Research Article

Effects of Placenta-Derived Mesenchymal Stem Cells on the Particulate Matter-Induced Damages in Human Middle Ear Epithelial Cells

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This study was aimed at investigating the effects of placenta-derived mesenchymal stem cells (PL-MSCs) on particulate matter (PM-) exposed human middle ear epithelial cells (HMEECs). HMEECs were treated with 300 μg/ml PM for 24 hours. The PL-MSCs were cocultured with PM-treated HMEECs. Cells were harvested on days 0, 1, and 4, and the expression of the inflammatory genes TNFα, COX2, IL1β, IL6, and MUC5B in HMEECs and anti-inflammatory genes PTGES, TGFβ, and VEGF in PL-MSCs was examined by qRT-PCR. The culture media were collected to measure the secreted PGE2 level using an enzyme-linked immunosorbent assay. The mRNA expression of TNFα, COX2, IL1β, IL6, and MUC5B in HMEECs increased following PM treatment. PM-treated HMEECs cocultured with PL-MSCs showed alleviated inflammatory reactions represented by lower mRNA expression levels of MUC5B, TNFα, IL1β, and IL6 compared to monocultured PM-treated HMEECs. The mRNA expression levels of PGE2, TGFβ, and VEGF were elevated in cocultured PL-MSCs compared to those of control PL-MSCs. The medium of PM-treated HMEECs cocultured with PL-MSCs exhibited increased PGE2 levels. The increased inflammatory response in PM-treated HMEECs was reversed using PL-MSCs. The PGE2, TGFβ, and VEGF were the mediators of the anti-inflammatory effects of PL-MSCs.

1. Introduction

Particulate matter (PM) is a major airway pollutant that has increased in recent years. Enormous adverse health outcomes have been documented for airway, cardiovascular, neurological, and metabolic disorders [1, 2]. It has been suggested that PM elevates inflammation and oxidative stress and impairs cell proliferation [3, 4]. The middle ear is connected to the upper airway via the Eustachian tube. Thus, PM could enter directly into middle ear epithelial cells and cause injury. In addition, the blood supply of the middle ear epithelium could be a route for PM-induced effects. Prior studies have demonstrated inflammation in human middle ear epithelial cells (HMEECs) following PM exposure [5, 6]. The 24 hours of diesel exhaust particle (DEP) increased the mucus-producing gene of MUC5AC in HMEECs [7]. However, few studies have explored the temporal changes in PM effects on the middle ear. In addition, the anti-inflammatory response of stem cells following PM exposure in the middle ear has not been well-described.

Several natural compounds are believed to relieve PM-induced inflammation [8]. However, the rescue effects from PM-induced inflammation are questionable. Moreover, the active compounds and their mechanisms of action have not been delineated. Mesenchymal stem cells (MSCs) secrete several trophic factors with immunomodulatory, antiapoptotic, and anti-inflammatory effects [9–11]. For instance, an in vitro study reported that the adipose-derived mesenchymal stem cells (AD-MSCs) exerted anti-inflammatory effects on osteoarthritic chondrocytes and synoviocytes after 3 days of coculture [9]. Several animal studies have revealed the therapeutic roles of MSC secretomes in the immune system...
HMEECs were cultured in a mixture of Dulbecco’s Modified Eagle’s Media (DMEM; Invitrogen, Carlsbad, CA, USA) and bronchial epithelial basal media (1:1; Lonza, Walkersville, MD, USA). Cells were maintained at subconfluence in a CO₂-enriched (95% air, 5% CO₂) humidified atmosphere at 37°C. PL-MSCs were obtained from CHA Biotech Co. Ltd. (South Korea). PL-MSCs were cultured in alpha-MEM (Gibco, Gaithersburg, MD, USA) containing 10% fetal bovine serum (Gibco), 100 U/ml penicillin, and 100 μg/ml streptomycin in a CO₂-enriched (95% air, 5% CO₂) humidified atmosphere at 37°C. PM (Standard Reference Material (SRM) 2975; National Institute of Standards and Technology, Gaithersburg, MD, USA) was suspended in sterile saline (0.9% NaCl) containing Triton X (0.001%). Prior to the experiments, PM was sonicated for 15 min and vortexed to achieve a 6-well plate at a 1:1 cell ratio (Figure 1). We set day 0 for the initial day of coculture. Before 300 μg/ml PM treatment, HMEECs were cocultured with PL-MSCs without direct contact. The preliminary results were presented at the International Congress of ORL-HNS 2017 (ICORL 2017) [15]. In addition, the expression of anti-inflammatory and proliferative genes was measured. The inflammatory changes in MSCs after coculture with PM-injured HMEECs were also examined.

2. Materials and Methods

2.1. Cell Culture. HMEECs were obtained from Dr. David J. Lim at the House Ear Institute (Los Angeles, CA, USA). HMEECs were cultured in a mixture of Dulbecco’s Modified Eagle’s Media (DMEM; Invitrogen, Carlsbad, CA, USA) and bronchial epithelial basal media (1:1; Lonza, Walkersville, MD, USA). Cells were maintained at subconfluence in a CO₂-enriched (95% air, 5% CO₂) humidified atmosphere at 37°C. PL-MSCs were obtained from CHA Biotech Co. Ltd. (South Korea). PL-MSCs were cultured in alpha-MEM (Gibco, Gaithersburg, MD, USA) containing 10% fetal bovine serum (Gibco), 100 U/ml penicillin, and 100 μg/ml streptomycin in a CO₂-enriched (95% air, 5% CO₂) humidified atmosphere at 37°C. PM (Standard Reference Material (SRM) 2975; National Institute of Standards and Technology, Gaithersburg, MD, USA) was suspended in sterile saline (0.9% NaCl) containing Triton X (0.001%). Prior to the experiments, PM was sonicated for 15 min and vortexed to minimize aggregation. HMEECs were treated with 200 or 300 μg/ml PM for 24 hours. Next, coculture was performed by incubating HMEECs in transwells (0.4 μm pore size; Corning® Transwell® polyester membrane cell culture inserts (Cat. No. COP-3450)) and PL-MSCs in the lower chamber of the lower chamber of a 6-well plate at a 1:1 cell ratio (Figure 1). We set day 0 for the initial day of coculture. Before 300 μg/ml PM-treated HMEECs were cocultured with PL-MSCs with transwells, HMEECs were washed three times using PBS. HMEECs and PL-MSCs were cocultured in the media used for HMEEC culture. Control cells were monocultures of PL-MSCs and HMEECs. To analyze the direct effects of PM on PGE2 secretion of PL-MSCs, the PL-MSCs were monocultured with treatment of 300 μg/ml PM. Coculture of PL-MSCs and HMEECs was repeated four times. On days 0, 1, and 4, both monocultures and cocultures were detached with trypsin-EDTA, and viability was determined using the trypan blue exclusion method and the MTT assay to measure the cytotoxic effects of PM in HMEECs. Cells were harvested on days 0, 1, and 4 for quantitative reverse transcription polymerase chain reaction (qRT-PCR) analysis of mucin 5B (MUC5B), tumor necrosis factor alpha (TNFa), interleukin 1 beta (IL1β), interleukin 6 (IL6), cyclooxygenase 2 (COX2), prostaglandin E synthase (PTGES), transforming growth factor beta (TGFβ), and vascular endothelial growth factor (VEGF), and supernatants were stored at -80°C for enzyme-linked immunosorbent assay (ELISA) of prostaglandin E2 (PGE2).

2.2. Analysis of Gene Expression. Total RNA was extracted from human PL-MSC and HMEEC monocultures and cocultures using TRI Reagent® (Sigma-Aldrich, St. Louis, MO, USA) according to the manufacturer’s instructions. Reverse transcription was performed using TOPscript™ RT DryMix (dT 18 plus; Enzymonics Co. Ltd., Daejeon, South Korea) according to the manufacturer’s protocol. Forward and reverse oligonucleotides for PCR amplification of MUC5B, TNFa, IL1β, IL6, COX2, PTGES, TGFβ, and VEGF are shown in Table 1. RT-PCR was run in a Bio-Rad-CFX96 Touch™ Real-Time PCR Detection System using TOPreal™ qPCR 2x PreMIX (SYBR Green with low ROX; Enzymonics) with the following protocol: initial activation of HotStarTaq DNA polymerase at 95°C for 15 min, followed by 45 cycles of 95°C for 10 s and 60°C for 15 s. The amplification efficiency (E) of each amplicon was determined using 10-fold serial dilutions of positive control complementary DNAs (cDNAs) and calculated from the slopes of the log input amounts (from 20 ng to 2 pg of cDNA) plotted versus the crossing point values according to the formula $E = 10^{1/slope}$. All primer efficiencies were confirmed to be high (>90%) and comparable (Table 1). For each target gene, messenger RNA levels were calculated, normalized to GAPDH, and presented as a percentage of the reference gene. All RT-PCRs were replicated three times for each gene for four respective coculture sets.

Cell culture media were collected following 4 days of cultures and monocultures, and the secretion of prostaglandin E2 (PGE2) was measured using an ELISA (Prostaglandin E2 Parameter Assay Kit, R&D Systems, Minneapolis, MN, USA) according to the manufacturer’s instructions.

2.3. Statistical Analysis. Statistical analysis was performed using the Mann–Whitney U test. SPSS v.21.0 software (IBM, Armonk, NY, USA) was used for analysis. P values less than 0.05 were considered significant.

3. Results

3.1. Effects of PM on HMEECs. The cell viability was significantly decreased after 200 or 300 μg/ml of 24-hour PM exposures, compared to control HMEECs (Figure 2(a)) ($P = 0.03$ for 200 μg/ml PM and $P = 0.02$ for 300 μg/ml PM). Trypan blue staining also showed a significant decrease of viable HMEECs in the PM-exposed groups (Figure 2(b)). The mRNA expression of TNFa, IL6, MUC5B, IL1β, and COX2 was differentially increased in HMEECs following incubation with 300 μg/ml PM at different time points (Figure 3). The mRNA expression of TNFa, IL6, and MUC5B was significantly increased at day 4 (5 days after PM treatment). The IL1β and COX2 expression peaked on day 1 (2 days after PM treatment) and decreased on day 4.
Effects of PL-MSCs on PM-Treated HMEECs.

Coculture with PL-MSCs caused a significant decrease in the mRNA expression of TNFα, IL1β, IL6, and MUC5B in HMEECs compared to monocultured HMEECs at 4 days of culture with PL-MSCs (Figure 3). In contrast, COX2 mRNA expression was more elevated under coculture conditions than monocultured HMEECs. The mRNA expression of TNFα and COX2 was not significantly changed in PL-MSCs after coculture with PM-treated HMEECs for 0, 1, and 4 days (Supplementary Figure 1). TGFβ, PTGES, and VEGF mRNA expression was increased in PL-MSC after coculture with PM-treated HMEECs for 44 days (Figure 4). TGFβ and VEGF mRNA expression was not significantly changed in PM-treated HMEECs after coculture with PL-MSCs for 0, 1, and 4 days (Supplementary Figure 2). On the other hand, PTGES mRNA expression in PM-treated HMEECs

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer sequence (forward)</th>
<th>Primer sequence (reverse)</th>
<th>Annealing temperature (°C)</th>
<th>Product size (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MUC5B</td>
<td>5′-GCCTACGAGGACTTCAACG</td>
<td>5′-CCITTGATGACCCACACG</td>
<td>60</td>
<td>79</td>
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<tr>
<td></td>
<td>T-3′</td>
<td>GGTG-3′</td>
<td></td>
<td></td>
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<tr>
<td>TNFα</td>
<td>5′-CCCCATGGTGTAGCAACCCC</td>
<td>5′-TGAGGTACAGGCCTCTGA</td>
<td>60</td>
<td>132</td>
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<tr>
<td></td>
<td>T-3′</td>
<td>T-3′</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IL1β</td>
<td>5′-CATTGCTAAATGTCTGAAA</td>
<td>5′-CATGGCCACACAAACT</td>
<td>60</td>
<td>238</td>
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<tr>
<td></td>
<td>GC-3′</td>
<td>GACG-3′</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IL6</td>
<td>5′-ACTCACTCTCTCAGAACGA</td>
<td>5′-CCATCTTGGAGGTACAG</td>
<td>60</td>
<td>149</td>
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<tr>
<td></td>
<td>ATTG-3′</td>
<td>GTTG-3′</td>
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<tr>
<td></td>
<td>(COX2)</td>
<td></td>
<td></td>
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<tr>
<td>PGE2</td>
<td>5′-GAAGAAGGCTTTGCAAC</td>
<td>5′-AGGGGCTTCACATAAAGG</td>
<td>60</td>
<td>139</td>
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<td></td>
<td>C-3′</td>
<td>T-3′</td>
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<tr>
<td>TGFβ</td>
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<td>T-3′</td>
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<tr>
<td>VEGF</td>
<td>5′-TCCACCATGCCAAGTGCTC</td>
<td>5′-GTCACCAGGGGTTCAG</td>
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<tr>
<td></td>
<td>3′</td>
<td>ATTG-3′</td>
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</table>

3.2. Effects of PL-MSCs on PM-Treated HMEECs. Coculture with PL-MSCs caused a significant decrease in the mRNA expression of TNFα, IL1β, IL6, and MUC5B in HMEECs compared to monocultured HMEECs at 4 days of culture with PL-MSCs (Figure 3). In contrast, COX2 mRNA expression was more elevated under coculture conditions than monocultured HMEECs. The mRNA expression of TNFα and COX2 was not significantly changed in PL-MSCs after coculture with PM-treated HMEECs for 0, 1, and 4 days (Supplementary Figure 1). TGFβ, PTGES, and VEGF mRNA expression was increased in PL-MSC after coculture with PM-treated HMEECs for 44 days (Figure 4). TGFβ and VEGF mRNA expression was not significantly changed in PM-treated HMEECs after coculture with PL-MSCs for 0, 1, and 4 days (Supplementary Figure 2). On the other hand, PTGES mRNA expression in PM-treated HMEECs
increased 1 day after coculture with PL-MSCs. This increase decreased after 4 days of coculture with PL-MSCs in PM-treated HMEECs. In the culture medium, secreted PGE2 was not increased in monocultured PM-treated HMEECs, while it was significantly elevated under coculture conditions (Figure 5).

4. Discussion

4.1. Principle Findings. PM exposure decreased the proliferation of HMEECs. The inflammatory genes TNFα, IL1β, IL6, and COX2 and the mucin-producing genes of MUC5B were elevated in HMEECs following treatment with PM. After coculture with PL-MSCs, the mRNA expression of TNFα, IL1β, and IL6 was decreased in HMEECs and that of TGFβ, PGE2, and VEGF was increased in PL-MSCs. Few studies have reported the rescue effects of PL-MSCs in PM-induced epithelial injuries. In addition, the present study observed temporal changes in gene expression profiles in both HMEECs and PL-MSCs. The administration of MSCs into the middle ear could be a potential therapeutic tool for middle ear inflammation or otitis media based on this study. The optimal duration of administered MSCs needed to exert anti-inflammatory effects is expected to be 2-4 days from this study. There are no therapies for otitis media except for antibiotics for bacterial infection. Thus, the MSCs could be used to rescue the inflamed and injured middle ear epithelium in the otitis media of patients. Although the optimal MSC treatment conditions need to be explored in future studies, the local delivery of drug via the middle ear may be promising, as described in previous studies [16, 17].

4.2. Expression of Inflammatory Genes in PM-Treated HMEECs. PM initiated the inflammatory response in HMEECs in the present study. The mRNA expression of IL1β and COX2 was elevated 2 days after exposure to PM (day 1). Leukotrienes and prostaglandins are some of the earliest types of cytokines produced in response to antigenic stimulation. COX2 produces prostaglandin and elevates its expression in the otitis media [18]. After 5 days of PM exposure (day 4), levels of the inflammatory genes TNFα and IL6 were increased. TNFα is an inflammatory gene that induces inflammation in the middle ear and upregulates mucin gene expression and subsequent mucin secretion [19]. Indeed, the mRNA expression of MUC5B increased after 5 days of PM exposure (day 4). MUC5B is related to mucociliary clearance [20]. The middle ear epithelium of chronic otitis media patients expresses MUC5B [21]. These temporal changes of inflammatory responses of HMEECs after PM exposure may be in line with increased activation of reactive oxygen species on day 4 compared to day 2 in monocultured HMEECs (Supplementary Figure 3).

4.3. Attenuated Inflammatory Response in PM-Treated HMEECs. The inflammatory response following PM exposure was relieved under PL-MSC coculture conditions. Because there was no direct contact between HMEECs and PL-MSCs, the mediating cytokines could interact between cells. The mRNA levels of the inflammatory genes TNFα and IL6 were decreased in HMEECs after coculture with PL-MSCs for 4 days. The mRNA expression of TGFβ, PGE2, and VEGF was increased in PL-MSCs when cocultured with PM-treated HMEECs. Moreover, it was presumed that the PL-MSC effects are dependent on existing inflammation. The levels of anti-inflammatory genes were elevated in PL-MSCs along with the increase of inflammatory genes in PM-treated HMEECs. Small effects were observed in cells that produce low levels of inflammatory genes on day 1 of coculture. However, both the inflammatory and anti-inflammatory genes in HMEECs and PL-MSCs were markedly changed after 5 days. Previous studies reported that human MSCs require activation, and activating stimuli appear to include the proinflammatory genes IL1β and TNFα [22]. It was also reported that the anti-inflammatory effects of MSCs are independent of the tissue sources or MSC donors and result from the inflammatory status of target cells [9]. In the presence of inflammatory stimuli, PL-MSCs relieve the inflammation by releasing anti-inflammatory genes.

4.4. Modulation of the COX2/PGE2 Pathway by PL-MSCs. Although several inflammatory (TNFα and IL6) and anti-inflammatory (TGFβ and VEGF) genes altered their expression level in accordance with PM exposure and coculture conditions, the changes in COX2 and PGE2 were remarkable.
The high expression of PGE2 during coculture with PL-MSCs suggests that the COX2/PGE2 pathway acts as a modulator of PL-MSC actions. In contrast to other inflammatory genes, COX2 mRNA expression was more augmented under PL-MSC coculture conditions, which may have been caused by the increased mRNA expression of COX2 in response to elevated PGE2 production [23]. Previous studies have also suggested that the therapeutic immunomodulatory effects of MSCs correlate with COX2-derived PGE2 expression [9, 10, 14]. The paracrine secretion of PGE2 from umbilical cord MSCs primarily affects acute lung injuries in mice [14]. PGE2 plays a major role in the immunomodulatory actions of MSCs. Although MSCs constitutively secrete PGE2 without the need for activation signals [10, 24, 25], the present results demonstrated accelerated secretion of PGE2 following the increase in inflammatory genes. These results confirm that exogenous PGE2 is sufficient to inhibit TNFα secretion from inflammatory-activated cells [10, 26]. In summary, the increased secretion of PGE2 from cocultured PL-MSCs may decrease TNFα expression in PM-exposed HMEECs, thereby alleviating the inflammatory response.

4.5. Limitations of the Present Study. The PL-MSC coculture showed orchestrated changes in gene profiles in both

![Figure 3](image-url)
HMEECs and PL-MSCs. However, we did not conduct a long-term coculture experiment in this study due to the limitations of the transwell system. From day 4 of the coculture condition, PL-MSCs showed decreased proliferation and morphological changes. VEGF mRNA expression was decreased after day 4 with senescence of PL-MSCs under the coculture conditions. PL-MSCs and HMEECs were incubated in transwells with the culture medium of HMEECs to reflect the clinical conditions of the middle ear. Although there was senescence of PL-MSCs, they exhibited anti-inflammatory effects in coculture conditions. Although the increased protein level of PGE2 was demonstrated using ELISA assay, other trophic gene products were not assessed in the present study. These anti-inflammatory effects may also depend on other factors released by PL-MSCs. A proteomics analysis of coculture media will help reveal the role of other important trophic factors. Different cell-cell interactions could be possible if PL-MSCs are in physical contact with HMEECs. Finally, for applications in the clinical setting, in vivo studies will be needed to examine changes in PL-MSCs further, after long-term transplantation to the middle ear epithelium.

**Data Availability**

The raw data of experiments used to support the findings of this study are available from the corresponding author upon request.

**Conflicts of Interest**

The authors declare that they have no conflict of interest.

**Acknowledgments**

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**Supplementary Materials**

. Supplementary Figure 1: changes in TNFα and COX2 mRNA expression levels of placenta-derived mesenchymal stem cells (PL-MSCs) after coculture with particulate matter- (PM-) treated human middle ear epithelial cells (HMEECs). The mRNA levels of TGFβ, VEGF, and PTGES were increased after coculture with PM-treated HMEECs (transforming growth factor beta (TGFβ), vascular endothelial growth factor (VEGF), and prostaglandin E synthase (PTGES)).

**References**


