Review Article

Targeting the Insulin-Like Growth Factor 1 Receptor in Ewing’s Sarcoma: Reality and Expectations

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Ewing’s sarcoma family of tumours comprises a group of very aggressive diseases that are potentially curable with multimodality treatment. Despite the undoubted success of current treatment, approximately 30% of patients will relapse and ultimately die of disease. The insulin-like growth factor 1 receptor (IGF-1R) has been implicated in the genesis, growth, proliferation, and the development of metastatic disease in Ewing’s sarcoma. In addition, IGF1-R has been validated, both in vitro and in vivo, as a potential therapeutic target in Ewing’s sarcoma. Phase I studies of IGF-1R monoclonal antibodies reported several radiological and clinical responses in Ewing’s sarcoma patients, and initial reports of several Phase II studies suggest that about a fourth of the patients would benefit from IGF-1R monoclonal antibodies as single therapy, with approximately 10% of patients achieving objective responses. Furthermore, these therapies are well tolerated, and thus far severe toxicity has been rare. Other studies assessing IGF-1R monoclonal antibodies in combination with traditional cytotoxics or other targeted therapies are expected. Despite, the initial promising results, not all patients benefit from IGF-1R inhibition, and consequently, there is an urgent need for the identification of predictive markers of response.

1. Introduction

Sarcomas represent a diverse group of tumours that arise from connective tissue, and account for 12% of paediatric malignancies and approximately 1% of all adult tumors [1–3]. Significant progress has been made in the classification, staging, and multimodal treatment of these heterogeneous conditions including: surgical advances in functional preservation, the use of radiotherapy as adjunct to other modalities, and the identification of active systemic therapies for certain sarcoma subtypes [4–6].

Ewing’s sarcoma family of tumours (ESFTs) comprise an aggressive group of sarcomas which can arise in soft tissue or bone and include classical Ewing’s sarcoma, Askin tumour, and primitive neuroectodermal tumour (PNET) [7]. These tumours are most commonly diagnosed in adolescence [8], although increasing numbers are being identified in adults [9], have a slight male predominance, and are more common in Caucasian populations [8, 9]. Approximately a quarter of patients present with metastatic disease at the time of diagnosis [10], and the most common metastatic sites are lung (50%), bone (25%), and bone marrow (20%). Over the last 30 years, the prognosis for patients with localised disease has improved dramatically. The introduction of combination chemotherapy has improved survival from 20%–30% (with surgery alone) to 60%–70% with multi-modality management [11]. Yet, the prognosis for patients with metastatic or recurrent disease is very poor with only 30%–40% achieving a cure. Furthermore, the prognosis for relapsing patients is very poor, with a 5-years survival rate between 20% to 25% [12], and it is even worse in those who relapse during the first 24 months [13, 14]. Therefore, there is an urgent need for developing better therapies to treat these patients with very poor prognosis.
The ESFTs family is characterised by specific chromosomal translocations involving the fusion of the EWS gene and ETS family of transcription factors. The translocation t(11;22) which results in the EWS-FLI1 fusion gene is the commonest [15]. Work by Scotlandi and colleagues revealed that IGF-1R system was activated in Ewing’s sarcoma cell lines and tumours by an autocrine loop [16]. Subsequently, Prieur and colleagues demonstrated the potential role of the EWS-FLI1 fusion protein in Ewing’s sarcoma in the IGF-1R pathway activation by repressing IGF-binding proteins [17]. The aim of this manuscript is to review the preclinical and clinical data supporting the use of IGF-1R inhibitors in ESFTs.

2. The IGF-1R Pathway

The IGF-1R pathway is more than a simple growth factor receptor, its ligands and a downstream signalling cascade. In vertebrates, it plays a key role in the growth and development of normal tissues and regulates the overall growth of organisms [23–25]. This pathway is also part of a more complex insulin-related signalling network. In the evolutionary process, the insulin-like growth factor receptors and IGF system have developed from a single, common ancestral receptor [26, 27] to a more complex system which involves three ligands (IGF-I, IGF-II, and insulin) and at least four receptors (IGF-1R, IGF-IIR, the insulin receptor (IR), and hybrid receptors) [28]. A diagram of the endocrine, paracrine, and autocrine regulation of this pathway is represented in Figure 1.

The IGF-1R is a transmembrane receptor that is activated by IGF-1 and by the related growth factor IGF-2. It is a tetrameric transmembrane receptor tyrosine kinase composed of two α and two β subunits linked by disulfide bonds. The extracellular α subunit is responsible for ligand binding, whereas the β subunit consists of a transmembrane domain and a cytoplasmic tyrosine kinase domain [29, 30]. The receptor is primarily activated by its cognate ligands,
through the transmembrane domain to the β-subunit. The β-subunit responds by undergoing a conformational change that causes stimulation of tyrosine kinase activity, followed by autophosphorylation of a cluster of tyrosine residues of the IGF-1R [31]. Then, IGF-1R activates alternative pathways for protection from apoptosis, cell proliferation, and differentiation. One of these pathways leads to the activation of PI3K-AKT-mTOR, while another pathway results in MAPKs (mitogen-activated protein kinases) activation. All these pathways, however, result in maintenance of cell survival by antagonizing the processes and proteins involved in apoptosis. This multiplicity of signalling pathways used by the IGF-1R may explain why this receptor has such powerful and widespread antiapoptotic activity [32–34].

3. Biological Implication of the IGF-1R Pathway in Ewing’s Sarcoma

The involvement of the IGF system in sarcoma initiation and progression has been associated with postnatal development [35, 36], primarily in those tumours that occur in younger patients. During this growing period, the function of growth hormone (GH) is mediated by IGF1. This is important, since IGFBPs are important regulators of growth and development. GH or IGF signalling may favour cell growth, thus increasing the risk of tumorigenesis.

In the case of Ewing’s sarcoma, IGF-1R is ubiquitously expressed and its activation is sustained by the autocrine production of IGF1 by tumour cells [38, 39]. In vitro studies have shown that IGF-1R is directly involved in Ewing’s sarcoma cell proliferation and survival [16, 40–42]. It has also been shown that EWS-FLI1, the genetic hallmark of Ewing sarcoma, is only capable of transformation in the presence of IGF-1R [43] and, more recently, that this fusion product directly affects IGF-1R signalling either by downregulating IGFBP3 [17], increasing IGF1 promoter, or both [44]. Additionally, it has been shown that forced expression of EWS-FLI in mesenchymal stem cells resulted in transformation into a phenotype similar to Ewing’s sarcoma. The cells transformed by the fusion gene expressed high levels of IGF1 and were dependent on IGF-1R signalling for growth and survival [45]. Similarly, in mouse fibroblasts, IGF-1R expression was necessary for EWS-FLI1-mediated transformation [43]. These in vitro results have been confirmed with the finding of IGF-1R expression in clinical samples of Ewing’s sarcoma and the demonstration that lower levels of IGF-1R expression correlate with a lower tumor proliferative rate and a better prognosis [46]. However, the limitations of this study in terms of its retrospective nature and the antibodies used should be noted. Despite such limitations, this observation is important when planning clinical trials, where stratification of patients for biological variables may be important.

The evidence described above supports a role for drugs targeting IGF-1R signalling in Ewing’s sarcoma. Blockade of IGF-1R has been shown to cause inhibition of cancer cell proliferation, survival, and anchorage-independent growth in vitro, to inhibit tumourigenesis, and block tumour invasion and metastasis, and to sensitize cancer cells to chemotherapy and radiotherapy [47].

4. Preclinical Experience Targeting IGF-1R in Ewing’s Sarcomas

Despite the advances in the treatment of Ewing’s sarcoma, many patients still succumb due to the development of metastatic or recurrent disease, and there is recognition that the benefit achieved with conventional cytotoxic therapy has reached a plateau. The need to identify and validate biologically critical targets is, therefore, extremely urgent. To achieve this aim, a large number of targeted therapeutic approaches have been evaluated in Ewing’s sarcoma models, both in vitro and in vivo. Some of these targets, including IGF-1R, have been validated in preclinical studies and IGF-1R inhibitors are currently undergoing evaluation in clinical trials. Among the various strategies used to interfere with IGF-1R function in preclinical studies, monoclonal antibodies (mAbs) and small molecule tyrosine kinase inhibitors represent the best candidates for clinical development.

Monoclonal antibodies need the following properties to be effective: they must inhibit binding of IGF1 and IGF2, induce receptor downregulation, and have little or no effect on insulin receptor signalling. Promising in vitro and in vivo studies have shown antitumor activity of several mAbs, resulting in inhibition of proliferation, apoptosis induction, and tumour growth inhibition [16, 48, 49].

There are a number of oral small molecule tyrosine kinase inhibitors in development. In vitro studies with a number of these agents have demonstrated inhibition of IGF-1R, high level of growth inhibition, survival reduction, complete pathway blockade, and xenograft tumor growth reduction [41, 50–52]. However, receptor downregulation was not observed with tyrosine kinase inhibitors, and this may partly account for their cytostatic, rather than cytotoxic effects against Ewing’s sarcoma xenografts [53].

Whether or not complete IGF-1R selectivity should be achieved is still under debate. Depending on the mechanism, inhibition of IGF-1R may target not only IGF-1R itself but also the hybrid receptors (especially those containing the fetal isoform insulin receptor-A) which favour cancer cell proliferation and are activated by both IGFS. It has been shown that targeting IGF-1R increases the efficacy of other anticancer therapies. This is based on evidence that IGF-1R signalling protects tumour cells from many insults, including chemotherapeutic agents and ionizing radiation [54–56], thus limiting the efficacy of such therapy. Inhibition of IGF-1R signalling has been shown to increase the sensitivity of Ewing’s sarcoma cells to chemotherapy [51, 57, 58]. Combining IGF1-R with conventional therapy may have the advantage of lowering the effective dosage of radiotherapy and chemotherapy, minimizing side effects.
while maintaining efficacy. This is particularly important for paediatric patients. In addition to a potential role in combination with traditional cytotoxic regimens and with radiotherapy, there are data demonstrating involvement of IGF-1R in trastuzumab resistance [59, 60] and resistance to AKT/mTOR inhibitors [61]. It has been shown that IGF-1R blockade can restore sensitivity to these agents.

An important issue in developing agents that specifically target IGF-1R is its high level of homology with the insulin receptor. There is a complete homology at the ATP-binding pocket and 84% homology within the intracellular kinase domain [62]. It is important to determine not only overlapping but also different biological effects of both receptors. Although both similarly activate PI3K and MAPK pathways [63, 64], subtle differences exist in the recruitment of certain docking proteins and intracellular mediators. These differences may be exploitable in terms of developing specific IGF-1R inhibitors. However, currently, there are no published data specifically addressing the role of the insulin receptor in Ewing’s sarcoma.

5. Clinical Experience with IGF-1R Targeted Treatments in Ewing’s Sarcoma

At the time of this review, mAbs against IGF-1R represent the most clinically advanced means of inhibiting this pathway in the treatment of Ewing’s sarcoma patients. Several antibodies have been tested in Phase II studies. Other approaches for blocking or disrupting IGF-1R activity in Ewing’s sarcoma patients include (a) the reduction of ligand levels or bioactivity or (b) the inhibition of receptor function using small-molecule tyrosine-kinase inhibitors [82]. Examples of different strategies for targeting the IGF-1R pathway are represented in Figure 1.

Several anti-IGF-1R mAbs have been developed for clinical use through the humanization of mouse mAbs, immunization of genetically engineered mice that produce fully human antibodies, or the selection of specific antibodies from phage display libraries. These antagonistic IGF-1R mAbs work through two major mechanisms: first by immediate inhibition of ligand binding, and secondly by a delayed effect on the downregulation of IGF-1R. At present, eight different mAbs have been evaluated in clinical trials: figitumumab (CP-751,871), ganitumab (AMG479), robatumumab (R1507), cixutumumab (IMC-A12), dalotuzumab (MK0646), SCH-717454, AVE-1642, and BIIB-022. Other reviews have extensively discussed the differences and similarities of these antibodies [22, 83]. In general, these mAbs are IgG1 isotype [65, 73, 84–87] with the exception of figitumumab and BIIB022 which are IgG2 [88] and IgG4 [71] isotype, respectively. There are significant pharmacokinetic and immunologic differences between IgG1, IgG2 and IgG4 isotypes. IgG2 mAbs appear to have longer half-lives than IgG1 and IgG4 mAbs, while IgG1 mAbs are usually potent activators of the classical complement pathway, complement-dependent cell-mediated cytotoxicity and antibody-dependent cellular cytotoxicity [89]. Table 1 reviews all the IGF-1R antibodies in clinical development.

5.1. Early Clinical Studies with Anti-IGF-1R MAbs Involving Ewing Sarcoma Patients. To date, three early studies involving the evaluation of IGF-1R mAbs in Ewing’s sarcoma have been published. The larger study, by Olmos et al. [67], enrolled 29 patients with sarcoma, of which 15 had refractory Ewing’s sarcoma. These patients were treated with figitumumab at the recommended dose of 20 mg/kg every four weeks. These patients were heavily pretreated (median of 3 lines), and notably 6 adolescent/paediatric patients (over 12 years of age) were included in Ewing’s sarcoma expansion cohort. Fourteen Ewing’s sarcoma patients were evaluable for radiological response, and 2 durable and ongoing radiological objective responses were observed, which included a pathological complete response (CR) (currently, 36+ months) in a 12 year old male, and a partial response (PR) (currently, 23+ months) in a young adult male (both responses are illustrated in Figure 2). In addition, 6 and 4 Ewing’s sarcoma patients were free of disease progression at 3 and 6 months, respectively. Furthermore, five of these Ewing’s sarcoma patients with prolonged stable disease (SD) had shrinkage of the target tumour lesions. Overall, the nonprogression rate at 3 months was 53% (CI-95% 28–78) and at 6 months was 40% (CI-95% 15–65) for all Ewing’s sarcoma patients included in the study. However, as this was a Phase I expansion cohort, it was not powered to formally detect antitumour activity as a primary endpoint [67].

The second Phase I study, reported by Tolcher et al. [75], studied the mAb ganitumab. This study included 12 adult Ewing’s sarcoma patients who were treated with doses of 12 and 20 mg/kg every 2 weeks. Ewing’s sarcoma patients received ganitumab on days 1, 15, and 29; and this was followed by a 28-day treatment-free period before resuming the drug if tumour response was observed. One patient with Ewing’s sarcoma attained a radiological CR which was maintained for 30 months. An additional Ewing’s sarcoma patient achieved an unconfirmed PR but was withdrawn from the study due to a myelodysplastic syndrome (non ganitumab related). No other objectives responses or prolonged disease stabilisation were reported [75].

A third mAb, R1507, has shown promising preliminary activity in Ewing’s sarcoma. The Phase I study of a weekly schedule of R1507 enrolled 9 Ewing’s sarcoma patients [73]. These patients were treated with doses ranging from 1 mg/kg to 9 mg/kg weekly. Two Ewing’s sarcoma patients had durable PRs (lasting 11 and 26+ months), and a further 2 had SD lasting for 4.3 and 6 months respectively.

Finally, a preliminary report of SCH-717454 was presented by Anderson et al. in the 2008 Annual Connective Tissue Oncology Society (CTOS) Meeting [66]. This study demonstrated radiological responses in patients with Ewing’s sarcoma [66]. This ongoing study included patients with refractory/resistant Ewing’s sarcoma, as well as patients with other sarcoma subtypes who were treated at a dose of 9 mg/kg every week.

5.2. Phase II Studies with Anti-IGF-1R mAbs Involving Ewing Sarcoma Patients. The exciting preliminary results with anti-IGF-1R mAbs led to the development of a Phase II
<table>
<thead>
<tr>
<th>Name</th>
<th>Class</th>
<th>Route</th>
<th>Company</th>
<th>Phase</th>
<th>Remarks in Ewing sarcomas</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Anti-IGF 1-receptor monoclonal antibodies</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AVE1642</td>
<td>Humanized IgG1</td>
<td>IV</td>
<td>Sanofi-Aventis</td>
<td>I-II</td>
<td>(i) Nil.</td>
<td>[65]</td>
</tr>
<tr>
<td>SCH-717454</td>
<td>Fully human IgG1</td>
<td>IV</td>
<td>Schering Plough</td>
<td>II</td>
<td>(i) PRs seen in EWS patients. (ii) Phase II in relapsed EWS or osteosarcoma.</td>
<td>[66]</td>
</tr>
<tr>
<td>CP-751,871figitumumab</td>
<td>Fully human IgG2</td>
<td>IV</td>
<td>Pfizer</td>
<td>II-III</td>
<td>(i) CR and PR in 2 pts with EWS. Prolonged SD in patients with EWS, synovial sarcoma and fibrosarcoma. (ii) Phase II trial in refractory paediatric and adult EWS: 15 PRs and 21 SD by RECIST.</td>
<td>[67, 68]</td>
</tr>
<tr>
<td>IMC-A12</td>
<td>Fully human IgG1</td>
<td>IV</td>
<td>ImClone systems</td>
<td>I-II</td>
<td>(i) Preclinical activity in EWS models. (ii) Phase II with 5 tiers: one for EWS.</td>
<td>[69, 70]</td>
</tr>
<tr>
<td>BIIB022</td>
<td>Fully human IgG4</td>
<td>IV</td>
<td>Biogen Idec</td>
<td>I</td>
<td>(i) Phase I dose-escalation ongoing in all solid tumors, no EWS enrolled yet.</td>
<td>[71]</td>
</tr>
<tr>
<td>MK-0646</td>
<td>Humanized IgG1</td>
<td>IV</td>
<td>Merck</td>
<td>II</td>
<td>(i) Phase I dose-escalation studies completed. (ii) SD in 2 patients: EWS and DSRCT respectively.</td>
<td>[69, 72]</td>
</tr>
<tr>
<td>R1507Robatumumab</td>
<td>Fully human IgG1</td>
<td>IV</td>
<td>Hoffman-La Roche</td>
<td>II</td>
<td>(i) Several PRs and prolonged SD in pts with EWS. (ii) Phase II in multiple sarcoma subtypes halted since 12/22/2009.</td>
<td>[73, 74]</td>
</tr>
<tr>
<td>AMG-479</td>
<td>Fully human IgG1</td>
<td>IV</td>
<td>Amgen</td>
<td>II</td>
<td>(i) CR and PR in 2 EWS patients. (ii) Phase II in relapsed Ewing and DSRCT. In EWS 1 PR and 1 SD &gt;6 months.</td>
<td>[75, 76]</td>
</tr>
<tr>
<td><strong>Anti-IGF-1-receptor small molecule/tyrosine kinase Inhibitors</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>INSM-18</td>
<td>Small molecule TKI</td>
<td>PO</td>
<td>Insmed</td>
<td>I-II</td>
<td>(i) An ATP competitive and reversible TKI, which also inhibits HER2.</td>
<td>[52]</td>
</tr>
<tr>
<td>BMS-754807</td>
<td>Small molecule TKI</td>
<td>PO</td>
<td>Bristol Myers Squibb</td>
<td>I</td>
<td>(i) An ATP competitive and reversible TKI. (ii) In vitro activity against in EWS, RMS, liposarcoma cell lines.</td>
<td>[51]</td>
</tr>
<tr>
<td>OSI-906</td>
<td>Small molecule TKI</td>
<td>PO</td>
<td>OSI pharmaceuticals</td>
<td>I</td>
<td>(i) An ATP competitive and reversible TKI. (ii) Phase I enrolled 2 Ewing sarcoma patients without benefit.</td>
<td>[77]</td>
</tr>
<tr>
<td>XL-228</td>
<td>Small molecule TKI</td>
<td>IV</td>
<td>Exelixis</td>
<td>I</td>
<td>(i) An ATP competitive TKI of IGF-1R, Aurora, FGFR and Src. (ii) Prolonged SD in 2 pts with leiomyosarcoma (≥7 m) and liposarcoma (≥5 m) respectively.</td>
<td>[78]</td>
</tr>
<tr>
<td>BVP-51004cycolignan,PPP</td>
<td>Small molecule TKI</td>
<td>PO</td>
<td>Biovitrum</td>
<td>I</td>
<td>(i) A non-ATP competitive TKI. (ii) In vitro activity in multiple sarcoma resistant cell lines, including osteosarcoma.</td>
<td>[79]</td>
</tr>
<tr>
<td>A-947864</td>
<td>Small molecule TKI</td>
<td>PO</td>
<td>Abbott</td>
<td>Preclinical</td>
<td>(i) An ATP competitive TKI.</td>
<td>[80]</td>
</tr>
<tr>
<td>BMS-554417</td>
<td>Small molecule TKI</td>
<td>PO</td>
<td>Bristol Myers Squibb</td>
<td>Preclinical</td>
<td>(i) An ATP-competitive and reversible TKI. (ii) In vitro activity in EWS and osteosarcoma cell lines.</td>
<td></td>
</tr>
<tr>
<td>NVP-AEW541NVP-ADW742</td>
<td>Small molecule TKIs</td>
<td>PO</td>
<td>Novartis</td>
<td>Preclinical</td>
<td>(i) ATP competitive and reversible TKIs. (ii) In vitro and in vivo activity in EWS.</td>
<td>[51, 53]</td>
</tr>
<tr>
<td>GSK1904529AGSK1838705A</td>
<td>Small molecule TKIs</td>
<td>PO</td>
<td>GlaxoSmithKline</td>
<td>Preclinical</td>
<td>(i) ATP-competitive, reversible, TKIs of IGF-1R and IR. (ii) In vitro efficacy in EWS cell lines.</td>
<td>[50, 81]</td>
</tr>
<tr>
<td>AG1024(Tyrphostin)</td>
<td>Small molecule TKI</td>
<td>N.A.</td>
<td>Merck</td>
<td>Preclinical</td>
<td>(i) Used mainly in preclinical drug testing. Non-ATP competitive.</td>
<td></td>
</tr>
</tbody>
</table>

study in a variety of sarcoma subtypes, including Ewing’s sarcoma, conducted by the Sarcoma Alliance for Research through Collaboration (SARC) study group. This ambitious study had 5 arms for specific sarcoma subtypes and had a planned recruitment of approximately 300 patients. The results of this study were reported during the 2010 American Society of Clinical Oncology (ASCO) annual meeting [74]. A Green and Dahlberg two-stage design was employed and the study included 111 Ewing’s sarcoma patients from 30 centres across North America and Europe. Patients were treated with 9 mg/kg weekly of R1507 and stratified in two different cohorts at study entry: poor prognosis cohort (relapse/refractory disease <24 months and/or ≥2 chemotherapy regimens) which included 67 patients and a good prognosis cohort (relapse ≥24 months and <2 prior chemotherapy regimens) which included 44 patients. A total of 10 confirmed objective responses were observed using WHO criteria [90], 1 CR, and 9 PRs. A further 7 patients achieved unconfirmed partial responses but progressed rapidly after the first radiological evaluation. Objective
Table 2: Responses in clinical trials.

<table>
<thead>
<tr>
<th>Drug</th>
<th>N</th>
<th>CR</th>
<th>PR</th>
<th>SD</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Figitumumab</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Phase I [67]</td>
<td>15</td>
<td>1</td>
<td>1</td>
<td>6</td>
</tr>
<tr>
<td>Phase II [68]</td>
<td>106</td>
<td>0</td>
<td>15</td>
<td>25</td>
</tr>
<tr>
<td><strong>R1507 Phase I</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Phase I [73]</td>
<td>9</td>
<td>0</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>Phase II [74]</td>
<td>111</td>
<td>1</td>
<td>7</td>
<td>17</td>
</tr>
<tr>
<td><strong>Ganitumumab</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Phase I [75]</td>
<td>12</td>
<td>1</td>
<td>1</td>
<td>NA/NR</td>
</tr>
<tr>
<td>Phase II [76]</td>
<td>19</td>
<td>0</td>
<td>1</td>
<td>7</td>
</tr>
</tbody>
</table>

$N =$ number of Ewing's patients; $CR =$ confirmed complete response; $PR =$ confirmed partial responses; $SD =$ stable disease (best response); NA/NR: nonavailable/nonreported.

responses were equally distributed between the poor and good prognosis cohorts (approximately 9% in both). The median duration of response in these patients was 25 weeks (range 12–47). A further 17 patients had confirmed SD as the best response, 3 of these would have been defined as PRs if RECIST rather than WHO criteria had been employed [91]. The median overall survival (OS) for patients treated in this study was 6.9 months.

A Phase II study of the IGF-1R mAb, ganitumab (AMG479), in Ewing's sarcoma and desmoplastic small round cell tumour (DSRCT) patients was also presented at the 2010 ASCO annual meeting [76]. The principal objective of this study was to determine the objective response rate (ORR) in patients who had not received prior therapy with an IGF-1R inhibitor; however, there was an exploratory cohort evaluating patients who had previously received another anti-IGF-1R targeted therapy. All patients received ganitumab at 12 mg/kg every 2 weeks. A total of 19 Ewing's sarcoma patients entered the primary cohort, and 3 were recruited to the exploratory cohort (no further data are currently available). One Ewing's sarcoma patient attained a PR, and a further 7 Ewing's patients achieved SD as best response; however, only one of these remained progression free beyond 24 weeks. The median progression-free survival (PFS) for Ewing's sarcoma patients included in this trial was 7.9 weeks.

More recently, Juergens et al. [68] presented the preliminary results of a Phase 2 study of figitumumab in paediatric (10 years or older) and adult patients with refractory Ewing's sarcomas. In this study, 106 patients were evaluable for objective response (RECIST), 15 patients had PRs, and 25 had stable disease. The median PFS for the overall population was poor 1.9 months (CI-95% 1.8–2.1), and the median overall survival was 8.9 months (CI-95% 7.2–10.8). However, in those patients with elevated blood IGF-1 levels ($>$110 ng/mL) at baseline, there was a significant advantage ($P < .001$) in OS compared with those with low IGF-1 ($<110$ ng/mL), that is, 10.5 months and 4.5 months, respectively.

To our knowledge, there are two further Phase II studies of IGF1-R inhibition in Ewing's sarcoma and osteosarcoma patients ≥4 years of age (http://www.clinicaltrials.gov/, NCT00617890) has a planned recruitment of 190 patients and (2) a study of cixutumumab (http://www.clinicaltrials.gov/, NCT00668148) in 185 patients (≥12 years) and five arms: Ewing's sarcoma, rhabdomyosarcoma, leiomyosarcoma, adipocytic sarcomas and synovial sarcoma.

Despite the preclinical data and promising early clinical results in Ewing’s sarcoma, the recent Phase II results with anti-IGF-1R mAb as monotherapy have been less impressive than initially hoped (Table 2). Preliminary data for the mTOR inhibitor, radiforolimus (previously known as deferolimus), have shown a nonprogression rate of 30% at 16 weeks in bone sarcomas [21]. The mTOR inhibitor was deemed active and a Phase III trial comparing radiforolimus with placebo, as maintenance therapy, has recently completed recruitment. Other targeted agents have also been explored, and in a recent trial of imatinib in various sarcoma subtypes, no clinical activity was seen in patients with Ewing's sarcoma [92].

The results of Phase II studies published to date have been disappointing, and the clinical development pathway for this class of agents is currently very uncertain. Furthermore, the poor results observed with these agents in lung cancer have led to Roche halting further development of R1507 in all tumours [93, 94]. However, there is still the promise that these agents may have a role in the management of Ewing’s sarcoma, either as monotherapy in selected patients or in combination regimens.

5.3. Toxicity with IGF-1R Monoclonal Antibodies. In general, IGF-1R mAbs are well tolerated, with the most common toxicities being mild and occasionally moderate. Severe (grade 3) or life-threatening (grade 4) adverse events are rare. Potential grade 3 and 4 hematologic adverse events reported in the Phase II trial with ganitumumab and R1507 included thrombocytopenia [74, 76], anemia [74, 76], neutropenia [76], pain at the time of administration [74], hyponatremia [74], and hyperglycemia [74, 76]. Thrombocytopenia was also reported in Phase I studies [69, 73, 75].
and 4 nonhematologic adverse events with figitumumab in sarcoma patients included deep venous thrombosis ($n = 1$), vomiting ($n = 1$), and back pain ($n = 1$). Grade 3 fatigue was also reported with figitumumab in nonsarcoma patients [95, 96]. Other relevant grade 3-4 nonhematological adverse events described with other IGF-1R mAbs include fatigue [72, 75, 97], arthralgia [75], chills [72], pneumonia [69], nausea or vomiting [72], rash and/or pruritus [72], pain [66, 72], and gastrointestinal bleeding [69].

Hyperglycaemia is a common toxicity of all the mAbs, with grade 3 hyperglycaemia seen in several studies [66, 72, 97]. The mechanism for hyperglycaemia is unclear although IGF-1R may be involved in glucose metabolism via crosstalk and heterodimerisation with the insulin receptor [98–101]. This observation, and the increased plasma insulin levels reported after treatment with IGF-1R mAbs [96, 102], suggests compensatory insulin secretion and associated insulin resistance, the latter possibly secondary to increased IGF-1 and growth-hormone levels [82, 103]. Other severe laboratory abnormalities observed in sarcoma patients include uric acid elevation and transaminitis [67].

Interestingly, despite the expression of IGF-1R in vascular smooth muscle and endothelial cells [104] and the potential cardiotoxicity associated with mAbs, no cardiac toxicity has been reported to date. In the case of sarcoma patients treated with figitumumab, it is noteworthy that three-quarters of the patients were pretreated with anthracyclines and none developed cardiotoxicity [67].

Theoretically, IGF-1R mAbs would be expected to have an inhibitory effect on IGF and growth hormone-mediated growth. Thus, IGF-1R blockade could cause linear and somatic growth delay in a childhood and teenage population, as supported by the identification of patients with genetic defects in the IGF-1 axis such as IGF-1 deficiency [105]. This potential long-term adverse event is extremely important in the management of young sarcoma patients [1]. The current clinical experience is too limited to definitively address this question [67]. Detailed assessments of growth and hormone levels have been included in ongoing Phase II trials recruiting paediatric and prepubertal teenage patients, and it is hoped that these studies will provide insights to the effect of IGF-1R targeted therapy on growth during childhood and puberty.

5.4. Early Experience with Tyrosine Kinase Inhibitors of IGF1-R. There are a number of small molecule tyrosine kinase inhibitors (TKIs) of IGF1-R that are currently being, or have been, evaluated (Table 1). Some of these small molecules also inhibit IR-A, a component of IGF-R hybrid receptors [83]. Although this can potentially result in greater antitumour activity, it may also be associated with a higher incidence of metabolic toxicity. From the results of clinical trials of monoclonal antibodies and tyrosine kinase inhibitors in other tumour types, it is apparent that predicting differences in efficacy between these two classes can be difficult [106]. Notably, small molecule tyrosine kinase inhibitors do not directly activate the immune response against tumour cells, but they may be more effective when activated receptors are localised in cytoplasmic caveosomes and/or endosomes.

Some of these novel IGF-1R TKIs (i.e., picropodophyllin (PPP), GSK183870A, GSK1904529A, BMS-536924, NVP-AEW541) have already shown promising preclinical activity as single agents or in combination in different sarcoma models [50, 51, 53, 79, 81, 107–109]. At the present time, only OSI-906 has been tested in Ewing’s sarcoma patients ($n = 2$) although no antitumor activity was seen in these two cases [77]. However, currently there are insufficient data to define any difference in clinical benefit in patients treated with these two classes of IGF1-R inhibitors.

5.5. Combination Therapy with IGF-1R mAbs. IGF-1R activation has been associated with chemoresistance in multiple cancers [110], including some sarcomas such as Ewing’s sarcoma [39]. Indeed, modulation of IGF signalling has been shown to enhance the antitumor activity of cytotoxic drugs in laboratory sarcoma models [58]. Thus, a strategy based on the combination of first- or second-line sarcoma chemotherapy with IGF-1R mAbs seems a rational approach in the utilisation of these agents. Currently, there are a number of ongoing or planned studies evaluating such combinations, including a Phase I/II trial of cixutumumab in combination with doxorubicin for advanced and unresectable soft-tissue sarcomas (http://www.clinicaltrials.gov/, NCT00720174), sponsored by the National Cancer Institute and a Phase I of SCH-717454 in combination with different commonly used chemotherapies in sarcoma such as vincristine, doxorubicin, and cyclophosphamide (CAV) or ifosfamide and etoposide (http://www.clinicaltrials.gov/, NCT00960063).

Furthermore, clinical studies of rational combinations of IGF-1R mAbs with other targeted therapies are in progress. Examples of such regimens are the use of mTOR inhibitors in combination with IGF-1R antibodies [49, 111]. Studies evaluating this approach include a trial of RAD001 (everolimus) in combination with figitumumab sponsored by the Dana-Faber Cancer Institute [112]. This study enrolled a total of 21 sarcoma patients one of whom had Ewing’s sarcoma. The reported toxicity profile for this combination was not significantly different from that of single agent everolimus. Grade 3 toxicity occurred in ≤10% of patients, and included mucositis, nausea, vomiting, and diarrhoea. One patient with Ewing’s sarcoma maintained stable disease for six months. In addition, a trial of temsirolimus with cixutumumab (http://www.clinicaltrials.gov/, NCT01016015) is actively recruiting sarcoma patients. Other rational combinations could include regimens with heat shock protein 90 [113] or EGFR/HER2 inhibitors [107], as these have been implicated in potential mechanisms of resistance to IGF-1R inhibition in sarcoma cell lines.

5.6. Patient Selection. Despite robust preclinical evidence supporting the role of IGF1-R-targeted agents in Ewing’s sarcoma, clinical results show that only a proportion of patients derive significant benefit, with many progressing early, even after an initial response. Although initial reports suggested an association between the EWS/FLI-1 type 1 translocation and response in Ewing’s sarcoma [75], the
purported predictive value of translocation type has not been observed consistently [67, 74, 76]. Clinical data in nonsmall cell lung cancer patients have suggested that circulating free IGF-1 may identify patients who derive clinical benefit from figitumumab [114]. Similar data has also been reported in the Phase II trial of figitumumab in refractory Ewing’s sarcoma, in which patients with elevated IGF-1 at baseline achieved longer OS [68]. However, it still remains unclear if an elevated IGF-1 level at baseline is a predictive factor for response to IGF-1R antibodies or simply a prognostic factor. Nonetheless, as IGF system and the activation of the IGF1-R are complex, response and resistance mechanisms are unlikely to be entirely dependent on or explained by circulating IGF-1 [115, 116].

6. Conclusions

During the last two decades, large amounts of preclinical data have been accumulated supporting the use of agents targeting IGF-1R in Ewing’s sarcoma. This rationale has been reinforced by the early reports of clinical activity with several IGF-1R antibodies in this disease. However, the benefit of this therapeutic approach clearly does not extend to all patients, with Phase II studies demonstrating less promising responses than initially anticipated. In addition to the exploration of IGF-1R in combination with chemotherapy and other targeted agents, there is an urgent need to identify predictive biomarkers to improve patient selection, as well as to elucidate the mechanisms of resistance to these drugs, thereby facilitating the development of rational combination regimens. Despite the disappointing Phase II data, this novel group of drugs does constitute an active treatment in a proportion of Ewing’s sarcoma patients.

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