Research Article

Essential Oils of Aromatic and Medicinal Plants as Botanical Biocide for Management of Coconut Eriophyid Mite (*Aceria guererronis* Keifer)

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The present study investigated the efficiency of essential oils extracted from different aromatic and medicinal plant sources on *Aceria guererronis* Keifer, one of the serious pests of coconut. The essential oils and the herbal extracts were prepared in two different formulations and were used both in laboratory and field conditions to assess the efficiency of the formulations against the coconut mite infestation. The field trial results showed that reduction in infestation intensity was found to vary between 73.44% and 44.50% at six different locations of trial farms with an average of 64.18% after four spells of treatment. The average number of live mites was higher in the third month old nuts both in the control as well as the treated nut samples. The laboratory experiments on the efficacy of botanical biocide showed that the time taken for dehydration and shriveling of body cells took only sixty seconds. The multilocational field trials revealed the overall efficiency of the biocide to significantly control the eriophyid mite in coconut crop in an ecofriendly and sustainable manner without adopting any chemical pesticide.

1. Introduction

The eriophyid mite (*Aceria guererronis* Keifer) is a microscopic organism that remains under the perianth of the coconut and has been one of the serious pests of coconut for the last three decades in major coconut growing countries [1–4]. These tiny mites aggregate in colonies in the inner and outer bracts and under the tepals and feed on the meristematic tissues on the nut surface. Due to mite congregation and feeding the meristematic tissue beneath the perianth becomes chlorotic and then cracks. *A. guererronis* infestation leads to surface scars, reduced fruit growth, and premature fruit fall [5]. The reported yield loss caused by *A. guererronis* was found to be 34% in India [6]. In the past few years, several studies have focused on the potential use of essential oil formulations in biological control of various insect pest and diseases. The essential oils which get more rapidly degraded into the environment than chemical compounds have been studied for their action against various insect pest of stored products [7, 8]. Recent studies have demonstrated the antilarval and antifeeding [9–11], delayed adult emergence and egg mortality [12], arrestance and repellent actions [13] of essential oils. The present study has been aimed to use these natural derivatives as an alternative ecofriendly means to control the eriophyid mite in coconut crop.
oils and extracts from Banatulsi (Hyptis suaveolens), Tulsi (Ocimum sanctum), Patcholi (Pogostemon cablin), Citronella (Cymbopogon winterianus), Kalmegh (Andrographis paniculata), Citrus (Citrus limon), and Soapnut (Sapindus marginatus) were used after analysis of their active principles against insect pest and diseases.

Field trials were conducted at five farms of coconut growing states of India (Karnataka, Kerala, West Bengal, Orissa and Andhra Pradesh) in replicated randomized block design. The botanical formulation in spray and herbal organic manure were applied to infested coconut plants at quarterly intervals by preparing rings to study its effectiveness against the mite. The botanical acaricide was applied on newly pollinated flowering bunches in two ways: one method consisted of spraying at the rate of 25 mL per flower using oil combination diluted in 250 mL water. Protocol two included application of 3 kg herbal organic manure per plant. The formulation was prepared by mixing coir pith and cow dung in a ratio of 1:2 along with 1% (w/v) each of Andrographis paniculata and Hyptis suaveolens kept in a pit for three months for composting. The efficacy of the botanical biocide was based on the infestation intensity of A. guererronis from different farms.

To analyze competence of the botanical biocide, laboratory experiments were also conducted. The number of live mites present in nuts of different ages was studied both for control and treated plants. Nut samples from five randomly selected bunches per replication from one to six month old were collected from each of the trial farms three months after biocide application. The nuts were then subjected to counting of mites by a method according to Lawson-Balagbo et al. [14]. The mite population inside the bracts and on the nut surface below the perianth were extracted using 50 mL of 1% cetrimide solution, and the mite suspension after collection in a beaker (100 mL) was agitated by blowing air into it for about 15–20 seconds using a pipette for uniform mixing. Immediately, 10 mL of the subsample of the suspension was transferred to De Grisse counting dish, and mites were counted (A). A second sample counting determined left out mites on bracts and nut surfaces (B). A third count was done by direct observation of the bracts and the surface below perianth using stereoscopic microscope to count any left out mites (C). Thus, the total number of mite (N) present per nut was calculated by the formulae: N = 5(A + B) + C. The samples from nuts of different ages were subjected to statistical analysis. The inflorescence was also examined for presence of eriophyid mite.

Essential oils of Hyptis suaveolens, Ocimum sanctum, Cymbopogon winterianus, Pogostemon cablin and Citrus limon, and botanical biocide formulation were placed in direct contact with eriophyid mite (A. guererronis). Ten live mites from each infested nut from different farms were transferred to a cavity slide under microscope, and 20 μL of the oils or formulation were applied to the slide. Time taken for dehydration and shriveling of body cells was recorded and analyzed statistically through F-test (Data not shown).

3. Results and Discussion

The botanical biocide treatments against the infestation intensity of the eriophyid mite produced interesting effects (Table 1), lowering the average infestation intensity to 69.37% after six treatments. The pretreatment eriophyid intensity averaged 80.8% (range 58.2–100%) at six trial farms. After first treatment count the botanical biocide exhibited significant effect in reducing infestation by 23.25%. The percentage reduction was essentially similar at the test sites (Orissa (35.16%), Kerala (29.96%), Phillips farm (23.28%), DSP farm (22.12%), West Bengal (15.16%), and Karnataka (13.83%) after first treatment. After second treatment, intensity of mite in the nuts was observed to be reduced at all sites by 33.24%. However, in trial farms of Karnataka and Kerala, mite infestations were higher compared to levels after previous treatments. This may be attributed to the secondary infestation of pathogens. The third treatment also increased reduction by 59.33% with a similar trend followed in the fourth treatment. Results were not significantly different at subsequent treatments.

The reduction of infestation intensity in Orissa and DSP farm is significantly higher than the other trial farms (Orissa (73.44%), DSP (71.5%), Phillips farm (67.51%), West Bengal (63.94%), and Karnataka (44.5%)). The statistical analysis of fifth and sixth round of botanical biocide applications showed results at par with the fourth application, thus inferring that the application dosage of the biocide for mite control in coconut can be standardized as four applications per year for most farming locations.

The laboratory experiment for number of live mites present in nuts of different age revealed that inflorescences of coconut carried no mite. This supported the result of Moore and Alexander [15] which reports that mites do not infest the meristematic zones of unfertilized coconut flowers. The presence of mites on the nuts collected from one to six-month-old bunches after fertilization supported the findings.
### Table 1: Efficacy of botanical biocide formulations against infestation intensity of *Aceria guerreronis*.

<table>
<thead>
<tr>
<th>Treatments</th>
<th>DSP farm Andhra Pradesh</th>
<th>Phillips farm Andhra Pradesh</th>
<th>Karnataka</th>
<th>Kerala</th>
<th>Orissa</th>
<th>West Bengal</th>
<th>Average infestation reduction (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Infestation (%)</td>
<td>Infestation reduction (%)</td>
<td>Infestation (%)</td>
<td>Infestation reduction (%)</td>
<td>Infestation (%)</td>
<td>Infestation reduction (%)</td>
<td>Infestation (%)</td>
</tr>
<tr>
<td>Before treatment</td>
<td>80.70 (63.94)</td>
<td>76.96 (61.34)</td>
<td>58.20 (49.72)</td>
<td>99.52 (86.03)</td>
<td>99.98 (89.19)</td>
<td>69.46 (56.48)</td>
<td>80.80</td>
</tr>
<tr>
<td>T1</td>
<td>62.85 (52.48)</td>
<td>22.12 (28.04)</td>
<td>59.04 (50.18)</td>
<td>23.28 (28.86)</td>
<td>50.15 (45.11)</td>
<td>13.83 (21.72)</td>
<td>69.70 (56.60)</td>
</tr>
<tr>
<td>T2</td>
<td>38.85 (38.59)</td>
<td>51.86 (46.09)</td>
<td>51.87 (34.82)</td>
<td>32.60 (29.77)</td>
<td>70.90 (57.35)</td>
<td>29.27* (32.77)</td>
<td>73.25 (58.89)</td>
</tr>
<tr>
<td>T3</td>
<td>28.00 (31.95)</td>
<td>65.30 (53.91)</td>
<td>35.00 (36.27)</td>
<td>54.52 (47.58)</td>
<td>20.00 (26.56)</td>
<td>65.63 (54.09)</td>
<td>NA</td>
</tr>
<tr>
<td>T4</td>
<td>23.00 (28.66)</td>
<td>71.50 (57.67)</td>
<td>25.00 (30)</td>
<td>67.51 (55.18)</td>
<td>13.70 (21.72)</td>
<td>44.50 (41.84)</td>
<td>NA</td>
</tr>
<tr>
<td>T5</td>
<td>38.85 (38.59)</td>
<td>51.86 (46.09)</td>
<td>NA</td>
<td>NA</td>
<td>5.63 (13.69)</td>
<td>52.57 (46.49)</td>
<td>38.85 (38.59)</td>
</tr>
<tr>
<td>T6</td>
<td>34.69 (36.09)</td>
<td>57.01 (49.02)</td>
<td>NA</td>
<td>NA</td>
<td>19.64 (26.28)</td>
<td>66.25 (54.51)</td>
<td>28.09 (32.01)</td>
</tr>
<tr>
<td>SE ± (m)</td>
<td>0.915</td>
<td>4.88</td>
<td>2.47</td>
<td>2.87</td>
<td>1.37</td>
<td>1.78</td>
<td></td>
</tr>
<tr>
<td>CD (P = .05)</td>
<td>1.275</td>
<td>10.40</td>
<td>4.79</td>
<td>5.57</td>
<td>2.65</td>
<td>3.45</td>
<td></td>
</tr>
</tbody>
</table>

*The percentage damage increased as compared to the previous treatments because of the secondary infestation by pathogens.
NA: materials sent but data not available because of local problems.
of [16] where they report that after fertilization, the coconut fruits of all ages are susceptible to mite attack but in general peak mite populations occur in 3–7 months old coconuts. The highest population density of the mites was observed on the three-month-old nuts (Figure 1). Similar results on occurrence of highest number of mites on nuts of third month were reported by [17, 18]. The laboratory studies revealed subsequent decline in number of mites with increase in age of nuts however, lowest number of mite per nut was recorded from nuts of six months. Similar report was furnished by [19] wherein mite population showed rapid increase up to an age of three months followed by steep reduction. The minimum population was being observed on nuts of age seven to eight months. However, the incidence of mites on nine to ten-month-old nuts was reported from several areas [20, 21]. Such di

differences in mite population as per age of the nuts may be attributed to differences in ecological factors, age, and varieties of palms under study.

The F-test for time of dehydration of cells of E. mite showed that the treatments were significantly different from each other. The botanical biocide formulation showed best response (60 sec) in order to cause dehydration and shriveling of cells thereby leading to death of mite (Figure 2). However, essential oil of Hyptis suaveolens (80 sec) showed a par results with Cymbopogon winterianus (92 sec). The time required by the essential oils of Citrus limon (132 sec) and Ocimum sanctum (135 sec) were significantly not different from each other. The essential oil of Pogostemon cablin (227 sec) took maximum time to inactivate the mite. With the treatment of biocide, the eggs were also found to degenerate along with the mite (Figure 2(b)).

4. Conclusion

From the significant results of the study, it can be concluded that essential oils and their constituents have varying degrees of pest controlling activities. The present study shows the possibilities of encouraging the use of botanical biocides as future pest management strategies of coconut mite.

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References


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