

Research Article

Optimization and Validation of Indirect ELISA Using Truncated TssB Protein for the Serodiagnosis of Glanders amongst Equines

Harisankar Singha,¹ Praveen Malik,^{1,2} Sachin K. Goyal,¹ Sandip K. Khurana,¹
Chiranjay Mukhopadhyay,³ Vandana K. Eshwara,³ and Raj K. Singh¹

¹ National Research Centre on Equines, Sirsa Road, Hisar, Haryana 125 001, India

² Veterinary Type Culture Collection, National Research Centre on Equines, Sirsa Road, Hisar, Haryana 125 001, India

³ Department of Microbiology, Kasturba Medical College, Manipal University, Manipal, Karnataka 576104, India

Correspondence should be addressed to Praveen Malik; malikphisar@hotmail.com

Received 14 August 2013; Accepted 18 December 2013; Published 3 February 2014

Academic Editors: A. C. Manna and G. A. Rocha

Copyright © 2014 Harisankar Singha et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Objective. To express truncated TssB protein of *Burkholderia mallei* and to evaluate its diagnostic efficacy for serological detection of glanders among equines. **Materials and Methods.** In an attempt to develop recombinant protein based enzyme-linked immunosorbent assay (ELISA), N-terminal 200 amino acid sequences of *B. mallei* TssB protein—a type 6 secretory effector protein—were expressed in prokaryotic expression system. Diagnostic potential of recombinant TssB protein was evaluated in indirect ELISA using a panel of glanders positive ($n = 49$), negative ($n = 30$), and field serum samples ($n = 1811$). Cross-reactivity of the assay was assessed with equine disease control serum and human melioidosis positive serum. **Results.** In comparison to CFT, diagnostic sensitivity and specificity of ELISA were 99.7% and 100%, respectively. **Conclusions.** The indirect ELISA method using the truncated TssB offered safer and more rapid and efficient means of serodiagnosis of glanders in equines. These data highlight the use of TssB as potential diagnostic antigen for serological diagnosis of glanders.

1. Introduction

Glanders is a fatal infectious disease mainly of horses, donkeys, and mules and also communicable to man. The disease is caused by the Gram-negative nonmotile bacillus *Burkholderia mallei*, a close relative of *Burkholderia pseudomallei*—the causal agent of melioidosis in humans. The glanders in equines is characterized by nodular lesions of the lungs and other organs as well as ulcerative lesions of the skin and mucous membranes of the nasal cavity and respiratory passages [1]. Horses are the primary carriers of disease and are largely responsible for transmitting the infection to healthy animals and humans [2]. Lack of prophylactic vaccine together with challenging treatment regime renders *B. mallei* a pathogen of choice for potential biological weapon.

Isolation and identification of the organism from fresh clinical specimens (cutaneous lesions and nasal exudates) is the “gold standard” for diagnosis of glanders. However,

clinical and bacteriological identification of causative agent from equines during the early stages of infection or in clinically inapparent cases is not always attainable. Although complement fixation test (CFT) and mallein test are officially recognized by OIE for international trade of equidae [3], species specific biasness of the test and need of technical expertise are the major limitations of these assays. Different ELISA formats including avidin-biotin dot ELISA [4], cELISA [5], and indirect ELISA [6] have been developed for diagnosis of glanders. However, these assays either use crude or purified bacterial fractions as antigens thereby affect the test specificity.

The type six secretion system (T6SS) is conserved in numerous Gram-negative pathogens and symbionts that interact closely with eukaryotic cells [7, 8]. The *B. mallei* T6SS gene cluster 1 (BMAA0730 to BMAA0744) and homologous gene in *B. pseudomallei* (BPSS1496–1512) are important for actin-based motility, multinucleated giant cell (MNGC)

formation, intracellular growth in murine macrophages, and virulence in hamsters [7, 9]. The TssB protein encoded by BMAA0743 locus of *B. mallei* T6SS-1 is a secretory effector-like protein and thus the TssB protein may be used as diagnostic antigen for serological detection of glanders. Therefore, the objectives of present study were to express immunodominant epitope of TssB protein of *B. mallei* and to evaluate its diagnostic efficacy for serological detection of glanders among equines.

2. Materials and Methods

2.1. Customization and Synthesis of tssB Gene. The 1500 bp sequences of *tssB* gene of *B. mallei* (locus tag BMAA0743) were retrieved from National Center for Biotechnology Information (NCBI) database and antigenicity plot of the translated sequence was determined by Kolaskar and Tongaonkar method [10] using CLC work bench v 6.1 (<http://www.clcbio.com/>). Based on antigenicity index, first 600 bases of the *tssB* gene encoding N-terminal 200 amino acid sequences were selected for expression. The codons of the selected sequences were optimized according to codon usage in *E. coli* K-12 strain. The *Bam*HI and *Hind*III restriction sites were placed at 5' and 3' end of the sequences, respectively, for ease of downstream application. The customized *tssB* gene was commercially synthesized in pUC57 vector (BioBasic Inc., Canada).

2.2. Production of Recombinant TssB Protein. The gel purified *Bam*HI and *Hind*III restricted 600 bp *tssB* gene was directionally ligated to pQE30 expression vector (Qiagen, Germany) and transformed into chemically competent *E. coli* M15 cells. Positive clones expressing recombinant TssB protein were screened by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE). Single clone showing maximum expression of TssB protein after 4 h of induction by 1 mM IPTG (isopropyl-D-thiogalactopyranoside) was selected for production of recombinant protein. The N-terminal histidine-tagged TssB protein was purified by Ni⁺-NTA agarose (Qiagen, Germany) as per manufacturer's instruction. Eluates containing recombinant TssB protein were pooled and dialyzed against 0.01 M phosphate-buffered saline (PBS; pH 7.2). The protein concentration was determined using a bicinchoninic acid (BCA) protein assay kit (Merck Millipore, India). Protein in 1 mL aliquots of 0.5 mg/mL concentration was kept in -20°C for future use.

2.3. Serum Panel. Diagnostic performance of the recombinant TssB protein was evaluated using a panel of glanders positive and negative equine serum samples maintained in the serum repository of National Research Centre on Equines (NRCE), Hisar, India. A total of 49 glanders positive equine serum samples collected during glanders outbreak between 2006 and 2011 were used as positive control [11]. Negative control consists of 40 serum samples of healthy equines which did not show any clinical signs of glanders and were consistently negative by CFT. The cross-reactivities of recombinant TssB protein were assessed with melioidosis

positive human serum. Serum samples from human patient suffering from melioidosis ($n = 6$) and septicemia caused by other infectious bacteria ($n = 10$) were used as melioidosis positive control and disease control serum, respectively. The human serum samples were collected at Kasturba Medical Hospital, Manipal, Karnataka, India. The serum panel also contained 14 samples from equine infected with bacteria like *Streptococcus equi* subspecies *equi* ($n = 5$), *Streptococcus equi* subspecies *zooepidemicus* ($n = 5$), and *Pseudomonas aeruginosa* ($n = 5$) for determination of the specificity of the ELISA. Equine serum samples ($n = 1811$) received from different race clubs and private farms or collected from field for serosurveillance of equine disease were used as test samples for diagnostic evaluation of the indirect ELISA.

2.4. Determination of TssB Specific Antibody by Western Blot. Specific reactivity of recombinant TssB protein to *B. mallei* antibody was determined by western blot. Approximately, 600 ng TssB protein/well was transferred to PVDF membrane. The unbound surface of the membrane was blocked by PBS containing 10% skimmed milk for overnight at 4°C. The membrane was cut into strips, washed with PBS containing 0.05% Tween 20 (PBS-T), and incubated with 1:200 dilution of serum for 2 h at 37°C. The strips were washed with PBS-T and incubated with 1:10000 dilutions of HRP conjugated rabbit anti-horse or anti-human IgG antibody as per need (Sigma-Aldrich, USA) for 1 h at 37°C. The reaction was developed with diaminobenzidine (6 mg/10 mL 50 mM Tris buffer pH 7.6) and 30 μ L of 30% H₂O₂. The reaction was stopped by washing the membrane in distilled water and results were recorded.

2.5. Optimization of Indirect ELISA. The optimal concentration of ELISA reagents (TssB protein, serum, and secondary antibody dilution) was determined by checkerboard titration method using known glanders positive and negative equine serum. The optimal ELISA reagents concentration was assumed to be for those showing the highest discrimination between positive and negative serum. After optimization of the assay, 1811 equine serum samples were assayed by ELISA. Pool of four positive and four negative serum samples were included in each ELISA plate along with field serum samples for monitoring the accuracy of the assay.

In brief, 96-well microtiter plates (Greiner Bio One, USA) were coated with TssB protein for overnight at 4°C followed by washing with PBS-T and blocking with 6% skimmed milk in PBS-T for 1 h at 37°C. Serum samples were diluted in dilution buffer (2% (w/v) skimmed milk in PBS-T) and 100 μ L of diluted serum was added in each well. The plates were incubated for 1 h at 37°C and washed five times with PBS-T. The diluted anti-horse conjugate (100 μ L) was added in each well and incubated for 1 h at 37°C. Following washing steps, 100 μ L of substrate solution containing 200 μ mol of orthophenylene-diamine in citrate perborate buffer (Sigma-Aldrich, USA) was added in each well and plate was kept in dark at 37°C for 10 min. Finally, the reaction was stopped by addition of 50 μ L of 1 M H₂SO₄ and the absorbance was read

```

BM ATCC23344 atggaaggcgaacacctgcaatccccgaagcagcagcagcgcgccgagcgcagcgcgccagtcccccgcctcgtctgctcgacgagctg 90
BPS K96243 ..... 90
Synthetic tssB ..... 90
BM ATCC23344 atcgaggccgcgcgctgaagcgcgacgaagcgcataccgatcacgcgccacggcatccaggcgttcgctcgcgcatctggcgaagccc 180
BPS K96243 ..... 180
Synthetic tssB ..t..a...c...c...t..t...t..c...t..t...t...t...a...a...t...a..g 180
BM ATCC23344 aagcgcggatcgagaccgtgagccaggcgcagatcgacgacatgatcgccgagatcgaccgaagctgtgccggcagatcgacgagcgc 270
BPS K96243 ..... 270
Synthetic tssB ..a..t.....a.....atc.....t..c..t..t.....t.....t..a.....t.....t..... 270
BM ATCC23344 ctccatgacccggcattccagcaactcgaatcgacgtggcgtcgtcgaagtttctcgtcgatcgaacggatttccgcgagaacgtgaag 360
BPS K96243 ..... 360
Synthetic tssB ..g.....g...g...agc..t.....c...a..c..g..t..c..c...c...t...a..t..t..a 360
BM ATCC23344 gttcagattctcgacgtcggcaaacggcgtgttcgacgacttcgaggattcgcggacatcacgaaatccgggctgtaccagaaggtc 450
BPS K96243 ..... 450
Synthetic tssB ..g.....c..g..t..t..t...c..t.....t.....c..t..g..t.....gag...t.....a..t 450
BM ATCC23344 tatacggccgagtagcggcaattcggcggccagccgatcggcgcgatcgtcgcgaactacacgttcgggcccggcgcgcagacgtcaag 540
BPS K96243 ..... 540
Synthetic tssB .....t..t..a..t...g..t..t...a...t.....t..t.....t..t..g..t.....t..g..a 540
BM ATCC23344 ctgctgcagtagcgtcggcagcagctcggcgcgatggcgcatacccggttcacgcggcggcg 600
BPS K96243 ..... 600
Synthetic tssB .....g..a...c..t.....c..c..a.....a..t 600
    
```

FIGURE 1: Sequence alignment of 5' terminal 600 bp tssB gene of *B. mallei*, *B. pseudomallei*, and customized tssB gene. Identical sequences are represented by dot. Nonidentical sequences at third base of codon have been modified during the process of codon optimization.

at 492 nm in a microtiter plate reader (Titertek Multiskan, Finland).

2.6. Complement Fixation Test (CFT). ELISA results were compared with CFT which was performed as per standard protocol described by the Office International des Épipizooties (OIE) [3] using commercial kits (Biovet, a.s., Czech Republic) for detection of *B. mallei* antibodies. The CFT titer of 1:8 and above was considered positive for glanders.

2.7. Data Analysis. The optimum cutoff point of ELISA was determined by receiver operative curves (ROC) analysis using normalized OD₄₉₂ value (PP%) as described elsewhere [12, 13]. For determination of the cutoff point, absorbance data of ELISA (O.D. values) were analyzed using Windows 7 Microsoft Excel programme (Microsoft Corp, Redmond, WA) and were normalized using the following formula:

$$\begin{aligned}
 &\text{Percent positivity (PP\%)} \\
 &= \frac{(\text{OD}_{492} \text{ sample serum} - \text{OD}_{492} \text{ negative control})}{(\text{OD}_{492} \text{ positive control} - \text{OD}_{492} \text{ negative control})} \\
 &\times 100\%.
 \end{aligned}
 \tag{1}$$

Test samples with PP% value greater than the cutoff point were treated as positive. Relative sensitivity and specificity of the assay were determined by following calculation and the results were expressed as percentages:

$$\begin{aligned}
 \text{Sensitivity} &= \frac{\text{True positive}}{(\text{True positive} + \text{False negative})} \\
 \text{Specificity} &= \frac{\text{True negative}}{(\text{True negative} + \text{False positive})}.
 \end{aligned}
 \tag{2}$$

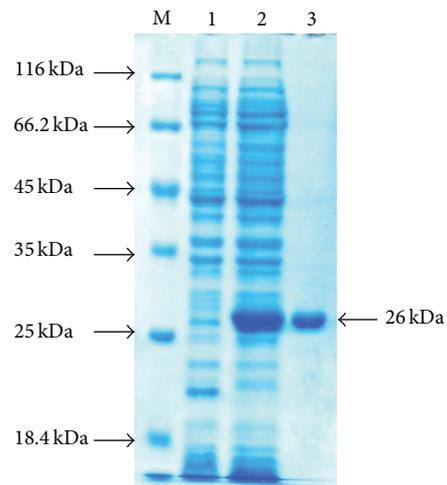


FIGURE 2: SDS-PAGE analysis of recombinant TssB protein. Lane M: protein molecular weight marker (Fermentas, number SM0431), Lane 1: uninduced control, 2: IPTG induced cell lysate, and 3: Nickel-NTA affinity column purified protein.

3. Result

3.1. Expression and Purification of Recombinant TssB Protein. The codon optimized 600 bp immunodominant fragment of tssB gene is shown in Figure 1. To reduce the G + C content of the targeted gene fragment, only third base of the codons was changed which results in changes in DNA sequence but the amino acid sequence remains unaltered. Besides *B. mallei*, the deduced amino acid sequence of truncated TssB also showed 100% identity with the protein homologue present in *Burkholderia pseudomallei* (data not shown). The recombinant *E. coli* carrying pQE30: tssB cassette expressed 26 kDa proteins upon IPTG induction (Figure 2 Lane 2). SDS-PAGE analysis of eluted fractions revealed that protein

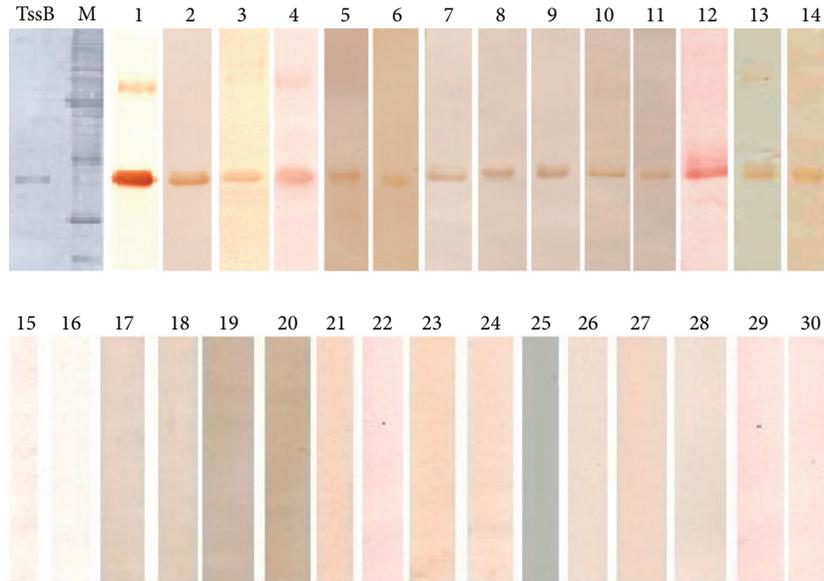


FIGURE 3: Specific immune reactivity of recombinant TssB protein in western blot. Lanes TssB and M: amido black stained recombinant TssB protein and protein molecular weight marker, respectively, upon transfer on PVDF membrane. Lanes 1–14: Glanders positive equine serum, Lanes 15–18: negative control serum, Lanes 19–22: melioidosis positive human serum. Equine disease control serum: Lanes 23–25: *Streptococcus equi* infection (strangles); Lane 26–28: *Streptococcus zooepidemicus* infection; Lane 29–30: and *Pseudomonas aeruginosa* infection.

was purified to homogeneity and no contaminating protein band was found.

3.2. Western Blot Analysis. In western blot, glanders positive equine serum gave a positive reaction band at 26 kDa position. Negative control serum, melioidosis (human), and other equine disease control serums showed no reactivity with TssB protein (Figure 3).

3.3. Diagnostic Efficacy of Indirect ELISA. Optimal concentration of ELISA reagents was found as follows: 150 ng of purified TssB protein/well, 1/200 serum dilution, and 1/15000 conjugate dilution. The percent positivity percentage (PP%) value of 30.0 was calculated as cutoff point (Figure 4); serum samples showing PP% greater than or equal to 30.0 and less than that were regarded as positive and negative, respectively. Out of 1811 equine serum samples ($n = 1500$ horses, $n = 150$ donkeys, and $n = 161$ mules), 4 samples were found false positive in ELISA (Figure 5). However, there were none that were negative in ELISA but positive in the CFT. The PP value of melioidosis and other equine disease control serum samples was below the cutoff. The relative specificity and sensitivity of the ELISA were 99.7% and 100%, respectively.

4. Discussion

Glanders is a notifiable disease in India and presently follows the “test and stamp-out policy” as the only control measure. The OIE approved mallein test and complement fixation test (CFT)—commonly used for the diagnosis of glanders—suffer from lack of sensitivity particularly in clinically advanced

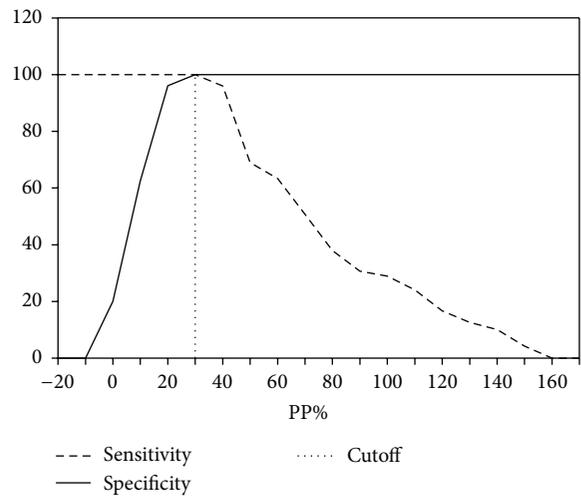


FIGURE 4: Determination of cutoff point of indirect enzyme-linked immunosorbent assay.

cases [14, 15], and additionally mallein test causes transient seroconversions in healthy uninfected animals [16]. The CFT for glanders is so far the only officially recognized serological laboratory test for international trade of equidae and remains the preferred diagnostic tool in eradication programme. However, complement fixation test (CFT) has drawback of producing notable number of nonspecific results especially with donkeys and mules serum [14, 17].

Partially purified or crude antigen preparations of *B. mallei* were evaluated in Rose Bengal plate agglutination test

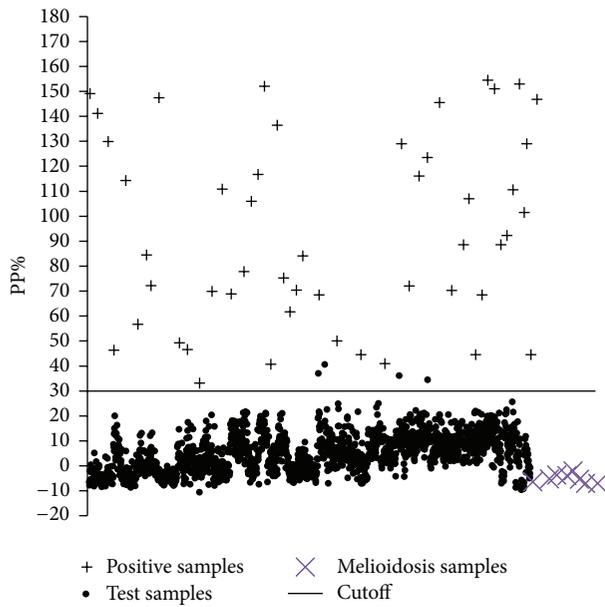


FIGURE 5: Distribution of PP% value of positive and negative serum samples. The horizontal line represents the cutoff value [percent positivity (PP% = 30)]. Glanders positive serum is indicated by + sign, dot represents the test serum samples, and melioidosis positive human serum is indicated by × symbol.

(RBT) [15], western blot assays [17], a competitive ELISA [5], and indirect ELISA [6]. The presence of cross-reacting epitopes in antigen preparation affects the specificity of these assays. Further, requirement of strict biosafety measures and the possible laboratory-acquired infection of *B. mallei* during the preparation of antigens is still a serious drawback. Although many PCR techniques for detection of *B. mallei* DNA have been described [18, 19], specificity of PCR detection method has not been tested in clinical samples so far. Moreover, PCR techniques have got limited acceptance in surveillance of equine populations, owing to the need for containment laboratory facilities, trained personnel, and financial resources.

Previously, Defence Research and Development Establishment (DRDE), Gwalior, and our lab in collaborative research project have reported the recombinant protein based ELISA for glanders diagnosis [20, 21]. In a continuous effort, we describe here the homogeneous production of truncated TssB protein of *B. mallei* and evaluated the diagnostic potentiality of the recombinant TssB protein in indirect ELISA. Cross-reactivity experiment with human melioidosis serum and equine disease control serum showed that the TssB protein is highly specific to detect *B. mallei* antibodies. In view of the high percentage of sequence identity of *B. mallei* TssB protein with *B. pseudomallei* protein homologue, nonreactivity of melioidosis positive serum with TssB observed in this study is really surprising. The similar findings were previously observed by Pal et al. [21] where *B. mallei* hypothetical protein BMA10229.0375 did not cross-react with melioidosis serum despite showing considerable sequence identity (~91%) with *B. pseudomallei* homologue proteins. Nonreactivity of TssB

protein with melioidosis serum of Indian origin may be substantiated by two facts: (i) genetic diversity of Indian *B. pseudomallei* isolates from other strains originated from melioidosis endemic region like Thailand and Australia [22] and (ii) differential regulation and function of TssB in *B. mallei* and *B. pseudomallei* infection [9, 23]. Therefore, on the basis of information obtained from literature and findings of the present experiment, it appears that expression and secretion pattern of TssB may be different in clinical infection of *B. mallei* and *B. pseudomallei* which may lead to variation in antibody production against the TssB protein.

The indirect ELISA method using recombinant TssB protein offered safer and more rapid and efficient means of serodiagnosis of glanders in equines. Further validation of the assay with equine serum from glanders endemic and glanders-free countries will be useful for wider acceptance of the TssB protein based ELISA.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

The research was funded by Life Science Research Board, Defence Research and Development Organisation, Ministry of Defence, Government of India (LSRB-218/SH&DD/2010). The authors thank Sh. Sitaram, Technical Officer, and Sh. Gurudutt, SSS, for their technical help in processing of serum samples.

References

- [1] D. DeShazer and D. Waag, "Glanders: new insights into an old disease," in *Biological Weapons Defense: Infectious Diseases and Counterbioterrorism*, L. Lindler, F. Lebeda, and G. W. Korch, Eds., pp. 209–237, The Humana Press, Totowa, NJ, USA, 2004.
- [2] E. E. Galyov, P. J. Brett, and D. Deshazer, "Molecular insights into *Burkholderia pseudomallei* and *Burkholderia mallei* pathogenesis," *Annual Review of Microbiology*, vol. 64, pp. 495–517, 2010.
- [3] Office International des Epizooties (OIE), *Glanders*, Chapter 2. 5. II. *OIE Terrestrial Manual*, OIE, Paris, France, 2008.
- [4] R. D. Verma, J. K. Sharma, K. S. Venkateswaran, and H. V. Batra, "Development of an avidin-biotin dot enzyme-linked immunosorbent assay and its comparison with other serological tests for diagnosis of glanders in equines," *Veterinary Microbiology*, vol. 25, no. 1, pp. 77–85, 1990.
- [5] L. D. Sprague, R. Zachariah, H. Neubauer et al., "Prevalence-dependent use of serological tests for diagnosing glanders in horses," *BMC Veterinary Research*, vol. 5, article 32, 2009.
- [6] J. A. A. Teles, A. C. Campos, K. P. C. Silva et al., "Standardization and evaluation of an indirect ELISA for the serological diagnosis of glanders in horses," *Pesquisa Veterinária Brasileira*, vol. 32, no. 9, pp. 838–842, 2012.
- [7] M. A. Schell, R. L. Ulrich, W. J. Ribot et al., "Type VI secretion is a major virulence determinant in *Burkholderia mallei*," *Molecular Microbiology*, vol. 64, no. 6, pp. 1466–1485, 2007.

- [8] S. Schwarz, T. E. West, F. Boyer et al., “*Burkholderia* type vi secretion systems have distinct roles in eukaryotic and bacterial cell interactions,” *PLoS Pathogens*, vol. 6, no. 8, Article ID e1001068, 2010.
- [9] Y. Chen, J. Wong, G. W. Sun, Y. Liu, G.-Y. G. Tan, and Y.-H. Gan, “Regulation of type VI secretion system during *Burkholderia pseudomallei* infection,” *Infection and Immunity*, vol. 79, no. 8, pp. 3064–3073, 2011.
- [10] A. S. Kolaskar and P. C. Tongaonkar, “A semi-empirical method for prediction of antigenic determinants on protein antigens,” *FEBS Letters*, vol. 276, no. 1-2, pp. 172–174, 1990.
- [11] P. Malik, H. Singha, S. K. Khurana et al., “Emergence and re-emergence of glanders in India: a description of outbreaks from 2006 to 2011,” *Veterinaria Italiana*, vol. 48, no. 2, pp. 167–178, 2012.
- [12] S. Siegel and N. J. Castellan, *Nonparametric Statistics for the Behavioral Sciences*, McGraw-Hill, New York, NY, USA, 2nd edition, 1998.
- [13] I. A. Gardner and M. Greiner, *Advanced Method for Test Validation and Interpretation in Veterinary Medicine*, Freie Universitat, Berlin, Germany, 2000.
- [14] H. Neubauer, L. D. Sprague, R. Zacharia et al., “Serodiagnosis of *Burkholderia mallei* infections in horses: state-of-the-art and perspectives,” *Journal of Veterinary Medicine B*, vol. 52, no. 5, pp. 201–205, 2005.
- [15] A. Naureen, M. Saqib, G. Muhammad, M. H. Hussain, and M. N. Asi, “Comparative evaluation of Rose Bengal plate agglutination test, mallein test, and some conventional serological tests for diagnosis of equine glanders,” *Journal of Veterinary Diagnostic Investigation*, vol. 19, no. 4, pp. 362–367, 2007.
- [16] J. M. Hagebock, L. K. Schlater, W. M. Frerichs, and D. P. Olson, “Serologic responses to the mallein test for glanders in solipeds,” *Journal of Veterinary Diagnostic Investigation*, vol. 5, no. 1, pp. 97–99, 1993.
- [17] M. C. Elschner, H. C. Scholz, F. Melzer et al., “Use of a Western blot technique for the serodiagnosis of glanders,” *BMC Veterinary Research*, vol. 7, article 4, 2011.
- [18] J. R. Bowers, D. M. Engelthaler, J. L. Ginther et al., “BurkDiff: a real-time PCR allelic discrimination assay for *Burkholderia pseudomallei* and *B. mallei*,” *PLoS ONE*, vol. 5, no. 11, Article ID e15413, 2010.
- [19] S. F. Koh, S. T. Tay, R. Sermswan, S. Wongratanacheewin, K. H. Chua, and S. D. Puthucheary, “Development of a multiplex PCR assay for rapid identification of *Burkholderia pseudomallei*, *Burkholderia thailandensis*, *Burkholderia mallei* and *Burkholderia cepacia* complex,” *Journal of Microbiological Methods*, vol. 90, no. 3, pp. 305–308, 2012.
- [20] S. Kumar, P. Malik, S. K. Verma et al., “Use of a recombinant *Burkholderia* intracellular motility a protein for immunodiagnosis of glanders,” *Clinical and Vaccine Immunology*, vol. 18, no. 9, pp. 1456–1461, 2011.
- [21] V. Pal, S. Kumar, P. Malik, and G. P. Rai, “Evaluation of recombinant proteins of *Burkholderia mallei* for serodiagnosis of glanders,” *Clinical Vaccine and Immunology*, vol. 19, no. 8, pp. 1193–1198, 2012.
- [22] C. Mukhopadhyay, M. Kaestli, K. E. Vandana et al., “Molecular characterization of clinical *Burkholderia pseudomallei* isolates from India,” *American Journal of Tropical Medicine and Hygiene*, vol. 85, no. 1, pp. 121–123, 2011.
- [23] G. Shalom, J. G. Shaw, and M. S. Thomas, “*In vivo* expression technology identifies a type VI secretion system locus in *Burkholderia pseudomallei* that is induced upon invasion of macrophages,” *Microbiology*, vol. 153, no. 8, pp. 2689–2699, 2007.



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

