

Research Article

Impaired Mitochondrial Bioenergetics Function in Pediatric Chronic Overlapping Pain Conditions with Functional Gastrointestinal Disorders

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Background. Fatigue is often the primary complaint of children with functional gastrointestinal disorders (FGDI) and other chronic overlapping pain disorders (COPC). The basis for this symptom remains unknown. We evaluated mitochondrial function in the white blood cells of these patients. Methods. This prospective Children's Wisconsin IRB approved study recruited subjects aging 10-18 years from pediatric neurogastroenterology clinics and healthy comparison subjects (HC). Environmental and oxidative stressors can damage the mitochondrial respiratory chain. The known low-grade inflammation in COPC could, therefore, impact the respiratory chain and theoretically account for the disabling fatigue so often voiced by patients. Mitochondrial energy generation can be easily measured in peripheral mononuclear cells (PMC) as a general marker by the Seahorse XF96 Extracellular Flux Analyzer. We measured 5 parameters of oxygen consumption using this methodology: basal respiration (BR), ATP linked oxygen consumption (ATP-LC), maximal oxygen consumption rate (max R), spare respiratory capacity (SRC), and extracellular acidification rate (ECAR), which reflect non-electron chain energy generation through glycolysis. In health, we expect high ATP linked respiration, high reserve capacity, low proton leak, and low non-mitochondrial respiration. In disease, the proton leak typically increases, ATP demand increases, and there is decreased reserve capacity with increased non-mitochondrial respiration. Findings and clinical data were compared to healthy control subjects using a Mann-Whitney test for skewed variables, Fisher's exact test for dichotomous variables, and regression tree for association with functional outcome (functional disability inventory, FDI). Results. 19 HC and 31 COPC showed no statistically significant difference in age. FGID, orthostatic intolerance, migraine, sleep disturbance, and chronic fatigue were present in the majority of COPC subjects. BR, ECAR, and ATP-LC rates were lower in the COPC group. The low BR and ATP-LC suggest that mitochondria are stressed with decreased ability to produce ATP. Tree analysis selected SRC as the best predictor of functional disability: patients with SRC >150 had a greater FDI (more disability) compared to patients with SRC <=150, p-value = 0.021. Conclusion. Subjects with COPC have reduced mitochondrial capacity to produce ATP. Predisposing genetic factors or reversible acquired changes may be responsible. A higher SRC best predicts disability. Since a higher SRC is typically associated with more mitochondrial reserve, the SRC may indicate an underutilized available energy supply related to inactivity, or a "brake" on mitochondrial function. Prospective longitudinal studies can likely discern whether these findings represent deconditioning, true mitochondrial dysfunction, or both.

1. Introduction

The term chronic overlapping pain conditions (COPC) emphasizes the cooccurrence of several common chronic pain syndromes without ascribing pathophysiologic assumptions. Previous names for these disorders include functional pain disorders, central sensitization disorders, and medically unexplained syndromes. Several disorders fit this classification, such as migraine headache, functional gastrointestinal disorders (FGID), fibromyalgia, vulvodynia, temporomandibular joint disorder, and interstitial cystitis/ bladder pain syndrome [1]. These disorders may also coexist with chronic fatigue and functional autonomic disorders like postural tachycardia syndrome (POTS). These ailments likely involve at least 10-20% of the pediatric population [2–4] and significantly impact daily activity in 40% of those affected [5]. In particular, FGIDs clearly cooccur with other painful disorders such as fibromyalgia, migraine, and chronic fatigue [6]. More importantly, the associated comorbid disorders and their symptoms, not the gastrointestinal issues, drive both the cost and the disability of FGIDs [7, 8].

In adults, fatigue is the third most common comorbidity of irritable bowel syndrome (IBS) [9]. Higher fatigue correlates with greater somatic symptoms (more severe IBS symptoms, larger number of comorbid medical disorders) and psychological comorbidities (anxiety and depression) [9]. Self-rating of health in IBS subjects relates to number of comorbidities and fatigue, not GI symptoms [10]. In turn, morning fatigue correlates with chronic pain [11], and people with chronic fatigue syndrome have lower pain thresholds [12]. Fatigue also generally correlates with psychiatric comorbidities without any implied cause-effect relationship [13].

Despite fatigue's prevalence in so many medical disorders, including COPCs, the pathophysiology of fatigue is unknown. Hypotheses include microinflammation [14, 15] and psychiatry comorbidities [16, 17]. One possibility is that fatigue reflects actual loss of bioenergetic sources, such as reduced ability to generate energy from mitochondria due to oxidative stress from low grade inflammation in the body, as it has been described in FGID [18]. Chacko et al. have developed a Bioenergetic Health index (BHI) based on mitochondrial bioenergetics that reflects the health of the circulating mitochondria in leukocytes or platelets, acting as early instruments to detect metabolic stress in tissues, and therefore, also in the mitochondria [19]. Chacko et al. describe metabolic stress from chronic illness as the combined damage of reactive oxygen species and systemic inflammation on the mitochondria of leukocytes and platelets [19].

In health, based on the BHI, we expect to see high ATP linked respiration, high reserve capacity, low proton leak, and low non-mitochondrial respiration, while, in disease, the proton leak increases, ATP demand increases, and there is decreased reserve capacity with increased non-mitochondrial respiration [19]. Utilizing the Seahorse XF96 Extracellular Flux Analyzer (North Billerica, MA) to assess mitochondrial bioenergetics is not new [20]. Divakaruni et al. wrote an extensive description of the interpretation of the data [21]. In depressed subjects, impaired mitochondrial bioenergetics including routine and uncoupled respiration, spared respiratory capacity (SRC), coupling efficiency, and ATP turnover-related respiration correlated with depressive symptom severity, loss of energy, difficulty concentrating, and fatigue [22]. Extensive work has also been done in cancer to suggest that these findings truly reflect the state of the mitochondria [23, 24]. These manifestations clearly recapitulate the severe fatigue and "brain fog" seen in 90% of pediatric FGID patients [25]. The aim of this study was to determine if, similar to the findings in depression [22], mitochondrial bioenergetics in children with COPC are reduced.

2. Methods

This prospective Children's Hospital of Wisconsin IRB approved study recruited subjects between June 2015 and May of 2018 in children aged 10-18 years from the pediatric neurogastroenterology and autonomic disorders clinic. Blood was obtained from youth with COPC and in carefully screened healthy comparison group (HC) with no functional disorder or chronic medical condition of any type. The comparison group was enrolled based on advertisement without aiming at matching for age or gender. The Oxygen Consumption Rate (OCR) and Extracellular Acidification Rate (ECAR) measurements were obtained utilizing the Seahorse XF96 Extracellular Flux Analyzer (North Billerica, MA). Peripheral Blood Mononuclear Cells (PMBCs) were seeded in a PS V7 cell culture plate at a density of 3.25×10^5 cells per well in a assay media supplemented with 1 mM pyruvate and 80 uL of RPMI, centrifuged for 10 min, and then placed in a non-CO₂ incubator for 1 hr @37C prior to analysis to avoid any temperature variances on the plate that could affect experimental results, followed by prompt analyzed using the Seahorse XF96 Extracellular Flux Analyzer. Oligomycin (1 ug/ml), carbonyl cyanide-4-phenyl-hydrazone (FCCP) (1 uM), and a cocktail of 1 uM Antimycin A and 1 uM rotenone to block complex 3 and 1, respectively, were added to properly determine mitochondrial and nonmitochondrial function. We measured OCR-linked basal respiration (BR), ATP linked oxygen consumption rate (ATP-LC), maximal respiration oxygen consumption rate (max R), non-mitochondrial linked oxygen consumption rate, and spare respiratory capacity (SRC) (Figure 1).

We collected the following data from a systematic chart review: (1) demographics including age and sex and medical history; (2) comorbid symptoms including symptoms of chronic fatigue (>6 months), dizziness, syncope >3 times/ life, GI symptoms with functional GI diagnoses based on fulfillment of Rome IV criteria, migraine headaches as per the 2013 International Classification of Headaches Disorders [26] and fibromyalgia assessed by tender-point score [27], pelvic pain, bladder pain syndrome, chronic regional pain syndrome (CRPS), and temporomandibular joint disorder (TMJ). We also recorded any history of posttraumatic stress disorder (PTSD) and panic disorder. To assess the clinical state of the subjects, we utilized a functional questionnaire, the Functional Disability Inventory (FDI) [28], thought to be an excellent measure of overall clinical status and a better



FIGURE 1: Schematic description of the Seahorse analysis. FCCP = carbonyl cyanide-4-phenyl-hydrazone; OCR = oxygen consumption rate.

assessment than simple pain reports. The FDI measures daily functions on a 5-point Likert scale such as ambulation, recreation, capacity for social interaction, performing chores, going to school, sleep, and eating. Validated for children with chronic abdominal pain, it shows excellent test-retest reliability within 2 weeks, and moderate at 3 months. It correlates well with school-related disabilities, and both the child-report and the parent-report scores correlate with abdominal pain measures and other somatic symptoms [29]. The FDI was only obtained in the COPC group, not in the HC.

Continuous variables are reported as median and interquartile range (IQR) and categorical variables as n (%). To compare differences between groups, a Mann–Whitney test was used for continuous, and a Fisher's exact test for categorical variables. Spearman correlation test was used to examine the correlations among the mitochondrial bioenergetic variables, and their correlations with FDI as well. To investigate the relationship of age, gender, BR, ATP-LC, max R, SRC, and ECAR with functional status as reflected in the FDI, a regression tree, optimized by least absolute deviation with 10% leave out samples for cross validation, was performed using Salford systems CART. Significance was attributed with a two-sided p value <0.05. All other data analyses were performed using SAS 9.4.

3. Results

Of 74 patients enrolled, 24 were excluded due to lack of mitochondrial bioenergetic data (assay malfunctioning or too few cells to run assay). To perform the Seahorse analysis, we need a total of 3.25×105 PBMCs when performing the final resuspension. We found many samples with an insufficient concentration of PBMCs, which could have been due to loss of cells through sample handling and isolation of PBMCs. This is similar to a standard clinical assay returning a result of "quantity not sufficient".

Of the remaining 50 subjects, 19 were HC and 31 had a COPC. Neither age (HC: 15.1 (14.3–16.1) yrs; COPC 16.5

(14.6–17.7) yrs; *p* = 0.06) nor gender differed (HC 16 (84%) females; COPC 22 (71%) females; p = 0.33) between the groups. The clinical characteristics of the COPC group included an FGID in 22 (71%) with chronic idiopathic nausea in 14 (48%), cyclic vomiting syndrome (CVS) 5 (17%), irritable bowel syndrome (IBS) (not subclassified, but the vast majority are IBS-C) 17 (57%), functional abdominal pain NOS 13 (45%), and functional dyspepsia 11 (38%). Extragastrointestinal comorbidities included orthostatic intolerance in 26 (90%), migraine in 21 (72%), chronic fatigue in 13 (43%), syncope in 11 (38%), TMJ in 6 (21%), fibromyalgia in 6 (21%), complex regional pain in 3 (10%), interstitial cystitis/painful bladder syndrome in 2 (7%), and myofascial pelvic pain in 1 (3%). In relation to the psychiatric comorbidities, 4 (14%) reported a panic disorder, and 6 (21%) reported PTSD.

Non-mitochondrial linked oxygen consumption rate, BR, ATP-LC, and ECAR were significantly lower in the COPC group (Table 1 and Figure 2) compared to HC. SRC statistically was not different in the 2 groups (p > 0.28).

In both COPC and HC groups, BR correlated with SRC, ATP-LC, and MaxR (Table 2). We investigated whether the severity of the patient's clinical syndrome in the COPC group, as reflected in our best measure, the FDI, might correlate with each of the respiratory impairments. No correlation emerged between FDI and BR (r=0.002, p > 0.99), MaxR (r=0.19, p = 0.34), and ATP production (p=ATP-LC (r=-0.01, p=0.95), nor SRC (r=0.22, p=0.25) and ECAR (r=0.22, p=0.26). Interestingly, ECAR showed a trend for correlation with SRC (r=0.46, p=0.06) and ATP-LC (r=0.41, p=0.08) in the HC group, while other correlations were stronger in the COPC group (Table 2).

Within the COPC group, females had a higher SRC than males (p = 0.048).

In a tree analysis, a possible threshold of 150 for SRC was found. Patients who had SRC >150 has a higher FDI than others with SRC <=150, p-value = 0.021.

4. Discussion

The main finding in this study is that youth with COPC have a reduced mitochondrial bioenergetic capacity compared to healthy control subjects, carrying important clinical and pathophysiologic implications. Clinically, these children and adolescents are often considered as having school avoidance and psychiatric issues and sometimes treated as having no physiologic basis for their complaints. We do not know if this finding demonstrates a basis for the frequent complaint of severe fatigue associated with these disorders, or whether it might simply reflect deconditioning, but it certainly merits further investigation.

Since we did not know if any putative mitochondrial defect might be inherited or acquired, to further understand the role of mitochondria in COPC, we examined the function of the mitochondria rather than looking for a structural mutation by looking at mitochondrial bioenergetics utilizing PMBC. Chacko et al. [19] reported a Bioenergetic Health Index (BHI). A healthy individual should

TABLE 1: Comparison in mitochondrial bioenergetic between healthy controls and youth with chronic overlapping pain conditions.

(pmol/min)	HC, median (IQR) $n = 19$	COPC, median (IQR) $n = 31$	<i>p</i> -value
Non mitochondrial respiration	13.0 (7.9, 18.0)	9.3 (5.3, 11.7)	0.016
BR	52.3 (44.2, 69.7)	43.4 (27.1, 51.8)	0.010
SRC	117.8 (100.3, 209.7)	118.33 (69.4, 161.1)	0.281
ATP production	49.9 (43.2, 65.4)	42.7 (27.3, 50.2)	0.013
ECAR	17.1 (14.7, 24.0)	16.2 (12.1, 19.5)	0.040

ECAR = extracellular acidification rate; OCR = oxygen consumption rate; BR = basal respiratory rate; SRC = spare respiratory capacity. All mitochondrial bioenergetics measurements are in pmol/min; HC = healthy controls; COPC = chronic overlapping pain conditions; IQR = interquartile range.



FIGURE 2: Oxygen consumption rate in non-mitochondrial respiration, basal respiration rate, ATP production (ATP-LC), and extracellular acidification rate (ECAR) in healthy controls and COPC.

TABLE 2: Correlation across bioenergetic variables in COPC and HC.

Correlation	COPC	HC
BR with SRC	R = 0.76, p < 0.0001	R = 0.89, p < 0.0001
BR with ATP-LC	R = 0.98, p < 0.0001	R = 0.99, p < 0.0001
BR with maximal	R = 0.83, p < 0.0001	R = 0.93, p < 0.0001
BR with ECAR	R = 0.69, p < 0.0001	R = 0.46, p = 0.06
ECAR with SRC	R = 0.72, p < 0.0001	R = 0.51, p = 0.025
ECAR with ATP-LC	R = 0.68, p < 0.0001	$R = 0.41, \ p = 0.08$

have high ATP production (ATP-LC), high SRC, low nonmitochondrial OCR, and low proton leak. Under increasing oxidative stress, these parameters reverse [19]. Blood leukocytes, as well as platelets, are exposed to circulating factors associated with metabolic stress. Since these cells contain mitochondria, they may provide a functional marker in diseases where inflammation or metabolic stress might play a significant role, such as the COPCs migraine, irritable bowel syndrome, and fibromyalgia [30–32].

It is unclear whether the present finding of reduced bioenergetic capacity reflects a consequence of a predisposition for a COPC or simply deconditioning. A predisposition might reflect genetic or epigenetic changes. In contrast, a consequence could reflect a change in mitochondrial control or modulation rather than any genetic process. For example, it is known that cardiac myocyte mitochondrial function is strongly modulated by the vagus [33]. Whether a similar modulation might impact peripheral blood monocytes is unknown. Since the majority of subjects with COPC demonstrate low vagal modulation [34–36], we believe an acquired (and, therefore, potentially reversible) mechanism may be more likely. This in turn might explain why mitochondrial studies focusing on genetic etiologies have been somewhat inconclusive. Further studies of bioenergetic capability once subjects with COPC improve clinically will help better define whether the observed deficits are reversible, and thus likely acquired.

While the most affected functions were basal respiration and ATP linked oxygen consumption, which depend on the respiratory chain itself, non-mitochondrial energy generation and extra-cellular acidification rates, neither of which depend on the respiratory chain, were also affected for reasons that are unclear (Figure 2). However, these findings favor an acquired defect over an inherited defect, which would have had a more specific impact. Interestingly, in contrast to the BHI index described by Chacko et al., our COPC subjects showed lower non-mitochondrial oxygen consumption, a marker of health [19]. It is not clear why the tree selected a higher spare respiratory capacity as the best predictor of higher functional disability. A higher spare respiratory capacity is generally thought to be healthy [19]. However, in the COPC population, it is possible that higher spare capacity results from less actual energy generation (lower BR in the COPC group) or, perhaps more likely, that the mitochondria in the cells in COPC are not as affected as in patients with dysfunctional bioenergetics such as in diabetes or cardiovascular diseases [19].

Abnormal mitochondrial bioenergetic function also occurs in some psychiatric disorders. Findings nearly identical to those in this study were present in patients with depression. The severity of bioenergetic dysfunction correlated with clinical fatigue [37]. In veterans with PTSD, mtDNA copy numbers are decreased, unrelated to any associated Major Depressive Disorder [38]. Supporting the hypothesis that fatigue most closely reflects the finding of abnormal mitochondrial bioenergetics, higher ventricular cerebrospinal fluid lactate measured by proton magnetic resonance spectroscopy imaging in chronic fatigue syndrome subjects correlated directly with their fatigue level [39]. Future work will need to include a direct assessment of fatigue through existing validated questionnaires in our population to determine if a similar relationship holds.

The exploratory nature of this study directly leads to many of its limitations. Not assessing lactate in our subjects reduced the richness of the interpretation of the anaerobic glycolysis findings. Second, since the FDI did not correlate with the mitochondrial bioenergetics, a measure of fatigue may more accurately reflect mitochondrial bioenergetics. Given that abnormal mitochondrial bioenergetics are present in psychiatric conditions, future studies need to include structured evaluation of PTSD, depression, and anxiety. In this study, we have only had formal records for PTSD. Furthermore, the finding of abnormal bioenergetics may not be specific to COPC. Future studies will require a "sick" comparison group with another chronic disorder of a different type (e.g., Crohn's or other inflammatory disorder) to determine the specificity of these findings to COPC vs. just "being sick" and will benefit from a measure of deconditioning such as maximal oxygen uptake.

In conclusion, subjects with COPC have decreased mitochondrial bioenergetics when compared to healthy age matched controls. The mechanism of the bioenergetic impairment could involve predisposing genetic factors or acquired changes that reverse as disease improves. Prospective longitudinal studies compared to other nonpainful chronic disorders will distinguish these possibilities.

Abbreviations

- BR: Basal respiration
- COPC: Chronic overlapping pain conditions
- ECAR: Extracellular acidification rate
- FDI: Functional disability inventory
- FGID: Functional gastrointestinal disorders
- HC: Healthy comparison group
- IBS: Irritable bowel syndrome
- OCR: Oxygen consumption rate
- PMBC: Peripheral blood mononuclear cells
- POTS: Postural tachycardia syndrome
- PTSD: Post traumatic stress disorder
- SRC: Spared respiratory capacity.

Data Availability

The deidentified data may be available by request after obtaining the appropriate approval of our IRB.

Disclosure

The sponsor had no role in the study design, data collection or analysis, or decision to publish.

Conflicts of Interest

Thomas and Gisela Chelimsky are co-owners of PainSTakers LLC (no income yet).

Authors' Contributions

The first draft of the manuscript was written by Dr. Gisela Chelimsky.

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References

- C. D. Veasley, D. Clauw, T. Cowley et al., "Impact of chronic overlapping pain conditions on public healthy and the urgent need for safe and effective treatment," in 2015 Analysis and Policy Recommendations, C. P. R. Alliance, Ed., The TMJ Association, Milwaukee, WI, USA, 2015.
- [2] J. Apley and N. Naish, "Recurrent abdominal pains: a field survey of 1,000 school children," *Archives of Disease in Childhood*, vol. 33, no. 168, pp. 165–170, 1958.

- [3] C. Boey, S. Yap, and K.-L. Goh, "The prevalence of recurrent abdominal pain in 11- to 16-year-old Malaysian school children," *Journal of Paediatrics and Child Health*, vol. 36, no. 2, pp. 114–116, 2000.
- [4] J. S. Hyams, P. Davis, F. A. Sylvester, D. K. Zeiter, C. J. Justinich, and T. Lerer, "Dyspepsia in children and adolescents: a prospective study," *Journal of Pediatric Gastroenterology and Nutrition*, vol. 30, no. 4, pp. 413–418, 2000.
- [5] M. Saps, P. Adams, S. Bonilla, A. Chogle, and D. Nichols-Vinueza, "Parental report of abdominal pain and abdominal pain-related functional gastrointestinal disorders from a community survey," *Journal of Pediatric Gastroenterology & Nutrition*, vol. 55, no. 6, pp. 707–710, 2012.
- [6] G. Chelimsky, S. Safder, and T. Chelimsky, "FGIDs in children are associated with many nonpsychiatric comorbidities," *Journal of Pediatric Gastroenterology & Nutrition*, vol. 54, no. 5, pp. 690-691, 2012.
- [7] P. A. Johansson, P. G. Farup, A. Bracco, and P. O. Vandvik, "How does comorbidity affect cost of health care in patients with irritable bowel syndrome? a cohort study in general practice," *BMC Gastroenterology*, vol. 10, no. 1, p. 31, 2010.
- [8] V. L. Michalsen, P. O. Vandvik, and P. G. Farup, "Predictors of health-related quality of life in patients with irritable bowel syndrome. A cross-sectional study in Norway," *Health and Quality of Life Outcomes*, vol. 13, no. 1, p. 113, 2015.
- [9] J. M. Lackner, G. D. Gudleski, J. DiMuro, L. Keefer, and D. M. Brenner, "Psychosocial predictors of self-reported fatigue in patients with moderate to severe irritable bowel syndrome," *Behaviour Research and Therapy*, vol. 51, no. 6, pp. 323–331, 2013.
- [10] J. M. Lackner, G. D. Gudleski, E. R. Thakur, T. J. Stewart, G. J. Iacobucci, and B. M. Spiegel, "The impact of physical complaints, social environment, and psychological functioning on IBS patients' health perceptions: looking beyond GI symptom severity," *American Journal of Gastroenterology*, vol. 109, no. 2, pp. 224–233, 2014.
- [11] R. M. Ghandour, M. D. Overpeck, Z. J. Huang, M. D. Kogan, and P. C. Scheidt, "Headache, stomachache, backache, and morning fatigue among adolescent girls in the United States," *Archives of Pediatrics & Adolescent Medicine*, vol. 158, no. 8, pp. 797–803, 2004.
- [12] A. Winger, G. Kvarstein, V. B. Wyller et al., "Pain and pressure pain thresholds in adolescents with chronic fatigue syndrome and healthy controls: a cross-sectional study," *BMJ Open*, vol. 4, no. 9, Article ID e005920, 2014.
- [13] T. Pawlikowska, T. Chalder, S. R. Hirsch, P. Wallace, D. J. M. Wright, and S. C. Wessely, "Population based study of fatigue and psychological distress," *BMJ*, vol. 308, no. 6931, pp. 763–766, 1994.
- [14] M. C. Serra, A. S. Ryan, H. K. Ortmeyer, O. Addison, and A. P. Goldberg, "Resistance training reduces inflammation and fatigue and improves physical function in older breast cancer survivors," *Menopause*, vol. 25, no. 2, pp. 211–216, 2018.
- [15] A. V. Sardeli, C. M. Tomeleri, E. S. Cyrino, B. Fernhall, C. R. Cavaglieri, and M. P. T. Chacon-Mikahil, "Effect of resistance training on inflammatory markers of older adults: a meta-analysis," *Experimental Gerontology*, vol. 111, pp. 188– 196, 2018.
- [16] E. C. Corfield, N. G. Martin, and D. R. Nyholt, "Co-occurrence and symptomatology of fatigue and depression," *Comprehensive Psychiatry*, vol. 71, pp. 1–10, 2016.
- [17] C. M. Galvez-Sánchez, C. I. Montoro, S. Duschek, and G. A. Reyes del Paso, "Depression and trait-anxiety mediate the influence of clinical pain on health-related quality of life in

fibromyalgia," Journal of Affective Disorders, vol. 265, pp. 486-495, 2020.

- [18] L. Öhman and M. Simrén, "Pathogenesis of IBS: role of inflammation, immunity and neuroimmune interactions," *Nature Reviews Gastroenterology & Hepatology*, vol. 7, no. 3, pp. 163–173, 2010.
- [19] B. K. Chacko, P. A. Kramer, S. Ravi et al., "The bioenergetic health index: a new concept in mitochondrial translational research," *Clinical Science*, vol. 127, no. 6, pp. 367–373, 2014.
- [20] H. V. Vangapandu and V. Gandhi, "Extracellular flux assays to determine oxidative phosphorylation and glycolysis in chronic lymphocytic leukemia cells," *Methods in Molecular Biology*, vol. 1881, pp. 121–128, 2019.
- [21] A. S. Divakaruni, A. Paradyse, D. A. Ferrick, A. N. Murphy, and M. Jastroch, "Analysis and interpretation of microplatebased oxygen consumption and pH data," *Methods in Enzymology*, vol. 547, pp. 309–354, 2014.
- [22] A. Karabatsiakis, C. Böck, J. Salinas-Manrique et al., "Mitochondrial respiration in peripheral blood mononuclear cells correlates with depressive subsymptoms and severity of major depression," *Translational Psychiatry*, vol. 4, no. 6, p. e397, 2014.
- [23] R. Sánchez-Alvarez, E. M. De Francesco, M. Fiorillo, F. Sotgia, and M. P. Lisanti, "Mitochondrial fission factor (MFF) inhibits mitochondrial metabolism and reduces breast cancer stem cell (CSC) activity," *Frontiers in Oncology*, vol. 10, p. 1776, 2020.
- [24] S. Taha-Mehlitz, G. Bianco, M. Coto-Llerena et al., "Adenylosuccinate lyase is oncogenic in colorectal cancer by causing mitochondrial dysfunction and independent activation of NRF2 and mTOR-MYC-axis," *Theranostics*, vol. 11, no. 9, pp. 4011–4029, 2021.
- [25] K. Kovacic, T. C. Chelimsky, M. R. Sood, P. Simpson, M. Nugent, and G. Chelimsky, "Joint hypermobility: a common association with complex functional gastrointestinal disorders," *The Journal of Pediatrics*, vol. 165, no. 5, pp. 973–978, 2014.
- [26] Headache Classification Committee of the International Headache Society (IHS), "The international classification of headache disorders, 3rd edition (beta version)," *Cephalalgia*, vol. 33, no. 9, pp. 629–808, 2013.
- [27] F. Wolfe, H. A. Smythe, M. B. Yunus et al., "The American college of rheumatology 1990 criteria for the classification of fibromyalgia. Report of the multicenter criteria committee," *Arthritis and rheumatism*, vol. 33, no. 2, pp. 160–172, 1990.
- [28] L. S. Walker and J. W. Greene, "The functional disability inventory: measuring a neglected dimension of child health status," *Journal of Pediatric Psychology*, vol. 16, no. 1, pp. 39–58, 1991.
- [29] R. L. Claar and L. S. Walker, "Functional assessment of pediatric pain patients: psychometric properties of the functional disability inventory," *Pain*, vol. 121, no. 1-2, pp. 77–84, 2006.
- [30] E. Sinagra, G. C. Morreale, G. Mohammadian et al., "New therapeutic perspectives in irritable bowel syndrome: targeting low-grade inflammation, immuno-neuroendocrine axis, motility, secretion and beyond," *World Journal of Gastroenterology*, vol. 23, no. 36, pp. 6593–6627, 2017.
- [31] M. Lukacs, J. Tajti, F. Fulop, J. Toldi, L. Edvinsson, and L. Vecsei, "Migraine, neurogenic inflammation, drug development - pharmacochemical aspects," *Current Medicinal Chemistry*, vol. 24, no. 33, pp. 3649–3665, 2017.
- [32] I. Coskun Benlidayi, "Role of inflammation in the pathogenesis and treatment of fibromyalgia," *Rheumatology International*, vol. 39, no. 5, pp. 781–791, 2019.

- [33] X. He, M. Zhao, X. Bi et al., "Novel strategies and underlying protective mechanisms of modulation of vagal activity in cardiovascular diseases," *British Journal of Pharmacology*, vol. 172, no. 23, pp. 5489–5500, 2015.
- [34] P. Mork, J. Nilsson, H. Lorås, R. Riva, U. Lundberg, and R. Westgaard, "Heart rate variability in fibromyalgia patients and healthy controls during non-REM and REM sleep: a casecontrol study," *Scandinavian Journal of Rheumatology*, vol. 42, no. 6, pp. 505–508, 2013.
- [35] G. Chelimsky, S. Rausch, D. Bierer et al., "Cardiovagal modulation in pediatric functional gastrointestinal disorders," *Neuro-Gastroenterology and Motility*, vol. 31, no. 5, Article ID e13564, 2019.
- [36] J.-H. Kang, H.-S. Chen, S.-C. Chen, and F.-S. Jaw, "Disability in patients with chronic neck pain," *The Clinical Journal of Pain*, vol. 28, no. 9, pp. 797–803, 2012.
- [37] N. J. Klinedinst and W. T. Regenold, "A mitochondrial bioenergetic basis of depression," *Journal of Bioenergetics and Biomembranes*, vol. 47, no. 1-2, pp. 155–171, 2015.
- [38] F. S. Bersani, C. Morley, D. Lindqvist et al., "Mitochondrial DNA copy number is reduced in male combat veterans with PTSD," *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, vol. 64, pp. 10–17, 2016.
- [39] J. W. Murrough, X. Mao, K. A. Collins et al., "Increased ventricular lactate in chronic fatigue syndrome measured by 1H MRS imaging at 3.0 T. II: comparison with major depressive disorder," *NMR in Biomedicine*, vol. 23, no. 6, pp. 643–650, 2010.