

Clinical Study

Clinical and Genetic Study of Algerian Patients with Spinal Muscular Atrophy

Y. Sifi,^{1,2} K. Sifi,^{2,3} A. Boulefkhad,^{1,2} N. Abadi,^{2,3} Z. Bouderdia,⁴ R. Cheriet,⁴ M. Magen,⁵
J. P. Bonnefont,⁵ A. Munnich,⁵ C. Benlatreche,^{2,3} and A. Hamri^{1,3}

¹ Service of Neurology CHU of Constantine, Algeria

² Laboratory of Biology and Molecular Genetics CHU and University of Constantine, Algeria

³ Laboratory of Biochemistry CHU of Constantine, Algeria

⁴ Service of Pediatrics CHU de Constantine, Algeria

⁵ Genetic Department of the Necker Hospital and Paris Descartes University, Paris, France

Correspondence should be addressed to Y. Sifi; sifimina@yahoo.fr

Received 20 December 2012; Revised 12 February 2013; Accepted 18 February 2013

Academic Editor: Haluk Topaloglu

Copyright © 2013 Y. Sifi et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Spinal muscular atrophy (SMA) is the second most common lethal autosomal recessive disorder. It is divided into the acute Werdnig-Hoffmann disease (type I), the intermediate form (type II), the Kugelberg-Welander disease (type III), and the adult form (type IV). The gene involved in all four forms of SMA, the so-called survival motor neuron (SMN) gene, is duplicated, with a telomeric (tel SMN or SMN1) and a centromeric copy (cent SMN or SMN2). SMN1 is homozygously deleted in over 95% of SMA patients. Another candidate gene in SMA is the neuronal apoptosis inhibitory protein (NAIP) gene; it shows homozygous deletions in 45–67% of type I and 20–42% of type II/type III patients. Here we studied the SMN and NAIP genes in 92 Algerian SMA patients (20 type I, 16 type II, 53 type III, and 3 type IV) from 57 unrelated families, using a semiquantitative PCR approach. Homozygous deletions of SMN1 exons 7 and/or 8 were found in 75% of the families. Deletions of exon 4 and/or 5 of the NAIP gene were found in around 25%. Conversely, the quantitative analysis of SMN2 copies showed a significant correlation between SMN2 copy number and the type of SMA.

1. Introduction

Spinal muscular atrophies (SMAs) are a group of motor neuron disorders characterized by degeneration of spinal cord anterior horn cells, leading to muscular wasting and atrophy [1]. SMA is the most common autosomal recessive disorder after cystic fibrosis, with an estimated 1/10,000 incidence and a 1/60 carrier frequency [2]. Affected patients are classified into four groups according to age at onset and phenotype severity [3, 4]. Type I SMA or the Werdnig-Hoffmann disease (OMIM No. 253300) is the most severe form, with an onset within the first 6 months of age, severe generalized muscle weakness with hypotonia, and death before two years of age. In type II SMA (OMIM No. 253550), affected children sit unassisted, may be able to walk for a short distance, and usually survive over 10 years of age. Type III SMA or the Kugelberg-Welander disease (OMIM No. 253400) has its

onset in the first to third decade. Though its course is highly variable, patients are constantly able to walk unassisted. Type IV SMA or adult-onset SMA (OMIM No. 271150) is quite rare.

The survival motor neuron (SMN) gene, implicated in the four forms of SMA, maps to chromosome 5q 11.2–13.3 [5–7] and is duplicated as telomeric and centromeric copies, so called SMN1 (OMIM No. 600354) and SMN2 (OMIM No. 601627), respectively [8, 9]. SMN1 and SMN2 comprising 8 exons are highly homologous, with only five base-pair differences within their 3' ends [8, 10], and thus encode nearly identical proteins. Two of these base-pairs, located in exons 7 and 8, allow SMN1 to be distinguished from SMN2 at DNA and RNA levels and are currently used for detection of SMN1 deletions [11]. A vast majority (90–98%) of SMA patients have homozygous deletions of SMN1 exons 7 and 8 [8, 12, 13], the remaining ones carrying SMN1 intragenic

TABLE 1: Classification of autosomal recessive proximal spinal atrophy as defined by Zerres et al. [26].

Type	Definition
I	Never able to sit
II	Able to sit but not to walk Able to walk
III	(a) Onset before 3 years (b) Onset 3–30 years
IV	Onset after 30 years

mutations [8, 14, 15], with a frequency higher in type I than in types II and III.

Conversely, *SMN2* homozygous inactivation is not directly responsible for SMA [8]. A number of studies have however shown that *SMN2* acts as a modulator of SMA severity, with an inverse correlation between the *SMN2* copy number and the disease severity [16, 17]. Failure of *SMN2* to fully compensate homozygous loss of *SMN1* is due to a sequence difference in exon 7 which causes alternative splicing of the *SMN2* gene, and subsequently lower amount of full-length protein [8, 18, 19].

The neuronal apoptosis inhibitory protein gene (*NAIP*) [20], close to the *SMN* genes (15, 5 kb) at 5q11–q13, was initially considered as a candidate gene for SMA [12, 20, 21]. While subsequent studies have ruled out its direct responsibility, for this disease [17, 22], *NAIP* has however shown to be more frequently mutated in SMA affected patients than in general population, with homozygous deletions in 45–67% of type I and 20–42% of type II/type III SMA patients [12, 20, 23–25].

Analysis of deletions encompassing both *NAIP* and *SMN* genes in a large number of SMA patients suggests that loss of *NAIP* may be associated with a higher disease severity [10, 20]. Here we investigated the clinical and molecular characteristics of 92 Algerian SMA patients from 57 families to assess the prevalence of *SMN1* deletions and the combined impact of *SMN2* copy number and *NAIP* deletions on clinical severity.

2. Materials and Methods

2.1. Patients. 92 patients from 57 Algerian families were diagnosed as having SMA on the basis of clinical findings and electromyoneurography. All patients fulfilled the diagnostic criteria for proximal SMA, as defined by the International SMA Collaboration [26] and by Zerres et al. [27] (Table 1). Inclusion and exclusion criteria were similar to those proposed by the International SMA Collaboration [26]. Patients with symmetrical, muscle weakness of trunk and limbs, proximal muscles weakness more than distal, lower limbs involvement more than upper limbs, and fasciculations of tongue and tremor of hands and in whom denervation was seen on EMG were included in our study. Patients who presented with CNS dysfunction, sensory loss, eye or facial muscle weakness, or involvement of other organs were excluded from this study. A complete

TABLE 2: Consanguinity in Algerian SMA Families.

Degree of inbreeding	No. of families
2nd degree	7 (12,28%)
3rd degree	10 (17,54%)
4th degree	8 (14%)
Distant relatives	3 (5%)
Unrelated	29 (51%)

TABLE 3: Frequency of SMA types.

SMA type	No. of cases	No. of families
I	20	14
II	16	10
III	53	31
IV	3	2
Total	92	57

clinical history was recorded with emphasis on age, sex, age at onset, course of the disease, perinatal history, parental consanguinity, and affected relatives. Clinical examination focused on neurological parameters, tone, power, reflexes, wasting and atrophy of muscles, and abnormal movements and sensations. Other investigations included serum creatine phosphokinase (CPK), electromyogram (EMG), and nerve conduction velocity.

2.2. Methods. After informed consent, DNA was extracted from peripheral blood samples according to a standard technique [28].

2.2.1. Molecular Analysis of *SMN* Genes. Search for *SMN1* exons 7 and 8 deletions was performed by PCR and restriction enzyme digestion, as described in [29]. The *SMN2* copy number was determined by Multiplex Ligation-dependent Probe Assay [30, 31].

2.2.2. Molecular Analysis of the *NAIP* Gene. All individuals were also tested for exons 4 and 5 deletion of the *NAIP* gene. PCR conditions and primers used to amplify exons 4 and 5 were identical to those of Roy et al. [20].

3. Results

3.1. Clinical and Genealogical Findings. The rate of consanguineous marriage in this study was approximately 47%. Degrees of consanguinity are listed in Table 2.

Twenty-two of the 57 families (39%) were multiplex. The most common type in our cohort was type III, with fifty-three (53) affected cases from 31 families (60%), followed by type I with 20 cases from 14 families (25%). Frequency of the different types is summarized in Table 3.

In the SMA type I group, the age at onset varied from birth to 6 months, with an average of 5 ± 2 , 5 months. All patients were mentally alert. The main symptom was severe hypotonia with poor limb mobility. Only two patients could

TABLE 4: Phenotypic and genotypic analysis of the 6 SMA type I index patients with prolonged survival.

Phenotypic and genotypic analysis	N° 1	N° 2	N° 3	N° 4	N° 5	N° 6
Sex	F	M	M	M	M	M
Consanguinity	–	–	–	+	–	+
Age at onset (months)	4	2	4	6	4	Birth
Age at diagnosis (year)	4	5	5	3	2	2
Age at last information (months)	50	60	72	39	48	41
Lifting of head	+	+	+	+	–	–
Hypotonia and muscle weakness	+	+	+	+	+	+
Deep tendon reflexes	–	–	–	–	–	–
Fasciculations	–	–	+	–	+	–
Bulbar symptoms	+	+	+	+	+	–
Breathing difficulties	+	–	–	–	–	–
Frequent pneumonia	+	+	+	+	+	+
Age at death (months)	54	–	–	–	–	–
SMN1 gene deletion	Yes	Yes	Yes	No	Yes	Yes
NAIP gene deletion	Yes	No	No	No	No	No
SMN2 copy number	2	2	3	–	3	3

hold up their heads, for a short period of time. Twelve patients (60%) died between 2 and 53 months of age, due to respiratory failure following respiratory tract infections. The remaining ones are still alive (the oldest patient is currently 72 months old). All patients with prolonged survival suffered from joint contractures caused by progressive muscular atrophy, spine deformation as scoliosis or kyphoscoliosis, and swallowing difficulties. The Bulbar symptoms were observed in 5 patients of them (Table 4). They were not able to cope with everyday routine and thus were totally family dependent. The DNA analysis showed that the *SMN1* gene is interrupted in five out of six SMA type I patients with prolonged survival, and only one patient showed homozygous deletion of *NAIP* gene (Table 4). The *SMN2* copy number was determined in 5 SMA type I patients carrying homozygous *SMN1* deletions, and two of them (2/5) had two *SMN2* copies, the remaining ones (3/6) carrying 3 *SMN2* copies (Table 4).

In the SMA type II group, age of onset ranged from 8 to 18 months (average 12, 7±3, and 3 months). They were defined by ability to sit alone. Some children experienced early difficulty for sitting or rolling over (2 patients), while 3 patients were able to crawl and stand with support at a mean age of 22 months for a period of 4 months, and two patients were able to walk with support for a period of 6 months. None walked unaided. Scoliosis and contractures constantly developed in the patients who all became wheelchair dependent (6/16). In the type II group all patients are still alive, and 4/16 patients (25%) survive beyond age 15. In the SMA type III group, clinical onset ranged from the first year of life to the 3rd decade. Twenty-two patients (41, 5%) were confined to a wheelchair at ages ranging from 10 to 34 years, the remaining ones being still able to walk, with support. Hand tremor was found in 26 out of the 53 patients type III. Distal muscle weakness and/or amyotrophy was associated with the classical proximal defect, with frequent spine deformities and

contractures in 39 patients. Life span was not significantly reduced.

SMA type II and III patients coexisted within 2 families. In the three adult-onset SMA patients (type IV), age of onset ranged from 20 to 41 years (mean age of onset 30 ± 8 years). Adult SMA patients, except for patient 2, had very mild phenotypes, compared with the childhood onset.

Blood CPK activity was normal in all SMA type I patients and was occasionally normal or slightly elevated in patients with type II (3 patients) or type III SMA (14 patients). In EMG examination the increased mean potentials, amplitude, duration, and area were stated. Maximal effort pattern in both proximal and distal muscles was reduced; spontaneous activity fibrillation and occasional fasciculations were present. Motor conduction velocity and sensory nerve conduction were normal.

3.2. Molecular Findings. Homozygous deletions of *SMN1* exon 7, exon 8, or both were observed in 43/57 families (75%) with the following distribution: type I 11/14, type II 7/10, type III 24/31, and type IV 1/2. Among the 43 families with deletions, 36 had both exons 7 and 8 deleted, while four had deletions only of exon 7, and 3 patients carried only homozygous deletion restricted to *SMN1* exon 8 (Table 5). Homozygous deletions of exons 4 and/or 5 of the *NAIP* gene were found in 4/14 type I, 2/10 type II, 9/31 type III, and 0/2 type IV families (Table 5). Homozygous *NAIP* deletions were constantly associated with homozygous *SMN1* deletions.

The *SMN2* copy number was determined in patients carrying homozygous *SMN1* deletions $n = 62$ (Table 6). 11/15 (73%) SMA type I patients had one or two *SMN2* copies, the remaining ones carrying 3 *SMN2* copies, 10/12 (83%) type II patients carried three or four *SMN2* copies, the remaining ones having 2 *SMN2* copies, and 32/33 (96%) type III patients

TABLE 5: Distribution of homozygous deletion of *SMN1* and *NAIP* genes according to the different types of *SMA*.

N of families	<i>SMN1</i> gene deletion			<i>NAIP</i> gene deletion
	Exon 7	Exon 8	Exons 7 and 8	Exon 4/5
<i>SMA</i> I (<i>n</i> = 14)	2 (14%)	1 (7%)	8 (57%)	4 (28%)
<i>SMA</i> II (<i>n</i> = 10)	1 (10%)	0	6 (60%)	2 (20%)
<i>SMA</i> III (<i>n</i> = 31)	1 (3%)	2 (6%)	21 (67%)	9 (29%)
<i>SMA</i> IV (<i>n</i> = 2)	0	0	1 (50%)	0

TABLE 6: Analysis of the *SMN2* copy number in the 62 patients with homozygous absence of the *SMN1* gene.

<i>SMA</i> type	N patients	<i>SMN2</i> copy number				
		1	2	3	4	5
I	15	2 (13%)	9 (60%)	4 (26%)	0	0
II	12	0	2 (16%)	7 (58%)	3 (25%)	0
III	33	0	0	5 (15%)	27 (81%)	1 (3%)
IV	2	0	0	0	0	2 (100%)

carried three or four *SMN2* copies. Finally, both adult onset patients carried at least 5 *SMN2* copies (Table 6).

4. Discussion

We analyzed three genes implicated in *SMA*, namely, *SMN1*, *SMN2*, and *NAIP*, in a cohort of 92 *SMA* affected patients from 57 Algerian families, in an attempt at phenotype/genotype correlation. All patients fulfilled the diagnostic criteria for proximal *SMA*, as defined by the International *SMA* Collaboration [27] and were classified into four subgroups according to the criteria of Zerres et al. [26]. Twenty patients had type I, 16 patients type II, 53 type III, and 3 patients type IV *SMA*. Though clinical classification of *SMA* is helpful in providing medical care and prognostic assessment; it is however based on subjective and arbitrary parameters which may still be controversial and subject to errors. Zerres and Rudnik-Schöneborn [32], in a retrospective study of 445 *SMA* patients, found 106 cases (24%) that could not be classified and suggested subdividing type III *SMA* into two groups, resulting in a total of four *SMA* types. In the present study, clinical classification of patients into four groups, based on criteria of the International *SMA* collaboration [27] and of Zerres and Rudnik-Schöneborn [32], was possible for most patients. In these classifications, age at onset is classically considered to be predictive of the outcome. However, in 11 cases (12%) age at onset and/or death and motor milestones (ability to walk independently) did clearly overlap between two subsets. It is thus important to keep in mind the possibility of long-standing disease courses with an early onset of weakness compatible with a prolonged survival. For example, 6 patients with *SMA* type I survived over age two. In 5 patients, age at onset was before 18 months, which is characteristic of type II *SMA*, while walking capacities were compatible with *SMA* type III. Coexistence of various types of *SMA* (II and III) within a given family occurred in our series (2/57 families), as reported elsewhere [33, 34], in favor of a continuous spectrum in childhood *SMA*. Additionally we found a predominance of males to females (17 female/36

males) in type III *SMA*, as previously reported by Rudnik-Schöneborn et al. who suggested the presence of a female sparing factor [35]. Tazir and Geronimi reported the same fact in a much larger Algerian series in which chronic cases were predominant [36].

Consanguinity rate was 47% in our cohort, that is above the average reported in the Algerian general population ($\approx 39\%$) [37]. Furthermore twenty-four families (42%) had a positive history of affected relatives. These data emphasize the importance of lowering the consanguinity rate and the value of genetic counseling and prenatal diagnosis for preventing *SMA* in our community.

From molecular point of view, the proportion of *SMN1* homozygous deletions was 75% in our study, lower than those found in several other previously reported population studies [38–42] (Table 7).

Deletions involving both exons 7 and 8 were observed in 36 families (63%), being much more frequent than deletions restricted to exon 7 (4 families, 7%) or 8 (3 families, 5%), in agreement with previous investigations [8, 23, 42, 43].

Several authors reported a frequency of large deletions, encompassing both *SMN* and *NAIP* genes, higher in *SMA* type I than in the other types [12, 25, 43–46].

In our study the frequency of *NAIP* gene deletions was 28%, 20%, and 16% for type I, II, and III, respectively, and did apparently not influence the disease severity.

Moreover, a great proportion of severely affected patients harboured no *NAIP* deletion, and the same pattern of deletions (involving *SMN* and *NAIP* genes) was found among affected sibs with different phenotypes (*SMA* II and *SMA* III). This supports the hypothesis that other factors may regulate the severity of the clinical course in addition to the extent of the deletion [17, 47]. The *SMN2* gene was consistently present as at least one copy in our series, thus contributing to some amount of *SMN* protein [46, 48]. It has previously been reported that most *SMN2* transcripts lack exon 7 and are thus functionally defective, reinforcing the view that the disease is the result of an insufficient amount of intact *SMN* protein [49]. Interestingly, no patient has been diagnosed

TABLE 7: Frequency of *SMN1* homozygous deletion in *SMA* around the world.

References	Countries	N patients	<i>SMN</i> Del (%)
[38]	Korea 2001	37	32,43%
[39]	Vietnam 2003	17	41,17%
[40]	Johannesburg 2007	92	51%
[41]	Egypt 2001	33	55%
[42]	Russia 2001	57	65%
[33]	Brasilia 1999	87	69%
<i>Our study</i>	<i>Algeria 2009</i>	92	75%
[43]	India 2005	45	76%
[44]	Saudi Arabia 1997	16	82%
[55]	Morocco 2003	54	83,33%
[56]	Turkey 2000	60	85%
[57]	Germany 1995	195	90%
[58]	Spain 1995	54	91%
[59]	Holland 1995	103	93%
[60]	Japan 2002	32	94%
[61]	Tunis 2006	60	95%
[62]	Iran 2004	22	95,40%
[24]	UK 1995	140	97,80%
[63]	Kuwait 2001	46	97,82%
[8]	France 1995	229	98%
[64]	Taiwan 1995	42	100%

with a homozygous absence of both *SMN1* and *SMN2* gene so far, suggesting that a total absence of *SMN* would be lethal in utero.

The results of our quantitative analysis of *SMN2* gene copies clearly show that the disease phenotype is influenced by the number of copies of the *SMN2* gene, consistent with previous studies indicating that type II and III patients have on average a larger number of *SMN2* copies than type I *SMA* patients [50–53]. In our series of 11 *SMA* type I patients who had a determination of the *SMN2* copy number, the two patients with one *SMN2* copy had a median survival of 5 months, whereas those with two and three *SMN2* copies survived 8 and 23 months, respectively.

It is classically admitted that the *SMN2* copy number is less than 3 in *SMA* type I and at least 3 in *SMA* type II, III, and IV [52–54]. Such a correlation between the number of *SMN2* genes and the clinical phenotype is however not conclusive.

In conclusion, our results are in agreement with the general consensus that there is no correlation between the size of *SMN1* deletions and the clinical severity of *SMA* and that there exists a close relationship between *SMN2* copy number and *SMA* disease severity, suggesting that the determination of *SMN2* copy number may be a good predictor of *SMA* disease type. We suggest that other still unknown factors may regulate the severity of the clinical course and influence phenotype expression. Our study additionally understanding the function of the *SMN* protein would probably be the key in unraveling the molecular basis of *SMA*.

Electronic Database Information

Accession numbers and the URL for data presented herein are as follows: Online Mendelian Inheritance in Man (OMIM), <http://www.ncbi.nlm.nih.gov/Omim/> (for type I *SMA* [MIM 253300], type II *SMA* [MIM 253550], type III *SMA* [MIM 253400], *SMN1* [MIM 600354], and *SMN2* [MIM 601627]).

Acknowledgments

The authors thank the *SMA* families and clinicians who collaborated with them in this study. They are also grateful to Professor Assia Benhabiles, Dr. Soumeya Lemai, Professor Louis Violet, and Professor Michel Koenig.

References

- [1] V. Dubowitz, "Muscle disorders in childhood," *Major Problems in Clinical Pediatrics*, vol. 16, pp. 1–282, 1978.
- [2] A. Czeizel and J. Hamula, "A Hungarian study on Werdnig-Hoffmann disease," *Journal of Medical Genetics*, vol. 26, no. 12, pp. 761–763, 1989.
- [3] J. Pearn, "Classification of spinal muscular atrophies," *The Lancet*, vol. 1, no. 8174, pp. 919–922, 1980.
- [4] I. Biros and S. Forrest, "Spinal muscular atrophy: untangling the knot?" *Journal of Medical Genetics*, vol. 36, no. 1, pp. 1–8, 1999.
- [5] T. C. Gilliam, L. M. Brzustowicz, L. H. Castilla et al., "Genetic homogeneity between acute and chronic forms of spinal muscular atrophy," *Nature*, vol. 345, no. 6278, pp. 823–825, 1990.
- [6] J. Melki, S. Abdelhak, P. Sheth et al., "Gene for chronic proximal spinal muscular atrophies maps to chromosome 5q," *Nature*, vol. 344, no. 6268, pp. 767–768, 1990.
- [7] T. L. Munsat, L. Skerry, B. Korf et al., "Phenotypic heterogeneity of spinal muscular atrophy mapping to chromosome 5q11.2-13.3 (*SMA* 5q)," *Neurology*, vol. 40, no. 12, pp. 1831–1836, 1990.
- [8] S. Lefebvre, L. Bürglen, S. Reboullet et al., "Identification and characterization of a spinal muscular atrophy-determining gene," *Cell*, vol. 80, no. 1, pp. 155–165, 1995.
- [9] U. R. Monani, C. L. Lorson, D. W. Parsons et al., "A single nucleotide difference that alters splicing patterns distinguishes the *SMA* gene *SMN1* from the copy gene *SMN2*," *Human Molecular Genetics*, vol. 8, no. 7, pp. 1177–1183, 1999.
- [10] L. Bürglen, S. Lefebvre, O. Clermont et al., "Structure and organization of the human survival motor neurone (*SMN*) gene," *Genomics*, vol. 32, no. 3, pp. 479–482, 1996.
- [11] H. Scheffer, J. M. Cobben, G. Matthijs, and B. Wirth, "Best practice guidelines for molecular analysis in spinal muscular atrophy," *European Journal of Human Genetics*, vol. 9, no. 7, pp. 484–491, 2001.
- [12] P. Bulet, L. Bürglen, O. Clermont et al., "Large scale deletions of the 5q13 region are specific to Werdnig-Hoffmann disease," *Journal of Medical Genetics*, vol. 33, no. 4, pp. 281–283, 1996.
- [13] F. Capon, C. Levato, S. Semprini et al., "Deletion analysis of *SMN* and *NAIP* gene in spinal muscular atrophy Italian families," *Muscle & Nerve*, vol. 19, pp. 378–380, 1996.
- [14] D. W. Parsons, P. E. McAndrew, S. T. Iannaccone, J. R. Mendell, A. H. M. Burghes, and T. W. Prior, "Intragenic tel*SMN* mutations: frequency, distribution, evidence of a founder effect, and modification of the spinal muscular atrophy phenotype by cen*SMN* copy number," *American Journal of Human Genetics*, vol. 63, no. 6, pp. 1712–1723, 1998.

- [15] B. Wirth, "An update of the mutation spectrum of the survival motor neuron gene (SMN1) in autosomal recessive spinal muscular atrophy (SMA)," *Human Mutation*, vol. 15, pp. 228–237, 2000.
- [16] S. Srivastava, M. Mukherjee, I. Panigrahi, G. S. Pandey, S. Pradhan, and B. Mittal, "SMN2-deletion in childhood-onset spinal muscular atrophy," *American Journal of Medical Genetics*, vol. 101, no. 3, pp. 198–202, 2001.
- [17] J. E. Taylor, N. H. Thomas, C. M. Lewis et al., "Correlation of SMNt and SMNc gene copy number with age of onset and survival in spinal muscular atrophy," *European Journal of Human Genetics*, vol. 6, no. 5, pp. 467–474, 1998.
- [18] M. Gennarelli, M. Lucarelli, F. Capon et al., "Survival motor neuron gene transcript analysis in muscles from spinal muscular atrophy patients," *Biochemical and Biophysical Research Communications*, vol. 213, no. 1, pp. 342–348, 1995.
- [19] C. Helmken, Y. Hofmann, F. Schoenen et al., "Evidence for a modifying pathway in SMA discordant families: reduced SMN level decreases the amount of its interacting partners and Htra2-beta1," *Human Genetics*, vol. 114, no. 1, pp. 11–21, 2003.
- [20] N. Roy, M. S. Mahadevan, M. McLean et al., "The gene for neuronal apoptosis inhibitory protein is partially deleted in individuals with spinal muscular atrophy," *Cell*, vol. 80, no. 1, pp. 167–178, 1995.
- [21] M. J. Somerville, A. G. Hunter, H. L. Aubry, R. G. Korneluk, A. E. MacKenzie, and L. C. Surh, "Clinical application of the molecular diagnosis of spinal muscular atrophy: deletions of neuronal apoptosis inhibitor protein and survival motor neuron genes," *American Journal of Medical Genetics*, vol. 69, pp. 159–165, 1997.
- [22] L. Campbell, A. Potter, J. Ignatius, V. Dubowitz, and K. Davies, "Genomic variation and gene conversion in spinal muscular atrophy: implications for disease process and clinical phenotype," *American Journal of Human Genetics*, vol. 61, no. 1, pp. 40–50, 1997.
- [23] J. G. Chang, Y. J. Jong, J. M. Huang et al., "Molecular basis of spinal muscular atrophy in Chinese," *American Journal of Human Genetics*, vol. 57, no. 6, pp. 1503–1505, 1995.
- [24] N. R. Rodrigues, N. Owen, K. Talbot, J. Ignatius, V. Dubowitz, and K. E. Davies, "Deletions in the survival motor neuron gene on 5q13 in autosomal recessive spinal muscular atrophy," *Human Molecular Genetics*, vol. 4, no. 4, pp. 631–634, 1995.
- [25] E. Velasco, C. Valero, A. Valero, F. Moreno, and C. Hernández-Chico, "Molecular analysis of the SMN and NAIP genes in Spanish spinal muscular atrophy (SMA) families and correlation between number of copies of (C)BCD541 and SMA phenotype," *Human Molecular Genetics*, vol. 5, pp. 257–263, 1996.
- [26] T. L. Munsat, "International SMA collaboration," *Neuromuscular Disorders*, vol. 1, no. 2, p. 81, 1991.
- [27] K. Zerres, S. Rudnik-Schöneborn, E. Forrest, A. Lusakowska, J. Borkowska, and I. Hausmanowa-Petrusewicz, "A collaborative study on the natural history of childhood and juvenile onset proximal spinal muscular atrophy (type II and III SMA): 569 patients," *Journal of the Neurological Sciences*, vol. 146, no. 1, pp. 69–72, 1997.
- [28] S. A. Miller, D. D. Dykes, and H. F. Polesky, "A simple salting out procedure for extracting DNA from human nucleated cells," *Nucleic Acids Research*, vol. 16, no. 3, p. 1215, 1988.
- [29] G. van der Steege, P. M. Grootsholten, P. van der Vlies et al., "PCR-based DNA test to confirm clinical diagnosis of autosomal recessive spinal muscular atrophy," *The Lancet*, vol. 345, no. 8955, pp. 985–986, 1995.
- [30] B. Wirth, T. Schmidt, E. Hahnen et al., "De novo rearrangements found in 2% index patients with spinal muscular atrophy (SMA): mutational mechanisms, parental origin, mutation rate and implications for prenatal diagnosis," *The American Journal of Human Genetics*, vol. 61, pp. 1102–1111, 1997.
- [31] B. Wirth, M. Herz, A. Wetter et al., "Quantitative analysis of survival motor neuron copies: identification of subtle SMN1 mutations in patients with spinal muscular atrophy, genotype-phenotype correlation, and implications for genetic counseling," *American Journal of Human Genetics*, vol. 64, no. 5, pp. 1340–1356, 1999.
- [32] K. Zerres and S. Rudnik-Schöneborn, "Natural history in proximal spinal muscular atrophy: clinical analysis of 445 patients and suggestions for a modification of existing classifications," *Archives of Neurology*, vol. 52, no. 5, pp. 518–523, 1995.
- [33] C. A. Kim, B. Passos, and A. Marie, "Clinical and molecular analysis of spinal muscular atrophy in Brazilian patients," *Genetics and Molecular Biology*, vol. 22, no. 4, pp. 1415–1475, 1999.
- [34] A. Belaid, "Amyotrophies Spinales," sous la direction de Cecile Jeager-Buet, Collection savoir et comprendre. AFM, 28, Juin 2006.
- [35] S. Rudnik-Schöneborn, D. Rohrig, G. Morgan, B. Wirth, and K. Zerres, "Autosomal recessive proximal spinal muscular atrophy in 101 sibs out of 48 families: clinical picture, influence of gender, and genetic implications," *American Journal of Medical Genetics*, vol. 51, no. 1, pp. 70–76, 1994.
- [36] M. Tazir and C. Geronimi, "Chronic childhood spinal muscular atrophies in Algeria. A genetic study," *Journal of the Neurological Sciences*, vol. 96, no. 1, pp. 89–101, 1990.
- [37] Enquête sur la consanguinité en Algérie, "Fondation nationale pour la promotion de la sante et le développement de la recherche (FOREM)," Septembre 2007.
- [38] K. Cho, K. Ryu, E. Lee et al., "Correlation between genotype and phenotype in Korean patients with spinal muscular atrophy," *Molecules and Cells*, vol. 11, no. 1, pp. 21–27, 2001.
- [39] N. Duc Bach, A. Hamim Sadewa, Y. Takeshima et al., "Deletion of the SMN1 and NAIP genes in Vietnamese patients with spinal muscular atrophy," *Kobe Journal of Medical Sciences*, vol. 49, no. 3-4, pp. 55–58, 2003.
- [40] R. Labrum, J. Rodda, and A. Krause, "The molecular basis of Spinal Muscular Atrophy (SMA) in South African black patients," *Neuromuscular Disorders*, vol. 17, no. 9-10, pp. 684–692, 2007.
- [41] R. M. Shawky, K. Abd El Aleem, M. M. Rifaat, and A. Moustafa, "Molecular diagnosis of spinal muscular atrophy in Egyptians," *Eastern Mediterranean Health Journal*, vol. 7, no. 1-2, pp. 229–237, 2001.
- [42] A. S. Glotov, A. V. Kiselev, T. E. Ivaschenko, and V. S. Baranov, "Analysis of deletions in SMN1, SMN2, and NAIP genes in spinal muscular atrophy patients from the northwestern region of Russia," *Russian Journal of Genetics*, vol. 37, no. 8, pp. 968–971, 2001.
- [43] A. Kesari, U. K. Misra, J. Kalita et al., "Study of survival of motor neuron (SMN) and neuronal apoptosis inhibitory protein (NAIP) gene deletions in SMA patients," *Journal of Neurology*, vol. 252, no. 6, pp. 667–671, 2005.
- [44] S. Al Rajeh, A. Ramanath Majumdar, A. Adetunji Adeyokunnu et al., "Molecular analysis of the SMN and NAIP genes in Saudi spinal muscular atrophy patients," *Journal of the Neurological Sciences*, vol. 158, no. 1, pp. 43–46, 1998.

- [45] N. R. Rodrigues, N. Owen, K. Talbot et al., "Gene deletions in spinal muscular atrophy," *Journal of Medical Genetics*, vol. 33, no. 2, pp. 93–96, 1996.
- [46] M. S. Watihayati, A. M. H. Zabidi-Hussin, T. H. Tang, H. Nishio, and B. A. Zilfalil, "NAIP-deletion analysis in Malaysian patients with spinal muscular atrophy," *Kobe Journal of Medical Sciences*, vol. 53, no. 4, pp. 171–175, 2007.
- [47] A. Amara, L. Adala, I. Ben Charfeddine et al., "Correlation of SMN2, NAIP, p44, H4F5 and Occludin genes copy number with spinal muscular atrophy phenotype in Tunisian patients," *European Journal of Paediatric Neurology*, vol. 16, no. 2, pp. 167–174, 2012.
- [48] D. D. Coovert, T. T. Le, P. E. McAndrew et al., "The survival motor neuron protein in spinal muscular atrophy," *Human Molecular Genetics*, vol. 6, no. 8, pp. 1205–1214, 1997.
- [49] E. Humphrey, H. R. Fuller, and G. E. Morris, "Current research on SMN protein and treatment strategies for spinal muscular atrophy," *Neuromuscular Disorders*, vol. 22, no. 2, pp. 193–197, 2012.
- [50] S. Lefebvre, P. Burlet, Q. Liu et al., "Correlation between severity and SMN protein level in spinal muscular atrophy," *Nature Genetics*, vol. 16, no. 3, pp. 265–269, 1997.
- [51] M. Feldkötter, V. Schwarzer, R. Wirth, T. F. Wienker, and B. Wirth, "Quantitative analyses of SMN1 and SMN2 based on real-time lightcycler PCR: fast and highly reliable carrier testing and prediction of severity of spinal muscular atrophy," *American Journal of Human Genetics*, vol. 70, no. 2, pp. 358–368, 2002.
- [52] M. D. Mailman, J. W. Heinz, A. C. Papp et al., "Molecular analysis of spinal muscular atrophy and modification of the phenotype by SMN2," *Genetics in Medicine*, vol. 4, no. 1, pp. 20–26, 2002.
- [53] S. Ogino, S. Gao, D. G. Leonard, M. Paessler, and R. B. Wilson, "Inverse correlation between SMN1 and SMN2 copy numbers: evidence for gene conversion from SMN2 to SMN1," *European Journal of Human Genetics*, vol. 11, no. 3, pp. 275–277, 2003.
- [54] B. Wirth, L. Brichta, B. Schrank et al., "Mildly affected patients with spinal muscular atrophy are partially protected by an increased SMN2 copy number," *Human Genetics*, vol. 119, no. 4, pp. 422–428, 2006.
- [55] A. Bouhouche, A. Benomar, N. Birouk et al., "High incidence of SMN1 gene deletion in Moroccan adult-onset spinal muscular atrophy patients," *Journal of Neurology*, vol. 250, no. 10, pp. 1209–1213, 2003.
- [56] S. Savas, N. Gokgoz, H. Kayserili, F. Ozkinay, M. Yuksel-Apak, and B. Kirdar, "Screening of deletions in SMN, NAIP and BTF2p44 genes in Turkish spinal muscular atrophy patients," *Human Heredity*, vol. 50, no. 3, pp. 162–165, 2000.
- [57] B. Wirth, E. Hahnen, K. Morgan et al., "Allelic association and deletions in autosomal recessive proximal spinal muscular atrophy: association of marker genotype with disease severity and candidate cDNAs," *Human Molecular Genetics*, vol. 4, no. 8, pp. 1273–1284, 1995.
- [58] E. Bussaglia, O. Clermont, E. Tizzano et al., "A frame-shift deletion in the survival motor neuron gene in Spanish spinal muscular atrophy patients," *Nature Genetics*, vol. 11, no. 3, pp. 335–337, 1995.
- [59] J. M. Cobben, G. van der Steege, P. Grootsholten, M. de Visser, H. Scheffer, and C. H. C. M. Buys, "Deletions of the survival motor neuron gene in unaffected siblings of patients with spinal muscular atrophy," *American Journal of Human Genetics*, vol. 57, no. 4, pp. 805–808, 1995.
- [60] T. Akutsu, H. Nishio, K. Sumino et al., "Molecular genetics of spinal muscular atrophy: contribution of the NAIP gene to clinical severity," *Kobe Journal of Medical Sciences*, vol. 48, no. 1-2, pp. 25–31, 2002.
- [61] R. M'rad, I. Dorboz, L. B. Jemaa et al., "Molecular analysis of the SMN1 and naip genes in 60 Tunisian spinal muscular atrophy patients," *La Tunisie Medicale*, vol. 84, no. 8, pp. 465–469, 2006.
- [62] Y. Shafeghati, S. Teymourian, G. Babamohammadi et al., "Molecular diagnosis Iranian patients with spinal muscular atrophy," *Archives of Iranian Medicine*, vol. 7, no. 1, pp. 47–52, 2004.
- [63] M. Z. Haider, A. Moosa, H. Dalal, Y. Habib, and L. Reynold, "Gene deletion patterns in spinal muscular atrophy patients with different clinical phenotypes," *Journal of Biomedical Science*, vol. 8, no. 2, pp. 191–196, 2001.
- [64] C. H. Tsai, Y. J. Jong, C. J. Hu et al., "Molecular analysis of SMN, NAIP and P44 genes of SMA patients and their families," *Journal of the Neurological Sciences*, vol. 190, no. 1-2, pp. 35–40, 2001.



Hindawi

Submit your manuscripts at
<http://www.hindawi.com>

