MOLECULAR BREEDING OF GENES, PATHWAYS AND GENOMES BY DNA SHUFFLING

Willem P.C. Stemmer

VP R&D, Maxygen, Inc., 515 Galveston Drive, Redwood City, CA 94063

pim_stemmer@maxygen.com; www.maxygen.com

Genomics is yielding a wealth of sequence and functional information that impacts all areas of biotechnology, including pharmaceuticals, vaccines, chemicals/enzymes, and agriculture. However, developing products from these novel genes and pathways typically requires optimization of the gene(s) for the commercially relevant property. Maxygen uses its Molecular Breeding technology to optimize human genes for improved performance as protein pharmaceuticals; typical goals are increasing potency, reducing side effects and modulating specificity and half-life. We also improve the effectiveness of vaccines by breeding of related pathogen sequences. In the chemicals area, we breed genes encoding enzymes to yield better enzymes for a wide range of industrial and biosynthesis applications. We also breed multigene pathways and whole microbial genomes, resulting in microbes with improved fermentation properties. In agriculture, we have used molecular breeding to evolve genes with improved insect resistance and other properties of agricultural interest. In all of these examples functional optimization of the gene(s) was essential. Molecular breeding typically starts within a variety of sequences from natural diversity, such as the same gene from related species. This pool of starting genes is then shuffled by (mostly) homologous recombination, resulting in a large pool of chimeric genes, which is assayed for improved properties in a variety of functional screens. The libraries of chimeric genes that are created by molecular breeding have surprisingly high average functionality but also have a wide range of combinations of properties ranging well beyond that of the parental molecules. Screening of a relatively small number of clones (in some cases only few hundred) from such libraries has yielded commercially important improvements. The small number of screens required to obtain improvements allows us to screen directly in complex assays which closely mimic the final commercial application. We feel that the quality of the screen is essential for successful product development, more so than the number of clones screened. In turn, the quality of the library is critical for successfully using the highest quality screens, which are typically low throughput. When possible, we will follow a lower quality, high throughput screen with a low throughput, high quality screen. I will describe a broad range of examples of genes, pathways and genomes that have been improved by this approach.

REFERENCES

1. Raillard, S., Krebber, A., Chen, Y., Ness, J.E., Bermudez, E., Trinidad, R.,

- Fuller, R., Davis, C., Welch, M., Seffernick, J., Wackett, L.P., Stemmer, W.P.C., and Minshull, J. (2001) *Chem. Biol.* 125, 1–9.
2. Ness et al. (2000) Molecular Breeding: the natural approach to protein design. In *Evolutionary Approaches to Protein Design*. Arnold, F., Ed. *Advances in Protein Chemistry*, Vol. 55. Academic Press, San Diego. pp 261–292.
 3. Minshull, J. and Stemmer, W.P.C. (1999) *Curr. Opin. Chem. Technol.* 3, 284–290.
 4. Chang et al. (1999) *Nat. Biotechnol.* 17, 793–797.
 5. Ness et al. (1999) *Nat. Biotechnol.* 17, 893–896.
 6. Crameri, A., Raillard, S.-A., Bermudez, E., and Stemmer, W.P.C. (1998) *Nature* 391, 288–291.
 7. Patten, P.A., Howard, R.J., and Stemmer, W.P.C. (1997) *Curr. Opin. Biotechnol.* 8, 724–733.
 8. Stemmer, W.P.C. (1995) *Bio/Technology* 13, 549–553.
 9. Stemmer, W.P.C. (1994) *Nature* 370, 389–391.