. Chakraborty, D. Ghosh, and K. Jana, "Protective effect of resveratrol on benzo(a)pyrene induced dysfunctions of steroidogenesis and steroidogenic acute regulatory gene expression in Leydig cells," *Frontiers in Endocrinology*, vol. 10, pp. 1–14, 2019.
- [125] L. Qiu, H. Wang, T. Dong et al., "Perfluorooctane sulfonate (PFOS) disrupts testosterone biosynthesis via CREB/CRTC2/StAR signaling pathway in Leydig cells," *Toxicology*, vol. 449, article 152663, 2021.
- [126] Y. Li, Y. Zhang, R. Feng et al., "Cadmium induces testosterone synthesis disorder by testicular cell damage via TLR4/MAPK/NF- κ B signaling pathway leading to reduced sexual behavior in piglets," *Ecotoxicology and Environmental Safety*, vol. 233, article 113345, 2022.
- [127] A. Han, L. Zou, X. Gan et al., "ROS generation and MAPKs activation contribute to the Ni-induced testosterone synthesis disturbance in rat Leydig cells," *Toxicology Letters*, vol. 290, pp. 36–45, 2018.
- [128] Y. Chen, J. Wang, X. Chen, D. Li, and X. Han, "Microcystin-leucine arginine mediates apoptosis and engulfment of Leydig cell by testicular macrophages resulting in reduced serum testosterone levels," *Aquatic Toxicology*, vol. 199, pp. 116–126, 2018.
- [129] A. M. Abdel-Aziz, S. M. Abozaid, R. K. Yousef, M. M. Mohammed, and H. M. Khalaf, "Fenofibrate ameliorates testicular damage in rats with streptozotocin-induced type 1 diabetes: role of HO-1 and p38 MAPK," *Pharmacological Reports*, vol. 72, pp. 1645–1656, 2020.
- [130] B. S. Pan, Y. K. Wang, M. S. Lai, Y. F. Mu, and B. M. Huang, "Cordycepin induced MA-10 mouse Leydig tumor cell apoptosis by regulating p38 MAPKs and PI3K/AKT signaling pathways," *Scientific Reports*, vol. 5, pp. 1–17, 2015.
- [131] N. Yildizbayrak and M. Erkan, "Therapeutic effect of curcumin on acrylamide-induced apoptosis mediated by MAPK signaling pathway in Leydig cells," *Journal of Biochemical and Molecular Toxicology*, vol. 33, pp. 1–8, 2019.
- [132] A. Simsek, E. Ozbek, Y. O. Ilbey, M. U. Cekmen, A. Somay, and A. I. Tasci, "Potential role of p38-mitogene-activated protein kinase and nuclear factor-kappa B expression in testicular dysfunction associated with varicocele: an experimental study," *Andrologia*, vol. 44, pp. 94–101, 2012.
- [133] J. L. Yuan, Y. T. Zhang, and Y. Wang, "Increased apoptosis of spermatogenic cells in cryptorchidism rat model and its correlation with transforming growth factor beta type II receptor," *Urology*, vol. 75, no. 4, pp. 992–998, 2010.
- [134] T. Lei, S. Moos, J. Klug et al., "Galectin-1 enhances TNF α -induced inflammatory responses in Sertoli cells through activation of MAPK signalling," *Scientific Reports*, vol. 8, pp. 1–15, 2018.
- [135] F. Aslani, H. C. Schuppe, V. A. Guazzone et al., "Targeting high mobility group box protein 1 ameliorates testicular inflammation in experimental autoimmune orchitis," *Human Reproduction*, vol. 30, no. 2, pp. 417–431, 2015.
- [136] M. A. E. Ahmed, A. A. E. Ahmed, and E. M. El Morsy, "Acetyl-11-keto- β -boswellic acid prevents testicular torsion/detorsion injury in rats by modulating 5-LOX/LTB4 and p38-MAPK/JNK/Bax/Caspase-3 pathways," *Life Sciences*, vol. 260, article 118472, 2020.
- [137] Y. Zhang, Y. Lv, Y. J. Liu et al., "Hyperbaric oxygen therapy in rats attenuates ischemia-reperfusion testicular injury through blockade of oxidative stress, suppression of inflammation, and reduction of nitric oxide formation," *Urology*, vol. 82, no. 2, pp. 489.e9–489.e15, 2013.
- [138] L. Minutoli, P. Antonuccio, F. Polito et al., "Involvement of mitogen-activated protein kinases (MAPKs) during testicular ischemia-reperfusion injury in nuclear factor- κ B knock-out mice," *Life Sciences*, vol. 81, no. 5, pp. 413–422, 2007.
- [139] N. Al-Kandari, F. Fadel, F. Al-Saleh, F. Khashab, and M. Al-Maghrebi, "The thioredoxin system is regulated by the ASK-1/JNK/p38/survivin pathway during germ cell apoptosis," *Molecules*, vol. 24, pp. 1–18, 2019.
- [140] M. P. Allen, D. A. Linseman, H. Udo et al., "Novel mechanism for gonadotropin-releasing hormone neuronal migration involving Gas6/ark signaling to p38 mitogen-activated protein kinase," *Molecular and Cellular Biology*, vol. 22, no. 2, pp. 599–613, 2002.
- [141] S. Salian-Mehta, M. Xu, and M. E. Wierman, "AXL and MET crosstalk to promote gonadotropin releasing hormone (GnRH) neuronal cell migration and survival," *Molecular and Cellular Endocrinology*, vol. 374, no. 1-2, pp. 92–100, 2013.
- [142] G. L. Kim, S. S. Dhillon, and D. D. Belsham, "Kisspeptin directly regulates neuropeptide Y synthesis and secretion via the ERK1/2 and p38 mitogen-activated protein kinase signaling pathways in NPY-secreting hypothalamic neurons," *Endocrinology*, vol. 151, pp. 5038–5047, 2010.
- [143] X. Li, J. Xiao, K. Li, and Y. Zhou, "MiR-199-3p modulates the onset of puberty in rodents probably by regulating the expression of *_Kiss1_* via the p38 MAPK pathway," *Molecular and Cellular Endocrinology*, vol. 518, article 110994, 2020.