Research Article

Lymphocyte Oxidative Stress/Genotoxic Effects Are Related to Serum IgG and IgA Levels in Coke Oven Workers

Meili Gao,¹ Yongfei Li,² Aquan Zheng,³ Xiaochang Xue,⁴ Lan Chen,⁵ and Yu Kong¹

¹ Department of Biological Science and Engineering, Institute of Mitochondrial Biology and Medicine, The Key Laboratory of Biomedical Information Engineering of Ministry of Education, School of Life Science and Technology, Xi'an Jiaotong University, Xianning West Road 28, Xi'an, Shaanxi 710049, China

² School of Materials and Chemical Engineering, Xi'an Technological University, Xi'an 710032, China

³ School of Science, Xi'an Jiaotong University, Xi'an, Shaanxi 710049, China

⁴ Department of Biopharmaceutics School of Pharmacy, State Key Laboratory of Cancer Biology, Fourth Military Medical University, Xi'an, Shaanxi 710032, China

⁵ Center of Shared Experimental Facilities, The Key Laboratory of Biomedical Information Engineering of Ministry of Education, School of Life Science and Technology, Xi'an Jiaotong University, Xi'an, Shaanxi 710049, China

Correspondence should be addressed to Yu Kong; kynk@sina.com

Received 17 January 2014; Revised 7 June 2014; Accepted 9 June 2014; Published 20 July 2014

Academic Editor: Raimo Pohjanvirta

Copyright © 2014 Meili Gao et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

We investigated oxidative stress/genotoxic effects levels, immunoglobulin levels, polycyclic aromatic hydrocarbons (PAHs) levels exposed in 126 coke oven workers and in 78 control subjects, and evaluated the association between oxidative stress/genotoxic effects levels and immunoglobulin levels. Significant differences were observed in biomarkers, including 1-hydroxypyrene levels, employment time, percentages of alcohol drinkers, MDA, 8-OHdG levels, CTL levels and CTM, MN, CA frequency, and IgG, IgA levels between the control and exposed groups. Slightly higher 1-OHP levels in smoking users were observed. For the dose-response relationship of IgG, IgA, IgM, and IgE by 1-OHP, each one percentage increase in urinary 1-OHP generates a 0.109%, 0.472%, 0.051%, and 0.067% decrease in control group and generates a 0.312%, 0.538%, 0.062%, and 0.071% decrease in exposed group, respectively. Except for age, alcohol and smoking status, IgM, and IgE, a significant correlation in urinary 1-OHP and other biomarkers in the total population was observed. Additionally, a significant negative correlation in genotoxic/oxidative damage biomarkers of MDA, 8-OH-dG, CTL levels, and immunoglobins of IgG and IgA levels, especially in coke oven workers, was found. These data suggest that oxidative stress/DNA damage induced by PAHs may play a role in toxic responses for PAHs in immunological functions.

1. Introduction

Coke oven workers are constantly exposed to coke oven emissions which are toxic chemicals, especially polycyclic aromatic hydrocarbons (PAHs). PAHs are formed during combustion of fossil fuels and typified by the 1-hydroxypyrene levels. The 1-hydroxypyrene level has been shown to be a good marker for total PAHs exposure [1, 2]. Some of PAHs are carcinogenic due to their metabolites and their ability to generate genetic damage and further oxidative DNA damage through the production of reactive oxygen species during metabolism [3, 4].

Of many indicators for oxidative DNA damage, 8-hydroxy-2'-deoxyguanosine (8-OHdG) represents an

important product from oxidative damage to DNA. 8-OHdG is formed in a promutagenic DNA lesion induced by the reaction of hydroxyl radicals with guanosine at the C8 site in DNA. A growing number of surveys and occupational studies indicated that elevated levels of 8-OHdG in DNA from leukocytes or excretion in urine have been observed in PAHs exposure of smokers and workers [5–8]. Malondialdehyde (MDA), which is an end product of the oxidation of polyunsaturated fatty acids and can determine the degree of lipid peroxidation, has been used as a marker for oxidative stress [9].

It has been reported that low level PAH exposure causes DNA single strand breakage, the formation of DNA damages,

and immunotoxicity [10–12]. Immunotoxicity can change lymphocytic subpopulation in peripheral blood and serum immunoglobulin levels in coke oven workers exposed to PAHs [12, 13]. As for genotoxic risk factors, the comet assay, micronucleus (MN) assay, and chromosomal aberrations (CA) assay have been used to evaluate the biomarkers of early biological effects [14]. The comet assay has been found to be a very sensitive method for measuring DNA damage. The MN test was found to provide a cytogenetic parameter and allowed the detection of both clastogenic and aneugenic agents [14]. Chromosomal damage has also been found to provide CA such as chromosome breakage, chromosome deletion, and chromosome polyploid [13, 14].

In the present study, we investigated if there was any relation between the levels of MDA, 8-OHdG, and genotoxic damages and immunoglobulin levels in serum and lymphocytes of workers exposed to coke oven emission.

2. Materials and Methods

2.1. Study Subjects. The 126 coke oven workers and 78 non-coke-oven workers who were all males and worked in the same steel company in northern China were studied in this paper. These 126 coke oven workers were in active service at the time of the study, were employed for at least 6 months, and were recruited as the exposed group. The 78 non-coke-oven workers were staff members of the offices and hospitals of the same steel company and served as the control group. The workers exposed to known mutagenic agents, such as radiotherapy and chemotherapy in the last 3 months, were excluded. Questionnaires were administered by trained interviewers to collect information on demographic information, including age, length of employment, smoking, and alcohol habits. Individuals who had smoked for 3 months were considered as smokers. Those who drank more than twice a week in the last six months were classified as drinkers. Blood samples were collected at the end of these days. In the morning, 5 mL fasting venous blood and 10 mL urine samples were collected from each subject for further analysis. The study was approved by the Ethics Committee of Xi'an Jiaotong Medical College and was performed in accordance with the Helsinki Declaration (1964).

2.2. 1-Hydroxypyrene in Urine Assay. Urine 1-hydroxypyrene (1-OHP) was measured by the method described by Jongeneelen and Anzion [15] and Siwińska et al. [16]. Briefly, aliquots of 10 mL urine samples were enzymatically deconjugated and transferred to primed C18 octadecyl cartridges (Beckman, USA). Then the samples were washed with 10 mL of water and eluted with 9 mL of methanol. The components of the eluate was subjected to a high pressure liquid chromatography of the Waters Alliance 2695 (Waters, USA) with the XAqua C18, 150 × 4.1 mm column. A fluorescence detector of F1000 (Hitachi, Japan) was quantitatively assayed 1-hydroxypyrene concentration. The wavelengths of excitation and emission were 229 nm and 400 nm, respectively. The concentrations of 1-OHP were normalised to urinary creatinine. Urinary creatinine concentrations were assayed by

a standard colorimetric method with the picric acid reaction and absorption at 520 nm [17].

2.3. Serum Biomarkers Assay. Blood samples were obtained after an overnight fast. Samples (3 mL) of venous blood were incubated in a water bath for 30 minutes at 37°C and centrifuged at 4500 rpm for 10 minutes. The supernatants were stored at –70°C. The levels of serum MDA represented lipid peroxidation product were measured spectrophotometrically by a modification of the method described by Buege and Aust [18]. The spectrophotometric measurements were done with Shimadzu UV-1208 spectrophotometer (Japan). The MDA concentration was calculated using its extinction coefficient of 1.56×10^5 L/mol cm at 535 nm. Enzyme linked immunosorbent assay (ELISA) was used to measure the concentrations of 8-OHdG, in accordance with the procedures described in the assay kit (Sigma, USA). Serum IgA, IgE, IgG, and IgM levels were measured by a nephelometric method [19].

2.4. Comet Assay, Micronuclei Test, and Chromosomal Aberrations Assay. Venous blood was collected from all subjects using heparinized syringes. Lymphocyte cultures were set up by adding 0.5 mL of heparinised blood to 4.5 mL of chromosome medium (RPMI 1640, Gibco) supplemented with 20% heat inactivated fetal bovine serum (Gibco), antibiotics (penicillin and streptomycin), and L-glutamine. Lymphocytes were stimulated by 1% phytohaemagglutinin (Gibco). The comet assay, micronuclei test, and CA assay were performed to assay the genotoxic damage of lymphocytes in workers. DNA damage to lymphocytes was assayed by the alkaline single-cell gel technique [20, 21]. Images of 25 randomly selected lymphocytes were analyzed for each sample. The slides were examined using a Comet 4.0 image analysis system fitted with an Olympus BX50 fluorescence microscope. For the collected cell sample of each subject, images of 50 randomly selected cells were analyzed for each sample. The mean level of comet tail length (CTL) and comet tail moment (CTM) of the 50 cells was as the value of the sample. The determination of micronuclei in lymphocytes was performed as described by Fenech and Morley [22]. The cultures were incubated at 37°C for 72 hours, and 44 hours after the initiation of cultures, cytochalasin-B (Sigma) at a concentration of 6 mg/mL was added to arrest cytokinesis. MN slides were stained with 10% Giemsa in phosphate buffer. A total of 1000 binucleates cells with well-preserved cytoplasm were examined by a well-trained research assistant under double blindness for each subject on coded slides throughout the study. The data are reported as the occurrence rate of ‰ micronucleated cells for each subject. The CA assay was according to the method described by Carrano and Natarajan [23]. Briefly, colcemid was added to arrest the cells in metaphase after incubating the cultures for 48 hours at 37°C; then cells were collected by centrifugation, resuspended in a prewarmed hypotonic solution (0.075 M KCl) for 25 minutes, and fixed in acetic acid : methanol (1 : 3, v/v). Air dried preparations were made and the slides were stained with Giemsa. A total of 100 well spread metaphases containing 46 ± 1 chromosomes was examined for each subject on coded slides; otherwise the

TABLE 1: Demographic characteristics of workers in the control and exposed groups.

Variables	Control group (78)	Exposed group (126)	P value
1-OHP ($\mu\text{mol/mol creatine}$)	0.55 ± 0.28	$9.36 \pm 2.14^{***}$	$<0.001^a$
Age (year)	32.4 ± 4.9	33.6 ± 5.2	0.103^a
Employment time (year)	12.7 ± 2.5	$14.1 \pm 2.7^{***}$	$<0.001^a$
Current smokers, yes (%)	41 (52.6%)	79 (62.7%)	0.153^b
Alcohol users, yes (%)	18 (23.1%)	57 (45.2%)	0.001^b
MDA (nmol/mL)	1.23 ± 0.37	$1.97 \pm 0.61^{***}$	$<0.001^a$
8-OH-dG (nmol/mol creatinine)	1.29 ± 0.15	$2.20 \pm 0.19^{***}$	$<0.001^a$
CTL (μm)	3.16 ± 0.58	$7.49 \pm 0.89^{***}$	$<0.001^a$
CTM	0.85 ± 0.23	$10.48 \pm 3.52^{***}$	$<0.001^a$
MN (‰)	1.83 ± 0.51	$2.78 \pm 0.63^{***}$	$<0.001^a$
CA (N, AR%)	10, 12.8%	36, 28.6%	0.009^b
IgG (g/L)	0.36 ± 0.11	$0.12 \pm 0.03^{***}$	$<0.001^a$
IgA (g/L)	2.09 ± 0.44	$1.66 \pm 0.44^{***}$	$<0.001^a$
IgM (g/L)	1.76 ± 0.69	1.62 ± 0.72	0.172^a
IgE (IU/mL)	173.90 ± 30.24	167.76 ± 43.16	0.273^a

1-OHP: 1-hydroxypyrene; CTL: comet tail length; CTM: comet tail moment; MN: micronucleus; CA: chromosomal aberrations. Values are shown mean $\pm$ SD except where indicated. ^awhen compared with control by Mann-Whitney test, ^bwhen compared with control by chi-square test.

*** $P < 0.001$.

TABLE 2: 1-OHP levels by smoking use status in control and exposed group.

	Control group		Exposed group	
	Smoking (No)	Smoking (yes)	Smoking (No)	Smoking (yes)
<i>n</i>	37	41	47	79
1-OHP level	0.48 ± 0.21	0.57 ± 0.31	8.62 ± 2.12	9.58 ± 3.65
<i>P</i> value ^a	0.142		0.096	

^awhen compared between nonsmoking and smoking users control and exposed group by chi-square test.

data were discarded. The data are reported as the occurrence number and occurrence rate of % CA for each group.

2.5. Statistical Analysis. All analyses were carried out using the Statistical Package for Social Sciences (SPSS13.0). We used Mann-Whitney and Pearson chi-square tests to compare the demographics and lifestyle variables between the exposure and control groups. We used the Mann-Whitney test to compare values of biomarkers between the exposure and control groups. Spearman's rank correlation coefficient was calculated to evaluate the relations between MDA, CTL, IgG, and the impact of independent variables (occupational PAHs exposure, age, length of employment, smoking, and alcohol drinking) on dependent variables. Pearson correlation was calculated to evaluate the relations between MDA and 8-OH-dG level, 8-OH-dG and CTL level, MDA, and IgG, IgA level. All statistical tests were two-sided with a significant level of $P < 0.05$.

3. Results

3.1. Demographic Characteristics of Study Subjects. Table 1 shows the characteristics of study subjects by work site.

Exposed workers were 1 year older (mean age) than control subjects. Significant differences were observed in 1-OHP levels, employment time, percentages of alcohol drinkers, MDA, 8-OHdG levels, CTL level and CTM, MN, CA frequency, IgG, and IgA between the control and exposed groups, but no significance for age, current smokers distribution, and IgM and IgE levels.

Table 2 shows the results of urinary 1-OHP relative to smoking status in exposed workers and controls, after adjustment for exposure concentration. A slight higher of 1-OHP levels in smoking users were observed both in control and exposed groups, but this was not significant ($P = 0.142$ in control and $P = 0.096$ in exposed group, resp.).

3.2. Dose-Response Relationships of Urinary 1-OHP with Immunoglobins and Correlation between 1-OHP and Other Biomarkers. The dose-response relationship of IgG, IgA, IgM, and IgE by 1-OHP was analyzed as in Table 3. The values of IgG, IgA, IgM, and IgE and 1-OHP were all ln-transformed in the multiple linear regression models. For the control group, each one percentage increase in urinary 1-OHP generates a 0.109% ($P = 0.353$), 0.472% ($P < 0.001$), 0.051% ($P = 0.658$), and 0.067% ($P = 0.565$) decrease in IgG, IgA,

TABLE 3: Dose-response relationship with the levels of 1-OHP with immunoglobins.

1-OHP level	IgG (g/L)	IgA (g/L)	IgM (g/L)	IgE (IU/mL)
	β	β	β	β
	(95% CI)	(95% CI)	(95% CI)	(95% CI)
	<i>P</i>	<i>P</i>	<i>P</i>	<i>P</i>
Control group	−0.109	−0.472	−0.051	−0.067
	(−0.359–0.130)	(−0.438–0.296)	(−0.445–0.283)	(−0.941–0.518)
	0.353	<0.001	0.658	0.565
Exposed group	−0.312	−0.538	−0.062	−0.071
	(−0.3455–0.176)	(−0.463–0.211)	(−0.903–0.465)	(−0.233–0.098)
	<0.001	<0.001	0.592	0.420

TABLE 4: Correlations of 1-OHP levels and the other studied variables in total subjects.

Variables	r^a	<i>P</i> -value ^b
Age (year)	0.157	0.233 ^a
Employment time (year)	0.455	<0.001 ^a
Current smokers, yes (%)	0.329	0.073 ^b
Alcohol users, yes (%)	0.165	0.209 ^b
MDA (nmol/mL)	0.623	<0.001 ^a
8-OH-dG (nmol/mol creatinine)	0.668	<0.001 ^a
CTL (μ m)	0.571	<0.001 ^a
CTM	0.682	<0.001 ^a
MN (‰)	0.459	<0.001 ^a
CA (N, AR%)	0.422	0.009 ^b
IgG (g/L)	−0.732	<0.001 ^a
IgA (g/L)	−0.677	<0.001 ^a
IgM (g/L)	−0.209	0.127 ^a
IgE (IU/mL)	−0.187	0.211 ^a

^aSpearman rank correlation coefficient. ^bSpearman rank correlation coefficient for the comparisons between each of the studied variables.

IgM, and IgE, respectively. For the exposed group, each one percentage increase in urinary 1-OHP generates a 0.312% ($P < 0.001$), 0.538% ($P < 0.001$), 0.062% ($P = 0.592$), and 0.071% ($P = 0.420$) decrease in IgG, IgA, IgM, and IgE, respectively. Correlation between 1-OHP and other indices was assayed as in Table 4. Except for age, alcohol status, current smokers, IgM, and IgE, a significant correlation in urinary 1-OHP and other biomarkers in the total population (exposed and controls) ($P < 0.001$ for other biomarkers, $P = 0.009$ for CA, resp.) was observed in the studied subjects.

3.3. Correlation between Oxidative/Genotoxic Damage Biomarkers and Immunoglobulin Levels. We observed a significant correlation in MDA and 8-OH-dG levels, 8-OH-dG, and CTL levels in the control group ($r = 0.556$, $P < 0.001$; $r = 0.644$, $P < 0.001$; resp.) (Figures 1(a) and 1(c)) and in the exposed group ($r = 0.640$, $P < 0.001$; $r = 0.794$, $P < 0.001$; resp.) (Figures 1(b) and 1(d)). Also, we examined the correlations in genotoxic/oxidative damage biomarkers and immunoglobins of exposed workers and control workers next to further evaluate a possible role

and a potent biomedical significance of immunoglobulin levels in genotoxic/oxidative damage. The correlation between MDA, 8-OH-dG, CTL, and IgG, IgA of workers exposed to coke oven emission is plotted in Figures 2(b), 2(d), 2(f), 2(h), 2(j), and 2(l). As can be seen, there is a significant negative correlation between MDA, 8-OH-dG, CTL, and IgG, IgA levels in exposed workers ($P < 0.001$). A similar significant negative correlation between MDA and IgG, IgA level, 8-OH-dG, and IgG in control workers (Figure 2(a), $r = -0.498$, $P < 0.001$; Figure 2(c), $r = -0.363$, $P = 0.001$; Figure 2(e), $r = -0.229$, $P = 0.004$, resp.) was observed. However, in the control group, 8-OH-dG levels were not significantly correlated with IgA levels (Figure 2(g), $r = -0.080$, $P = 0.486$). Also, CTL levels were not significantly correlated with IgG and IgA in control workers (Figure 2(i), $r = -0.126$, $P = 0.270$; Figure 2(k), $r = -0.110$, $P = 0.339$, resp.). Further, PAH exposure enhances this negative correlation. Interestingly, although most individuals with low IgG, IgA levels tended to have more MDA, 8-OH-dG, and CTL levels, some with high IgG, IgA also had more MDA, 8-OH-dG, and CTL levels as can be seen particularly in Figures 2(b), 2(f), 2(h), 2(j), and 2(l), respectively.

4. Discussion

The working environment at a coke plant can negatively affect the employed workers who are exposed to coke oven emissions containing PAHs [24]. PAHs are introduced to the environment in numerous ways and are ubiquitously distributed. They are taken up by humans at the workplace, by smoking, by breathing polluted air, and by consuming food and medicines. They are potent oxidative/genotoxic damage and immunotoxic compounds. Current occupational exposure to PAHs was investigated in China coke oven plants in order to establish which resulted in oxidative/genotoxic DNA damages and immunotoxic effects which are considered to be indicators for long term adverse health effects. Biological monitoring of the internal dose was based on the analysis of urinary 1-hydroxypyrene. Biological effect monitoring including MDA, 8-OH-dG, CTL, CTM, MN, CA and IgA, IgE, IgG, and IgM levels as well as the correlation among these markers were evaluated.

FIGURE 1: Correlation between MDA level and 8-OH-dG level in lymphocytes of workers in control group ((a), $r = 0.556$, $P < 0.001$) and exposed group ((b), $r = 0.640$, $P < 0.001$), and correlation between 8-OH-dG and CTL level in lymphocytes of workers in control group ((c), $r = 0.644$, $P < 0.001$) and exposed group ((d), $r = 0.794$, $P < 0.001$).

In the present study, the coke oven workers seemed to have been exposed to a significant high level of PAHs based on the urinary 1-OHP. The PAH exposure of coke oven workers in the present study was similar with that in Siwińska et al.'s [16] and in Zhang et al.'s [25] study conducted in coke oven workers and much higher than that reported by Pavanello et al. [26] in CMBN assay in coke oven workers. Though the length of exposure is not, therefore, the only risk factor, Yang et al. reported that the length of time spent outdoors had marginal positive associations with urinary 1-OHP levels in nonoccupationally exposed Koreans [27]. So, significantly higher length of employment time may promote a synergistic effect on excretion of 1-OHP in PAH exposed coke oven workers. Significant distributions for alcohol users and no significant distributions for smoking status in exposed coke oven workers were observed in the present study. On the contrary, no significant difference was found in alcohol users between control and coke oven workers in Leng et al.'s [28] and Zhang et al.'s studies [25]. Other studies [29, 30]

indicated that the smoking of cigarettes significantly increases the urinary 1-OHP concentration, while alcohol consumption does not affect the urinary 1-OHP concentration. We further analyzed the smokers and nonsmokers in control and exposed group. The findings indicated the smoking was actually increase 1-OHP levels but not significance. This may be partially due to the relatively higher exposure to PAHs, which may hide the effect of smoking on 1-OHP concentrations [25]. Hence, the smoking life style may not significantly affect the urinary 1-OHP concentration in this study.

Metabolic transformation of PAHs generates reactive electrophilic metabolites, causing DNA damage and triggering the production of reactive oxygen species that lead to oxidative stress [31]. The analysis of urinary excretion of 8-OH-dG is a useful approach to assess individual cancer risk due to oxidative stress [32]. Data from the present study also showed that levels of MDA, 8-OHdG was associated with exposure to carcinogenic PAHs among coke oven workers.

FIGURE 2: Continued.

FIGURE 2: Correlation between MDA and IgG, IgA level in lymphocytes of workers in control group ((a), $r = -0.498$, $P < 0.001$; (c), $r = -0.363$, $P = 0.001$) and exposed group ((b), $r = -0.606$, $P < 0.001$; (d), $r = -0.814$, $P < 0.001$), respectively; correlation between 8-OH-dG and IgG, IgA level in lymphocytes of workers in control group ((e), $r = -0.229$, $P = 0.004$; (g), $r = -0.080$, $P = 0.486$) and exposed group ((f), $r = -0.405$, $P < 0.001$; (h), $r = -0.523$, $P < 0.001$), respectively; correlation between CTL and IgG, IgA level in lymphocytes of workers in control group ((i), $r = -0.126$, $P = 0.270$; (k), $r = -0.110$, $P = 0.339$) and exposed group ((j), $r = -0.312$, $P < 0.001$; (l), $r = -0.350$, $P < 0.001$), respectively.

Significant differences were observed for CTL levels, MN, and CA frequency of PAH exposed workers. In agreement with our results, Siwińska et al. reported that the level of urinary 1-OHP was correlated with genotoxic effects in coke oven workers as determined by a number of assays, including micronuclei frequency and DNA damage measured by sister-chromatid exchange and comet assays [16]. Significant correlation in MDA and 8-OH-dG levels, 8-OH-dG, and CTL levels in all subjects was observed in the present study. It is well known that reactive oxygen species generated by PAHs may induce damage to DNA. It is implied that oxidative DNA modification and oxidative DNA damage generated by PAHs exposed in coke oven workers [33]. High levels of MDA, 8-OHdG, and DNA damage were also indicated by several occupational studies [3, 8, 34]. Additionally, accumulating evidence suggested that ethanol can induce oxidative DNA damage in humans [3, 35]. The significant distributions for alcohol users in exposed coke oven workers indicates that alcohol drinking may synergetically increase levels of MDA, 8-OHdG, and DNA damage in alcohol users of coke oven workers.

A general observation has been that the immune system of humans is the most sensitive to modulation by PAHs. Four kinds of immunoglobulins were lower in coke oven workers, but this was significant only for IgG and IgA ($P < 0.001$). The results of this study regarding immunoglobulin levels also show consistent with the data of Szczeklik et al. [36] who demonstrated biosynthesis of IgG and IgA was markedly depressed immunoglobulins in serum of coke oven workers. Oh et al. [13] found that the four kinds of immunoglobulin types were lower in automobile emission inspectors, but this was significant only for IgG ($P = 0.047$). On the other hand, Winker et al. [37] did not find any alterations of the serum immunoglobulin levels in PAH-exposed workers. Similarly, the four kinds of immunoglobulin types were found in lower amounts in the waste incineration workers, but this disparity was not significant one [38]. By contrast, there was also a significant enhancement in serum IgG levels and the percentage of monocytes in the PAH exposed asphalt workers compared to the control group [12]. In addition, Szczeklik et al. also found that serum IgE had a trend toward increased values in serum of coke oven workers [36]. These may be due

to the fact that the place of adaptive responses or changes may come to play in situation of chronic exposure [39]. Generally, IgG plays a major role in humoral immune response through complement fixation and neutralization, and reduction in IgG may lead to downregulation of antibody-mediated host resistance, which could enhance influence of infection or tumor progression [40, 41]. The decrease in IgA production suggests that PAHs may compromise protection of mucosal surfaces in the respiratory tract [42]. There has been reported that many PAHs are known to be potent suppressors of the immune response. Additionally, the immunosuppressive potential of PAHs is linked to their carcinogenic potency [43]. Accordingly, in some extent, the PAH exposure in coke oven workers showed the immune depression and the increase in potential carcinogenic effect through the significant reduction of IgG and IgA levels. Further, except age, alcohol status, smoking status, IgM, IgE, and other markers were significantly associated with PAH exposure as indicated by 1-OHP levels. As previous descriptions, the markers of employment time, MDA, 8-OH-dG, CTL, CTM, MN (%), CA, IgG, and IgA may be more important markers in coke oven workers.

Further, a significant negative correlation in genotoxic/oxidative damage biomarkers of MDA, 8-OH-dG, CTL levels, and immunoglobins of IgG, IgA levels, especially in coke oven workers, was found in the present study. Studies have suggested that oxidative stress-induced lipid peroxidation may play a role in toxic responses to PAH in immunological systems. For example, Zhigacheva et al. found that the increased lipid peroxidation products were linked to modification of leucocyte membranes and reduced activity of lymphocyte enzymes in animals after 6 months of PAH inhalation as compared with controls [44]. In addition, moderate activation of lipid peroxidation and oxidative damage as determined by plasma MDA and 8-OHdG levels were detected simultaneously with significant alterations in IgA and IgE levels described by Jeng et al. [42]. Also, a significant difference was found regarding the levels of IgE and the levels of DNA damage in polynuclear cells were found in waste incineration workers described by Oh et al. [38]. Further, PAH exposure enhances this negative correlation (Figure 2). Generally speaking, PAHs are cytotoxic at high doses, while lower doses of PAHs frequently result in alteration of immunological responses. It is likely that heavier PAH species increased redox activity and subsequently induced generation of ROS [42] which altered IgE and IgA levels in workers exposed to PAHs. Further research should clarify the effects of high molecular weights of PAHs on immunological function.

The study also has some limitations. First, our sample size is relatively small and this is one of the most common reasons for the failure to replicate reported associations across studies [45]. This may be due to no significance for 1-OHP levels in smokers and nonsmokers, IgM, IgE levels between the control and exposed group. In particular, the number of subjects in CA assay was only 10 for control and 36 in exposed group. So, further investigation still recommends large sample population in the future. On the other hand,

though alcohol consumption does not affect the urinary 1-OHP concentration [29, 30], excess alcohol consumption may result in the enhancement of PAH metabolites [25] and the mechanism is needed further study. Finally, staff members of the offices and hospitals of the same steel company served as the control group. There may be differences in economics and nutrition status between control and coke oven workers. This may be affected by food intakes which are also known as an important route of exposure to PAHs [9]. To better understand the factors related to the increase of 1-OHP, the food consumption, social activities, and nutrition status should be included in the questionnaire and further analyzed in the future.

5. Conclusion

In conclusion, the present study provided yet more evidence that PAHs cause significant oxidative stress/DNA damage through the MDA, 8-OHdG levels and CTL level, CTM, MN, and CA frequency in coke oven workers. Moreover, our immunoglobulin study showed that IgG and IgA were significantly reduced by PAHs. In addition, our investigation of the association between MDA and 8-OH-dG levels, 8-OH-dG and CTL levels, MDA, 8-OH-dG, CTL levels, and immunoglobins of IgG, IgA levels was proven to be a meaningful or relevant association. It is implied that oxidative stress/DNA damage induced by PAHs may play a role in toxic responses for PAH in immunological systems.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

The authors extend special thanks to Dr. Fei Yan of the Fourth Military Medical University for his assistance in statistical analysis. The research was supported by the Fundamental Research Funds for the Central Universities (2012jdhz48) and by the Shaanxi Postdoctoral Science Foundation funded project (18420024).

References

- [1] H. Wang, W. Chen, H. Zheng et al., "Association between plasma BPDE-Alb adduct concentrations and DNA damage of peripheral blood lymphocytes among coke oven workers," *Occupational and Environmental Medicine*, vol. 64, no. 11, pp. 753–758, 2007.
- [2] G. Castaño-Vinyals, A. D'Errico, N. Malats, and M. Kogevinas, "Biomarkers of exposure to polycyclic aromatic hydrocarbons from environmental air pollution," *Occupational and Environmental Medicine*, vol. 61, no. 4, article e12, 2004.
- [3] J. Zhang, M. Ichiba, T. Hanaoka et al., "Leukocyte 8-hydroxydeoxyguanosine and aromatic DNA adduct in coke-oven workers with polycyclic aromatic hydrocarbon exposure," *International Archives of Occupational and Environmental Health*, vol. 76, no. 7, pp. 499–504, 2003.

- [4] M. Gao, Y. Li, Y. Sun et al., "A common carcinogen benzo[a]pyrene causes p53 overexpression in mouse cervix via DNA damage," *Mutation Research: Genetic Toxicology and Environmental Mutagenesis*, vol. 724, no. 1-2, pp. 69-75, 2011.
- [5] M. Lodovici, C. Casalini, R. Cariaggi, L. Michelucci, and P. Dolara, "Levels of 8-hydroxydeoxyguanosine as a marker of DNA damage in human leukocytes," *Free Radical Biology and Medicine*, vol. 28, no. 1, pp. 13-17, 2000.
- [6] B. Marczynski, H. P. Rihs, B. Rossbach et al., "Analysis of 8-oxo-7,8-dihydro-2'-deoxyguanosine and DNA strand breaks in white blood cells of occupationally exposed workers: comparison with ambient monitoring, urinary metabolites and enzyme polymorphisms," *Carcinogenesis*, vol. 23, no. 2, pp. 273-281, 2002.
- [7] D. Kuang, W. Zhang, Q. Deng et al., "Dose-response relationships of polycyclic aromatic hydrocarbons exposure and oxidative damage to DNA and lipid in coke oven workers," *Environmental Science & Technology*, vol. 47, no. 13, pp. 7446-7456, 2013.
- [8] A. L. Liu, W. Q. Lu, Z. Z. Wang et al., "Elevated level of urinary 8-hydroxy-2'-deoxyguanosine, lymphocytic micronuclei, and serum glutathione S-transferase in workers exposed to coke oven emissions," *Environmental Health Perspectives*, vol. 114, no. 5, pp. 673-677, 2006.
- [9] S. Bae, X. Pan, S. Kim et al., "Exposures to particulate matter and polycyclic aromatic hydrocarbons and oxidative stress in schoolchildren," *Environmental Health Perspectives*, vol. 118, no. 4, pp. 579-583, 2010.
- [10] R. Crebelli, P. Carta, C. Andreoli et al., "Biomonitoring of primary aluminium industry workers: detection of micronuclei and repairable DNA lesions by alkaline SCGE," *Mutation Research—Genetic Toxicology and Environmental Mutagenesis*, vol. 516, no. 1-2, pp. 63-70, 2002.
- [11] S. J. Kwack and B. M. Lee, "Correlation between DNA or protein adducts and benzo[a]pyrene diol epoxide I-triglyceride adduct detected in vitro and in vivo," *Carcinogenesis*, vol. 21, no. 4, pp. 629-632, 2000.
- [12] A. Karakaya, B. Yücesoy, A. Turhan, O. Erdem, S. Burgaz, and A. E. Karakaya, "Investigation of some immunological functions in a group of asphalt workers exposed to polycyclic aromatic hydrocarbons," *Toxicology*, vol. 135, no. 1, pp. 43-47, 1999.
- [13] E. Oh, H. Im, H.-S. Kang et al., "Comparison of immunological and genotoxicological parameters in automobile emission inspectors exposed to polycyclic aromatic hydrocarbons," *Environmental Toxicology and Pharmacology*, vol. 21, no. 1, pp. 108-117, 2006.
- [14] S. Sellappa, B. Mani, and K. S. Keyan, "Cytogenetic biomonitoring of road paving workers occupationally exposed to polycyclic aromatic hydrocarbons," *Asian Pacific Journal of Cancer Prevention*, vol. 12, no. 3, pp. 713-717, 2011.
- [15] F. J. Jongeneelen and R. B. M. Anzion, "Analyses of hazardous substances in biological materials," *Deutsche Forschungsgemeinschaft*, vol. 3, pp. 151-169, 1990.
- [16] E. Siwińska, D. Mielżyńska, and L. Kapka, "Association between urinary 1-hydroxypyrene and genotoxic effects in coke oven workers," *Occupational and environmental medicine*, vol. 61, no. 3, article e10, 2004.
- [17] R. C. Baselt, *Biological Monitoring Methods for Industrial Chemicals*, Biomedical Publications, Davis, Calif, USA, 1980.
- [18] J. A. Buege and S. D. Aust, "Microsomal lipid peroxidation," *Methods in Enzymology*, vol. 52, pp. 302-310, 1978.
- [19] J. Brostoff, G. K. Scadding, D. Male, and I. M. Roitt, *Clinical Immunology*, Gower Medical Publishing, London, UK, 1991.
- [20] N. P. Singh, M. T. McCoy, R. R. Tice, and E. L. Schneider, "A simple technique for quantitation of low levels of DNA damage in individual cells," *Experimental Cell Research*, vol. 175, no. 1, pp. 184-191, 1988.
- [21] V. J. McKelvey-Martin, M. H. L. Green, P. Schmezer, B. L. Pool-Zobel, M. P. de Meo, and A. Collins, "The single cell gel electrophoresis assay (comet assay): a European review," *Mutation Research—Fundamental and Molecular Mechanisms of Mutagenesis*, vol. 288, no. 1, pp. 47-63, 1993.
- [22] M. Fenech and A. A. Morley, "Measurement of micronuclei in lymphocytes," *Mutation Research—Genetic Toxicology and Environmental Mutagenesis*, vol. 147, no. 1-2, pp. 29-36, 1985.
- [23] A. V. Carrano and A. T. Natarajan, "Consideration for population monitoring using cytogenetic techniques," *Mutation Research*, vol. 204, no. 3, pp. 379-406, 1988.
- [24] M. T. Wu, I.-F. Mao, C. K. Ho et al., "Urinary 1-hydroxypyrene concentrations in coke oven workers," *Occupational and Environmental Medicine*, vol. 55, no. 7, pp. 461-467, 1998.
- [25] J. Zhang, M. Ichiba, K. Hara et al., "Urinary 1-hydroxypyrene in coke oven workers relative to exposure, alcohol consumption, and metabolic enzymes," *Occupational and Environmental Medicine*, vol. 58, no. 11, pp. 716-721, 2001.
- [26] S. Pavanello, L. Kapka, E. Siwinska, D. Mielżyńska, C. Bolognesi, and E. Clonfero, "Micronuclei related to Anti-B[a]PDE-DNA adduct in peripheral blood lymphocytes of heavily polycyclic aromatic hydrocarbon-exposed nonsmoking coke-oven workers and controls," *Cancer Epidemiology Biomarkers and Prevention*, vol. 17, no. 10, pp. 2795-2799, 2008.
- [27] M. Yang, S. Kim, E. Lee et al., "Sources of polycyclic aromatic hydrocarbon exposure in non-occupationally exposed Koreans," *Environmental and Molecular Mutagenesis*, vol. 42, no. 4, pp. 250-257, 2003.
- [28] S. Leng, Y. Dai, Y. Niu et al., "Effects of genetic polymorphisms of metabolic enzymes on cytokinesis-block micronucleus in peripheral blood lymphocyte among coke-oven workers," *Cancer Epidemiology Biomarkers and Prevention*, vol. 13, no. 10, pp. 1631-1639, 2004.
- [29] M. Yang, J. Jang, S. Kim et al., "Genetic effects on urinary 1-hydroxypyrene levels in a Korean population," *Carcinogenesis*, vol. 24, no. 6, pp. 1085-1089, 2003.
- [30] H. Nan, H. Kim, H. Lim et al., "Effects of occupation, lifestyle and genetic polymorphisms of CYP1A1, CYP2E1, GSTM1 and GSTT1 on urinary 1-hydroxypyrene and 2-naphthol concentrations," *Carcinogenesis*, vol. 22, no. 5, pp. 787-793, 2001.
- [31] J. Wang, E. H. Weyand, and R. G. Harvey, "Synthesis of suspected carcinogenic metabolites of 7H-benzo[c]fluorene, a coal tar component implicated in causation of lung tumors," *Journal of Organic Chemistry*, vol. 67, no. 17, pp. 6216-6219, 2002.
- [32] B. Halliwell, "Free radicals, antioxidants, and human disease: curiosity, cause, or consequence?" *The Lancet*, vol. 344, no. 8924, pp. 721-724, 1994.
- [33] M. S. Cooke, M. D. Evans, R. Dove et al., "DNA repair is responsible for the presence of oxidatively damaged DNA lesions in urine," *Mutation Research—Fundamental and Molecular Mechanisms of Mutagenesis*, vol. 574, no. 1-2, pp. 58-66, 2005.
- [34] X. Yang, J. Zheng, Y. Bai et al., "Using lymphocyte and plasma Hsp70 as biomarkers for assessing coke oven exposure among steel workers," *Environmental Health Perspectives*, vol. 115, no. 11, pp. 1573-1577, 2007.

- [35] R. Guo and J. Ren, "Alcohol and acetaldehyde in public health: from marvel to menace," *International Journal of Environmental Research and Public Health*, vol. 7, no. 4, pp. 1285–1301, 2010.
- [36] A. Szczeklik, J. Szczeklik, Z. Galuszka, J. Musial, E. Kolarzyk, and D. Targosz, "Humoral immunosuppression in men exposed to polycyclic aromatic hydrocarbons and related carcinogens in polluted environments," *Environmental Health Perspectives*, vol. 102, no. 3, pp. 302–304, 1994.
- [37] N. Winker, H. Tuschl, R. Kovac, and E. Weber, "Immunological investigations in a group of workers exposed to various levels of polycyclic aromatic hydrocarbons," *Journal of Applied Toxicology*, vol. 17, pp. 23–29, 1996.
- [38] E. Oh, E. Lee, H. Im et al., "Evaluation of immuno- and reproductive toxicities and association between immunotoxicological and genotoxicological parameters in waste incineration workers," *Toxicology*, vol. 210, no. 1, pp. 65–80, 2005.
- [39] A. O. Odewabi, O. A. Ogundahunsi, and M. Ekor, "Effect of exposure to solid wastes in relation to employment duration on some important markers of health and disease in waste management workers of Ogun State in southwest Nigeria," *Human and Experimental Toxicology*, vol. 32, no. 12, pp. 1231–1244, 2013.
- [40] H. A. Kim, E. M. Kim, Y. C. Park et al., "Immunotoxicological effects of agent orange exposure to the Vietnam war Korean veterans," *Industrial Health*, vol. 41, no. 3, pp. 158–166, 2003.
- [41] R. Jefferis and D. S. Kumararatne, "Selective IgG subclass deficiency: quantification and clinical relevance," *Clinical and Experimental Immunology*, vol. 81, no. 3, pp. 357–367, 1990.
- [42] H. A. Jeng, C. Pan, N. Diawara et al., "Polycyclic aromatic hydrocarbon-induced oxidative stress and lipid peroxidation in relation to immunological alteration," *Occupational and Environmental Medicine*, vol. 68, no. 9, pp. 653–658, 2011.
- [43] P. Rajendran, T. Jayakumar, I. Nishigaki et al., "Immunomodulatory effect of mangiferin in experimental animals with Benzo(a)pyrene-induced lung carcinogenesis," *International Journal of Biomedical Science*, vol. 9, no. 2, pp. 68–74, 2013.
- [44] I. V. Zhigacheva, E. B. Burlakova, L. S. Evseenko et al., "Humoral immunity: polycyclic aromatic hydrocarbons and nitrosamines," *Doklady Biological Sciences*, vol. 383, no. 1–6, pp. 120–122, 2002.
- [45] K. Fagerström, "Time to first cigarette: the best single indicator of tobacco dependence?" *Monaldi Archives for Chest Disease*, vol. 59, no. 1, pp. 91–94, 2003.